

RESEARCH ARTICLE

Transcriptomic changes associated with maternal care in the brain of mouthbrooding cichlid Astatotilapia burtoni reflect adaptation to self-induced metabolic stress

Joshua J. Faber-Hammond and Suzy C. P. Renn*

ABSTRACT

Parental care in Astatotilapia burtoni entails females protecting eggs and developing fry in a specialized buccal cavity in the mouth. During this mouthbrooding behavior, which can last 2-3 weeks, mothers undergo voluntary fasting accompanied by loss of body mass and major metabolic changes. Following release of fry, females resume normal feeding behavior and quickly recover body mass as they become reproductively active once again. In order to investigate the molecular underpinnings of such dramatic behavioral and metabolic changes, we sequenced whole-brain transcriptomes from females at four time points throughout their reproductive cycle: 2 days after the start of mouthbrooding, 14 days after the start of mouthbrooding, 2 days after the release of fry and 14 days after the release of fry. Differential expression analysis and clustering of expression profiles revealed a number of neuropeptides and hormones, including the strong candidate gene neurotensin, suggesting that molecular mechanisms underlying parental behaviors may be common across vertebrates despite de novo evolution of parental care in these lineages. In addition, oxygen transport pathways were found to be dramatically downregulated, particularly later in the mouthbrooding stage, while certain neuroprotective pathways were upregulated, possibly to mitigate negative consequences of metabolic depression brought about by fasting. Our results offer new insights into the evolution of parental behavior as well as revealing candidate genes that would be of interest for the study of hypoxic ischemia and eating disorders.

KEY WORDS: Behavior, Hypoxia, Parental care, Feeding, Reproductive strategy, Starvation, Transcriptome

INTRODUCTION

Parental behavior has evolved multiple times in animals. Although it is a costly reproductive strategy for parents, it increases fitness through higher offspring survival rate. These parental costs come in the form of time and energy, as time spent caring for and protecting offspring must be balanced against competing behavioral interests such as feeding and mating (Numan and Woodside, 2010; Olazábal and Young, 2006). For most species, the diverse set of activities that compose 'feeding' and 'parental care' must be co-regulated through partially overlapping neuroendocrine circuits, and evidence indicates substantial cross-talk between these circuits in at least some species (e.g. Olszewski et al., 2010; Crossin et al., 2012; Shahjahan et al., 2014). While some species increase feeding during

Department of Biology, Reed College, Portland, OR 97202-8199, USA.

*Author for correspondence (renns@reed.edu)

D J.J.F., 0000-0002-0954-0025; S.C.P.R., 0000-0003-0122-9934

parental care to compensate for the added demands, others inhibit feeding behavior to favor parental care, a tradeoff that comes at the expense of parental condition (Fischer and O'Connell, 2017; O'Rourke and Renn, 2015).

The neural circuits and molecular mechanisms regulating parental care and feeding behaviors are well studied, primarily in mammalian systems. Ongoing work provides inference regarding homology across vertebrates at both the circuit (Goodson, 2005; Goodson and Kingsbury, 2013; O'Connell and Hofmann, 2011) and molecular signaling (Volkoff, 2016) levels. The close connection between reproduction and energy balance is well documented in mammals with several nuclei of the 'maternal brain' receiving either direct or indirect input from the arcuate nucleus feeding regulatory neurons (Schneider, 2004). Within the arcuate nucleus of the hypothalamus, two specific populations of neurons, one producing orexigenic neuropeptide Y (NPY) and agouti related peptide (AgRP), and the other producing anorexigenic pro-opiomelanotcortin (POMC) and cocaine- and amphetamine-regulated transcript (CART), are directly responsive to the peripheral signals of satiety, leptin (Cone, 2005) and ghrelin (Cowley et al., 2003).

Mouthbrooding, a form of parental care seen in several species of fish in which the parent holds the developing eggs and fry in the buccal cavity, represents an extreme example of food restriction and parental investment. In an ultimate sense, mouthbrooding is an adaptation in response to predation pressure (Kuwamura, 1997) and has evolved independently in cardinalfishes, gobies, betas (Rüber et al., 2004) and several clades of cichlid fishes (Barlow, 2000; Fryer and Iles, 1972), where it is thought to have promoted rapid speciation (Salzburger et al., 2005). In a proximate sense, mouthbrooding severely limits food intake, such that most species completely abstain from food intake while others obtain food only for provisioning the fry (Yanagisawa et al., 1996). This brooding phase is plastically adjusted in response to predation threat (Taborsky and Foerster, 2004), noise-induced perturbations (Butler and Maruska, 2021) and parental energy reserve as well as the quality and availability of mates (Iwao et al., 2004).

