

Diet mediates thermal performance traits: implications for marine ectotherms

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Keywords: temperature, thermal acclimation, fish, thermal limits, *Girella nigricans*, omnivore

Summary Statement: Diet impacted thermal performance traits in a temperate marine teleost fish, opaleye (*Girella nigricans*). Generalist ectotherms that can choose their diet may be able to regulate their performance at different environmental temperatures.

Abstract

Thermal acclimation is a key process enabling ectotherms to cope with temperature change. To undergo a successful acclimation response, ectotherms require energy and nutritional building blocks obtained from their diet. However, diet is often overlooked as a factor that can alter acclimation responses. Using a temperate omnivorous fish, opaleye (*Girella nigricans*), as a model system, we tested the hypotheses that 1) diet can impact the magnitude of thermal acclimation responses and 2) traits vary in their sensitivity to both temperature acclimation and diet. We fed opaleye a simple omnivorous diet (*ad libitum* *Artemia* sp. and *Ulva* sp.) or a carnivorous diet (*ad libitum* *Artemia* sp.) at two ecologically relevant temperatures (12 and 20°C) and measured a suite of whole animal (growth, sprint speed, metabolism), organ (cardiac thermal tolerance), and cellular-level traits (oxidative stress, glycolytic capacity). When opaleye were offered two diet options compared to one, they had reduced cardiovascular thermal performance and higher standard metabolic rate under conditions representative of the maximal seasonal temperature the population experiences (20°C). Further, sprint speed and absolute aerobic scope were insensitive to diet and temperature, while growth was highly sensitive to

temperature but not diet, and standard metabolic rate and maximum heart rate were sensitive to both diet and temperature. Our results reveal that diet influences thermal performance in trait-specific ways, which could create diet trade-offs for generalist ectotherms living in thermally variable environments. Ectotherms that alter their diet may be able to regulate their performance at different environmental temperatures.

Introduction

Understanding the full range and maximum capacity of ectotherm physiological responses to environmental change is essential to predict species vulnerability to global climate change (Huey et al., 2012; Somero, 2011; Stillman, 2003). Temperature is a critical environmental factor governing ectotherm physiology, behavior, and ecology (Brett, 1971). The current paradigm suggests that ectotherms have three options when faced with unfavorable temperatures: they can move to a more suitable habitat, adapt over multiple generations, or acclimate to the new conditions (Daufresne et al., 2009; Glanville and Seebacher, 2006; Hofmann and Todgham, 2010; Somero, 2011). Thermal acclimation is an essential survival mechanism for ectotherms living in variable environments and a critical coping mechanism against global climate change (Bernhardt and Leslie, 2013; Jackson et al., 2021; Seebacher et al., 2015). During thermal acclimation, ectotherms undergo reversible phenotypic changes that improve their performance at a given temperature (Figure 1; e.g., enzyme activity, membrane composition, mitochondrial density, oxygen transport, organ morphology and function; Anttila et al., 2014; Chung and Schulte, 2020; Ekström et al., 2016; Little et al., 2020; Seebacher et al., 2015). It is often assumed that ectotherms will achieve the same level of performance after repeated exposures to a temperature, as long as all other environmental conditions (i.e., salinity, pH, dissolved oxygen) are held the same (Sinclair et al., 2016). To undergo a successful acclimation response, ectotherms require energy and nutritional building blocks obtained from their diet. Diets vary considerably in nutritional and energetic content, which suggests that different diets may mediate distinct thermal acclimation responses (Figure 1).

Food quality and availability can change seasonally, with global climate change, and across habitats (Alton et al., 2020; Arnold et al., 2010; Birnie-Gauvin et al., 2017; Ho et al., 2010). Many ectotherms are also generalists and vary their diet to meet their nutritional requirements or maximize energy-use efficiency (Jobling, 2016; Johnson et al., 2017; Kaiser and

Hughes, 1993; Raubenheimer et al., 2005; Rubio et al., 2003, 2009; Sánchez-Vázquez et al., 1998). Some ectotherms also change their diet with temperature (Boersma et al., 2016a; Carreira et al., 2016; Jang et al., 2015; Rho and Lee, 2017; Schmitz and Rosenblatt, 2017; Vejříková et al., 2016). For example, multiple omnivorous fishes consume higher proportions of algae as water temperatures increase (e.g., Behrens and Lafferty, 2012; Emde et al., 2016; González-Bergonzoni et al., 2016; Guinan et al., 2015; Prejs, 1984). The exact reasons for these diet shifts are unknown, with some suggesting that the optimal dietary protein: carbohydrate ratio for ectotherms differs across temperatures (Lee et al., 2015; Rho and Lee, 2017; Zhang et al., 2020), or that cold temperature constrains the digestive physiology of herbivores and omnivores (Floeter et al., 2005; González-Bergonzoni et al., 2012). These proposed explanations hint at a broader hypothesis; that omnivores consume different proportions of plants and animals to regulate their physiological responses to changing temperatures. More broadly, any changes in an ectotherm's diet that coincide with a change in environmental temperature (either through differences in dietary preference, availability, or nutrient composition) might alter its thermal performance.

To understand the interaction between diet and temperature, we must consider how traits critical to survival may be differentially affected (Figure 1). Measuring thermal limits in conjunction with vital biological rates provides comprehensive insight into ectotherm thermal biology in variable and changing environments (Magozzi and Calosi, 2015). A common assumption in thermal biology is that biological rates have the same thermal sensitivity and that aerobic capacity and baseline metabolism can be used as proxies for many performance traits (Clark et al., 2013; Fry, 1947; Brett, 1971; Claireaux and Lefrançois, 2007; Pörtner, 2001; Pörtner, 2010; for critique: Clark et al., 2013; Schulte, 2015). However, there is growing support for a multiple-performance, multiple- optima model, which states that thermal sensitivity differs across biological rates (i.e., absolute aerobic scope, standard metabolic rate, growth rate, sprint speed) and is not always predictable based on aerobic capacity or baseline metabolism (Clark et al., 2013; Dell et al., 2011; Kellermann et al., 2019; Seebacher et al., 2015). For example, Healy and Schulte (2012) demonstrated that specific growth rate was negative at temperatures where absolute aerobic scope (i.e., maximum – standard metabolic rate) was maximal in killifish. This model has been challenging to test empirically as few performance traits are usually measured per study and these traits are often considered separately from upper

and lower thermal limits (Magozzi and Calosi, 2015). If diet and temperature together influence biological rates and thermal limits, ectotherms may be incentivized to make diet choices that improve their thermal responses. However, if fitness-enhancing traits are differentially affected by diet and temperature, there could be important performance consequences associated with an ectotherm's ultimate diet choices.

Opaleye (*Girella nigricans*) are temperate omnivorous fish that consume a greater proportion of algae in warmer water across their geographic range (Behrens and Lafferty, 2012), which makes them an ideal model for exploring diet effects on thermal acclimation responses. Here, we tested the hypothesis that when offered a simple choice omnivorous diet (*ad libitum* *Artemia* sp. and *Ulva* sp.) versus a carnivorous diet (*ad libitum* *Artemia* sp. only) at two ecologically relevant temperatures (12 and 20°C), opaleye would make diet choices that altered their thermal acclimation responses in trait-specific ways. As juveniles, opaleye live in the intertidal where they face many challenges, including escaping predators, maintaining growth rates, and dealing with high daily thermal variation (Somero, 2010). Therefore, we adopted an integrative approach and assessed the opaleye's thermal acclimation responses at the whole animal (growth, sprint speed, metabolism, critical thermal limits), organ (cardiac thermal limits), and cellular (glycolytic capacity, oxidative stress) levels to compare ecologically and physiologically relevant performance traits and thermal limits for the fish in their juvenile life stage and identify any trade-offs associated with the treatment diets. We hypothesized that biological rates would increase with temperature but have different thermal sensitivities depending on the diet treatment. Specifically, we predicted that there would be costs to consuming the omnivorous diet (e.g., higher digestive infrastructure costs resulting in higher maintenance metabolism or reduced growth from lower protein diet) that would be offset by increases in the performance of other traits (e.g., thermal limits, sprint performance, glycolytic capacity, oxidative stress).

Methods:

Fish Collection

Juvenile opaleye were collected in Spring 2019 (Exp #1: respirometry, sprint, growth, critical thermal maxima (CT_{max}/CT_{min}); N=144; body mass (BM): 14.75 ± 3.53 g and total length (TL): 9.42 ± 0.76 cm; mean \pm SD) and in Winter 2020 (Exp #2: Arrhenius breakpoint test;

N=126; BM: 19.5 ± 6.1 g and TL: 10.5 ± 1.1 cm; mean \pm SD) by seine or hook and line from Santa Barbara Harbor, California, USA (34.40829, -119.691389). Fish were transported in coolers (>70% air saturation) to the University of California, Santa Barbara and held in 95 L fiberglass flow-through seawater tanks (9-12 fish per tank). Prior to the start of acclimation, fish were held at ambient conditions (Exp 1: $13.9 \pm 1.1^\circ\text{C}$ and Exp 2: $16.2 \pm 0.6^\circ\text{C}$ mean \pm SD) and fed *ad libitum* omnivorous diets (*Ulva* sp. and *Artemia* sp.). All protocols were approved by the Institutional Animal Care and Use Committee at the University of California, Santa Barbara.

Acclimation and Diet Treatments

Fish were randomly assigned to one of two ecologically relevant temperatures (12 and 20°C , representative of the low and high seasonal temperatures experienced in Santa Barbara, CA (Figure 2) and fed one of two *ad libitum* diets (omnivorous: *Artemia* sp. and *Ulva* sp. ; and carnivorous: *Artemia* sp.) in a factorial design with 3-4 replicate tanks per treatment. *Ulva* sp. (collected by hand from Goleta Beach, Goleta, CA) and *Artemia* sp. (brineshrimpdirect.com) were replaced every morning. Diets were selected based on stomach content information from Barry and Ehret (1993), which demonstrated that both *Ulva* sp. and small crustaceans constitute a significant portion of wild opaleye diets and Behrens and Lafferty (2007), which used *Ulva* sp. as a representative herbivorous diet for a lab study on opaleye. Preliminary data demonstrated that *Ulva* sp. only diets were not nutritionally sufficient. Dietary proximate analysis is provided in Table S1.

