# Acceleration-triggered animal-borne videos show a dominance of fish in the diet of female northern elephant seals

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**Summary statements:** Seal-borne acceleration-triggered video cameras showed that female northern elephant seals feed mainly on fish rather than squid, in the mesopelagic zones across eastern North Pacific.

#### **ABSTRACT**

Knowledge of the diet of marine mammals is fundamental to understanding their role in marine ecosystems and response to environmental change. Recently, animal-borne video cameras have revealed the diet of marine mammals that make short foraging trips. However, novel approaches that allocate video time to target prey capture events is required to obtain diet information for species that make long foraging trips over great distances. We combined satellite telemetry and depth recorders with newly developed date/time-, depth-, and acceleration-triggered animal-borne video cameras to examine the diet of female northern elephant seals during their foraging migrations across the eastern North Pacific. We obtained 48.2 hours of underwater video, from cameras mounted on the head (n = 12) and jaw (n = 3) of seals. Fish dominated the diet (78% of 697 prey items recorded) across all foraging locations (range: 37-55°N, 122-150°W), diving depths (range: 238-1167 m) and water temperatures (range: 3.2-7.4 °C), while squid comprised only 7% of the diet. Identified prey included fishes such as myctophids, Merluccius sp., and Icosteus aenigmaticus, and squids such as Histioteuthis sp., Octopoteuthis sp., and Taningia danae. Our results corroborate fatty acid analysis, which also found that fish are more important in the diet and contrasts to stomach content analyses that found cephalopods to be the most important component of the diet. Our work shows that in-situ video observation is a useful method for studying the at-sea diet of long-ranging marine predators.

**KEY WORDS:** bio-logging, diving behavior, foraging, marine mammal, mesopelagic zone.

#### **INTRODUCTION**

Marine mammals play an important role in marine ecosystems as major consumers of a wide variety of prey (Estes et al., 2016). Knowledge regarding when, where, and on what type of prey marine mammals feed is fundamental to understand their roles in marine ecosystems and their responses to marine environmental changes (Costa et al., 2012; Miloslavich et al., 2018; Bax et al., 2019). Recent developments in bio-logging technologies have allowed us to obtain information about when and where marine mammals feed on their prey. For example, animal-borne satellite transmitters or GPS loggers are routinely used to identify hotspots where marine mammals focus their foraging (Block et al., 2011; Costa et al., 2012; Hussey et al., 2015). In addition, instruments such as stomach temperature loggers or accelerometers attached to the head or jaw of animals identify when animals capture and consume prey, at a fine temporal scale (Kuhn et al., 2009; Naito et al., 2013; Guinet et al., 2014). However, these technologies are not able to identify what type of prey marine mammals consume in the open ocean.

Marine mammal diet has been studied by various techniques, such as stomach content analysis, scat content or scat DNA analysis, stable isotopes, and fatty acid analyses (reviewed in Tollit et al., 2010). Stomach content analysis allows the direct assessment of prey that was consumed by marine mammals (Antonelis et al., 1987, 1994; Field et al., 2007). However, this method has inherent biases toward prey with hard parts and recently-consumed items (Harvey and Antonelis, 1994). Similarly, scat content analysis has been widely used but also presents challenges to identify prey items that are highly digested as well as inherent biases toward prey items with hard parts (Gales and Cheal, 1992; Tollit et al., 2010). More recently, scat DNA analysis has been developed to help address the identification and bias issues, but there are still unavoidable limitations as only recently consumed items will be present in scat. Stable isotope (Cherel et al., 2008; Hückstädt et al., 2012) and fatty acid analyses (Bradshaw et al., 2003) allow us to estimate the diet that is consumed (and incorporated) over a relatively long period, though, some necessary information such as isotopic and fatty acid signatures of prey are often difficult to obtain, limiting the ability to precisely estimate the diet composition (but see Goetsch et al., 2018). Hence, new developments are still necessary to identify the types of prey that marine mammals feed on at sea, and

these should operate at a temporal resolution that matches the behavioral data on where and when they feed.

Recently, animal-borne videos have been used successfully to quantify at-sea diet compositions of relatively short-ranging pinnipeds, such as Weddell seals *Leptonychotes weddellii* (Davis et al., 1999), harbor seals *Phoca vitulina* (Bowen et al., 2002), Antarctic fur seals *Arctocephalus gazella* (Hooker et al., 2002) and Australian fur seals *Arctocephalus pusillus doriferus* (Kernaléguen et al., 2016). This approach has allowed us to examine detailed at-sea diet at a fine temporal scale. However, the duration of the video recordings is limited to several hours or days by battery/memory capacity, unlike the behavioral data obtained from satellite transmitters or time-depth recorders that can last for several months. Moreover, successful video recordings of prey captures are often limited, because animal-borne cameras do not target areas where prey capture events occur (e.g. depth), especially in situations where rates of prey encounters are relatively low. Hence, use of video to estimate diet has largely been confined to breeding seasons, when it is possible to recover videos after a few days. It has not been applied, however, to study the diet of the wide-ranging species that spend prolonged periods of time (weeks to months) feeding at sea.

Northern elephant seals *Mirounga angustirostris* are upper-trophic level predators of the North Pacific, and their colonies extend from California (USA) to Mexico. Female northern elephant seals feed primarily at depths between 400 to 600 m (Le Boeuf et al., 2000; Robinson et al., 2012), with maximum dive depths over 1000 m (Robinson et al., 2012; Naito et al., 2013). They move over long distances during biannual foraging migrations that last approximately 75 or 220 days (Le Boeuf et al., 2000; Robinson et al., 2012). Their migration routes and diving behaviors have been widely studied by using satellite transmitters or time-depth recorders (Simmons et al., 2010; Robinson et al., 2012). However, relatively few studies have examined the diet of northern elephant seals while they are at sea. Previous studies using stomach content analysis (Antonelis et al., 1987, 1994) suggested that seals fed more on squid than fish. Yet, more recently, Naito et al. (2013, 2017) proposed that northern elephant seals feed on myctophids and ragfish in the mesopelagic zone, based on several images and videos obtained from animal-borne still and video cameras. Similarly, Goetsch et al. (2018)

reported that energy-rich mesopelagic fishes, such as myctophids, are more important in the diet than squid, based on quantitative fatty acid signature analyses.

