

1 **Interplay between group size, huddling behavior and basal metabolism: an**  
2 **experimental approach in the social degus**

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16 **plasticity**

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23 **Abstract**

24 Mammals exposed to low temperatures increase their metabolic rate to maintain constant  
25 body temperature and thus compensate heat loss. This high and costly energetic demand  
26 can be mitigated through thermoregulatory behavior such as social grouping or huddling,  
27 which helps to decrease metabolic rate as function of the numbers of individuals grouped.  
28 Sustained low temperatures in endothermic animals produce changes over time in rates of  
29 energy expenditure, by means of phenotypic plasticity. However, the putative modulating  
30 effect that huddling exerts on the flexibility of the basal metabolic rate (BMR) due to  
31 thermal acclimation remains unknown. We determined BMR values in *Octodon degus*,  
32 an endemic Chilean rodent, after being acclimated either to 15 °C or 30 °C during 60  
33 days, both alone and in groups of 3 and 5 individuals. At 15 °C, BMR of huddling  
34 individuals was 40 % lower than that of animals housed alone. Moreover, infrared  
35 thermography revealed a significant increase in local surface temperatures in huddled  
36 animals. Furthermore, individual thermal conductance was lower in individuals  
37 acclimated to 15 °C than at 30 °C, but no differences were observed between single and  
38 grouped animals. Our results indicate that huddling prevent an increase in BMR when  
39 animals are acclimated to cold conditions and that this effect is proportional to the number  
40 of animals grouped.

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50 **Introduction**

51 Endothermy is defined as the ability to produce endogenous heat, allowing individuals to  
52 maintain a positive temperature differential with the environment and remain in  
53 homeothermic condition (Hill et al., 2004). Heat production may be modulated through  
54 behavioral and physiological changes at different scales and across a wide range of  
55 ambient temperatures (Gilbert et al., 2010). For example, animals exposed to temperatures  
56 below the thermoneutral zone (TNZ) must compensate heat losses by increasing their  
57 metabolic rate in order to remain homeothermic (Canals, 1998b). Thus survival of small  
58 mammals at low temperatures may depend on their ability to reduce heat loss and/or to  
59 increase metabolic rate, which in many cases involves a large energy cost (Kauffman et  
60 al., 2003).

61 To compensate for the increased energy expenditure caused by exposure to low  
62 temperatures individuals may exhibit behavioral responses such as social grouping or  
63 huddling (Canals, 1998b; Gilbert et al., 2010). Recently Gilbert et al. (2012) documented  
64 that local heating is crucial in reducing the extent of the cold challenge in huddling rabbit  
65 pups. Through thermal images they demonstrated that at 14 °C, the mean surface  
66 temperature of the huddle was higher than the mean temperature of isolated pups. This  
67 study demonstrated that local heating when huddle provided each pup with an ambient  
68 “public warmth” in the cold. Thus, huddling behavior reduces energy costs by reducing  
69 the metabolic rate and average thermal conductance of each individual in the group, due  
70 mainly to the reduction of surface area and altering the thermal environment experienced  
71 by animals exposed to the cold. In this sense, however, whilst in some species the benefit  
72 is shared in an equitable manner, in others it has been reported that some individuals may  
73 benefit more than others when huddling (Bustamante et. al., 2002). Apparently, this  
74 asymmetry would be the result that some animals would preferably occupy the best  
75 location in the group (i.e., the center), while others would be relegated to occupy a larger  
76 proportion of time to the periphery (Shank and Alberts, 1997). Moreover, this reduction is  
77 proportional to the number of individuals in the group to the power of -0.33 (Gilbert et al.,  
78 2010; Canals and Bozinovic, 2011). For example, for *Octodon degus*, a rodent that dwells  
79 in semi-arid areas of northern and central Chile, the huddling effectiveness (He, the

80 maximum energy saving during huddling, Canals et al. 1997) when these organisms are  
81 exposed to temperatures of 0, 5 and 10 °C reaches 43%, which constitutes a significant  
82 fraction of the body's energy budget (Canals et al., 1998a). Therefore, at temperatures at  
83 least 5 °C below the lower limit of the TNZ the He is constant, a phenomenon that was  
84 also reported for other mammal species. However, with increasing temperature above this  
85 critical value He tends to decrease; therefore huddling effectiveness decreases at high  
86 temperatures (Canals et al., 1997).