Food consumption rates were assessed during preliminary trials. All tanks were fasted for 24 h, and then the fish were offered their pre-weighed diet treatment for 1 h. The remaining *Artemia* sp. and *Ulva* sp. was removed and weighed. However, we were unable to obtain any measurable estimates of *Ulva* sp. consumption during these brief 1 h trials. At 20°C , *Artemia* sp. consumption was ~23% lower in the omnivorous treatment (7.39% body mass) than the carnivorous treatment (9.58% body mass), suggesting that the fish in the omnivorous treatment were supplementing their diet with *Ulva* sp. This is consistent with our visual observations that the fish readily consumed *Ulva* sp. in the warm treatment (though they consistently ate more *Artemia* sp. compared to *Ulva* sp.). In contrast, at 12°C *Artemia* sp. consumption was only ~10% lower in the omnivorous treatment (3.65% body mass) compared to the carnivorous (4.07% body mass) and we did not personally observe the fish consuming *Ulva* sp. at this temperature. This

suggests that the fish either were not consuming *Ulva* sp. or were doing so in small amounts at the cold acclimation treatment. Further, opaleye ate less at the cold acclimation treatment compared to the warm acclimation treatment. Overall *Artemia* sp. consumption was 55% lower in the cold treatment compared to the warm.

Temperature and dissolved oxygen content were monitored 1-2 times daily by hand using a Digi-sense Traceable Single RTD thermometer (Cole Palmer, IL, USA) and an Oxyguard handy Polaris 2 (OxyGuard International A/S, Farum, Denmark). Oxygen was maintained at >80% air saturation throughout the study. The average temperature per treatment was $20.0 \pm 0.4^\circ\text{C}$ and $12.2 \pm 0.4^\circ\text{C}$ in both experiments (mean \pm SD; determined from in tank Thermochron 4K iButtons programmed to record every 20 minutes). Fish were acclimated to treatment conditions for 3 weeks prior to experimentation (14:10 h light: dark cycle). All individuals were fasted for 36-40 hours prior to respirometry, thermal limit, and sprint testing. All tests were performed at acclimation temperatures unless otherwise noted.

Intermittent Flow Respirometry

Respirometry was conducted using 12 respirometers in one of three sizes: 349 ml, 579 ml and 711 ml. Water was flushed and recirculated through the chamber at a rate of 2.5L min^{-1} (Eheim Universal 300 pumps; Eheim, Germany). Dissolved oxygen was measured continuously in each respirometer using a robust oxygen probe and Firesting optical oxygen meter (Pyroscience, Germany).

Fish were transferred with minimal air exposure (<10 seconds), to a cylindrical chase tank (20 L), where they were chased by hand for five minutes and then immediately placed in the respirometers to obtain an estimate of their maximum metabolic rate (MMR) (this chase protocol was the most effective at eliciting MMR in opaleye; data not shown). Chases occurred between 10:30-14:30 and were followed by ~20 h of automated measurement cycles (15-minute total flush/recirculation cycle). Tanks were covered in shade cloth to minimize potential disturbance. Fish were held at either $20.1 \pm 0.6^\circ\text{C}$ or $11.8 \pm 0.4^\circ\text{C}$ (mean \pm SD) across all tests. Background respiration was measured before and after each test for ≥ 3 full measurement cycles.

After 20 h in the respirometers, fish were removed and anesthetized in 80 mg L^{-1} MS-222 buffered with 80 mg L^{-1} NaHCO_3^- (Sigma Aldrich Co., St. Louis, MO, USA). Each fish was

weighed (mass in g), measured for total length (TL), and tagged with a unique color code using Visible Implant Elastomer Tags (Northwest Marine Fisheries INC, WA, USA).

Data Analysis for respirometry data

All oxygen consumption data were analyzed in R (version 3.5.1) using best practices as outlined in Rosewarne et al. (2016) and Chabot et al. (2016) (R core team, 2018). The data was used to calculate four metabolic rate metrics which define an individual's aerobic energy budget. Standard metabolic rate (SMR) is the baseline metabolic rate needed to survive, and MMR is the maximum rate of energy expenditure. The difference between these two metabolic rates is the absolute aerobic scope (AAS = MMR - SMR), which is representative of the aerobic energy budget a fish has to perform all critical biological processes (Clark et al., 2013). Factorial aerobic scope (FAS = MMR / SMR) is another estimate of the aerobic energy budget and represents the scope for increasing metabolic rate proportional to SMR (Clark et al., 2013). Only fish where >75% of measurements followed a linear decrease with an $R^2 > 0.9$ were included in SMR analysis (i.e., >60 MO_2 measurements total per fish; N = 6-10 per treatment). SMR was calculated as the lowest 15% quantile of all recorded measurement cycles (Chabot et al., 2016)). MMR was calculated as the steepest 120s slope during the first measurement period (Little et al., 2020; N = 6-11 per treatment). All presented metabolic rate measurements are body mass specific (i.e., presented as $\text{mg O}_2 \text{ kg}^{-1} \text{ min}^{-1}$). We tested for the need to account for body size scaling but given the small size range in the study (8-39g), we did not find evidence of a scaling relationship when the data were plotted on log-log plots (data not shown). Given lack of knowledge on a species-specific scaling exponent, we did not scale the metabolism to a common body size using an allometric scaling slope. Background respiration was assessed and when applicable, a linear regression was fitted between the pre- and post-background measurements and subtracted from the corresponding MO_2 values (Rodgers et al., 2016; Rosewarne et al., 2016).

Sprint Speed

We used a modified protocol based on Kraskura and Nelson (2018). The setup included a custom-built acrylic sprint chamber (128 cm X 30 cm X 30 cm (LxWxH); water height = 10 cm; water volume = 41.6 L) with a camera (Canon EOS Rebel T4i) positioned above the sprint

chamber to ensure full view of the work area. Each sprint trial was recorded at 60 fps and saved for digital analysis. Fish were placed behind a gated and shaded area in a sprint chamber and habituated to the chamber for 10 minutes. The gate was removed and the fish were motivated to sprint by manual chasing. Each fish performed ≤ 7 trials with 5 minutes of recovery between trials. Only trial videos where the fish had undergone a sprint with 1) a straight path of > 20 cm 2) an unobstructed view of 50 cm of the chamber 3) active and continued bursts 4) no back and forth swimming around the chamber were used in subsequent analysis. Temperature was monitored throughout all trials and remained within $\pm 1^\circ\text{C}$ of the test temperature. All videos used in analysis were visualized and validated by ≥ 2 researchers. Videos were tracked in ImageJ/MtrackJ and subsequently analyzed in R to determine the fastest 5 frames of sprinting (~ 10 cm) per trial. 2-3 trials were analyzed per fish and averaged to determine each fish's sprint performance (cm s^{-1}) (Figure S1 and S2; $N = 8-10$ per treatment). All fish were given at least 3 weeks of recovery between respirometry or thermal limit experiments and the sprint test.

Thermal Limits: CT_{\max} and CT_{\min} testing

We used a standardized critical thermal maxima and minima tests (CT_{\max} and CT_{\min} , respectively) as described in Beitinger and Lutterschmidt (2011). CT_{\max} and CT_{\min} represent the most extreme temperatures a fish can survive at for a short period of time. Briefly, 3-4 fish per tank were transferred to the testing tank (78 cm X 61 cm X 18 cm (LxWxH); water height = 12 cm, water volume = 57 L) at their treatment temperature with minimal air exposure (<10 seconds; total $N = 8-12$ per treatment). After a brief habituation period (~ 5 min), the test tank was heated (CT_{\max} ; 1500-Watt immersion heater, Process Technology, Ohio, USA) or cooled (CT_{\min} ; 1 HP AquaEuro chiller, AquaEuroUSA, CA, USA) at a rate of $\sim 0.3^\circ\text{C min}^{-1}$ until fish lost the ability to maintain their righting response. Temperature was recorded at the loss of equilibrium and the fish were immediately placed in a recovery tank at an intermediate temperature before being returned to their treatment tanks. All fish fully recovered following the critical thermal tests.

Growth

All fish were weighed and measured for length prior to acclimation and again after 8 weeks under treatment conditions (note that some had been randomly removed for tissue sample collection, see below) to estimate growth across tanks (i.e., average weight gain per week).

Dissection and Enzyme Analysis

Fish from each treatment (N = 6 per treatment) were euthanized by cerebral concussion followed by severing the spinal cord on day 27-34 of acclimation after the aerobic scope trials were complete. Morphometrics (body mass and total length) were measured, and a white muscle and liver sample were flash frozen in liquid nitrogen and stored at -80°C for future analysis. Stomach and remaining gut contents were emptied and leftover fish remains were stored at -20°C for proximate analysis (See supplemental methods and table S1). *Lactate dehydrogenase assays*: lactate dehydrogenase activity assays were performed as outlined in Little et al. (2020) as a proxy for glycolytic capacity in white muscle, where high levels of lactate dehydrogenase activity indicate that the animal has a high capacity to support anaerobic ATP production to fuel burst swimming or hypoxia tolerance (Little et al., 2020). Briefly, white muscle samples were homogenized in homogenization buffer (0.1% Triton, 50mM Hepes, 1mM EDTA, pH 7.4 buffer) before being run in triplicate (intra-assay CV% <15%) on a SpectraMax iD3 Multi-Mode Microplate Reader (Molecular Devices, USA) using a wavelength of 340 nm to measure the disappearance of NADH. The assay was repeated at 8, 12, 20, 26, and 32°C. *Lipid peroxidation assays*: Thiobarbituric acid reactive substances were quantified in the liver to estimate lipid peroxidation as a proxy of oxidative stress using a commercially available Fluorometric assay kit (Caymen Chemical, USA). Here, higher levels of lipid peroxidation are indicative of greater oxidative damage to cellular components (Castro et al., 2012). Samples were homogenized in RIPA buffer and treated according to manufacturer's instructions before being run in duplicate at an excitation wavelength of 544 nm and emission wavelength of 590 nm.

Thermal Limits: Arrhenius Breakpoint Test

Arrhenius breakpoint tests on the heart (ABT test) were conducted (N = 6-14 per test and per treatment) as outlined in Casselman et al. (2012) and Gilbert et al. (2020). Briefly, fish were anesthetized in seawater containing 80 mg L⁻¹ MS-222 buffered with 1g L⁻¹ NaHCO₃⁻ before

being placed ventral side up in an experimental sling in the test tank (10 L seawater containing buffered 65mg L⁻¹ MS-222). Water was circulated past the gills to irrigate them. Stainless Steel Needle Tip Electrodes (ADInstruments INC, Colorado Springs, CO, USA) were inserted just under the skin to detect an ECG signal, which was amplified using Dual Bio Amp (ADInstruments INC, Colorado Springs, CO, USA) and filtered (Filters: 60hz Notch filter; Mains filter; Low-Pass: 2Kz; High Pass: 10hz; Range: 2mV).