In the African cichlid fish Astatotilapia burtoni, it is the female alone that provides parental care in the form of mouthbrooding, abstaining from food intake for 2-3 weeks. This leads to a dramatic reduction in body mass (Grone et al., 2012; Renn et al., 2009), and reduced gonadosomatic index associated with lowered estradiol and 11-KT (a fish-specific androgen) (Maruska et al., 2020b). Unlike a state of involuntary starvation, feeding motivation is decreased during mouthbrooding even when the developing eggs are removed from the buccal cavity (Mrowka, 1984), and thus the female does not anticipate caloric intake. The capacity to withstand and recover from prolonged periods of food deprivation requires behavioral, physiological, morphological and molecular adaptations

conserve energy and maintain immune function (Secor and Carey, 2016). We have previously identified an enhanced reduction in gut epithelial turnover during mouthbrooding as one such potentially adaptive mechanism for energy conservation (Faber-Hammond et al., 2019). While the full extent of metabolic changes during mouthbrooding is not known, it may share similarities with changes seen in animals that engage in hibernation or other related extreme adjustments when caloric intake is not anticipated. During hibernation, shifts in mitochondrial ATP production and calcium uptake may improve energy efficiency (Barger et al., 2003), neuroprotective mechanisms may mitigate oxidative damage (Yin et al., 2016), and changes to brain water homeostasis can also impact neuronal signal transduction (Osborne and Hashimoto, 2008).

The expression of genes encoding some neuropeptides and their receptors is known to differ between brooding, non-brooding and starved females, suggesting that different regulatory mechanisms control this special case of feeding inhibition (Grone et al., 2012). Furthermore, recent work examining neuronal activation across the brain during mouthbrooding in *A. burtoni* provides evidence for a large degree of shared circuitry involved in the regulation of maternal care, food intake and energy balance with nuanced differences in specific brain regions (Maruska et al., 2020a). Such work predicts global changes in gene expression across the brain with specifically regulated genes in some regions.

In the current study, we analyzed whole-brain transcriptomes of female A. burtoni from multiple time points across the reproductive cycle. The time points selected represent dramatically different stages in terms of nutrition state, energy requirements and parental care behaviors. We conducted differential expression analyses and parallel enrichment analyses for functional gene sets to identify patterns of upregulation or downregulation at either of two specific mouthbrooding stages relative to each other and to two post-release time points, one of which includes intensive maternal care. Genes that are differentially regulated across the maternal stages (both mouthbrooding and post-release) may underlie the specific behaviors performed at these time points, while genes showing trends that correlate with decreasing body condition may reflect energy reserves. In addition to providing valuable insight into the evolution of parental behavior, our results reveal previously unknown changes in the maternal cichlid brain that could have clinical impacts in the study of brain hypoxic ischemia, inflammation and neuroprotection. Our holistic approach using whole-brain samples aims to identify novel mechanisms beyond a candidate gene approach and capture global changes of a metabolically active tissue.

MATERIALS AND METHODS

Animal husbandry

Astatotilapia burtoni (Günther 1894) used in this experiment were derived from the same lineage as was sequenced for the reference genome assembly (GCA_000239415). Females of reproductive age were maintained in 30 gallon (~1141) mixed-sex stock tanks with 2–4 males and 8–12 females. Tanks were kept on a 12 h:12 h day: night cycle with 0.5 h dawn and dusk settings. The salinity and pH

levels of tanks were matched to the average levels of Lake Tanganyika. Tanks were checked daily, and when brooding females were captured and fry development stage was confirmed. Females were assigned to one of four treatment groups: 2 days mouthbrooding (B02), 14 days mouthbrooding (B14), 2 days post-release of fry (R02) and 14 days post-release (R14) (Fig. 1). To avoid confounding date with treatment groups, females were sequentially assigned one to each group, with the exceptions that only females confirmed to be holding stage 0 fry were assigned to the B02 group. Brooding females were placed into a 5 gallon (\sim 191) tank with a terracotta pot for shelter and gravel. A single male stimulus fish (\sim 3 g±1 g) was housed in the same tank, retained by a cylindrical mesh barrier (10 cm diameter) to allow visual and olfactory contact. The male stimulus fish was fed standard cichlid pellets daily within the confined cylinder. Because brooding females are not expected to eat, the addition of food would have fouled the tank. Therefore, females had visual and olfactory cues regarding food availability without actual access to it. Once females released the fry, they were fed standard fish flakes daily. All protocols complied with Renn lab IACUC approval #01-2017.

Sampling

Females were monitored daily for brooding or release of fry. All fry were removed 2 days after the initial release date. Any female that cannibalized her fry prior to 2 days post-release was reassigned to the R14 group, so that all R02 fish were actively caring for freeswimming fry. At B02, B14, R02 and R14 time points, females were anesthetized in MS-222, weighed, measured for standard length and rapidly decapitated. Whole brains were removed and preserved in 1 ml RNAlater (Ambion) and stored at -80°C. All dissections were done between 16:00 and 18:00 h. While wholebrain gene expression profiling avoids the technical variation introduced by brain punch techniques, this approach is not without caveats and limitations. Primarily, region-specific regulation and genes expressed in a limited number of cells are less likely to be detected. Certain genes are known to be differentially regulated in brain region-specific patterns that are lost with whole-brain expression profiling. For example, Butler et al. (2020) studied the role of galanin in maternal care and infanticide in A. burtoni, and found that expression in preoptic area (POA) neurons is associated with maternal care while expression in nucleus lateralis tuberis (NLT) neurons is associated with feeding. While this finer scale resolution is lost in analysis of whole brains, our method does capture a snapshot of the broad changes in the brain throughout the reproductive cycle associated with this metabolically demanding behavior.