This study examined the at-sea diet of female northern elephant seals using digital video, obtained from recently-developed video loggers with a feeding-triggered (acceleration) system (Naito et al., 2017). As described in Naito et al. (2017), the video camera remains in a power-saving mode until a predetermined date, at which time individual video recordings are then triggered when a predetermined depth and acceleration threshold are reached. This allows for the efficient video recording of prey capture events. Studies on the congeneric southern elephant seals (*M. leonina*) suggested that their diet varies spatially (Daneri et al., 2000). We, therefore, hypothesized that the diet of female northern elephant seals differs among foraging areas, foraging depths, or water temperatures, and we tested these hypotheses by using information obtained from satellite transmitters and time-depth/temperature recorders that were attached to elephant seals outfitted with video cameras.

### MATERIALS AND METHODS

## Field experiments and instruments

Fieldwork was conducted at Año Nuevo State Park, California, USA (37.12°N, 122.33°W). We used an intramuscular injection of Telazol (Tiletamine hydrochloride and Zolazepam hydrochloride, Fort Dodge Animal Health, Fort Dodge, IA, USA) at an estimated doze of 1 mg kg<sup>-1</sup> to chemically immobilize seals for the attachment of instruments. To investigate the at-sea diet, we attached depth- and acceleration-triggered video cameras (Little Leonardo Co., Tokyo, Japan; Naito et al., 2017) on the head or jaw of post-breeding female northern elephant seals, in February of each study year (Table 1, 2). The head- and jaw-mounted video instruments were designed to record surge acceleration signals at 32 Hz, depth at 0.2 Hz, and video at 30 fps, except two jaw-mounted video cameras used in 2016, which recorded three axes accelerations at 100 Hz, depth at 1 Hz, and video at 30 fps. To start recording when elephant seals feed, all except two instruments (Table 1) used a three step trigger. First, we implemented a start delay timer, which caused the loggers to start recording videos

at a designated date/time after deployment of the instruments, presumably when seals were well into their foraging migration. Second was a depth trigger; since elephant seals usually feed at 400-800m depths (Naito et al., 2013), we set the trigger depths at either 300m, 400m, or 800m. The instrument would only respond to feeding events on dives below those depths. The third trigger was a feeding signal based on acceleration. The camera would start recording only when the signal from the built-in acceleration sensor suggested signals consistent with foraging (within 10 seconds from receiving the acceleration signal, because of the inherent delay in starting video records). Feeding-attempts were identified as head or jaw-motions with a surge acceleration amplitude threshold of 0.3 g, based on Naito et al. (2013, 2017). In 2013, the first study year, we set video cameras to record for a duration of 9.7 and 4.7 min. for the depth triggers of 300 m and 800 m, respectively (Table 1). In 2015 and 2017, we set the camera to record for a duration of 1 min. and 4 min., for the depth triggers of 400 m and 800 m, respectively (Table 1), as we found that these recording durations would allow us to record video clips of feeding attempts more frequently, based on the analysis of depth and jaw motion events obtained prior to this study (Naito et al., 2013). After recording video for a predetermined duration, the video cameras stopped recording until the initiation of the next dive, the next depth threshold and feeding signal. This allowed us to obtain relatively short video clips for each dive across many dives. To examine the efficiency of this three step trigger, we also deployed two jaw video cameras that recorded continuously after an initial delay, with no depth or acceleration triggers (Table 1, 2). The visibility range of the video is about 60 cm from the mouth of a seal (Naito et al., 2017).

The at-sea locations of seals were recorded using satellite transmitters (Wildlife Computers, Redmond, WA, USA, or Sea Mammal Research Unit, St. Andrews, Scotland: Robinson et al., 2010). We recorded depth profiles using jaw-mounted accelerometers (KamiKami Logger, Little Leonardo Co., Tokyo, Japan: Naito et al., 2013). Water temperature was obtained every 4 seconds from time-depth recorders (MK9, Wildlife Computers, Redmond, WA, USA: Robinson et al., 2012) that were affixed to the back of the seals. All instruments were wrapped in rubber splicing tape and attached to high-tension mesh with cable ties. These packages were adhered to the seals' pelage using Loctite Quickset<sup>TM</sup> epoxy (Henkel Corp., Düsseldorf, Germany).

## **Data analysis**

All video data were analyzed using QuickTime Player (Ver.10.2: Apple Inc., CA, USA) to visually determine prey capture events. Prey items that were consumed by the seals were classified as fish, cephalopods or unidentified prey items. We identified prey items to the lowest possible taxonomic level based on available literature and expert opinion.

Satellite data were processed using the R package crawl (Johnson et al., 2008) to eliminate erroneous location estimates and interpolate between locations (see also Robinson et al., 2010; Robinson et al., 2012). To test possible regional differences in the proportion of prey types (fish versus squid), we categorized daily locations where seals captured prey into three regional categories (California Current, Gulf of Alaska, Eastern North Pacific offshore), based on boundaries of Large Marine Ecosystems of the Worlds (<a href="http://lme.edc.uri.edu/index.php/digital-data">http://lme.edc.uri.edu/index.php/digital-data</a>) (Supplemental figure 1). To examine possible differences in prey types between day and night, we assigned prey capture events to day or night, based on solar zenith angle calculated from the time and location, by using R package GeoLight (Lisovski and Hahn, 2013). Regional and day-night differences in the proportion of prey types (fish versus squid) were examined with G tests by pooling the data from all seals, by using R package DescTools (Signorell, 2019). Depth and water temperature data, where seals fed on prey, were extracted using IGOR Pro software (Ver. 6.22J; WaveMetrics, Inc., Lake Oswego, OR, USA). Water temperature data were unavailable for three of the 15 seals studied, due to temperature sensor malfunction of time-depth recorders. We tested the effect of prey types (fish versus squid) on depth and water temperature where seals fed on prey, with linear mixed effect models (LMM) with seal ID included as a random effect, by using R packages lme4 (Bates et al., 2018) and multcomp (Hothorn et al., 2008).

## **RESULTS**

#### Video observations

Seals in our study (n = 15) ranged widely over the eastern North Pacific (Fig. 1). From these seals, a total of 48.2 h of video records from 1467 dives were obtained from head-mounted (n = 12) and jaw-mounted cameras (n = 3) (Table 1). A total number of 697 prey items were observed (Supplemental movies 1, 2), across a total of 77 days

during the foraging trips of the 15 seals. Out of all prey captured, 543 (78%) were identified as fish, whereas 49 (7%) were identified as cephalopods. The remaining 105 items (15%) could not be identified. Unidentified prey items (referred to as unknown prey) likely included both fish and cephalopods.

Fish dominated the diet in most seals, across all sampling years and trigger depths (Fig. 2). The proportion of fish in the diet was >50% for all seals that had video records that included more than 19 prey items (n = 12). Fish and cephalopod captures were recorded on 72 and 24 days, respectively, of the 77 total days of foraging for which we had video records. Fish dominated the prey items both during the day and night (92.4 % and 90.6 % of 380 and 212 identified items, respectively), with no significant differences in prey type composition between day and night (G-test, G=0.57, df = 1, p = 0.45).