After a 30 min equilibration period at the acclimation temperature (Ferreira et al, 2014; Hansen et al, 2017), atropine sulfate was injected intraperitoneally (1.2 mg kg⁻¹ in 0.9% NaCl) to block vagal tone followed by isoproterenol (4 ug kg⁻¹ in 0.9% NaCl) to maximally stimulate β -adrenoreceptors. Any fish that did not respond to the drug injections or where experimental error occurred (e.g., water pump failure) were removed from the analysis and not considered further. These drug concentrations were tested prior to experimentation to ensure doubling the concentration did not further increase heart rate (f_H ; bpm). Fifteen minutes after isoproterenol injection, water temperature was heated (warm ABT test) or cooled (cold ABT test) at 1°C every 6 min (Polystat recirculating heater/chiller; Cole-Palmer, Vernon Hills, IL, USA). At each 1°C interval, f_H and temperature were stabilized to record a value for f_H . This procedure was repeated until the onset of cardiac arrhythmia (T_{ARR}), as indicated by a transition from rhythmic to arrhythmic beating or a missed QRS peak resulting in a precipitous drop in heart rate (Casselman et al., 2012) or until the known average CT_{max} for the species (generally 0-5°C higher than T_{ARR} , Chen et al., 2015; Muñoz et al., 2014; Safi et al., 2019), to ensure that curves could later be fit to the data for comparisons of acute thermal performance curves across treatments. All fish were immediately euthanized at the end of the test.

Data Analysis for Arrhenius Breakpoint Test

All ECG analyses were performed in LabChart software (AD Instruments; www.adinstruments.com). f_H was calculated for each temperature increment from 15 continuous seconds of measurements using automated ECG analysis software in lab chart (Gradil et al., 2016). The heart may set upper thermal tolerance temperatures in fishes (Anttila et al., 2014; Muñoz et al., 2014). The warm ABT test measures three sublethal thermal limits on cardiac function (T_{AB} , T_{PEAK} , T_{ARR}). Each of these limits indicate transition temperatures where the hearts capacity to transport oxygen, nutrients, and immune cells becomes compromised (Anttila

et al., 2014; Muñoz et al., 2014). The first thermal limit (T_{AB}) is highly correlated with the thermal optimum for aerobic scope in other teleosts (Anttila et al., 2013; Ferreira et al., 2014). T_{AB} was calculated by performing Arrhenius breakpoint tests on the rising phase of the thermal performance curve for f_H using *segmented* (v1.1-0; Muggeo, 2008) in R. The temperature corresponding to the breakpoint in f_H was defined as T_{AB} (warm ABT test) or $T_{AB-cold}$ (cold ABT test). Overall maximum heart rate (f_{Hmax}) was defined as the highest f_H recorded during the 15 s measurement phases in the warm ABT test and minimum heart rate (f_{Hmin}) was defined as the lowest f_H recorded during the 15 s measurement phases during the cold ABT test. Peak temperature (T_{PEAK}) was the temperature corresponding to f_{Hmax} .

Statistical Analysis

All data were statistically analyzed using R (version 3.5.1). All metrics were investigated for normality using Shapiro Wilk test and quantile-quantile plots, and for heteroskedasticity using Levene's test. Data that was not normally distributed was log-transformed before statistical analysis (only factorial aerobic scope). Data are displayed with untransformed values. All data were statistically analyzed (significance level $\alpha = 0.05$) using a 2-way ANOVA (*Car* v3.0-2; Fox and Weisberg, 2011) with post-hoc Tukey HSD, except for the thermal limits from the cold ABT, which was analyzed using a T-Test. Differences between treatments were also assessed using Cohens D tests (*effsize*, Torchiano, 2017). Note that 12°C fish were not tested for the cold ABT due to complications surrounding COVID-19 forcing the early shutdown of the experiment. In all 2-way ANOVA tests, significance of interaction between diet and temperature was tested for and excluded when non-significant. Polynomial curves were fit to heart rate data and compared using Akaike information criterion (AIC), where fit with the lowest AIC score was assigned the best fit model, but all models with $\Delta AIC < 2$ were considered (Burnham and Anderson, 2002). Measures of thermal sensitivity for all biological rates were calculated for each diet treatment using Q_{10} values, where $Q_{10} = \frac{R_2}{R_1} \left(\frac{10}{T_2 - T_1} \right)^{10}$, with R_1 = treatment mean at 12°C, R_2 = treatment mean at 20°C, $T_1 = 12^\circ\text{C}$, and $T_2 = 20^\circ\text{C}$.

Results

Metabolic Rates

Metabolism was influenced by diet and temperature, but each metabolic rate responded differently (Figure 3, 4). SMR was 28% higher in the 20°C omnivorous diet treatment compared to the carnivorous diet treatment, resulting in a significant interaction between diet and temperature (Figure 3A; Table 1). This was further supported by a large effect size between the 20°C treatments (Cohens D Test). In contrast, MMR significantly increased with temperature acclimation, but did not differ across diet treatments (Figure 3BC; Table 1). There was a marginal, but not significant diet and temperature effect on absolute aerobic scope (Figure 3C; Table 1). This was likely driven by the high individual variability in MMR, as the effect of diet on absolute aerobic scope had a p-value of 0.087 and medium effect sizes between diet treatments at 20°C. In contrast, factorial aerobic scope in the omnivorous diet treatment was significantly lower than the carnivorous diet treatment (diet: $df = 1$, $F = 5.796$, $p = 0.023$), which was largely driven by a 31% lower factorial aerobic scope (with a large effect size) in the 20°C omnivorous compared to 20°C carnivorous diet (Figure 3D; Table 1).

Other Biological Rates and Traits

Growth rate, sprint speed, lipid peroxidation, and lactate dehydrogenase activity were inconsistently affected by diet and temperature (Figure 4). Growth was significantly higher at 20°C but did not differ across diets (Figure 4, Table 1, Figure S2). Unexpectedly, the Q_{10} value for growth was higher than all other rates (28.05) and not close to any of the Q_{10} values for metabolic rate (ranged from 1.28-4.12; Figure 4, Table 1, Figure S2). Proximate analyses for whole tissue did not differ between treatment groups (Table S1).

In contrast with growth, maximum sprint speed did not differ in response to thermal acclimation ($Q_{10} = 1.04$) or diet treatment (Figure 4, Table 1, Figure S1, S2). Similarly, lipid peroxidation in liver tissue did not differ in response to temperature and showed a marginal, but insignificant effect of diet (Figure 4; Figure S3; temp: $df = 1$, $F = 0.318$, $p = 0.579$; diet: $df = 1$, $F = 3.260$, $p = 0.085$). Lactate dehydrogenase activity in white muscle was moderately affected by temperature acclimation ($Q_{10} = 2.10$), where lactate dehydrogenase activity was higher at 20°C compared to 12°C, but did not differ across diets (Figure 4, Figure S3). Lactate dehydrogenase activity also increased with acute temperature exposure (Figure S3). Overall, Q_{10} values for the

reaction norms differed dramatically across biological rates, with sprint speed having a Q_{10} of 1.04 (insensitive to temperature), while growth rate had a Q_{10} of 28.05 (highly sensitive to temperature) (Figure 4).

Thermal Tolerance

All thermal limits increased with acclimation to 20°C (Figure 5, Table 2). Upper thermal limits (CT_{max} , T_{AB} , T_{PEAK} , T_{ARR}) increased between 2.6-5.3°C and CT_{min} increased 2.1-2.3°C with warm acclimation (Figure 5, Table 2). Surprisingly, f_{Hmax} was significantly lower in the omnivorous treatments, which was driven by a 10% lower f_{Hmax} in the 20°C omnivorous treatment relative to the carnivorous treatment (Figure 5, Table 2). As expected, f_H followed the shape of an acute thermal performance curve (TPC), where it increased with acute temperature increase until T_{PEAK} , at which point f_H began declining with temperature until the onset of cardiac arrhythmia (T_{ARR} ; Figure 5A). Thermal acclimation to 20°C shifted the acute TPC for f_H to the right of the TPC at 12°C (Figure 5A). There was evidence to support an effect of diet on model selection for f_H in the warm ABT test, where the best fit model by AIC were 3rd order polynomial curves that incorporated an interaction of acclimation temperature, acute test temperature and diet (Figure 5A, Table S2). These models demonstrated that the acute TPC for f_H was lower across temperatures in the 20°C choice treatment compared to 20°C carnivorous treatment, which was consistent with the observed differences in f_{Hmax} (Figure 5A). Altogether, these results demonstrate that the omnivorous diet reduced overall cardiac function across temperatures at 20°C but did not affect the upper thermal tolerance limits on the heart.

The results of the cold ABT test showed similar evidence of an effect of diet on f_H and f_{Hmin} . Here, diet did not have a significant effect on any cold thermal limits (CT_{min} , $T_{AB-cold}$, $T_{ARR-cold}$), but f_{Hmin} was lower (16%) in the omnivorous treatment compared to the carnivorous treatment at 20°C. There was also evidence to support an effect of diet on model selection, where the best fit model by AIC was a 4th order polynomial curve that incorporated an effect of acute test temperature and diet (Figure 5C, Table S2).

Discussion

Using an omnivorous fish, opaleye, as a model species, we found evidence that diet can influence thermal performance in ectotherms and does so in trait-specific ways. This has critical

implications for our understanding of species responses to temperature change. Specifically, we examined three concepts outlined in Figure 1: 1) whether diet and temperature acclimation could affect thermal performance and limits, 2) whether ecologically important traits varied in their diet and temperature sensitivity (integrating multiple levels of biological organization), 3) if trait-specific variation in diet and temperature responses created performance trade-offs for opaleye fed an omnivorous diet compared to a carnivorous diet.

Thermal limits increased with temperature but did not differ across diet treatments

We assessed thermal limits using a standard and commonly used critical thermal (CT) test, as well as an Arrhenius breakpoint (ABT) test, which measures thermal limits of the heart (Casselman et al., 2012). The heart may be a primary regulator of functional thermal tolerance in fishes. It is responsible for oxygen, immune cell, metabolite, and waste transport around the body and is thought to be the first organ system to fail at extreme temperatures (Christen et al., 2018; Eliason and Anttila, 2017). The heart starts showing declines in performance (at T_{AB}) ~10 to 20°C lower and fails (T_{ARR}) at temperatures ~0 to 5°C lower than CT_{max} in fishes (Chen et al., 2015; Christen et al., 2018; Muñoz et al., 2014; Safi et al., 2019). As expected, all thermal limits increased with temperature acclimation and CT_{max} was 2.7-3.8°C higher than T_{ARR} . Opaleye showed a highly plastic acclimation response across all thermal limits, consistent with other temperate marine ectotherms (Vinagre et al., 2016). However, their thermal limits did not differ between diets.