RNA extraction/sequencing

Brains were removed from RNAlater and placed in 200 μ l of 1-Thioglycerol/Homogenization Solution from the MAxwell 16 LEV Simply RNA Tissue Kit (Promega AS1280). Samples were homogenized using a hand-held pestle motor and plastic pestle in a microcentrifuge tube. Homogenized samples were transferred to the MAxwell 16 LEV Simply RNA Tissue Kit cartridge and processed

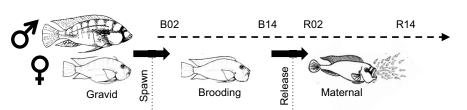


Fig. 1. Astatotilapia burtoni sample groups. Whole brains were harvested from females 2 days after the start of mouthbrooding (B02), 14 days after the start of mouthbrooding (B14), 2 days post-release of fry (R02) and 14 days post-release of fry (R14).

in the Maxwell 16 Instrument (Promega AS1000) using the High Strength LEV Magnetic Rod and Plunger Bar Adaptor (Promega SP1070). Standard DNAse treatment was included and samples were eluted in 50 μ l H₂O including RNAseOUT (Invitrogen). RNA quality was initially checked by electrophoresis and quantity by NanoDrop 1000 (Thermo Scientific). RNA (1–2 μ g per sample) was sent to Novogene (Sacramento, CA, USA) for library preparation and paired-end 150 bp sequencing (Illumina Platform PE150).

Bioinformatics

Paired-end RNA-seq reads were preprocessed with TrimGalore v0.6.4 (https://www.bioinformatics.babraham.ac.uk/projects/trim_galore/) for adaptor trimming and FastQC v0.11.3 (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/) for quality filtering. Remaining paired reads were aligned to the *A. burtoni* reference genome (GCF_000239415.1) with STAR v2.5.3 (Dobin et al., 2013), using the associated RefSeq GTF file to guide the alignment. Aligned reads were assembled and quantified using StringTie v1.3.4 (Pertea et al., 2015), and count tables were constructed as input for differential expression analysis. All methods, results and discussion of a WGCNA analysis (v1.69: Langfelder and Horvath, 2008) that was conducted in parallel are available from GitHub (github.com/jfaberha/cichlid_transcriptome/tree/main/lab_stock_analysis).

Data normalization and subsequent differential expression analysis were performed with DESeg2 v1.20.0 (Love et al., 2014) in R-studio v1.1.383 (http://www.rstudio.com/). A likelihood ratio test (LRT) was performed to generate a list of genes that showed significant expression differences between any of the four sampled time points, followed by pairwise binomial Wald tests to discover which pair of time points differ. As a supplementary analysis, brooding (B02, B14) and release (R02, R14) stages were pooled and a pairwise binomial Wald test was performed to discover candidate genes that differentiate by associated parental behaviors and response to food intake. Genes showing significant differential expression from both tests are reported at a false discovery rate (FDR)<0.1 and for exploratory purposes we report genes trending towards differential expression at an unadjusted P<0.05 (results reported in Table S1). This less stringent threshold provided a sufficient number of genes for downstream functional analyses. LRT differentially expressed genes (DE genes) were tested for gene ontology (GO) term enrichment with BiNGO v3.0.4 (Maere et al., 2005), implemented in Cytoscape v3.7.2 (Shannon et al., 2003). While A. burtoni transcripts were assigned names based on their NCBI RefSeq GTF annotation, they were reannotated by predicted protein sequence to obtain a comprehensive list of GO terms (Table S2). In order of priority, GO annotations were preferentially used from the Swiss-Prot, TrEMBL (The UniProt Consortium, 2019) and EggNOG (Huerta-Cepas et al., 2019) protein databases (accessed February 2018).

We performed hierarchical clustering analysis on subsets of genes showing significant differential expression and trending towards differential expression that address original and novel exploratory hypotheses with the goal of discovering co-expression and possible interaction of candidate genes. For all clustering analyses, we used variance stabilizing transformation (VST)-normalized count tables generated by DESeq2, and constructed heatmaps using the made4 v1.56.0 R-package (Culhane et al., 2005). This package utilizes the correlation similarity metric and average linkage clustering for expression data, which allows for clustering of expression patterns regardless of the overall magnitude of expression. Clustering was

performed on individuals to generate expression-based dendrograms, while the associated heatmaps display time-point medians for easier interpretation of expression data. Bootstrap values for dendrograms were generated with the PVclust v2.2-0 R-package (Suzuki and Shimodaira, 2006) following 10k clustering iterations.

In addition to the detection of individual DE genes, we aimed to identify changes in functional gene categories across time points; therefore, we ran Gene Set Enrichment Analysis (GSEA v4.0.3) for sets of genes representing GO categories (Subramanian et al., 2005). Pairwise comparisons were done with gene-level VST-normalized counts between each of the four time points in both directions, resulting in 12 possible comparisons. Only six comparisons are possible between four time points; however, reciprocal comparisons (e.g. B02 versus B14 and B14 versus B02) were performed because GSEA results slightly differ based on the direction of the test, due to significance being determined by ranking of sets, and the union is reported. For each GSEA comparison, 1000 permutations of gene set randomization were performed because there were fewer than 10 samples per group. Sets with FDR<0.05 were considered significantly enriched, and the GO categories had to meet this threshold in either of two reciprocal tests per time-point comparison. GSEA documentation recommends an FDR cutoff of 0.25 for determining significance in exploratory analyses because of false negatives being less desirable than false positives in such analyses; however, we chose the more restrictive alpha threshold of 0.05 to limit the number of significant GO sets. GSEA tests were followed by leading-edge analysis to determine both the direction of change and the overlap between enriched gene sets.