## Consistency of diet across regions, depths, and water temperature

Prey items were recorded in videos captured at a wide-range of locations across the eastern North Pacific (Fig. 1). Because the start of video recordings was delayed up to 36 days and triggered by depth (Table 1), video clips encompassed the portion of the post-breeding migration ranging from  $122^{\circ}E$  to  $150^{\circ}E$  in longitude and from  $36^{\circ}N$  to  $56^{\circ}N$  in latitude (Fig. 1). Both prey types were recorded across coastal and pelagic regions of eastern North Pacific. Cephalopods occurred more frequently in Eastern North Pacific offshore (12.7 % of 245 prey items of which prey type was identified) than in California Current and Gulf of Alaska (4.3 % and 6.0 % of 164 and 183 prey items) (G-test, G=10.909, df = 2, p < 0.01). Fish were recorded across all longitudinal and latitudinal ranges. Cephalopods were not seen in the records from the seals that moved north of  $50^{\circ}N$ , along the west coast of Canada (Fig. 1).

Prey items were recorded across a range of depths between 238 m and 1,167 m (Fig. 3). Prey captures were recorded most frequently at 400-500 m depths (33% of all captures recorded) because 6 out of 15 videos were set to start recording when the seal reached 400 m. Fish dominated the diet across depth ranges deeper than 400 m (Fig. 3). Within the depth range of 238-400 m, a total of 17 prey items were recorded, 53% of which were cephalopods. Depths where seals captured prey items did not differ

significantly between fish and squid prey (LMM, effect of prey type on depth; estimate =  $-11.62\pm11.64$  (s.e.), z = -0.99, p = 0.54).

Seals captured prey across temperatures that varied between  $3.2^{\circ}$ C and  $7.35^{\circ}$ C, although prey captures occurred more frequently between  $3.5\text{-}4.0^{\circ}$ C (Fig. 4), possibly reflecting the spatial distribution of video recordings (Fig. 1). Fish dominated the diet across all water temperature ranges (Fig. 4). Water temperatures where seals captured cephalopod prey were slightly ( $0.26^{\circ}$ C) higher than that for fish prey (LMM, effect of prey type on water temperature; estimate =  $0.26\pm0.06$  (s.e.), z = 4.26, p < 0.01).

## Prey identification and prey behavior

In most cases, it was not possible to identify the species or genus of all prey items. However, three fish (Fig. 5) and six squids (Fig. 6) were identified to the species or genus level, based on the shape and appearance of the overall body, fins, or tentacles.

Behavior of fish and squids, just before being captured, were highly variable among capture events. For example, some fish were motionless and appeared to be not responding to seals' approaches (clip 1-3 of Supplementary movie 1), but other fish swam vigorously to escape from the seal (clip 4). Some squid were motionless (clip 1 of Supplementary movie 2), but others showed escape movements (clip 2 and 3), occasionally emitting bioluminescence, to which the seal appeared to respond (clip 3).

## **DISCUSSION**

## Diet of northern elephant seals

Our results indicate that fish largely dominate the diet of post-breeding female northern elephant seals (Fig. 2). Our data offer a contrasting view to previous studies that, based on stomach contents, reported a higher occurrence of cephalopods than fish (cephalopods and teleosts found in 112 and 75 out of 193 stomach contents, respectively; Antonelis et al., 1987, 1994). Stomach content analysis has an inherent bias toward hard-parts like squid beaks that tend to be retained in the stomach, which likely explains the discrepancy (Harvey and Antonelis, 1994). More recently, Goetsch et al. (2018), based on quantitative fatty acid signature analysis, reported that fish and squid comprised 63.7% and 36.3% of population level diet of female northern elephant

seals, respectively, and that energy-rich mesopelagic fishes are important in the diet. This finding is consistent with our results.

Mesopelagic fish, such as myctophids, would be preferable prey for northern elephant seals, as they are energy-rich and highly abundant in the mesopelagic depth zone (200-1000 m) (Brodeur and Yamamura, 2005; Catul et al., 2011). New estimates suggest that the biomass of mesopelagic fish is one order of magnitude higher than previously thought, based on acoustic observations (Irigoien et al., 2014). High mesopelagic fish biomass (consisting mainly of myctophid fishes) was also reported using ship-based net sampling in the Transition Zone of the eastern North Pacific (Saijo et al., 2017), the primary foraging area of female northern elephant seals (Robinson et al., 2012). Female northern elephant seals generally show a large number of feeding-associated jaw motion events (around 2000 times per day) (Naito et al., 2013), which agrees with the hypothesized abundance of myctophids. Myctophidae fish often form dense aggregations (Catul et al., 2011), which would make them easier to detect by predators, and generally have high energy density (7-13 kJ g<sup>-1</sup> wet weight; Lea et al., 2002; Sinclair et al., 2015; Goetsch et al., 2018), and therefore should be preferentially targeted by northern elephant seals.

Our results confirm that post-breeding female northern elephant seals feed on fish across their foraging range (Fig.1) and main foraging depths (Fig. 3) in the eastern North Pacific. Slightly higher occurrence of cephalopod in Eastern North Pacific offshore might reflect regional differences in the distribution and abundance of mesopelagic squids (Brodeur and Yamamura, 2005). Although our depth-trigger system biased our records to 400 m and 800 m depths, our previous work using jaw motion event recorders shows these depths to be the primary foraging depths of female northern elephant seals (Naito et al., 2013, 2017; Adachi et al., 2019).

#### **Comparison with other marine mammals**

Previous studies suggested that southern elephant seals, the congeneric species of northern elephant seals in the southern hemisphere, feed on both fish and cephalopods (Daneri et al., 2000; Daneri and Carlini, 2002; Bradshaw et al., 2003; Field et al., 2007; Cherel et al., 2008). Daneri and Carlini (2002) found more cephalopods than fish in stomach content samples (98.1% and 14.8%, respectively) at King George Island, with

myctophids as the most dominant (76.6%) fish group. Similarly, Field et al. (2007) reported a higher occurrence of squid than fish remains in the stomach content samples (100 % and 75.9 %, respectively) at Macquarie Island. However, Cherel et al. (2008) concluded that southern elephant seals in Kerguelen Islands fed mainly on mesopelagic fish, especially myctophids, based on stable isotope analysis of blood samples. Similarly, northern elephant seals had a higher occurrence of cephalopods reported from stomach content analyses (Antonelis et al., 1987, 1994), in contrast to a higher occurrence of fish as determined by quantitative fatty acid (Goetsch et al., 2018) and video analyses (this study). Recently, McGovern et al. (2019) obtained head-mounted video records from five female southern elephant seals at Peninsula Valdes, Argentina, and reported a few fish species (myctophids, smelt, dragonfish), but no squid, in the video records. However, no quantitative descriptions on the composition of fish species were available in McGovern et al. (2019). These results suggest that the diet of northern and southern elephant seals is broadly similar. It appears that there may be some geographical variation in the relative importance of fish versus squid in the diet of southern elephant seals, though methodological differences between studies make quantitative assessments difficult.