While we did not observe a diet difference here, diet quality and quantity can alter thermal limits in ectotherms; (Fishes: Hoar and Cottle, 1952; Craig, Neill and Gatlin, 1995; Abdel-Ghany et al., 2019, Gomez et al, 2019; Lee et al., 2016; Turko *et al.*, 2020; Woiwode and Adelman, 1992). Most previous studies used formulated diets varying in lipid composition; thus, dietary lipid composition may be a primary factor affecting thermal limits. This is critical to consider in the context of aquaculture, where animal feeds should be designed to ensure farmed animals have adequate thermal performance and resistance to suboptimal temperatures. To our knowledge, this is the first study to test the effect of quasi-natural diets on the thermal acclimation of critical and cardiac thermal limits in ectotherms. Research with natural diets is ecologically relevant for ectotherms that consume broad diets and especially for fish like opaleye, that seem to change their diet in response to temperature (Behrens and Lafferty, 2007,

2012; Emde et al., 2016; González-Bergonzoni et al., 2016; Guinan et al., 2015; Vejříková et al., 2016). Climate change is altering the nutritional landscape for many aquatic ectotherms (Birnie-Gauvin et al., 2017; Huey and Kingsolver, 2019); managers and biologists should consider the effects of this on ectotherm thermal limits. Thus, future research should explore if other natural diets can influence thermal limits in ectotherms, as this has critical implications for how they may respond to global climate change.

Thermal acclimation responses did not scale with temperature and diet equally across biological rates

The biological rates assessed here did not scale with diet and temperature equally. Thermal sensitivity differed dramatically across biological rates, where sprint speed did not differ with diet and temperature acclimation ($Q_{10} = 1.04$), lactate dehydrogenase activity was moderately affected by temperature ($Q_{10} = 2.10$), but insensitive to diet, and growth was highly temperature sensitive ($Q_{10} = 25.74$), but also not sensitive to the diet treatments used in this study. These results are consistent with large scale meta-analyses, which have revealed that Q_{10} values differ across biological rates (Dell et al., 2011; Seebacher et al., 2015). Diet sensitivity was also inconsistent across rates, where SMR, and f_{Hmax} were sensitive to diet, but all other rates were not. This was surprising for rates that are known to be impacted by diet, such as growth rate. Diet has been shown to have interactive effects with temperature on growth in other ectotherms (Sengalese Sole: Guerreiro et al., 2012; Rohu: Mishra and Samantaray, 2004; Yellowtail Kingfish: Ilham and Fotedar, 2016; Crustaceans: Malzahn et al., 2016; Persson et al., 2011; Ruiz et al., 2020; Starke et al., 2020; Insects: Kingsolver et al., 2006; Lee and Roh, 2010). However, *Ulva* sp. supplementation has had mixed effects on growth in aquaculture fish, with some studies showing modest amounts of *Ulva* sp. reducing growth, while others finding positive or no effect of *Ulva* sp. supplementation on growth (Wan *et al.*, 2019). Future work should explore how broader diet differences affect growth in relation to other important biological rates.

The effects of temperature on sprint speed in comparison to lactate dehydrogenase activity were similarly unexpected. Sprinting in fish is primarily driven by glycolytic fast twitch white muscle (McDonald et al., 1998; Kraskura and Nelson, 2018). Lactate dehydrogenase is a critical enzyme in lactic acid fermentation during glycolysis. Thus, we expected that sprint performance would have a comparable thermal sensitivity to lactate dehydrogenase activity. Lactate

dehydrogenase activity increased with temperature acclimation, suggesting the opaleye had a greater anaerobic capacity at 20°C (McDonald et al., 1998). However, this did not translate to increases in sprint performance. Other enzymes in glycolysis could be rate limiting steps (e.g., phosphofructokinase; McDonald et al., 1998) and more highly correlated with the sprint speed. Sprint speed has been examined in one other diet x temperature study in fishes, which measured macronutrient selection and temperature effects on damselfish (Rowe et al., 2018). Macronutrient selection did not change with temperature, but sprint speed was highest in the colder temperature treatment (Rowe et al., 2018). Fish use two different swimming modes: anaerobically powered burst swimming and sustained aerobically powered swimming. These swimming modes may be differentially affected by diet and temperature. The effect of diet and temperature on aerobically powered swimming and maximum swim speeds (i.e., Maximum swimming speed, U_{\max} , and critical swimming speed, U_{crit}) have been assessed in grey mullet (Vagner et al., 2015; Vagner et al., 2019a), with no observed effect of diet, but an effect of temperature on U_{\max} and no effect of diet or temperature on U_{crit} . Given that two metabolic traits, SMR and factorial aerobic scope, were affected by diet and temperature in this study, future work should explore how aerobically powered swimming is impacted by different levels of omnivory.

The diverse diet and temperature responses that we observed across biological rates resulted in no consistent pattern in the thermal sensitivity of any metabolic rates or levels of biological organization. The small number of papers that have measured the interactive effects of diet and temperature on metabolic rates in other ectotherms have found mixed results (i.e., Insects: Alton et al., 2020; Schmitz and Rosenblatt, 2017; Fish: Pérez-Casanova et al., 2010; Vagner et al., 2015). For example, Vagner et al. (2015) found an interactive effect of dietary fatty acid composition and temperature on the absolute aerobic scope of juvenile grey mullet. In contrast, Pérez-Casanova et al. (2010) did not find any effects of macronutrient ratios on metabolism in juvenile Atlantic Cod and Haddock. Our results were especially surprising for SMR, as it is often assumed that maintenance metabolism is coupled to all other biological rates and should have the same thermal sensitivity (Gillooly et al., 2001; Brown et al., 2004). Aerobic scope is also considered a master physiological factor that determines the capacity for all other aerobically powered functions (Fry, 1947; Brett, 1971; Claireaux and Lefrançois, 2007; Pörtner, 2010); though, this idea has been heavily debated (Clark et al., 2013; Schulte, 2015). Here, absolute

aerobic scope did not differ in response to diet or temperature (though factorial aerobic scope did) and there was also no observed pattern between absolute aerobic scope, SMR, and other biological rates. These results indicate that caution be taken when using absolute aerobic scope and SMR as indicators of overall performance or proxies for other biological rates, especially when predicting the effects of multiple factors, such as diet and temperature.

Performance costs and benefits for diet choice at different acclimation temperatures

Contrary to our hypothesis, we did not find evidence of a performance trade-off to the omnivorous diet used in this study. Instead, opaleye that consumed the omnivorous diet displayed several higher costs: lower f_H across a thermal gradient, higher SMR, and reduced factorial aerobic scope when acclimated to 20°C. Given that the thermal limits of the heart did not change with diet, it was remarkable that diet downshifted the thermal performance curve for maximum heart rate. A reduction in maximum heart rate indicates a reduced capacity to transport oxygen, metabolites, immune cells, and waste around the body. There are several potential mechanisms that could have driven this reduction in maximum heart rate across acute temperatures. For example, differences in the lipid composition of the diet can impact membrane remodeling which can affect cardiac function in fish (Chatelier et al., 2006; McKenzie, 2001). Further, although *Ulva* sp. contains no known herbivore deterrents, our results suggest some sort of anti-nutrient effect of *Ulva* sp. supplementation which could have caused the observed reductions in cardiac thermal performance.

The higher SMR observed in the 20°C omnivorous treatment was expected as omnivorous and herbivorous animals generally have higher digestive infrastructure costs than carnivores (e.g., more broad digestive enzymes, higher gut surface area; Caruso and Sheridan, 2011; Clements et al., 2009; Horn, 1989). Consistent with the higher SMR at 20°C, the opaleye in the omnivorous treatment had a reduced factorial aerobic scope (3.43) that was less than half of that at 12°C (8.65). In contrast, the carnivorous diet maintained a 44% higher factorial aerobic scope at 20°C (4.94) compared to the omnivorous diet. Factorial aerobic scope is indicative of the amount of scope available to perform critical biological functions that scale proportional to SMR (Farrell, 2016). Digestion generally requires at least a doubling of SMR in many fishes (i.e., factorial aerobic scope of >2, Chabot et al., 2016; Farrell, 2016; McCue, 2006). This suggests that the opaleye in the omnivorous treatment at 20°C may have been on the threshold of having

limited scope for activities beyond digestion. As climate change increases the seasonal extreme temperatures and the frequency of marine heatwaves in the rocky intertidal (IPCC, 2019), the opaleye's factorial aerobic scope will decrease further, which could limit other important biological functions, like digestion, and exacerbate energetic tradeoffs associated with different diets. Thus, another avenue for future research will be to untangle diet x temperature effects on digestive costs (i.e. specific dynamic action) in relation to absolute aerobic scope and digestion efficiency (Jutfelt, 2021).

Given that opaleye and other omnivorous fishes eat more plants in warmer water in the wild, we expected there to be a performance advantage to the omnivorous diet. We measured a suite of traits to test for any performance benefits that may offset the costs of the omnivorous treatment. Specifically, we explored how diet and temperature affected 1) thermal tolerance, 2) maximum sprint speed 3) glycolytic capacity, and 4) oxidative stress. We predicted that because opaleye eat more plants in warmer water (Behrens and Lafferty, 2012), the omnivorous diet would result in higher thermal limits than the carnivorous diet at 20°C. However, we did not find evidence of any benefits to the omnivorous diet. We also expected that the higher lipid and protein content of the carnivorous treatment would raise oxidative stress relative to the omnivorous diet. Analysis of lipid peroxidation in liver tissue revealed a marginal trend of higher oxidative stress in the omnivorous diet treatment. However, antioxidant capacity could still have been higher in the omnivorous treatment. For example, Coggins et al. (2017) examined the effect of dietary glutathione supplementation on thermal limits, antioxidant capacity, and lipid peroxidation in *Daphnia* sp. As the glutathione concentration increased, so did total antioxidant capacity, but glutathione supplementation did not alter lipid peroxidation or thermal limits (Coggins et al., 2017). In contrast, Castro et al. (2012) examined the effect of macronutrient ratios (45% and 55% protein) and temperature acclimation (12 and 18°C) on multiple antioxidant enzymes and lipid peroxidation in juvenile Senegalese Sole. Lipid peroxidation differed across temperature treatments and was highest in the 55% protein diet (Castro et al., 2012). The limited amount of research on these interactions and inconclusiveness across studies indicate that more research is needed to elucidate the role of dietary antioxidants and macronutrient ratios in regulating oxidative stress across temperatures and taxa.