Significantly enriched GO gene sets are not fully independent in the gene accessions they contain, as single genes often have a number of associated GO terms. To account for this, we took steps to reduce complexity in the list of significant GO terms by merging terms with minimum 50% overlap of member genes into GO families. Overlap coefficients (CO) were calculated as the proportion of shared genes between pairs of terms divided by the smaller of the two gene sets. The inverse values of these coefficients (1–CO) were converted into a distance matrix for all significant GO terms from GSEA, then used to build a neighbor joining tree with PHYLIP v3.698 (http://evolution.genetics.washington.edu/phylip. html). TreeClust v1.00 (Balaban et al., 2019) was used to cluster nodes that had a maximum distance of 0.5 from each other, equivalent to a minimum 50% overlap coefficient between GO terms. Merged GO families were named using the most significant (lowest FDR) term from all GSEA time-point pairwise comparisons.

For hierarchical clustering of significant GO terms and families, GSEA was re-run to obtain normalized enrichment scores (NES) of each of four time points versus all other samples, with positive values representing upregulation and negative values representing downregulation of entire gene sets in an analog to \log_2 -fold change of individual genes. Hierarchical clustering was performed using the Ward.D2 clustering and Euclidean distance methods implemented by the Heatmap tool from the ComplexHeatmaps R package v2.6.2 (Gu et al., 2016; Ward, 1963).

RESULTS

The time points selected for the current study represent dramatically different states with regard to behavioral, feeding and metabolic activity. We used the R14 stage as a baseline in which there is active feeding behavior, adequate energy reserves and normal metabolic activity but no maternal care behavior. The early brooding stage,

B02, is characterized by modest buccal churning as maternal care, no feeding behavior, adequate energy reserves and potentially reduced metabolic activity to conserve energy; and while the later brooding stage, B14, shares the lack of feeding behavior, it is also characterized by more intense buccal-based maternal care and dramatically reduced energy reserves, which likely necessitates dramatic regulation of metabolic activity. The early release stage, R02, shares this increased maternal care but of a very different nature in that it includes aggressive territory defense, and while feeding behavior has resumed, energy stores remain depleted, again potentially requiring substantial metabolic regulation.

Transcriptome profile statistics

Twenty-two samples passed QC filters for analysis, with 5 samples representing B02 and R02 time points and 6 samples representing B14 and R14 time points. From ~610 million total high-quality reads (mean per sample ~27.7 million±7.8 million), 84.6% reads were aligned to the *A. burtoni* genome. These *A. burtoni* brain transcriptomes yielded hits to 25,572 out of the 44,653 RefSeq annotated genes (57.3%), which accounted for 92.9% of final gene counts. Conversely, 7.1% of gene counts represented novel unannotated transcripts within the genome assembly, and upon GO annotation for the entire transcriptome we found only a small portion (17.7%) of these novel transcripts to be putative protein coding genes, many of which appeared to code for transposable element machinery. Therefore, only annotated NCBI RefSeq genes in the assembly were used for downstream analyses.

Differential gene expression

We found 128 genes to be differentially expressed across the four reproductive time-points through DESeq2 LRT analysis with an FDR<0.1, making up 0.5% of RefSeq annotated genes expressed in the brain. For exploratory purposes, we also examined a secondary list of 1643 genes (1515 additional genes) trending toward differential expression at an unadjusted P<0.05 (Table S1). With post-hoc pairwise comparisons, we explored patterns among the less stringent set of genes trending toward differential expression (Table 1). The highest number of DE genes, 105, occurred between late brooding B14 and early release R02 stages in the brooding cycle (44 B14 biased: 61 R14 biased) when maternal behavior and feeding are dramatically changing. Conversely, the fewest number of DE genes, 20, was detected in the contrast between early brooding B02 and early release R02 (11 B02 biased and 9 R02 biased). Among the full set of DE genes identified for a given brooding stage (both upregulated and downregulated), only a small percentage were biased toward that brooding stage relative to all

Table 1. Summary of the number of differentially expressed genes

	B02	B14	R02	R14	All 3	% Specific
B02		16	11	5	0	2.56
B14	26		44	7	0	0.56
R02	9	61		7	0	1.36
R14	11	26	15		0	0
All 3	2	1	2	0		

Differentially expressed genes (DE genes) between stages were identified by pairwise comparison (binomial Wald tests, FDR<0.1). Red (above the diagonal) indicates upregulation in the stage named in the row versus the stage named in the column; blue indicates downregulation in the stage named in the column stage compared with that in the stage named in the row. Deeper shades represent DE genes found in all three pairwise comparisons and the percentage calculated for stage-specific regulation is shown.

three other stages (B02 2.56%, B14 0.56%, R02 1.36%, R14 0%), meaning that there were few stage-specific gene expression levels. The same dynamic pattern was seen among the larger list of genes trending toward differential expression, with a greater percentage of differential expression being stage specific (2.5-6.8%). While the two brooding stages are very different in terms of duration of starvation and other physiological factors, they share a mouthbrooding maternal component that differs from either of the release stages; therefore, we also combined the two brooding stages and the two release stages and performed a pairwise pooled brooding versus pooled release binomial Wald test. This analysis revealed 31 DE genes at FDR<0.1, 21 of which were significant DE genes, with the remaining 10 trending towards significance based on LRT analysis. An additional 1124 genes trended towards differential expression in the brooding versus release analysis at an unadjusted P<0.05 (Table S1). Functionally interesting candidate genes appearing in both LRT and brooding versus release differential expression lists are highlighted in expression heatmaps while the following discussion primarily reports on the LRT DE gene list across all four time points.