In the North Pacific, mesopelagic fish, such as myctophids, have been important in the diet of other marine mammals, such as dolphins and fur seals (Springer et al., 1999; Ohizumi et al., 2003; Brodeur and Yamamura, 2005). However, dolphins and fur seals are considered to feed on myctophids when the fish migrate to the surface during the night (Ohizumi et al., 2003), whereas northern elephant seals are feeding on myctophids much deeper in the water column. The density of myctophids was reported to be higher in deeper depths (250-550 m) than near the surface during the night, which may explain why northern elephant seals feed in deep depths despite increased diving costs (Saijo et al., 2018). Also, the size of individual fish tends to be larger in deeper depths (Frost and McCrone, 1979), which may bring further benefit to deep-diving seals. Mesopelagic fish are also consumed worldwide by beaked whales (~ 30% frequency of occurrence in stomach contents, MacLeod et al., 2003), with foraging presumed to occur at deep depths that have not been quantified. Northern elephant seals appear to occupy a unique trophic niche foraging mainly on mesopelagic fish at deep depths, by having a deep diving ability. Their relatively small body size (compared to beaked

whales that dive to similar depth zones) and relatively low metabolic rates (Maresh et al., 2015) might allow them to fulfill energy requirements by feeding on relatively small fish prey rather than on large squids (Naito et al., 2013). On the other hand, mesopelagic squid constitute an important component of the diet of toothed whales. Squid species including *Berryteuthis sp.*, *Octopoteuthis sp.*, and *Histioteuthis sp.* are reported in the diet of sperm whales (*Physeter macrocephalus*) and short-finned pilot whales (*Globicephala macrorhynchus*) (Kawakami, 1980; Sinclair, 1992), and are also found in the diet of northern elephant seals (Antonelis et al., 1987, 1994; Goetsch et al., 2018; this study). Northern elephant seals might feed opportunistically on mesopelagic squid species, complementing their feeding on mesopelagic fish. Given that substantial interannual variability and decadal-scale trends were reported on the relative abundance of myctophid fish (Springer et al., 1999; Brodeur and Yamamura, 2005), the foraging and reproductive success of northern elephant seals can potentially serve as a sentinel of the mesopelagic fish and squid abundance in the eastern North Pacific (Le Boeuf and Crocker, 2005; Crocker et al., 2006).

### Advantages and limitations of animal-borne video cameras

Animal-borne video cameras can provide insight into what animals feed on in the open ocean. By combining video cameras with satellite tags and time-depth recorders, we were able to identify prey items in the diet of northern elephant seals in relation to foraging location and depth. We have shown that animal-borne video cameras are a useful tool for studying the at-sea diet of marine predators. Video cameras with depth and acceleration triggers are more efficient in recording prey encounters, as those loggers with such triggers recorded 17 items per hour on average while those that only used a delay timer recorded 4.6 prey items per hour, on average (Table 1). Previous studies reported acceleration signals of prey encounters in a range of marine predators (Kokubun et al., 2011, Guinet et al., 2014, Nakamura et al., 2015, Sato et al., 2015, Foo et al., 2016), and these signals could be used to trigger the start of videos for efficient video recordings. There are still some limitations that need to be considered. The duration of video recordings was limited by the battery capacity (4-5 hours per device), and the video resolution and sensitivity made prey identification difficult. Furthermore, our feeding-signal triggers may have biased our sample toward prey found in schools

rather than solitary prey, as our cameras might not be able to record solitary prey items that induced the first feeding signal due to the delay (<10 s) in starting video recording after the trigger was reached. Further developments and adjustments will improve the utility of animal-borne cameras, making them more widely applicable to diet studies of marine predators.

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## **Competing interests**

None declared.

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## **Data availability**

Data associated with this article is available at ADS (Arctic Data archive System) of National Institute of Polar Research (https://ads.nipr.ac.jp/dataset/A20191218-001)

#### Reference

- Adachi, T., Hückstädt, L. A., Tift, M. S., Costa, D. P., Naito, Y. and Takahashi, A. (2019). Inferring prey size variation from mandible acceleration in northern elephant seals. *Mar. Mamm. Sci.* **35**, 893-908.
- Antonelis, G. A., Lowry, M. S., DeMaster, D. P. and Fiscus, C. H. (1987). Assessing northern elephant seal feeding habits by stomach lavage. *Mar. Mamm. Sci.* 3, 308-322.
- Antonelis, G. A., Lowry, M. S., Fiscus, C. H., Stewart, B. S. and DeLong, R. L. (1994). Diet of the northern elephant seal. In *Elephant seals: population ecology, behavior, and physiology* (ed. B. J. Le Boeuf and R. M. Laws), pp. 211-223. Berkley: University of California Press.
- **Bates, D., Maechler, M., Bolker, B. and Walker, S.** (2018). lme4: linear mixed-effects models using Eigen and S4. *R package version 1.1-18*. http://cran.r-project.org/web/packages/lme4
- Bax, N. J., Miloslavich P., Muller-Karger, F. E., Allain, V., Appeltans, W., Batten, S. D., Benedetti-Cecchi, L., Buttigieg, P. L., Chiba, S., Costa, D. P. et al. (2019). A response to scientific and societal needs for marine biological observations. *Front. Mar. Sci.* 6, 395.
- Block, B. A., Jonsen, I. D., Jorgensen, S. J., Winship, A. J., Shaffer, S. A., Bograd, S. J., Hazen, E. H., Foley, D. G., Breed, G. A., Harrison, A. -L. et al. (2011). Tracking apex marine predator movements in a dynamic ocean. *Nature* 475, 86-90.
- Bowen, W. D., Tully, D., Boness, D. J., Bulheier, B. M. and Marshall, G. J. (2002). Prey-dependent foraging tactics and prey profitability in a marine mammal. *Mar. Ecol. Prog. Ser.* **244**, 235–245.
- Bradshaw, C. J. A., Hindell, M. A., Best, N. J., Phillips, K. L., Wilson, G. and Nichols, P. D. (2003). You are what you eat: describing the foraging ecology of southern elephant seals (*Mirounga leonina*) using blubber fatty acids. *Proc. R. Soc. B Biol. Sci.* 270, 1283–1292.
- **Brodeur, R. and Yamamura, O.** (2005). Micronekton of the North Pacific. *PICES Sci. Rep.* **30**, 1-115.
- Catul, V., Gauns, M. and Karuppasamy, P. K. (2011). A review on mesopelagic fishes belonging to family Myctophidae. *Rev. Fish Biol. Fisheries* **21**, 339-354.
- Cherel, Y., Ducatez, S., Fontaine, C., Richard, P. and Guinet, C. (2008). Stable isotopes reveal the trophic position and meso-pelagic fish diet of female southern elephant seals breeding on the Kerguelen Islands. *Mar. Ecol. Prog. Ser.* **370**, 239–247.
- Costa, D. P., Breed, G. A. and Robinson, P. W. (2012). New insights into pelagic migrations: implications for ecology and conservation. *Annu. Rev. Ecol. Evol. Syst.* 43, 73-96.
- Crocker, D. E., Costa, D. P., Le Boeuf, B. J., Webb, P. M. and Houser, D. S. (2006). Impact of El Niño on the foraging behavior of female northern elephant seals. *Mar. Ecol. Prog. Ser.* **309**, 1-10.
- **Daneri, G. A., Carlini, A. R. and Rodhouse, P. G. K.** (2000). Cephalopod diet of the southern elephant seal, Mirounga leonina, at King George Island, South Shetland Islands. *Antarct. Sci.* **12**, 16-19.
- **Daneri, G. A. and Carlini, A. R.** (2002). Fish prey of southern elephant seals, Mirounga leonina, at King George Island. *Polar Biol.* **25**, 739-743.