87 Thus the capacity of endotherms inhabiting seasonal environments depends on their  
88 ability to develop physiological and behavioral mechanisms allowing physiological  
89 homeostasis (Pigliucci, 2001, Piersma and van Gils 2011). In this vein physiological  
90 flexibility, the ability to change and modify physiological traits in response to  
91 environment cues, is crucial for maintaining homeostasis in changing environments  
92 (Piersma and Drent, 2003; McKechnie et al., 2007) and has been demonstrated to occur in  
93 response to seasonal variations (Bozinovic and Contreras, 1990; Bacigalupe et al., 2004)  
94 and in laboratory thermal acclimation experiments in several mammal species (Nespolo  
95 and Rosenmann, 1997; Nespolo et al., 2002). This physiological flexibility, a particular  
96 case of phenotypic plasticity (see Garland and Adolph, 1991; Piersma and Drent , 2003),  
97 can cause variations in thermal insulation as well as changes in basal (BMR) and  
98 maximum cold- induced metabolic rates ( $M_{sum}$ ). Thus, changes in any of these parameters  
99 may be indicative of changes in energy expenditure rates (Piersma et al., 1996; Nespolo et  
100 al., 1999). In particular, BMR represents the minimum rate of energy necessary to  
101 maintain homeostasis and reflects the cost of maintaining high levels of sustained activity.  
102 This metabolic trait exhibits high flexibility, which has been demonstrated in experiments  
103 in the laboratory by thermal acclimation (Nespolo and Rosenmann, 1997; Speakman,  
104 2000). The reaction norm of BMR (i.e. a function that describes the changes of BMR as a  
105 function of acclimation temperature) exhibits a negative association, that is, endotherms  
106 acclimated to low temperatures commonly exhibited an increase in their BMR. Such  
107 increase in BMR is thought to be related to the increase in maintenance costs of  
108 metabolically active organs when animals are faced with the high energy requirements of  
109 thermoregulation in the cold (see Cruz-Neto et al., 2003; McKechnie et al., 2007).

110 Experimental evidence has shown the existence of i) decreased rates of individual  
111 energy expenditure (e.g. resting metabolic rate) in grouped organisms when exposed to  
112 temperatures lower than the TNZ and ii) a remarkable physiological flexibility of rates  
113 of energy expenditure when acclimated to different temperatures. In the former case  
114 huddling allows energy savings during the grouping behavior, whereas physiological  
115 flexibility modifies the rates of energy expenditure in the medium term (e.g. weeks) in  
116 order to cope with the different thermoregulatory needs. However, to date there is no  
117 evidence that the use of huddling may affect medium- to long-term rates of energy  
118 expenditure in endotherms, i.e., that huddling behavior affects the phenotypic response  
119 of individuals acclimated to different temperatures. The aim of this study was to  
120 estimate the effect of social grouping on flexibility in BMR, TEWL and thermal  
121 conductance in *Octodon degus* or degu, a social rodent that exhibits huddling behavior  
122 both in captivity and in the wild (Ebensperger and Wallen, 2002) and dwells in highly  
123 seasonal environments of central Chile (Di Castri and Hajek, 1976). We test the  
124 hypothesis that the presence of huddling and group size plays a modulating role in the  
125 acclimation capacity of BMR in adults. Specifically, we predict that huddling will  
126 decrease or prevent an increase in BMR when animals are acclimated to cold  
127 conditions and that this effect is proportional to the number of animals grouped.