The DE genes discussed below include several that are classically associated with parental care such as the teleost ortholog to oxytocin, isotocin neurophysin (itnp), and others that have more recently emerged as potential candidates in parental care such as neurotensin (nts). We found strong correlation of expression that clustered genes involved in parental care with others classically associated with feeding, such as galanin (gal). As expected, given the voluntary starvation phase, many of the known orexigenic genes and anorexigenic genes varied dramatically across the time points sampled, and we saw compensatory response genes that constitute metabolic regulatory mechanisms, suggesting hypoxic conditions and activation of neuroprotective genes.

GO enrichment

GO enrichment for the 128 genes determined to be differentially expressed across stages by LRT analysis (regardless of comparison or directionality) yielded only six significant terms (Table 2). However, none of the enriched GO terms were typically associated with maternal or voluntary fasting behavior in our searches of current literature. Instead, five were related to oxygen transport, and the sixth was 'negative regulation of protein phosphatase type 2B activity'. The oxygen transport GO categories were detected based on differential expression of four globin genes, and 'negative regulation of protein phosphatase type 2B activity' was represented by two DE genes, repressor of calcipressin1-like (rcan11) and peptidyl-prolyl cis-trans isomerase fkbp1-like. Although the number of significantly differentially expressed genes in both of these groups was low and significance levels may be inflated as a result, there were only 25 and three non-significant genes in these GO categories, respectively, out of the 22,924 annotated genes in the brain transcriptome. In addition, these GO groups are represented by genes that show dramatic changes across time points, with globin gene hemoglobin subunit beta-A-like as the third-most significant DE gene (FDR=4.68E-6) and rcan11 as the second-most significant gene (FDR=8.75E-14) in the dataset. When running GO enrichment analysis on the 1643 genes that trend toward differential expression for an unadjusted P < 0.05, we found a list of six different enriched terms (Table 2), including 'hormone activity' as the top GO term, represented by 36 accessions. These findings suggest that observed behavioral changes may not require vast changes in expression of entire gene pathways, but rather they could result from dramatic changes in a small number of genes or minor expression changes in larger gene networks.

Table 2. Identification of genes that differ in expression across experimental time points

Differential expression test set threshold	GO ID	Description	Correlated <i>P</i> -value	No. of test genes	No. of reference genes
FDR<0.1	GO:0005833	Hemoglobin complex	9.30E-03	4/124	22/22.924
FDR<0.1	GO:0005344	Oxygen transporter activity	9.30E-03	4/124	25/22,924
FDR<0.1	GO:0015671	Oxygen transport	9.30E-03	4/124	26/22,924
FDR<0.1	GO:0015669	Gas transport	9.30E-03	4/124	27/22,924
FDR<0.1	GO:0019825	Oxygen binding	9.30E-03	4/124	28/22,924
FDR<0.1	GO:0032513	Negative regulation of protein phosphatase type 2B activity	4.45E-02	2/124	3/22,924
P<0.05	GO:0005179	Hormone activity	3.46E-03	36/1563	213/22,924
P<0.05	GO:0033269	Internode region of axon	3.13E-02	6/1563	9/22,924
P<0.05	GO:0061077	Chaperone-mediated protein folding	3.86E-02	23/1563	126/22.924
P<0.05	GO:0005576	Extracellular region	4.09E-02	470/1563	5869/22,924
P<0.05	GO:0048200	Golgi transport vesicle coating	4.09E-02	5/1563	7/22,924
P<0.05	GO:0048205	COPI coating of Golgi vesicle	4.09E-02	5/1563	7/22,924

Genes were identified based on gene ontology (GO) enrichment of DE gene lists from likelihood ratio test (LRT) analyses performed in DESeq2.

GSEA

We used GSEA as a parallel approach to discover enriched pathways that differ between time points, which allowed us to detect differences in aggregate signal for groups of genes related in function even when individual member genes failed to meet significance thresholds in differential expression analysis. In fact, there were a number of enriched sets that failed to contain any identified DE genes. Because there was partial overlap of gene accessions between sets, we merged significant GO sets into families, and after this data reduction step we found a total of 23 enriched GO families composed of 107 GO sets and an additional 32 unmerged GO terms for a total of 55 independent sets (Fig. 2; Table S3). These enriched GO terms include several that are potentially involved in modifying behavior, including several related to neuronal growth/death and neuropeptide receptor activity. Other notable enriched families and sets are related to ion channel regulation, oxygen transport, energy metabolism and inflammatory response. Following NES clustering for these sets, the two dominant patterns that emerged were both highly dynamic, alternating between upregulation and downregulation across adjacent time points. The majority of independent GO sets and families (38/55) showed these patterns with 19 sets upregulated in B02 and R02 relative to B14 and R14, and another 19 sets showing the opposite pattern. Of the remaining 17/55 sets, 14 showed more gradual expression changes across the approximately cyclical time points, with adjacent pairs of time points having either higher or lower expression than the two distal time points; the other three sets were families composed of GO terms with inconsistent NES patterns between time points.