- Davis, R. W., Fuiman, L. A., Williams, T. M., Collier, S. O., Hagey, W. P., Kanatous, S. B., Kohin, S. and Horning, M. (1999). Hunting behavior of a marine mammal beneath the Antarctic fast ice. *Science* **283**, 993–996.
- Estes, J. A., Heithaus, M., McCauley, D. J., Rasher, D. B. and Worm., B. (2016). Megafaunal impacts on structure and function of ocean ecosystems. *Annu. Rev. Environ. Resour.* 41, 83-116.
- Field, I. C., Bradshaw, C. J. A., van den Hoff, J., Burton, H. R. and Hindell, M. A. (2007). Age-related shifts in the diet composition of southern elephant seals expand overall foraging niche. *Mar. Biol.* **150**, 1441-1452.
- Foo, D., Semmens, J. M., Arnould, J. P., Dorville, N., Hoskins, A. J., Abernathy, K., Marshall, G. J. and Hindell, M. A. (2016). Testing optimal foraging theory models on benthic divers. *Anim. Behav.* 112, 127-138.
- **Frost, B.W. and McCrone, L.E.** (1979). Vertical distribution, diel vertical migration, and abundance of some mesopelagic fishes in the eastern Sub-arctic Pacific Ocean in summer. *Fish. Bull.* **76**, 751-770.
- **Gales, N. J. and Cheal., A.J.** (1992). Estimating diet composition of the Australian sea-lion (*Neophoca cinerea*) from scat analysis an unreliable technique. *Wildl. Res.* **19**, 447-456.
- Goetsch, C., Conners, M. G., Budge, S. M., Mitani, Y., Walker, W. A., Bromaghin, J. F., Simmons, S.E., Reichmuth, C. and Costa, D. P. (2018). Energy-rich mesopelagic fishes revealed as a critical prey resource for a deep-diving predator using quantitative fatty acid signature analysis. *Front. Mar. Sci.* 5, 430.
- Guinet, C., Vacquié-Garcia, J., Picard, B., Bessigneul, G., Lebras, Y., Dragon, A. C., Viviant, M., Arnould, J. P. Y. and Bailleul, F. (2014). Southern elephant seal foraging success in relation to temperature and light conditions: insight into prey distribution. *Mar. Ecol. Prog. Ser.* 499, 285-301.
- **Harvey, J. T. and Antonelis, G. A.** (1994). Biases associated with non-lethal methods of determining the diet of northern elephant seals. *Mar. Mamm. Sci.* **10**, 178-187.
- Hooker, S. K., Boyd, I. L., Jessopp, M., Cox, O., Blackwell, J., Boveng, P. L. and Bengtson J. L. (2002). Monitoring the prey-field of marine predators: Combining digital imaging with datalogging tags. *Mar. Mamm. Sci.* 18, 680-697.
- **Hothorn, T., Bretz, F. and Westfall, P.** (2008). Simultaneous Inference in General Parametric Models. *Biometrical J.* **50**, 346-363.
- Hückstädt, L. A., Koch, P. L., McDonald, B. I., Goebel, M. E., Crocker, D. E. and Costa, D. P. (2012). Stable isotope analyses reveal individual variability in the trophic ecology of a top marine predator, the southern elephant seal. *Oecologia*, 169, 395-406.
- Hussey, N. E., Kessel, S. T., Aarestrup, K., Cooke, S. J., Cowley, P. D, Fisk, A. T., Harcourt, R. G., Holland, K. N., Iverson, S. J., Kocik, J. F. et al. (2015).
  Aquatic animal telemetry: A panoramic window into the underwater world. *Science*, 348, 1255642.
- Irigoien, X., Klevjer, T. A., Røstad, A., Martinez, U., Boyra, G., Acuña, J. L., Bode, A., Echevarria, F., Gonzalez-Gordillo, J. I., Hernandez-Leon, S. et al. (2014). Large mesopelagic fishes biomass and trophic efficiency in the open ocean. *Nat. Commun.* 5, 3271.
- Johnson, D. S., London, J. M., Lea, M. A. and Durban, J. W. (2008). Continuous-time correlated random walk model for animal telemetry data. *Ecology* 89, 1208-1215.