While it was not within the scope of this study, other important performance traits may have differed between the diet treatments. These include, but are not limited to, differences in

microbiome diversity and function, visual acuity, cognitive ability, digestion efficiency and digestive costs relative to aerobic scope, immune function, aerobic swimming performance, and cardiac stroke volume (Glencross and Rutherford, 2010; Koven et al., 2018; Vagner et al., 2014, 2019b). It remains unclear whether opaleye and other omnivorous ectotherms consume different proportions of plant: animal to regulate their thermal responses. Here, the opaleye's omnivorous diet did not maximize their performance compared to a more specialized carnivorous diet. However, many other factors govern diet choice in the wild, including life history (Zhang et al., 2020), competition (Pfenning, 1990), predation (Schmitz et al., 2016), food availability (MacArthur and Pianka, 1966), and habitat structure (Behrens and Lafferty, 2007). Therefore, the ecological benefits of consuming an omnivorous diet may outweigh the physiological costs.

Many ectotherms change their diets with temperature; either directly because their diet preference changes in response to temperature (Boersma et al., 2016b; Carreira et al., 2016; Devries and Appel, 2014; Lee et al., 2015; Lemoine et al., 2014; Rho and Lee, 2017; Schmitz et al., 2016; Vejřiková et al., 2016) or indirectly because food availability or the nutritional content of a diet item changes with temperature (Alton et al., 2020; Boersma et al., 2016b; Cross et al., 2015; Ho et al., 2010). In either scenario, generalist ectotherms that have the capacity to adjust their diet may be at an advantage compared to those with more specialized inflexible diets. However, the ultimate diet choices that ectotherms make may not always be “better” or “worse” because diet and temperature can interact and have trait-specific effects. Not all diet choices are necessarily adaptive. Irrespective of the reasons for *why* an ectotherm eats what it eats, our work here demonstrates that diet choices have consequences. These consequences have far-reaching implications, including whether diet choice can facilitate geographic range expansion, or colonization of warmer or cooler habitats. Further, if specialist diets constrain thermal niches or if diet can facilitate different acclimation rates or performances under fluctuating temperatures. Overall, diet should be treated as an interacting factor that has the capacity to modify the thermal responses of ectotherms.

Concluding Remarks

Thermal acclimation is a key mechanism that ectotherms employ to maintain performance across a range of temperatures. Acclimation requires energy and nutritional building blocks that ectotherms obtain from their diet. Here we explored whether different diets mediated distinct

thermal acclimation responses in an omnivorous fish, opaleye. We found clear evidence that diet influences thermal acclimation responses. However, there was no consistent pattern in how different biological rates responded to the temperature and diet treatments, with Q_{10} values ranging from 1-25. When confronted with a seasonal warm temperature (20°C), the opaleye in the omnivorous diet treatment had inferior performance (higher SMR, lower factorial aerobic scope, and lower cardiac performance) relative to the opaleye fed a carnivorous diet treatment. Global climate change is already changing the average and extreme temperatures that marine ectotherms experience as well as their nutritional landscape (IPCC, 2019; Birnie Gauvin, 2017). These environmental changes are likely to interact and alter many ectotherms' thermal performance in the wild. Incorporating multiple interacting factors into our understanding of species responses to global climate change is the next step in ensuring that researchers capture the resilience of different species and populations (Jackson et al., 2021). Accordingly, diet is essential to consider when predicting ectotherm performance in variable environments and in response to global climate change.

Symbols and Abbreviations

SMR: Standard metabolic rate

MMR: Maximum metabolic rate

ABT test: Arrhenius breakpoint test

T_{AB} : Breakpoint temperature on the heart

T_{PEAK} : Temperature corresponding to maximum heart rate

T_{ARR} : Temperature of the first cardiac arrhythmia

F_{Hmax} : Maximum overall heart rate

CT_{max} : Critical thermal maxima

CT_{min} : Critical thermal minima

LDH: Lactate dehydrogenase

LPO: Lipid peroxidation

Acknowledgements

We thank Mason Tittle, Vincent Han Lee, Samantha Csik, Bella Giglio, and Terra Dressler for assistance in the lab and with fishing. We thank Bartholomew Difiore, Joseph

Curtis, Sevan Esaian, Nicholas Lee, Hope Hardison, Osborne Hardison, Brendan Shanny, Dr. Alexander Lill, Terra Dressler, Dr. Alexander Little, Bella Giglio, Anhadh Jassal, Claire Anderson, Jennay Argiris, Garret Parsons, Tyler Parsons, and Alecia Dezzani for help fishing. We thank David Davis and the entire marine operations staff at the University of California, Santa Barbara. We thank Dr. Adrian Stier and Dr. Elizabeth Wilbanks for lending equipment. We thank Dr. Christopher Jerde, Dr. Gretchen Hofmann, Dr. Kevin Lafferty, Dr. Alexander Little, and Dr. Elizabeth Wilbanks for advice and feedback.

Author Contributions

EAH and EJE designed the experiment and wrote the manuscript. EAH, KK, JWV, TN, EJE collected the data. KK and TN ran the sprint trials and assisted in the sprint data analyses. EAH led all experiments, performed data analysis, and constructed the figures. KK wrote the R package used to analyze the aquatic respirometry data. KK, JWV, TN reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Funding

This work was supported by a Hellman Family Faculty Fellowship and the University of California, Santa Barbara. Additional funding for EAH and JWV was provided by the National Science Foundation Graduate Research Fellowships. Additional support for TN was provided by the University of California LEADS Program.

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Figures

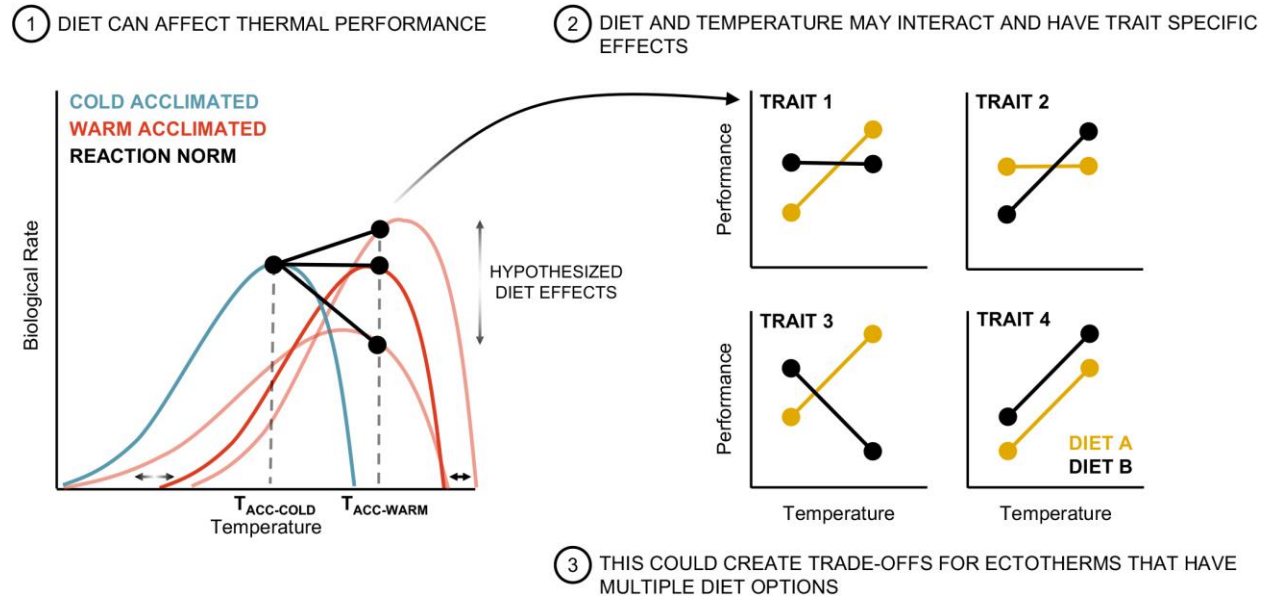


Figure 1. Conceptual graphs illustrating how diet may affect thermal acclimation responses and how those effects could be trait specific. The graph on the left side of the figure shows how acute thermal performance curves (TPC) shift towards the acclimation temperature, where blue is the acute TPC after acclimation to cold and red is the acute TPC after acclimation to warm conditions. Note that both cold and warm acclimation may be affected by diet, but only potential effects of warm acclimation are displayed for simplicity. (1) Diet may influence the shape (height and breadth) of those acute thermal performance curves (indicated by pink curves) or the location of the curve along the X axis (i.e., temperature of peak performance). These effects could influence the slope of the line between reaction norms (performance at acclimation conditions, indicated by black lines). The right side of the figure shows a series of hypothetical reaction norm plots for various traits. (2) These traits can have different diet and temperature sensitivities, (3) which could create performance trade-offs for ectotherms consuming different diets.

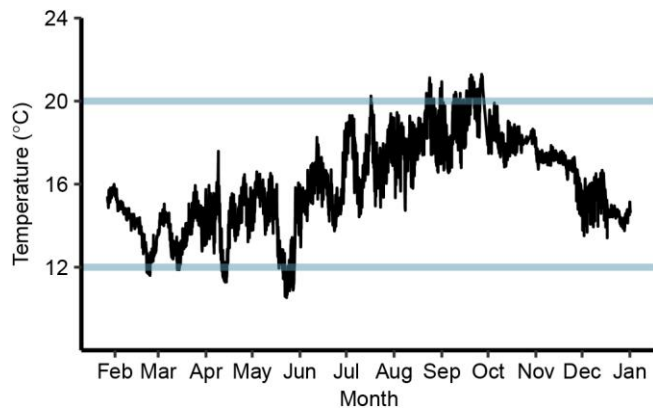


Figure 2. Temperature data collected in 2019 at Stearns Wharf in Santa Barbara, CA, USA (located next to SB Harbor where fish were collected). The temperature sensor was mounted at 2m depth and set to record every 4 min. Blue lines indicate treatment temperatures (12 and 20°C). Data source: Southern California Coastal Ocean Observing System (SCCOOS); https://www.sccoos.org/data/autoss/timeline/?main=single&station=stearns_wharf.