128

## 129 **Results**

130 After thermal acclimation, we found a significant effect of body mass on total BMR. The  
131 allometric equation relating BMR with body mass (bm) was:  $BMR = 7.9 \cdot bm^{0.53}$  ( $r^2 = 0.21$ ,  
132  $F_{(1, 25)} = 13.88$ ,  $p = 0.009$ ). We also found a significant effect of the acclimation  
133 temperature ( $F_{1, 25} = 6.98$ ,  $P = 0.014$ ), the number of individuals grouped ( $F_{2, 25} = 9.27$ ,  $P$   
134  $= 0.001$ ) and the interaction between these two factors on mass-adjusted BMR ( $F_{2, 25} =$   
135  $6.92$ ,  $P = 0.004$ ). The *post hoc* test revealed that mass-adjusted BMR of rodents  
136 acclimated individually at 15 °C was greater than that observed in rodents acclimated  
137 individually at 30 °C (Fig.1). In groups of three, *O. degus* decreased mass-adjusted BMR  
138 by 15% and 7% when acclimated at 15 °C and 30 °C, respectively (Figure 1).  
139 Additionally, cold-acclimated animals in groups of three exhibited higher mass-adjusted

140 BMRs than those of warm-acclimated animals in groups of three, but this difference  
141 appeared to be smaller than that observed in individual acclimated groups. Compared with  
142 those housed alone, in groups of five the decrease in mass-adjusted BMR was  
143 approximately 40% for degus acclimated to 15 °C. However, there were no significant  
144 energy reduction for degus acclimated to 30 °C. Finally, mass-adjusted BMRs of animals  
145 in groups of five individuals did not present significant differences between warm and  
146 cold-acclimated groups. Furthermore, the mass-adjusted BMR of cold-acclimated animals  
147 in groups of three was similar to that of cold-acclimated animals in groups of five  
148 individuals. The same was true for warm-acclimated animals for these two groups. Also,  
149 we found a significant effect of acclimation temperature on thermal conductance in *O.*  
150 *degus* (ANOVA  $F_{1, 29} = 6.47$ ,  $P = 0.016$ , Table 1). Specifically, we found that the average  
151 individual conductance of animals was greater in rodents acclimated to 30 °C than at 15 °  
152 C (Fig. 2). However, we found no effect of the group size ( $F_{2, 29} = 1.56$ ,  $p = 0.22$ ) or the  
153 interaction between factors ( $F_{2, 29} = 0.90$ ,  $P = 0.41$ ) on thermal conductance.

154 We found significant differences in measured temperatures between treatments  
155 ( $F_{9,71} = 165.9$ ,  $p < 0.0001$ ). The a posteriori analyses showed that maximum surface  
156 temperature and mean contour temperatures were higher in grouped than isolated animals  
157 at 15°C, but not at 30°C. Moreover, at 15°C the temperature of contact was higher than the  
158 contour temperatures of grouped and isolated animals (Figure 2). We also found that the  
159 spatial location of individuals in the group was different between individuals ( $X^2 = 16.94$ ,  
160 d.f. = 4;  $P = 0.02$ ). Some individuals spent more than 5 times in the center of the group  
161 than others. The 95% confidence intervals of the frequencies that each animal spent in the  
162 center of the group were: [0.06-0.24], [0.07-0.26], [0.03-0.17], [0.17-0.40] and [0.26-  
163 0.51]. Finally, we found solitary animals consumed significantly more food than grouped  
164 animals at 15 °C which in turn consumed more food than animals maintained at 30°C  
165 (Kruskall-Wallis ANOVA:  $H_{3,20} = 16.63$ ,  $P = 0.001$ ). In descending order the food intake  
166 per animal were the follows: solitary animals at 15 °C ( $29.98 \pm 4.87$  g/day per animal) >  
167 grouped at 15 °C ( $19.86 \pm 2.35$ ) > grouped at 30 °C ( $10.47 \pm 2.17$ ) = solitary at 30°C  
168 ( $8.02 \pm 1.70$ ).