- Kawakami, T. (1980). A review of sperm whale food. Sci. Rep. Whales Res. Inst. 32, 199-218.
- Kernaléguen, L., Dorville, N., Ierodiaconou, D., Hoskins, A. J., Baylis, A. M., Hindell, M. A., Semmens, J., Abernathy, K., Marshall, G. J., Cherel, Y. et al. (2016). From video recordings to whisker stable isotopes: a critical evaluation of timescale in assessing individual foraging specialisation in Australian fur seals. *Oecologia* **180**, 657-670.
- **Kokubun, N., Kim, J. H., Shin, H. C., Naito, Y. and Takahashi, A.** (2011). Penguin head movement detected using small accelerometers: a proxy of prey encounter rate. *J. Exp. Biol.* **214**, 3760-3767.
- Kuhn, C. E., Crocker, D. E., Tremblay, Y. and Costa, D. P. (2009). Time to eat: measurements of feeding behaviour in a large marine predator, the northern elephant seal Mirounga angustirostris. *J. Anim. Ecol.* **78**, 513-523.
- **Lea, M. A., Nichols, P. D. and Wilson, G.** (2002). Fatty acid composition of lipid-rich myctophids and mackerel icefish (Champsocephalus gunnari)-Southern Ocean food-web implications. *Polar Biol.* **25**, 843-854.
- **Le Boeuf, B. J. and Crocker, D. E.** (2005). Ocean climate and seal condition. *BMC Biol.* **3.** 9.
- Le Boeuf, B. J., Crocker, D. E., Costa, D. P., Blackwell, S. B., Webb, P. M. and Houser, D. S. (2000). Foraging ecology of northern elephant seals. *Ecol. Monogr.* 70, 353-382.
- **Lisovski, S. and Hahn, S.** (2013) GeoLight processing and analysing light-based geolocation in R. *Methods Ecol. Evol.* **3**, 1055-1059.
- **MacLeod, C. D., Santos, M. B. and Pierce, G. J.** (2003). Review of data on diets of beaked whales: evidence of niche separation and geographic segregation. *J. Mar. Biol. Assoc. UK* **83**, 651-665.
- Maresh, J. L., Adachi, T., Takahashi, A., Naito, Y., Crocker, D. E., Horning, M., Williams T. M. and Costa, D. P. (2015). Summing the strokes: energy economy in northern elephant seals during large-scale foraging migrations. *Mov. Ecol.* 3, 22.
- McGovern, K. A., Rodríguez, D. H., Lewis, M. N. and Davis, R. W. (2019). Diving classification and behavior of free-ranging female southern elephant seals based on three-dimensional movements and video-recorded observations. *Mar. Ecol. Prog. Ser.* **620**, 215-232.
- Miloslavich, P., Bax, N. J., Simmons, S. E., Klein, E., Appeltans, W., Aburto-Oropeza, O., Garcia, M.A., Batten, S.D., Benedetti-Cecchi, L., Checkley, D.M. et al. (2018). Essential ocean variables for global sustained observations of biodiversity and ecosystem changes. *Glob. Change Biol.* 24, 2416-2433.
- Naito, Y., Costa, D. P., Adachi, T., Robinson, P. W., Fowler, M. and Takahashi, A. (2013). Unravelling the mysteries of a mesopelagic diet: a large apex predator specializes on small prey. *Funct. Ecol.* 27, 710-717.
- Naito, Y., Costa, D. P., Adachi, T., Robinson, P. W., Peterson, S. H., Mitani, Y. and Takahashi, A. (2017). Oxygen minimum zone: An important oceanographic habitat for deep-diving northern elephant seals, Mirounga angustirostris. *Ecol. Evol.* 7, 6259-6270.
- Nakamura, I., Goto, Y. and Sato, K. (2015). Ocean sunfish rewarm at the surface after deep excursions to forage for siphonophores. *J. Anim. Ecol.* **84**, 590-603.

- **Ohizumi, H., Kuramochi, T., Kubodera, T., Yoshioka, M. and Miyazaki, N.** (2003). Feeding habits of Dall's porpoises (*Phocoenoides dalli*) in the subarctic North Pacific and the Bering Sea basin and the impact of predation on mesopelagic micronekton. *Deep Sea Res. I: Oceanogr. Res. Pap.* **50**, 593-610.
- Robinson, P. W., Simmons, S. E., Crocker, D. E. and Costa, D. P. (2010). Measurements of foraging success in a highly pelagic marine predator, the northern elephant seal. *J. Anim. Ecol.* **79**, 1146-1156.
- Robinson, P. W., Costa, D. P., Crocker, D. E., Gallo-Reynoso, J. P., Champagne, C. D., Fowler, M. A., Goetsch, C., Goetz, K. T., Hassrick, J. L., Hückstädt, L. A. et al. (2012). Foraging behavior and success of a mesopelagic predator in the northeast Pacific Ocean: insights from a data-rich species, the northern elephant seal. *PLoS ONE* 7, e36728.
- Saijo, D., Mitani, Y., Abe, T., Sasaki, H., Goetsch, C., Costa, D. P. and Miyashita, K. (2017). Linking mesopelagic prey abundance and distribution to the foraging behavior of a deep-diving predator, the northern elephant seal. *Deep Sea Res. II Top. Stud. Oceanogr.* **140**, 163-170.
- Sato, N. N., Kokubun, N., Yamamoto, T., Watanuki, Y., Kitaysky, A. S. and Takahashi, A. (2015). The jellyfish buffet: jellyfish enhance seabird foraging opportunities by concentrating prey. *Biol. Lett.* 11, 20150358.
- **Signorell, A.** (2019). DescTools: Tools for descriptive statistics. *R package version* 0.99, 31. <a href="https://cran.r-project.org/package=DescTools">https://cran.r-project.org/package=DescTools</a>.
- Simmons, S. E., Crocker, D. E., Hassrick, J. L., Kuhn, C. E., Tremblay, Y., Costa, D. P. and Robinson, P. W. (2010). Climate-scale hydrographic features related to foraging success in a capital breeder, the northern elephant seal Mirounga angustirostris. *Endang. Species Res.* **10**, 233-243.
- **Sinclair, E. H.** (1992). Stomach contents of four short-finned pilot whales (Globicephala macrorhynchus) from the southern California Bight. *Mar. Mamm. Sci.* **8**, 76-81.
- **Sinclair, E. H., Walker, W. A. and Thomason, J. R.** (2015). Body size regression formulae, proximate composition and energy density of eastern Bering Sea mesopelagic fish and squid. *PloS ONE* **10**, e0132289.
- Springer, A. M., Piatt, J. F., Shuntov, V. P., Van Vliet, G. B., Vladimirov, V. L., Kuzin, A. E. and Perlov, A. S. (1999). Marine birds and mammals of the Pacific Subarctic Gyres. *Progr. Oceanogr.* **43**, 443-487.
- **Tollit, D. J., Pierce, G. J., Hobson, K. A., Bowen, W. D. and Iverson, S. J.** (2010). Diet. In *Marine mammal ecology and conservation: a handbook of techniques* (ed. I.L. Boyd, W. D. Bowen and S. J. Iverson), pp. 191-221. Oxford: Oxford University Press.