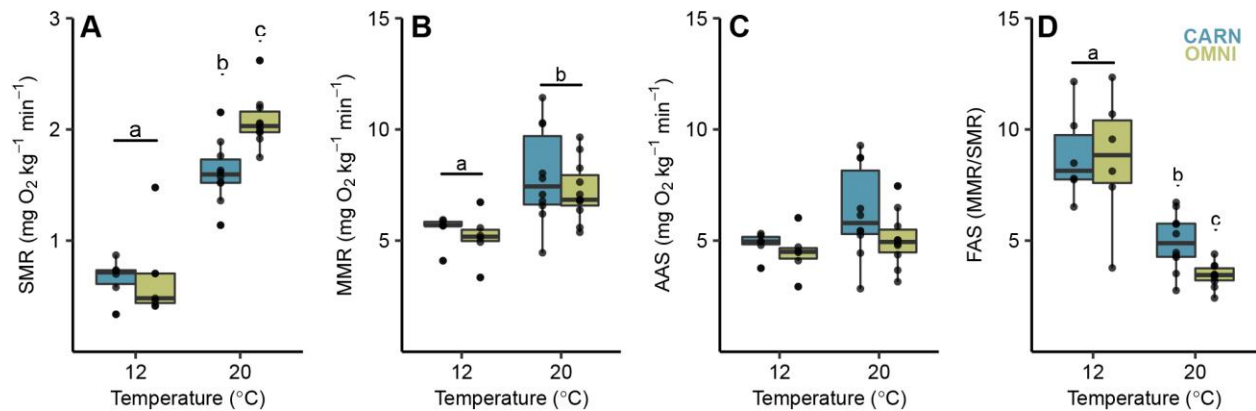


Figure 3. Oxygen uptake rate (MO_2 ; $\text{mg O}_2 \text{ kg}^{-1} \text{ min}^{-1}$) after an exhaustive chase protocol in opaleye acclimated to 12 °C or 20 °C and fed either a carnivorous (*Artemia* sp., represented as blue) or omnivorous (*Artemia* and *Ulva* spp., represented as green) diet. Presented are **A**) standard metabolic rate (SMR), **B**) maximum metabolic rate (MMR), **C**) absolute aerobic scope (AAS; $\text{MMR}-\text{RMR}$), and **D**) factorial aerobic scope (FAS; MMR/RMR). Lower case letters

indicate significant difference ($p < 0.05$) between treatment groups where applicable (see Table S1 for 2-way ANOVA outputs). Boxplots represent interquartile ranges (boxes and whiskers), median values (solid lines) and outliers (> 1.5 beyond interquartile range) plotted as data points outside the whiskers.

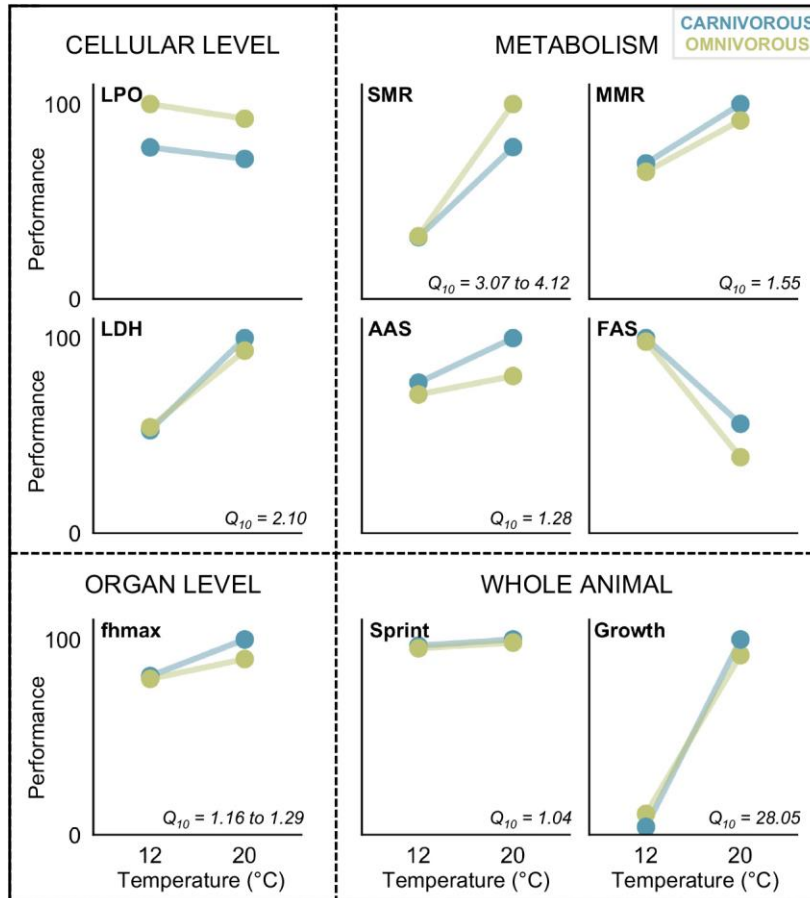


Figure 4. Reaction norms plotted across all measured biological rates and traits. Average data from each treatment is scaled to the max average treatment value (i.e., the max treatment value is equal to 100%). All values are from fish tested at their acclimation temperature. Graphs are organized by level of biological organization (cellular, organ, whole animal) and labelled by the trait (SMR = standard metabolic rate, MMR = maximum metabolic rate, AAS = absolute aerobic scope (MMR – SMR), FAS = factorial aerobic scope (MMR/SMR), f_{Hmax} = maximum overall heart rate, Sprint = maximum sprint speed, Growth = growth rate, LPO = lipid peroxidation, LDH = lactate dehydrogenase activity). Q10 values are listed below all biological rate

measurements (i.e., everything except LPO and FAS), and are listed as a range when the diet treatments were statistically different and an individual value when the diet treatments were not statistically different. Lines and circles indicate reaction norms and colors indicate diet treatment (*Artemia* sp., carnivorous diet in blue and *Artemia* and *Ulva* sp. omnivorous diet in green).

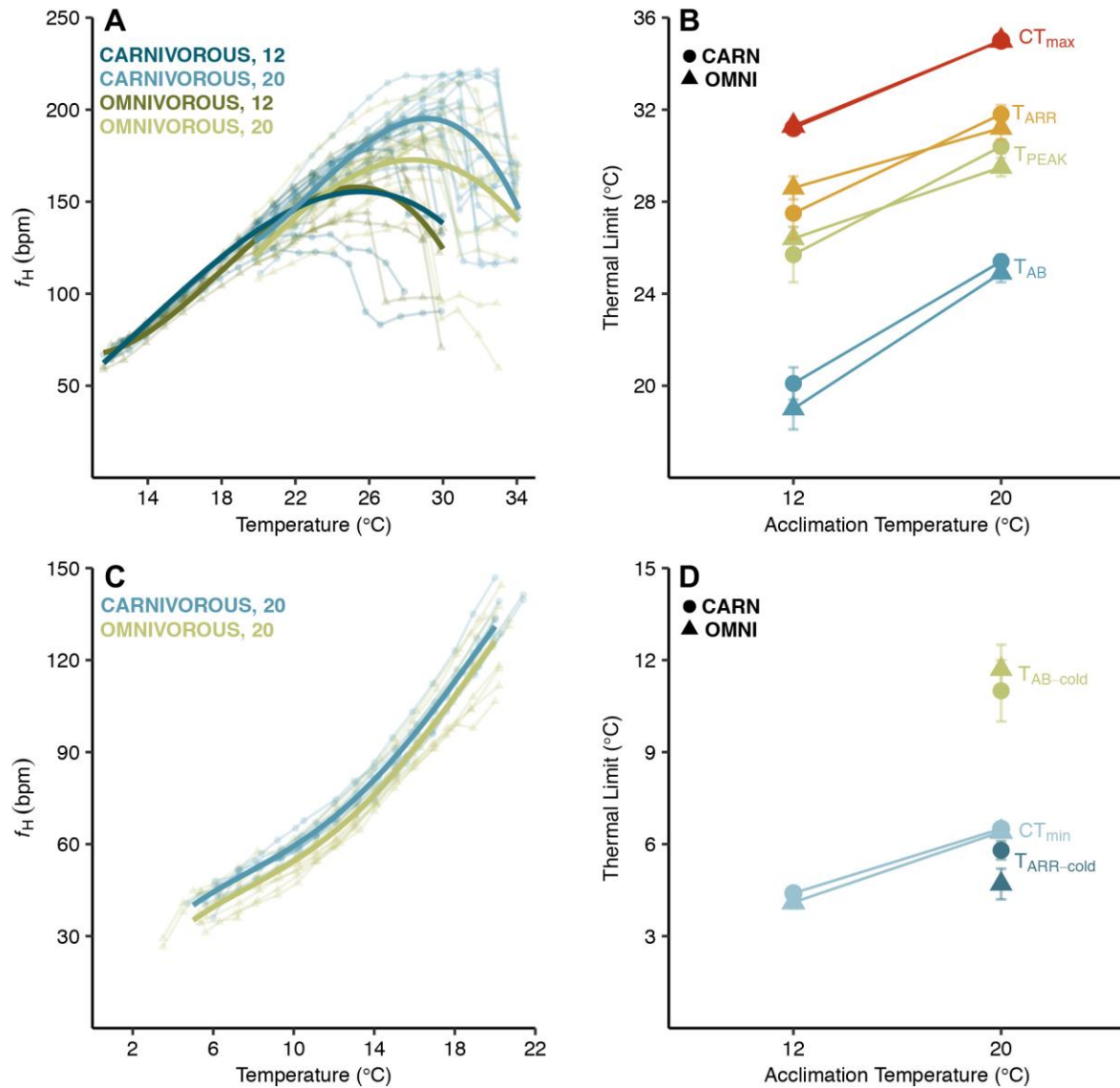


Figure 5. A) Individual and treatment level responses of maximum heart rate (f_H) of opaleye during acute warming from 12 $^{\circ}\text{C}$ (dark blue =carnivorous diet, N= 6 and dark green =omnivorous diet, N= 7) or 20 $^{\circ}\text{C}$ (light blue =carnivorous diet, N= 13 and light green

=omnivorous diet, N= 14). Curves are 3rd order polynomials that account for an interaction diet × acclimation temperature × acute temperature. This was determined to be the best fit model by AIC (Table S2). **B)** Upper thermal limits across diet and temperature treatments in opaleye. Circles and triangles indicate mean (\pm SEM) values for the carnivorous and omnivorous diet treatments, respectively. **C)** Individual and treatment level responses in f_H from opaleye during acute cooling from 20 °C (light blue (carnivorous diet; N= 9) and light green (omnivorous diet); N= 11). Curves are 4th order polynomials that account for a fixed effect of diet and acute temperature. This was determined to be the best fit model by AIC (Table S2). **D)** Lower thermal limits across diet and temperature treatments in opaleye. Circles indicate mean (\pm SEM) values for the carnivorous diet treatment and triangles indicate mean (\pm SEM) values for the omnivorous diet treatments. Note that f_H limits are missing from the 12 °C treatments due to issues surrounding COVID-19. T_{ARR} = temperature of the onset of cardiac arrhythmia, T_{AB} = breakpoint temperature of the heart, T_{PEAK} = temperature corresponding to maximum heart rate, CT_{max} = critical thermal maximum, CT_{min} = critical thermal minimum.