169

170 **Discussion**

171 Our results confirm the previously demonstrated effect of short-term thermal acclimation  
172 on energy expenditure rates in endotherms. In fact, the difference in BMR between cold-  
173 and warm solitary acclimated animals can reach ca 30%, which is in the range of the  
174 acclimation magnitudes reported for other rodent species in comparable studies (e.g.,  
175 Novoa et al. 2005; Nespolo et al., 2001). However, our results demonstrate for the first  
176 time that huddling behavior exerts a modulatory effect on thermal acclimation of BMR in  
177 endotherms. Increasing the number of individuals to three animals per group and allowing  
178 them to huddle during acclimation led to a reduction in the difference in BMR between  
179 temperatures. When animals were allowed to form groups of five individuals, BMR was  
180 lower than solitary animals when acclimated at 15 °C, suggesting that degus were  
181 acclimated to different effective temperatures. In fact, huddling promotes local heating  
182 and reduces the cold challenge, because single degus experienced colder conditions  
183 compared with degus huddling in a group of five (Fig. 2), a phenomenon that has been  
184 also recently demonstrated in rabbit pups (Gilbert et al. 2012). It is therefore likely that  
185 the individual minimum energy requirement decreases in degus which huddle for long  
186 periods, as occurs with the energy expenditure of this species when grouped (Canals et al.,  
187 1989; Kotze et al., 2008). In this vein, the observed reduction in the acclimation response  
188 of individual BMR in grouped degus (ca 40%) is comparable with the reduction of  
189 metabolic rate by means of huddling of grouped individuals in other mammal species  
190 (range: 11-50%, see Canals et al., 1989; Canals et al., 1998, Gilbert et al., 2010 and  
191 references therein). Moreover, the fact that individuals acclimatized at 30 °C and  
192 maintained in groups of five individuals did not save significant amounts of energy agrees  
193 with the idea proposed by Canals et al. (1998a), which states that the huddling efficiency  
194 decreases at temperatures near the TNZ of the animal, which for *O. degus* corresponds to  
195 the range between 24 °C and 32 °C (Rosenmann, 1977).

196           Thus huddling allows energy savings over at least two time scales. First,  
197 huddling induces metabolic depression of animals while grouped, as has been  
198 demonstrated previously (e.g., Canals et al., 1997) and second, our results strongly  
199 suggest that in degus acclimated to low temperatures and in groups of three or more

200 individuals, individual energy expenditure (BMR) decreases compared to degus subjected  
201 to the same temperatures but acclimated individually. Huddling therefore helps mitigate  
202 the excessive expenditure of energy to maintain homeothermy, buffering the decreasing  
203 ambient temperature by decreasing its thermogenic response. This is also supported by the  
204 observed decrease in food consumption when animals were grouped at low temperatures.  
205 Interestingly, the low food consumption may lead to a decrease of the masses of digestive  
206 organs (not measured), which suggests the possibility that BMR changes reflect changes  
207 in the mass of metabolically active organs. Accordingly, some studies have shown a  
208 significant increase in size of metabolically active organs such as the heart and liver when  
209 exposed to cold (Hammon et al., 2001; Naya et al., 2010). This possible proximal  
210 mechanism, not addressed in this paper, could be considered in future studies.

211 Energy economy may be beneficial for rodents inhabiting arid or semi-arid  
212 environments, where animals are exposed to large temperature variations and  
213 heterogeneous distribution of food (Scantlebury et al., 2006a; Scantlebury et al., 2006b).  
214 The energy savings observed in the laboratory probably can also have consequences in the  
215 field. Thus, decreasing thermogenic response by means of huddling may lead to a  
216 decrease in food needs and thus to allocate time to activities other than foraging.  
217 Alternatively, since energy is limited for animals, the energy saved by huddling may be  
218 allocated to perform other biological functions or activities, while acting as a selective  
219 pressure important for life in groups in rodents (Gilbert et. Al 2012; Ebensperger and  
220 Wallen, 2002; Shradin et al., 2006). For example, in *O. degus* some activities associated  
221 with foraging may be energetically costly, such as digging in dry areas with low humidity  
222 (Ebensperger and Bozinovic, 2000), exploration behavior in open spaces at low  
223 temperatures after foraging and building burrows (Ebensperger, 2000; Quispe et al.,  
224 2009). All these activities may increase the biological fitness of *O. degus*, particularly in  
225 periods of high-energy demand such as winter and during the breeding season.

226 In this way individuals can regulate the levels of energy expenditure to maintain their  
227 functional capabilities in variable thermal environment through plastic and adaptive  
228 changes in metabolic rate, and reassign the energy saved to other activities (Nespolo,  
229 2000). However, Bozinovic et al. (2004) reported that the BMR in *O. degus* in the wild is



230 not altered by seasonality in a Mediterranean environment of central Chile. Indeed, these  
231 authors found that the BMR remains unchanged along the year. Interestingly, the average  
232 BMR values reported for wild degus are very similar to the BMR of cold-acclimated  
233 degus in groups of three and five individuals. As *O. degus* is a social rodent that lives in  
234 groups of four or more individuals sharing the same burrow system and performs  
235 huddling in nature, one might infer that thermoregulatory behavior such as huddling and  
236 its long-term effects could be a compensatory mechanism that helps maintain this rodent  
237 BMR relatively stable over the year.