## **Figures**

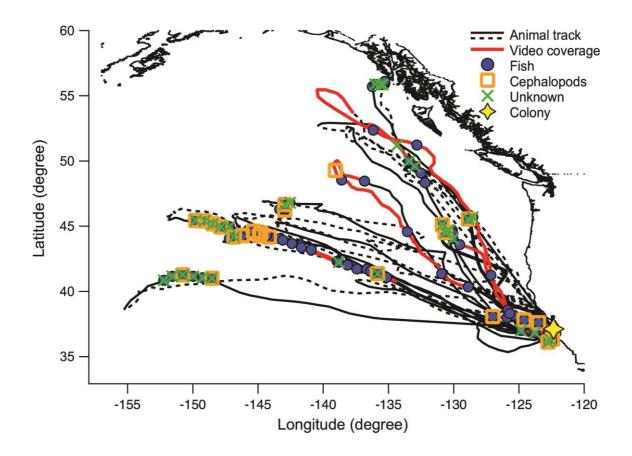


Fig. 1. At-sea movements of 15 female northern elephant seals from Año Nuevo, California, during 2013-2017. Black line indicated animal tracks (solid and dashed line: outbound and inbound part of each track). Red line sections indicated where video data were recorded. Blue circles indicate daily locations where video cameras recorded the seals catching fish. Orange squares indicate daily locations where cephalopods were caught, and green crosses indicate unknown prey items.

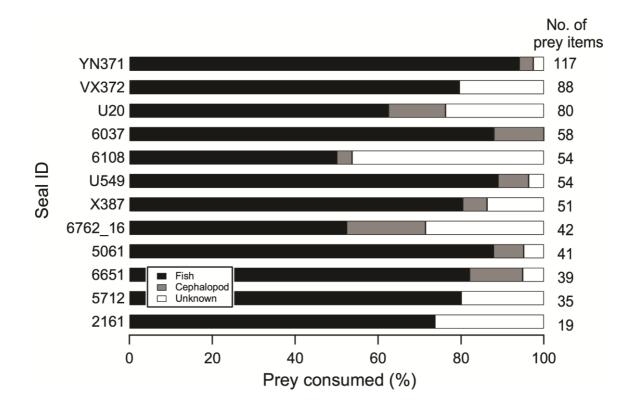


Fig. 2. Proportion of prey types consumed by adult female northern elephant seals from Año Nuevo, California, determined from animal-borne video records. Data from 12 seals with >19 observed prey capture events are shown. The shade of the bar indicates the percent of fish (black), cephalopod (grey) or unknown prey items (white).

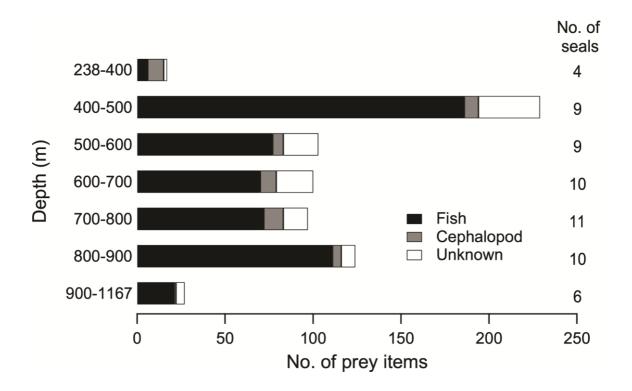


Fig. 3. Composition of prey types of adult female northern elephant seals from Año Nuevo, California, in relation to feeding depth, observed using head-mounted video cameras. Color codes indicate fish (black), cephalopod (grey) and unknown prey items (white).

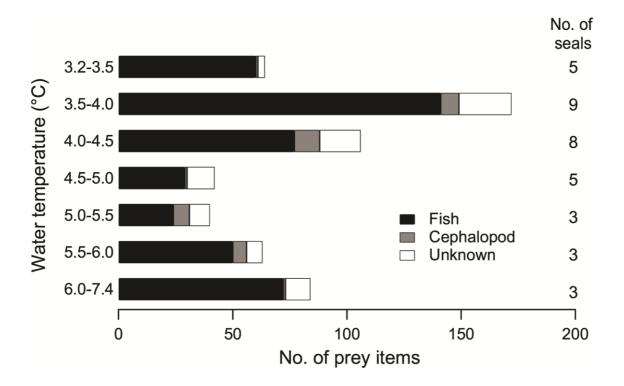


Fig. 4. Composition of prey types of adult female northern elephant seals from Año Nuevo, California, in relation to *in-situ* water temperature, observed using head-mounted video cameras. Color codes indicate fish (black), cephalopod (grey) and unknown prey items (white).

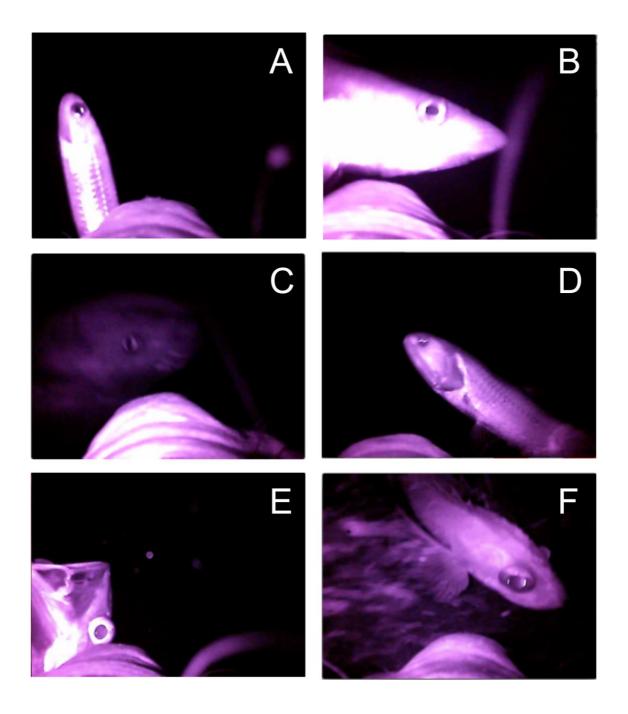


Fig. 5. Fish prey images from head-mounted video cameras on adult female northern elephant seals. A. *Myctophidae*, B. *Merluccius sp.*, C. *Icosteus aenigmaticus*, D-F. unidentified fishes. The seal's snout is visible at the bottom of the images. The seal's supraorbital vibrissae are also visible in B and E.