Tables

Table 1. Summary Statistics for Biological Rates

Biological Rate	Temp (°C)	Results				Statistical Parameters								
		Carnivorous		Omnivorous		Diet			Temperature			Diet X Temperature		
		n	mean ± SEM	n	mean ± SEM	df	F	P	df	F	P	df	F	P
Standard Metabolic Rate (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	0.66 ± 0.07	7	0.67 ± 0.14	1	6.846	0.007	1	130.9	<0.001	1	4.065	0.029
	20	10	1.62 ± 0.09	10	2.08 ± 0.07									
Maximum Metabolic Rate (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	5.49 ± 0.28	6	5.15 ± 0.45	1	1.364	0.311	1	19.85	<0.001			
	20	10	7.89 ± 0.69	11	7.23 ± 0.40									
Absolute Aerobic Scope (mg O ₂ kg ⁻¹ min ⁻¹)	12	6	4.84 ± 0.23	6	4.46 ± 0.41	1	3.704	0.087	1	4.677	0.066			
	20	10	6.27 ± 0.66	10	5.05 ± 0.40									
Factorial Aerobic Scope (MMR/SMR)	12	6	8.81 ± 0.83	6	8.65 ± 1.21	1	5.796	0.023	1	52.24	<0.001			
	20	10	4.94 ± 0.41	10	3.43 ± 0.17									
Growth (g week ⁻¹)	12	3	0.03 ± 0.19	3	0.08 ± 0.01	1	0.003	0.958	1	23.498	<0.001			
	20	3	0.75 ± 0.15	3	0.69 ± 0.16									
Sprint Speed (cm s ⁻¹)	12	10	112.36 ± 4.88	9	110.85 ± 5.26	1	0.099	0.756	1	0.456	0.504			
	20	8	116.10 ± 5.46	8	114.26 ± 6.01									

Represented are means and standard error (SEM) values for each test group, and ANOVA results. Temp = temperature, df = degrees of freedom, F = F-statistic, P = P-value.

Table 2. Summary Statistics for all Thermal Limits.

Thermal Limit	Temp (°C)	Results				Statistical Parameters					
		Carnivorous		Omnivorous		Diet			Temperature		
		n	mean ± SEM	n	mean ± SEM	df	F	P	df	F	P
CT _{min}	12	9	4.4 ± 0.2	10	4.1 ± 0.2	1	1.984	0.168	1	226.21	<0.001
	20	12	6.5 ± 0.1	8	6.4 ± 0.1						
T _{AB}	12	6	20.1 ± 0.7	7	19.0 ± 0.9	1	2.13	0.153	1	110.74	<0.001
	20	13	25.4 ± 0.3	14	24.9 ± 0.4						
T _{PEAK}	12	6	25.7 ± 1.2	7	26.4 ± 0.5	1	0.612	0.439	1	46.99	<0.001
	20	13	30.4 ± 0.3	14	29.5 ± 0.4						
T _{ARR}	12	6	27.5 ± 1.1	7	28.6 ± 0.5	1	0.0208	0.886	1	33.766	<0.001
	20	13	31.8 ± 0.4	14	31.2 ± 0.5						
CT _{max}	12	10	31.2 ± 0.2	10	31.3 ± 0.2	1	0.547	0.465	1	500.08	<0.001
	20	8	35.0 ± 0.2	10	35.0 ± 0.1						
overall <i>f</i> _{Hmax}	12	6	166.6 ± 12.5	7	163.5 ± 6.2	1	5.243	0.028	1	17.669	<0.001
	20	13	204.8 ± 4.4	14	184.4 ± 5.4						
Thermal Limit	Temp (°C)	n	mean ± SEM	n	mean ± SEM	df	T	P			
T _{ARR-cold}	12	NA	NA	NA	NA	18	1.79	0.09			
	20	9	5.8 ± 0.3	11	4.7 ± 0.5						
T _{AB-cold}	12	NA	NA	NA	NA	14	-0.493	0.6295			
	20	8	11.0 ± 1	8	11.7 ± 0.8						
overall <i>f</i> _{Hmin}	12	NA	NA	NA	NA	18	2.407	0.027			
	20	9	44.5 ± 1.9	11	37.4 ± 2.2						

Represented are means and standard error values for each test group, and ANOVA results. Note that T-Test results are reported on the results of the cold ABT test, as this test was only run on 20°C acclimation fish due to complications surrounding COVID-19. T_{ARR} = Temperature of the onset of cardiac arrhythmia, T_{AB} = breakpoint temperature of the heart, T_{PEAK} = temperature corresponding to maximum heart rate, *f*_{Hmax} = Maximum heart rate across entire hot ABT test, *f*_{Hmin} = minimum heart rate during cold ABT test, CT_{max} = critical thermal maximum, CT_{min} = critical thermal minimum, Temp = temperature, df = degrees of freedom, F = F-statistic, P = P-value.

Supplemental Methods

Proximate Analysis

Frozen fish remains were homogenized using a Fisher Brand Bead Mill 24 and subsamples of the homogenate were weighed and freeze dried (Labconco Lyophilizer). *Protein:* Protein content was estimated in triplicate (intra-assay CV% <10%) using a BCA assay with a 72% TCA precipitation (Pierce BCA kit, ThermoFisher Scientific, MA, USA), where absorbance was measured at 562 nm. *Lipids:* Lipid content was estimated using a chloroform:methanol extraction as described in Mann and Gallager, 1985 and Johnson et al, 2017. Lipids from 50 mg of freeze-dried homogenized sample were extracted using 100 ul milliQ water and 1.5 ml chloroform:methanol (1:2) (vortexed, incubated at 4°C, centrifuged at 4000 rpm for 5 min). The supernatant was removed and remaining sample was re-extracted in 1.5 ml chloroform:methanol (2:1). The supernatants were pooled, mixed with 950 ul NaCl (0.7%), incubated at 4°C for 30 min, then centrifuged (4000 rpm, 5 min), and the volume of the bottom layer was measured. Dried subsamples of the bottom layer were used to extrapolate lipid content to the entire sample. *Ash Content:* Ash content was determined by drying freeze-dried samples overnight at 100 °C to account for any moisture that returned during sample storage. Samples were then weighed (~30 mg) before being combusted in a muffle furnace at 450°C for 12 h and then re-weighed.

Table S1. Dietary and whole-body Proximate composition (% wet weight)

Dietary Proximate composition (% wet weight)				
	Experiment 1		Experiment 2	
	<i>Ulva</i>	<i>Artemia</i>	<i>Ulva</i>	<i>Artemia</i>
% Moisture	82.04 ± 1.63	87.48 ± 0.91	75.33 ± 3.81	86.83 ± 0.38
% Protein	1.47 ± 0.27	4.75 ± 0.51	1.95 ± 0.88	5.59 ± 0.62
% Lipid	0.55 ± 0.06	1.58 ± 0.18	0.71 ± 0.13	2.39 ± 0.10
% Ash	10.71 ± 1.87	1.44 ± NA	9.93 ± NA	1.78 ± 0.04
Whole body Proximate composition (% wet weight)				
	12°C		20°C	
	Carnivorous	Omnivorous	Carnivorous	Omnivorous
% Moisture	70.15 ± 1.15	72.25 ± 1.40	72.98 ± 0.74	71.79 ± 0.45
% Protein	13.40 ± 1.09	13.91 ± 1.21	12.61 ± 1.45	10.76 ± 0.85
% Lipid	5.01 ± 0.54	3.98 ± 0.33	4.80 ± 0.82	5.11 ± 0.20
% Ash	5.45 ± 0.84	4.31 ± 0.50	4.60 ± 0.60	5.26 ± 0.60

Represented are means and standard error values for dietary proximate composition in *Ulva* sp., *Artemia* sp., and proximate body composition from whole opaleye from experiments 1 and 2. Proximate body composition were statistically analyzed using 2-way ANOVA and no significant differences were found between treatment groups. When sample size <3 standard error was not calculated and is listed as NA.

Table S2. AIC Outputs for Polynomial Curves.

AIC outputs for warm ABT test f_{hmax} polynomial curves				
Model	Formula	df	AIC	Δ AIC
Model 1	poly(acute_temp, 3) * diet * temp + (1 fish_id)	18	5282.61153	0
Model 2	poly(acute_temp, 3) * temp + diet + (1 fish_id)	11	5295.12706	12.515531
Model 3	poly(acute_temp, 3) * temp + (1 fish_id)	10	5297.84122	15.2296905
Model 4	poly(acute_temp, 2) * diet * temp + (1 fish_id)	14	5331.73874	49.1272062
Model 5	poly(acute_temp, 3) * diet + temp + (1 fish_id)	11	5425.26489	142.653365
Model 6	poly(acute_temp, 4) + temp + diet + (1 fish_id)	9	5427.25882	144.64729
Model 7	poly(acute_temp, 4) + temp * diet + (1 fish_id)	10	5428.15216	145.540635
Model 8	poly(acute_temp, 3) + temp + diet + (1 fish_id)	8	5429.2756	146.664069
Model 9	poly(acute_temp, 3) + temp * diet + (1 fish_id)	9	5430.17554	147.564011
Model 10	poly(acute_temp, 3) + temp + (1 fish_id)	7	5432.27141	149.659875
Model 11	poly(acute_temp, 3) + diet + (1 fish_id)	7	5433.3873	150.775766
Model 12	poly(acute_temp, 3) + (1 fish_id)	6	5435.91176	153.300235
Model 13	poly(acute_temp, 2) + temp + diet + (1 fish_id)	7	5491.9684	209.356872
Model 14	poly(acute_temp, 2) + temp * diet + (1 fish_id)	8	5492.77923	210.167701
Model 15	acute_temp + temp + diet + (1 fish_id)	6	5742.45159	459.840058
Model 16	acute_temp + temp * diet + (1 fish_id)	7	5743.27212	460.660589
Model 17	acute_temp + temp + (1 fish_id)	5	5745.43438	462.822848
Model 18	acute_temp + diet + (1 fish_id)	5	5749.62894	467.017406
Model 19	acute_temp + (1 fish_id)	4	5751.89944	469.287913
AIC outputs for cold test f_{hmax} polynomial curves				
Model	Formula	df	AIC	Δ AIC
Model 1	poly(acute_temp, 4) + diet + (1 fish_id)	8	1620.06402	0
Model 2	poly(acute_temp, 4) * diet + (1 fish_id)	12	1623.74238	3.67835332
Model 3	poly(acute_temp, 3) + diet + (1 fish_id)	7	1636.74603	16.6820023
Model 4	poly(acute_temp, 3) * diet + (1 fish_id)	10	1639.36915	19.3051258
Model 5	poly(acute_temp, 3) + (1 fish_id)	6	1640.02832	19.9642973
Model 6	poly(acute_temp, 2) * diet + (1 fish_id)	8	1640.27173	20.2077023
Model 7	poly(acute_temp, 2) + diet + (1 fish_id)	6	1641.6658	21.6017758
Model 8	acute_temp + diet + (1 fish_id)	5	1992.02421	371.960185
Model 9	acute_temp + (1 fish_id)	4	1994.25531	374.191287

Represented are model formulas as input into R and AIC output results. df = degrees of freedom, AIC = Akaike Information Criterion Δ AIC = AIC(model) – AIC(min AIC value), acute_temp = acute temperature, fish_id = individual fish.