238 There are several factors that can affect metabolic expenditure of animals performing  
239 huddling. Among the most important are decreasing body area exposed to the  
240 environment (Canals et al., 1998) and the local microclimate (Hayes et al., 1992). In  
241 agreement with that, our results revealed that the surface temperature of the entire group  
242 exposed to 15 °C was higher than the surface temperature of solitary degus and that such  
243 difference is coupled with the increased temperature of the boundary of each animal when  
244 huddle (Fig. 2). An additional hypothesis the so-called socio-physiological effect  
245 (Speakman and Rossi, 1999), states that there would be a decrease in metabolic rate of  
246 animals grouped since they tend to decrease their levels of anxiety when perceiving  
247 conspecifics (Martin et al., 1980). However, the socio-physiological factor did not have a  
248 significant effect in this study, because individuals grouped in five and acclimated at 30  
249 °C did not exhibit a significant reduction compared to the BMR observed in solitary  
250 individuals (Fig.1).

251 Furthermore, the decrease in energy expenditure individually within the group  
252 conducting huddling also has a per capita cost since the fuel consumed by thermogenesis  
253 is generated individually, although the benefits are shared by the group (Haig, 2007). In  
254 this vein, it is possible that some individuals could benefit more than others in huddling.  
255 Our video record revealed that some degus of our group treatments may have benefited  
256 more than others and thus decreased their BMR more, which is consistent that the  
257 coefficient of variation in BMR seems to be higher in grouped than in non-grouped  
258 acclimated degus. According to Bustamante et al. (2002), small individuals of the rodent  
259 *Phyllotis darwini* are most favored by the larger individuals huddling, especially at

260 temperatures below the TNZ. Further experimental studies will need to assess whether all  
261 individuals in the group performing huddling receive the same benefits, as for example,  
262 individuals who are dominant competitors and can occupy the best location in the center  
263 of the group (Shank and Alberts, 1997). In this vein, experiments of Bautista et al. (2008)  
264 reported that the offspring of rabbits (*Orytolagus cuniculus*) share the thermal advantages  
265 by moving continuously within the group. It would be interesting to study the effect of  
266 body size, ontogeny and social hierarchy within the group on the energy benefits in the  
267 medium term (acclimation effect) of huddling in adult and juvenile of *O. degus*.

268 Furthermore, from the viewpoint of comparison it would also be interesting to perform  
269 acclimation experiments of other species that exhibit different degrees of sociability.

270 Individual thermal conductance of *O. degus* was lower only in individuals acclimated  
271 at 15 °C, irrespective of the group size. Thus the results of this study suggest that  
272 compared to the temperature factor, the grouping effect would not be so important. It is  
273 possible that the metabolic changes due to huddling are less rigid than the changes that  
274 occur at the morphological level such as increased length and density of the fur, which  
275 could in turn restrict their movement (Cutrera and Antinuchi, 2004). Future studies will  
276 then be necessary to determine the mechanisms that explain the association between  
277 metabolic changes and thermal conductance acclimatization *O. degus*.

278 The convergence of huddling behavior among animals is a well established evolutionary  
279 event, being emperor penguins a notable case (Gilbert et al 2010). In this vein it is  
280 possible that this phenomena might also occur in other endotherms including birds, which  
281 has been demonstrated can reduce energy expenditure by comparable amounts when  
282 huddle (MacKechnie and Lovegrove 2001, Wojciechowski et al. 2011). Summarizing,  
283 during huddling animals group together and maintain close bodily contact, being  
284 particularly important for decreasing thermoregulatory costs and for increasing survival  
285 times when environmental conditions are harsh. Huddling induces metabolic depression  
286 without hypothermia and is mainly attributed to the reduced surface area/volume ratio of  
287 the huddling group and to the increase of effective temperature. The novelty of our  
288 results are that huddling not only decreases metabolic rate in individuals while grouped,  
289 but also provides significant energy savings at the individual level that persists after

290 huddling. Finally, further studies are necessary to assess if huddling behavior could  
291 modulate the thermal acclimation in other species of endotherms.