DISCUSSION

Among cichlids, mouthbrooding is thought to have arisen 10–14 times (Goodwin et al., 1998) and once at the base of the remarkably diverse haplochromine clade (DeVos and Seegers, 1998). Thus, the mouthbrooding haplochromine cichlid species *A. burtoni*, an important model system for the physiological and genomics studies of complex social behavior (Maruska and Fernald, 2013; Brawand et al., 2014), offers a useful perspective to identify mechanisms underlying mouthbrooding that may have contributed to diversification in this cichlid clade and possibly also in others.

Across the mouth-brooding reproductive cycle, female *A. burtoni* undergo two important transitions: an initial transition to starvation during the obligatory mouthbrooding phase and a second transition to increased aggression and maternal defense following release of fry (Renn et al., 2009). The care given to the developing offspring

during the mouthbrooding phase includes metabolically demanding buccal churning to aerate the fry. As the obligatory phase of mouthbrooding continues, this metabolic demand is coupled with a dramatic reduction in body condition as a result of prolonged voluntary starvation. Following the release of fry from the buccal cavity, the metabolically demanding maternal care is coupled with the resumption of feeding behavior. From the perspective of parental care, while considering fundamental distinctions, these two transitions can be compared to initiation of mammalian pregnancy and parturition, or avian incubation of eggs, which involves an early decrease in activity for incubation followed by increased activity for provisioning of chicks. From the perspective of feeding behavior, these transitions are dramatically different from mammalian transitions of pregnancy and parturition given the complete lack of food intake and dramatic reduction in body condition during brooding.

The time points selected for the current study highlight these important transitions that encompass behavioral, feeding and metabolic changes. The late release time point, R14, provides a rough baseline during which extensive maternal care and aggression have subsided. The early brooding phase, B02, describes the initial transition to the maternal brooding stage, while late brooding stage, B14, describes an individual that has undergone substantial loss of body condition and yet retains brood care motivation while sustaining suppression of feeding behavior. The early release stage, R02, describes the social stages of maternal care as the female interacts with offspring, shows heightened aggression and has resumed feeding behavior. That we found the greatest number of differentially regulated genes between B14 and R02 likely reflects the dramatic differences with regard to both maternal behavior and body condition.

The results discussed below include both known and novel candidates for the regulation of maternal care, feeding behavior and metabolism. In addition, oxygen transport pathways were found to be dramatically downregulated throughout mouthbrooding while neuroprotective pathways and immune response were upregulated, likely to mitigate the negative consequences of metabolic depression brought about by fasting. These results offer new insights into the evolution of mouthbrooding behavior. The individual genes identified by differential expression analysis are corroborated by GO term and GSEA analysis implicating full pathways in these transitions.

Some of the GO and GSEA analysis results (e.g. 'equilibrioperception' and 'sensory perception of light') indicate biological processes known to be controlled by specialized brain regions (Kasumyan, 2004; Rosa Salva et al., 2014), thus pointing to the need for future studies focused on more localized gene expression in

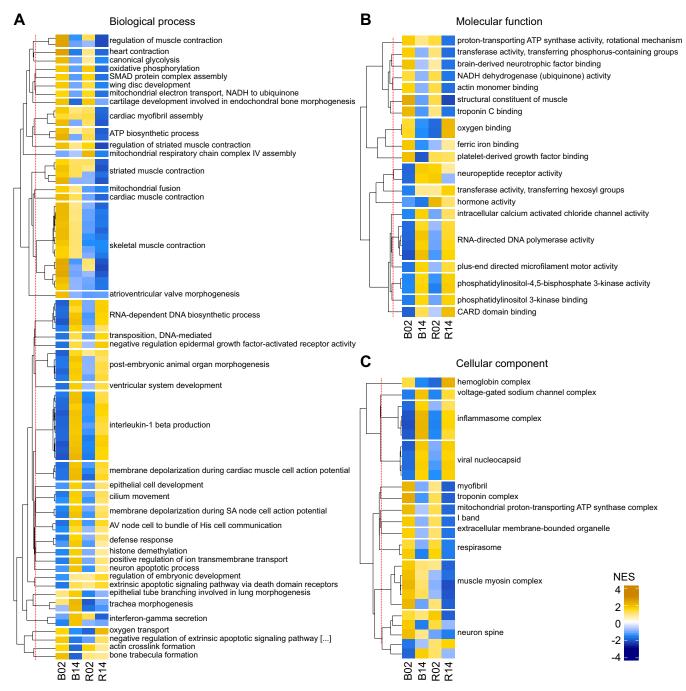


Fig. 2. Heatmap showing clustering of normalized enrichment scores (NES) for significant gene ontology (GO) gene sets identified by Gene Set Enrichment Analysis (GSEA). Terms from each of the three GO categories are shown separately: (A) biological process, (B) molecular function and (C) cellular component. Terms were merged if they had a minimum overlap coefficient of 50% (represented by the dotted red line) for the number of shared member genes, and hierarchical clustering of GO terms was performed using the Ward.D2 clustering method. GO terms on the right of each heatmap represent either unclustered GO terms or the most significant GO term within a merged cluster. For B02 and R02 time points, n=5; for B14 and R14 time points, n=6.

the brain using micropunch or single-cell approaches. With regard to maternal mouthbrooding, it is known that activation of galanin neuropeptide-expressing neurons in the POA plays a role in maternal care and infanticide while activation of neurons expressing the same peptide in the NLT brain region is associated with feeding regulation (Butler et al., 2020). To the extent that neuronal activation correlates with gene regulation, when not studied in isolation, gene expression profiles from these regions controlling different aspects of behavior will be lost.