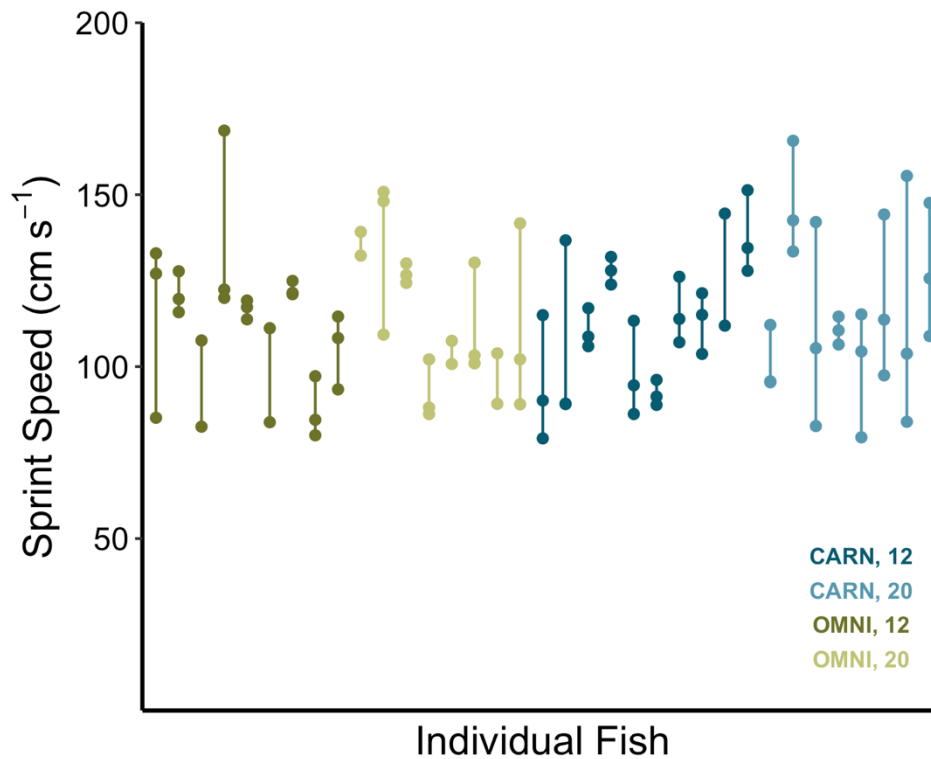


Fig. S1. Figure illustrating repeatability of sprint performance across individuals. Each dot indicates a max sprint performance (cm s^{-1}) calculated from an individual sprint trial. Colors indicate treatments with dark blue (carnivorous diet at 12 °C), dark green (omnivorous diet at 12 °C), light blue (carnivorous diet at 20 °C), light green (omnivorous diet at 20 °C).

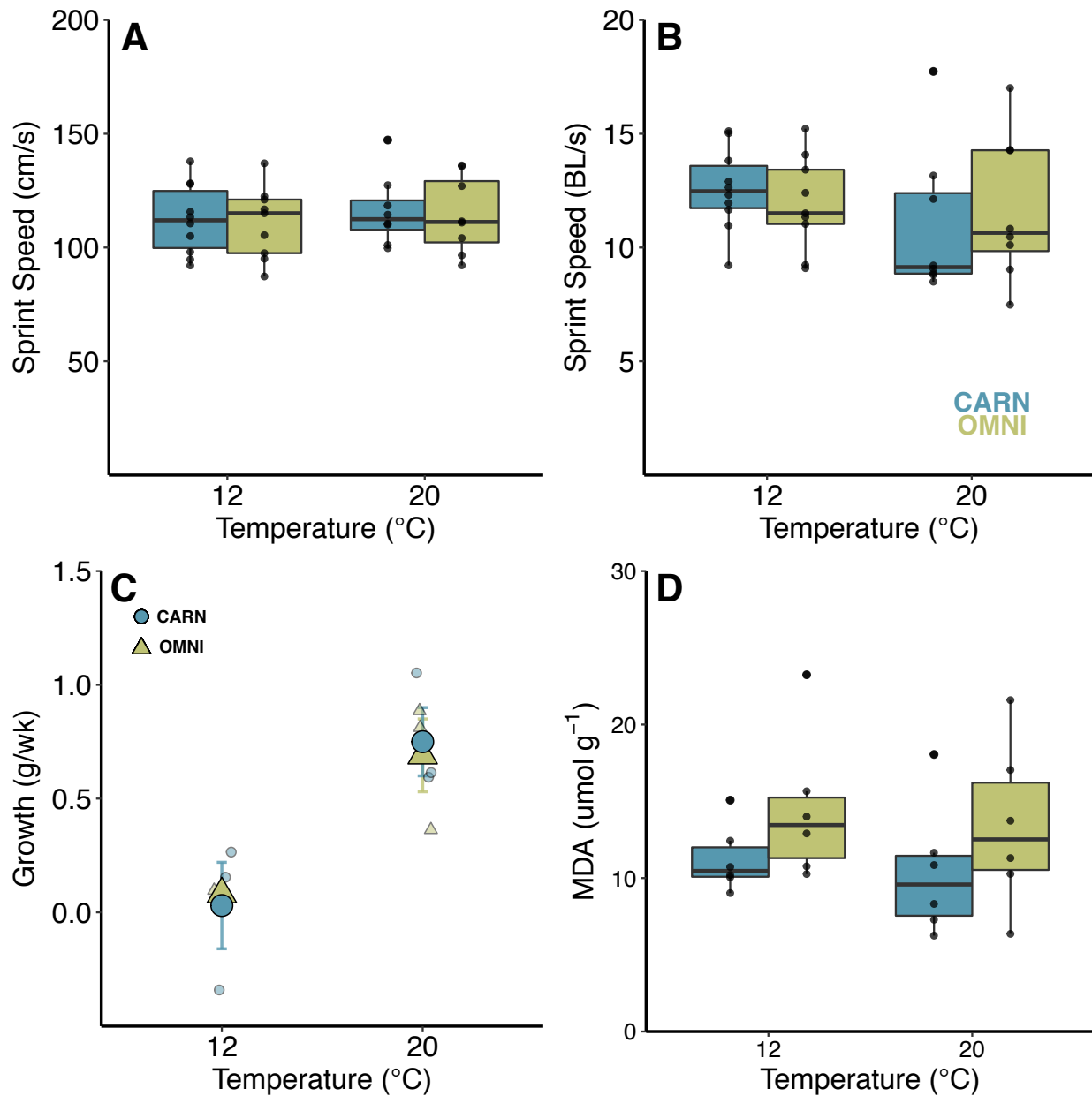


Fig. S2. Performance in opaleye acclimated to 12°C or 20°C and fed either a carnivorous (blue) or omnivorous (green) diet. Presented are **A**) sprints measured as speed in cm s⁻¹ and **B**) sprints measured as speed in BL s⁻¹, **C**) Growth rate (average fish mass (g) gained per week per tank) **D**) Lipid Peroxidation (LPO) in liver tissue measured as Malonhydride concentration (MDA) in umol gram⁻¹ of liver tissue. In panel A, B, D box plots represent interquartile ranges (boxes and whiskers), median values (solid lines) and outliers (> 1.5 beyond interquartile range) are plotted as data points outside the whiskers. In panel C, large circles and triangles indicate mean (± SEM) values for the carnivorous (*Artemia* sp.) and omnivorous diet treatments (*Artemia* sp. and *Ulva* sp.), respectively.

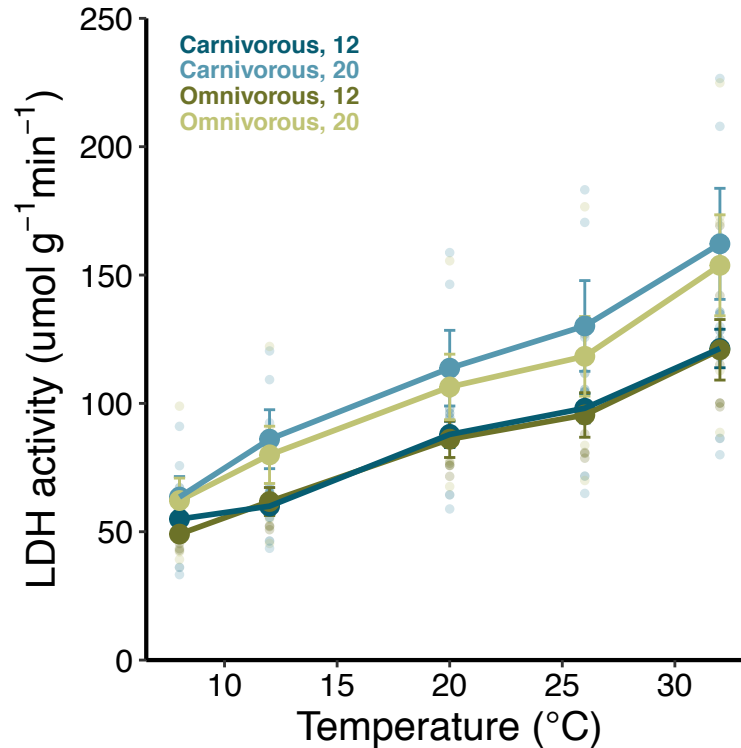


Fig. S3. Lactate dehydrogenase (LDH) activity in μmol per gram wet white muscle tissue weight in opaleye acclimated to 12°C (dark colors) or 20°C (light colors) and fed either a carnivorous (*Artemia* sp., represented as blues) or omnivorous diet (*Artemia* sp. and *Ulva* sp., represented as greens). Circles represent mean values and error bars indicate SEM. For each sample, LDH activity was measured at 5 different temperatures (8, 12, 20, 26, 32°C). Lactate dehydrogenase activity was higher at 20°C compared to 12°C but did not differ across diets. Lactate dehydrogenase activity also increased with acute temperature exposure. Acute temp: $df = 4$, $\chi^2 = 1061.711$, $p < 0.001$; acclimation temp: $df = 1$, $\chi^2 = 5.132$, $p = 0.023$; diet: $df = 1$, $\chi^2 = 0.172$, $p = 0.679$; acute temp \times acclimation temp: $df = 4$, $\chi^2 = 22.526$, $p < 0.001$.

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