292

### 293 **Methods & Techniques**

294 Thirty-two adult individuals of *O. degus* were trapped with Shermann traps in autumn-  
295 winter 2011 in the Quebrada de la Plata, Chile (33° 28'S, 70° 54'W). Individuals were  
296 transported to the laboratory in Santiago, Chile where they were kept in cages of 2.0 m x  
297 2.5 m x 2.0m (length x width x height) with food and water *ad libitum* for one month, to  
298 minimize the possible effects of the previous thermal experience (see Nespolo and  
299 Rosenmann, 1997). Then individuals were randomly divided into two groups of 16  
300 individuals. One group was acclimatized at 15 °C and the other at 30 °C for 2 months,  
301 with water and food *ad libitum* and with a photoperiod of 12L: 12D. Each acclimation  
302 group was divided into three treatments: solitary individuals (n = 5), two groups of 3 (n =  
303 6) and one group of 5 (n = 5) individuals in plastic cages 87 cm x 33 cm x 32 cm (length x  
304 width x height). We decided to use a maximum size group of 5 since five individuals  
305 exhibited the largest energy savings by huddling in this species (Canals et al., 1989). All  
306 observations and measurements were performed on adult males and females with a body  
307 mass (mean ± SD) of 173 ± 25.7 g (males) and 147.92 ± 23.76g (females). No differences  
308 in body mass between sexes (p=0.1) or between groups of different size were found  
309 (p=0.09).

310

### 311 *Respirometry*

312 Following acclimation, BMR was measured as the oxygen consumption rate using a  
313 computerized open-flow respirometry system (Sable Systems, Henderson, NV). For BMR  
314 determinations, post-absorptive animals were placed individually in stainless steel  
315 metabolic chambers (5000 mL) for 12 h in the rest phase (night) in the dark at 30.0 ± 0.5  
316 °C, which is within the thermoneutral zone of this species (Rosenmann, 1977). To  
317 calculate thermal conductance we determined oxygen consumption at two temperatures  
318 below TNZ (20 and 10 ± 1 °C) for at least 2 h or until a visual inspection of the

319 recorded data allowed us to determine when steady- state conditions had been  
 320 achieved. With these values of oxygen consumption we estimated heat loss using the  
 321 following equation:  $f = \frac{M}{\Delta T}$ , (eqn. 1) where C is in  $J g^{-1} h^{-1} ^\circ C^{-1}$ . Values of oxygen  
 322 consumption were transformed into Joules using caloric equivalent of oxygen of 20.17 J  
 323 per mL  $O_2$ . Briefly, external air ( $700 mL min^{-1}$ ) was drawn into the metabolic chamber by  
 324 negative pressure created by a downstream vacuum pump controlled by a Sierra mass  
 325 flowmeter/ controller (Sierra Instruments, Monterey, CA), which was calibrated monthly  
 326 with a volumetric flowmeter. Before arriving in the chamber the air was dried using  
 327 Drierite desiccant and passed through Bev-A-Line tubing (Thermoplastic Processes). The  
 328 excurrent air from the metabolic chamber passed through Drierite, Baralyme, and Drierite  
 329 to remove water vapor and  $CO_2$  gas before being passed through the  $O_2$  analyzer (model  
 330 FoxBox, Sable Systems). The open-flow respirometry system was calibrated with a  
 331 known mixture of oxygen (20%) and nitrogen (80%) that was certified by  
 332 chromatography (INDURA, Chile). Because  $CO_2$  and water vapour was scrubbed before  
 333 entering the  $O_2$  analyzer, oxygen consumption was calculated according to Withers (1977  
 334 p. 122):

$$335 \quad \dot{V}O_2 = FR * 60 * \frac{F_i O_2 - F_e O_2}{1 - F_i O_2}, \quad (\text{eqn. 2}) \text{ where FR is the flow rate}$$