Parental and reproductive behavior

Numerous studies show a positive correlation between parental care and expression of the teleost ortholog of *oxytocin*, *isotocin neurophysin* (*itnp*) (DeAngelis et al., 2017; Keverne and Kendrick, 1992; O'Connell et al., 2012; Pedersen et al., 1982), making it a candidate gene for regulation in the current study. When compared with the R14 baseline, late stage brooding females in B14 showed significantly lower expression of *itnp* (FDR=7.32E-2) (Fig. 3). This negative, rather than positive, correlation between *itnp*

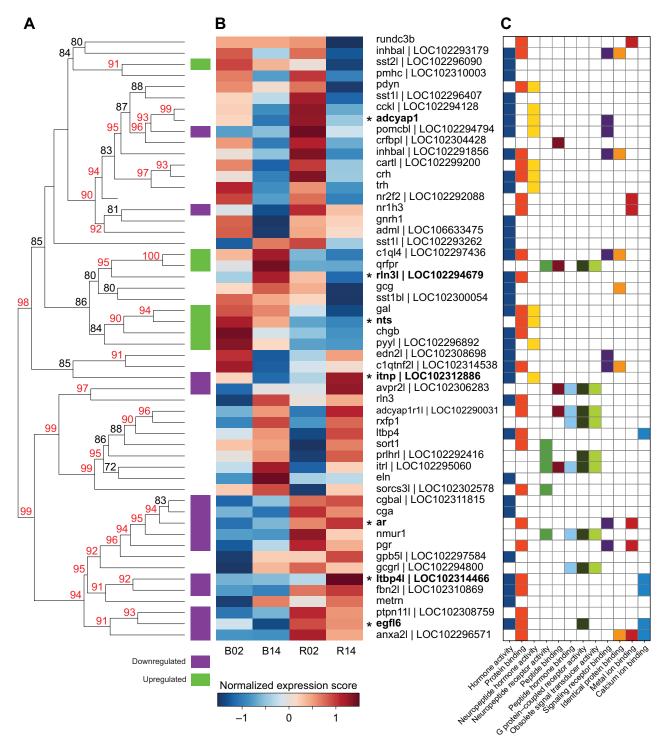


Fig. 3. Hierarchical clustering of gene expression patterns of neurohormones, neuropeptides and receptors selected from the list of genes trending toward differential expression based on their GO annotations. (A) Gene hierarchical clustering was performed using the average linkage method with a correlation metric distance. Bootstrap values were generated in the PVclust R package with red values highlighting nodes conserved in ≥90% of iterations. (B) Heatmap values represent mean variance stabilizing transformation (VST) expression of samples within groups scaled by row for visualization in the made4 R package. Gene abbreviations were manually assigned based on reference gene names for those that include *A. burtoni* accessions. Asterisks and bold indicate genes that are differentially expressed based on likelihood ratio test (LRT) analysis at FDR<0.1, while all other genes show near significance at a less stringent threshold with an unadjusted P<0.05. Genes marked with colored bars on the left are either upregulated (green) or downregulated (purple) in pooled mouthbrooding time points relative to pooled release time points based on DESeq2 binomial Wald results at an unadjusted P<0.05. (C) The top 12 molecular function GO terms, selected by the abundance of represented genes within this set, are displayed in a binary heatplot with columns clustered by the average linkage method and Euclidean distance. For BO2 and RO2 time points, n=5; for B14 and R14 time points, n=6.

and care may be analogous to the reduced expression seen in female stickleback following spawning (Kulczykowska and Kleszczyńska, 2014), associated with oocyte maturation and ovulation, or may simply reflect the low social engagement during this phase and suggest it is not a primary regulator of maternal mouthbrooding behavior in *A. burtoni*.

The neuropeptide *neurotensin* (*nts*) has emerged as a candidate for involvement in parental care, showing significantly elevated expression (FDR=4.1E-2) throughout the mouthbrooding stages, with a sharp decrease after release of fry (Figs 3 and 4). Expression of *nts* is positively and negatively associated with different forms of parental behavior in different taxa. In mice, injected *nts* inversely modulates aggressive behavior, potentially through interaction with its receptor *neurotensin receptor 1* (*ntsr1*; Fig. 4) (Gammie et al., 2009), and causes increased neuronal activity in several brain regions including the lateral septal nucleus (LS) and bed nucleus of the stria terminalis (BNST), which are involved in social behavior and anxiety (Clauss et al., 2019; Deng et al., 2019). Localized expression studies in mice suggest a complex relationship with

elevated *nts* expression in postpartum females in the medial preoptic area (MPOA) and lower expression in the LS (Driessen et al., 2014). Meanwhile, data from birds (domesticated Japanese quails) show higher *nts* expression in the female hypothalamus associated with parental behavior including reduced aggression towards juveniles (Lopes and de Bruijn, 2021).