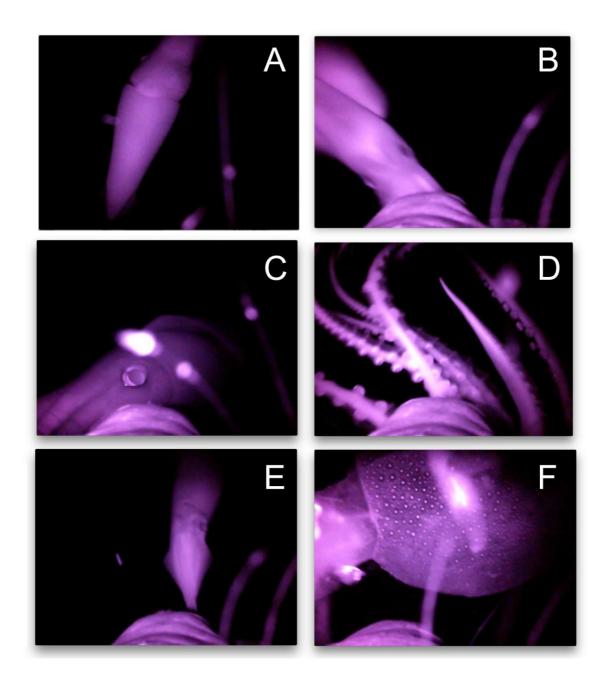


Fig. 6. Squid prey images from head-mounted video cameras on adult female northern elephant seals. A. Histioteuthis sp., B. Octopoteuthis sp., C. Taningia danae, D. Belonella sp., E. Berryteuthis sp., F. Japetella sp.. The seal's snout is visible at the bottom of the images. The seal's supraorbital vibrissae are also visible in A-C and E-F.

Table 1. Summary of video recordings from adult female northern elephant seals from Año Nuevo, California, USA.

Seal ID	Year	Position of video	Flash type	Delay timer duration	Trigger depth (m)	Scheduled duration of one	Total video duration	Total no. of prey items	No. of prey items / h
				(days)		video		recorded	
X387	2013	Head	Infrared	18	300	0:10:00	3:41:17	51	13.8
2161	2013	Head	Infrared	15	800	0:04:00	1:05:23	19	17.4
5061	2015	Head	Infrared	36	400	0:01:00	3:35:00	41	11.4
U549	2015	Head	Infrared	36	400	0:01:00	3:35:25	54	15
4176	2015	Head	Infrared	0	800	0:04:00	2:42:16	8	3
6037	2015	Head	Infrared	21	800	0:04:00	3:31:04	58	16.5
6651	2015	Head	Infrared	21	800	0:04:00	3:25:28	39	11.4
6762*	2015	Head	Infrared	21	800	0:04:00	0:51:46	6	7
YN371	2015	Head	Infrared	36	400	0:01:00	3:17:13	117	35.6
VX372	2015	Jaw	Near-red	21	800	0:04:00	2:31:31	88	34.8
6762*	2016	Jaw	Near-red	21	0	continuous**	5:05:01	42	8.3
T28	2016	Jaw	Near-red	21	0	continuous**	5:40:28	5	0.9
5712	2017	Head	Infrared	36	400	0:01:00	2:09:47	35	16.2
6108	2017	Head	Infrared	36	400	0:01:00	3:29:10	54	15.5
U20	2017	Head	Infrared	36	400	0:01:00	3:29:47	80	22.9

<sup>\*</sup>Video attached on the same seal

**TABLES** 

<sup>\*\*</sup> Continuous recording once video recording started

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Table 2. Summary of video instruments deployed on female elephant seals from Año Nuevo, California, USA.

Туре	Size and weight	Resolution	Start tiggers	
Head-mounted videos	30mm (diameter) x 151mm (length), 158g	640 x 480 pixels	Delay timer, depth, acceleration	
Infrared flash unit for head-mount video	30mm (diameter) x 130mm (length), 187g			
Jaw-mounted video (2015)	28mm (diameter) x 106mm (length), 118g	640 x 480 pixels	Delay timer, depth, acceleration	
Near-red flash unit for jaw-mount video (2015)	28mm (diameter) x 93mm (length), 117g			
Jaw-mounted video (2016)*	23 x 21x 72mm, 53.2 g	1280 x 960 pixels	Delay timer only	

<sup>\*</sup>Near-red flash was built in with video

## **Supplementary Information**

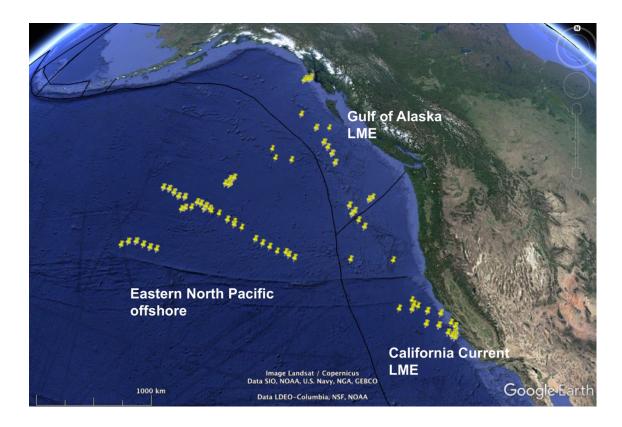


Fig. S1. Assignment of prey capture locations into three regions, 'California Current', Gulf of Alaska', and 'Eastern North Pacific offshore'. The regional boundaries (black lines) were derived from 'Large Marine Ecosystems of the World' (<a href="http://lme.edc.uri.edu/index.php/digital-data">http://lme.edc.uri.edu/index.php/digital-data</a>).



Movie 1 (Fish capture.mp4): Video clips upon fish captures by adult female northern elephant seals. Clip 1: The seal fed on an unidentified fish (depth: 475 m). Clip 2: The seal fed on unidentified schooling fish (depth: 455 m). Clip 3: The seal encountered and fed on the fish *Merluccius* sp. (depth: 515 m). Clip 4: The seal fed on an unidentified fish (depth: 910 m). In Clip 1-3, supraorbital vibrissae of the equipped seal are visible at the right-hand side of the movies.



Movie 2 (Squid capture.mp4): Video clips upon squid captures by female northern elephant seals. Clip 1: The seal fed on the squid *Histioteuthis* sp. (depth: 656 m). Clip 2: The seal fed on the squid *Octopoteuthis* sp. (depth: 415 m). Clip 3: The seal encountered the squid *Taningia danae* (at time 00:18; depth 662 m), but the squid escaped (00:21). After following the lights from the tentacles of the squid (00:26-00:34), and encountering with squid ink (00:39-00:49), the seal captured the squid (00:51), as indicated by the quick movements of the snout. In all video clips, supraorbital vibrissae of the equipped seal are visible at the right-hand side of the movies.