336 ( $mL min^{-1}$ ) and  $F_i$  and  $F_e$  are the fractional concentrations of  $O_2$  entering and leaving the  
 337 metabolic chamber, respectively. In order to confirm that animals were euthermic after the  
 338 metabolic trials ( $T_b = 37-38 ^\circ C$ ), we recorded their rectal body temperature ( $T_b$ ) with a  
 339 Cole Palmer 24-gauge copper-constantan thermocouple attached to a Digisense  
 340 thermometer (model 92800- 15). Outputs from the oxygen analyzer (%) were digitized  
 341 using a Universal Interface II (Sable Systems) and recorded on a personal computer using  
 342 data acquisition software (EXPEDATA, Sable Systems). Our sampling interval was 5 s.  
 343 We averaged  $O_2$  concentration of the excurrent air stream over an entire record period of  
 344 30 min after the lower steady state was reached (following Bozinovic et al., 2009). To  
 345 assess if changes in thermal/huddling conditions were accompanied with changes in  
 346 energy intake, we measured food consumption of grouped (5) and solitary animals at 15  
 347 and  $30 ^\circ C$ . At the end of the acclimation period, animals were placed in metabolic cages

348 to estimate food intake in 48-h trials. We fed animals with a known amount of pellet  
349 (rabbit food, Champion<sup>®</sup>) and after 48 h we collected the uneaten food. Samples were  
350 dried at 90 °C to constant weight ( $\pm 0.001$  g), and the intake was calculated by subtracting  
351 the uneaten to the total offered food.

352

### 353 *Thermography and behavior*

354 In a second experiment, the surface temperature of solitary and degus in groups of 5, was  
355 recorded at the end of acclimation periods using a thermal imaging camera FLIRi40  
356 calibrated at FLIR Systems Brasil (2011, www.FLIR.com). Images were recorded at a  
357 height of 1 m above the chamber and at two ambient temperatures of 15 and 25 °C .  
358 Thermal images were analyzed using the ad hoc software FLIR QuickReport 1.3 SP1;  
359 with a fur emissivity 0.98. Surface temperatures were averaged from 10 images for each  
360 treatment taken at the end of the respirometry periods. The contour surface temperature of  
361 degus was determined by fitting a polygon around the individual animal in the case of  
362 isolated pups and around the entire huddle for grouped animals by using the option  
363 “isotherm” of the software. This isotherm is the average temperature along the polygon,  
364 which defines the limit between above and below temperatures observed in the thermal  
365 image. The maximum temperature of degus was determined by using the options area of  
366 the software. After fitting the area of huddled or solitary animals, a unique maximum  
367 temperature value was recorded for each thermography. Furthermore, we determined the  
368 temperature of contact between pairs of individuals in grouped animals using the option  
369 mobile, by fitting a line of contact between paired animals, dividing it in five sections and  
370 using the four points inside the sections in order to get the average surface temperature of  
371 contact between two animals. Then we averaged those temperatures to calculate the  
372 average temperature of contact of the entire group. Finally, in order to assess whether  
373 grouped animals constantly changed positions, we determine the proportion of time that  
374 each animal spent in various positions within the group. For this, we marked each  
375 individual with hair dye on the back and performed 10 two-hour films each in a group of  
376 5 animals kept at 15 ° C. The measurements were performed at 1 m height using a Sony  
377 NP-FV50. The films were analyzed on a personal computer by recording the time each

378 individual spent in the center, on the periphery of the group or isolated while doing  
379 huddling in groups of five individuals. In order to maximize the time that each animal was  
380 analyzed and because the groups are not always formed with the maximum number of  
381 individuals (i.e., 5), video recordings were analyzed only when the animals spent more  
382 than 15 min clustered in groups of at least 4 individuals. The proportion of time that  
383 animals was in such behavior approached 87%.

384

### 385 *Statistical Analysis*

386 We performed a General linear Model (GLM) to compare BMR between treatments using  
387 Log BMR and thermal conductance as the dependent variables, and sex, temperature and  
388 the number of individuals in the group as fixed factors and log body mass as covariate.  
389 Because our analyses exhibited a non-significant effect of sex ( $F_{1,19} = 0.84$ ,  $r^2 = 0.01$ ,  $p =$   
390  $0.77$  and  $F_{1,19} = 0.84$ ,  $r^2 = 0.01$ ,  $p = 0.77$  respectively) this term was dropped from the  
391 models. Our analysis also revealed that thermal conductance was not related to body mass  
392 in degus ( $F_{1,30} = 0.99$ ,  $r^2 = 0.03$ ,  $p = 0.33$ ), thus, we only performed an ANOVA to test  
393 differences among experimental groups. Then a *post hoc* Fisher test was performed to  
394 determine significant differences in mass-adjusted BMR (least square means calculated  
395 from ANCOVA) and thermal conductance between specific groups. To analyze the  
396 association between physiological variables and body mass, a simple linear regression  
397 was performed. All the data met the assumptions of the ANOVA. Surface and contour  
398 temperatures of the different experimental conditions (grouped and solitary at 15 and 30  
399 °C) were compared with one-way ANOVAs and then a post-hoc Tukey test was  
400 performed to check for specific differences among treatments. Proportion of time each  
401 animal spent in the center of the group was evaluated by means of counting the times each  
402 animal was viewed in this position in random analyses. Each tape record was analyzed  
403 through random selection of five periods of 6 min by trial (two hour each). We hence  
404 analyzed a total of 50 records for which we obtained 239 individual observations. We  
405 count as positive when an animal were surrounded by at least two animals and negative  
406 when where the subject was located at the periphery of the group. Then, we calculated the  
407 observed frequency of times that each animal spent in the center of the group and the

408 expected frequency by chance (i.e., total number of observations divided by the number of  
409 animals). A Chi-square test was used to estimate significant differences among  
410 individuals. Food intake was evaluated by means of a nonparametric Kruskal-Wallis test  
411 and then a multiple comparison of mean rank was performed. All results are reported as  
412 mean  $\pm$  SE. All analyses were performed using Statistica 7.0 software (StatSoft Inc.,  
413 Tulsa, OK, USA).

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635 **Figure legend**

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637 Figure 1. Mass-adjusted basal metabolic rate in *Octodon degus* as a function of the  
638 number of individuals grouped after a thermal acclimation of two months to two  
639 temperature conditions. The black bars represent the mean  $\pm$  SE of BMR animals  
640 acclimated to 15 ° C, and the white to 30 °C. Similar letters denote non-significant  
641 difference between treatments by a Fisher *a posteriori* test.

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643 Figure 2. Lower panel: Surface temperatures of the back (maximum), perimeter and  
644 contact of isolated and huddling *Octodon degus*, at 23°C and 15 °C and 30 °C. Similar  
645 letters denote non-significant difference between treatments by a Fisher *a posteriori* test.  
646 Upper panel: Thermal images are shown for isolated and huddling degus, exposed to an  
647 ambient temperature of 15 °C. Examples of measurements of the temperature of the  
648 perimeter (Tp), maximum temperature (Tm) and temperature of the contact between  
649 individuals (Tc) are shown.

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664 Table 1. Body mass (bm), whole animal BMR and wet thermal conductance (Ct) of  
665 *Octodon degus* acclimated (Ta) either at 15 or 30 °C for two months in groups of 3, 5 and  
666 solitary individuals. Data are presented as mean  $\pm$  SD. In parenthesis is shown the  
667 coefficient of variation in percentage.

Ta	Group size	bm (g)	BMR (kJ h <sup>-1</sup> )	Ct (J g <sup>-1</sup> h <sup>-1</sup> ° C <sup>-1</sup> )
	1	152.27 $\pm$ 12.20	2.78 $\pm$ 0.24 (8.7)	0.67 $\pm$ 0.25
15 °C	3	176.55 $\pm$ 9.80	2.59 $\pm$ 0.25 (9.6)	1.25 $\pm$ 0.84
	5	148.37 $\pm$ 17.70	1.96 $\pm$ 0.45 (23.1)	1.30 $\pm$ 0.38
	1	192.27 $\pm$ 10.37	2.60 $\pm$ 0.23 (8.2)	1.50 $\pm$ 0.33
30 °C	3	157.44 $\pm$ 8.16	1.92 $\pm$ 0.36 (18.8)	1.72 $\pm$ 0.17
	5	146.33 $\pm$ 13.60	2.19 $\pm$ 0.32 (14.6)	1.97 $\pm$ 0.42

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