The involvement of the *nts* pathway, beyond differential expression of the neuropeptide itself, is corroborated by our GSEA enrichment results that reveal the GO set 'G protein-coupled neurotensin receptor activity' to be increased in B14 compared with B02 fish (FDR=1.18E-2), suggesting receptor expression may be subject to feedback from early expression of the neuropeptide (Fig. 2B; Table S3). This GO set represented many genes of low expression level that failed to meet significance thresholds independently, though three of the 17 *nts* receptors or receptor-interacting ligands did reach an unadjusted threshold (*P*<0.05) (Fig. 4). Two of these genes, *sortilin 1/neurotensin receptor 3 (sort1)* and *sortilin-related VPS10 domain containing receptor 3-like (sorcs31)* follow a pattern similar to that of *nts* itself,

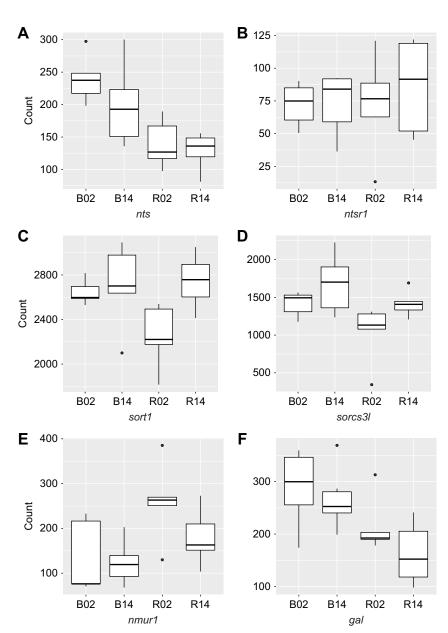


Fig. 4. Boxplots showing expression by time point for candidate genes involved in parental care. (A–E)
Expression of the candidate behavioral gene neurotensin (nts; A), and its receptors neurotensin receptor 1 (ntsr1; B), sortilin 1 (sort1; C), sortilin-related VPS10 domain containing receptor 3-like (sorcs3l; D) and neuromedin-U receptor 1 (nmur1; E). (F) Expression of correlated behavioral neurohormone gene galanin (gal). Expression is represented as DESeq2 normalized counts. Box plots indicate median (horizontal line inside boxes) values, upper and lower quartiles (box), 1.5× interquartile range (whiskers) and outliers (dots).

although their highest expression was in B14 instead of B02. Spatial co-expression of *nts* and *sort1* is also well documented in mammals, with both genes upregulated in the MPOA while being downregulated in the LS brain region of postpartum female mice relative to virgins (Driessen et al., 2014). These region-specific changes in mice further highlight the need for more targeted analysis in the future in A. burtoni in order to disentangle the interactive role of nts and its receptors in the regulation of maternal behavior. In our study, a third nts receptor, neuromedin U receptor 1 (nmur1) followed an inverted pattern relative to nts with its highest expression at release, R02. This gene's functional relationship to nts signaling is not well studied as its primary affinity is for the pleiotropic peptide neuromedin U (nmu), which has a brain expression profile that parallels that of nmur1, with highest expression in R02, suggesting they are co-expressed in the maternal brain (Martinez and O'Driscoll, 2015).

Among the differentially expressed neuropeptides, hormones and receptors shown in Fig. 4, hierarchical clustering revealed that expression of nts was most strongly correlated with gal, with both expressed at significantly higher levels in brooding time points relative to release time points (Fig. 4; Table S1). Studies of fish, mammals and amphibians have found gal to be associated with parental behavior (Butler et al., 2020; Fischer et al., 2019; Wu et al., 2014). Specific MPOA galanin neurons are activated during parental behavior in male and female mice, and optogenetic activation induces parental pup-grooming while genetic ablation suppresses parental behavior (Wu et al., 2014). In A. burtoni, galanin neuron activity is associated with maternal care in the POA and feeding in the arcuate nucleus (Butler et al., 2020). Correlated increased expression of gal and nts is also reported in POA of Dendrobates tinctorius males involved in tadpole transport (Fischer et al., 2019). That expression of *nts* and *gal* wanes in *A. burtoni* as maternal aggression increases, just prior to and following the release of fry (Renn et al., 2009), suggests a conserved role in recognition and protection of offspring rather than overt defense. Additional parallels with paternal care in frogs include the upregulation of myelin basic protein-like (mbpl) (unadjusted P=1.51E-2) and downregulation of progesterone receptor (pgr) (unadjusted P=8.83E-3) and prolactin receptor b (prlrb) (unadjusted P=8.80E-3) at the two mouthbrooding time points in A. burtoni and in parental D. tinctorius males, yet prolactin (prl) itself was not differentially expressed in either study (Renn et al., 2009; Fischer et al., 2019). Several genes associated with parental behavior in both studies show opposite regulation, including *neuroligin-3* (*nlgn3*) and secretogranin II (scg2), which are downregulated in D. tinctorius parental males and upregulated in A. burtoni mouthbrooding females, while androgen receptor (ar) shows the opposite pattern in each species. Assuming these normalized read counts correlate with translated peptide levels, the opposing patterns of differential expression between A. burtoni and D. tinctorius may reflect the species-specific demands of parental behavior or result from parental behavior being expressed in the opposite sex.

While inconsistent patterns of regulation between *D. tinctorius* and *A. burtoni* may result from the different techniques (brain punch versus whole brain) or taxa-specific parental strategies, the similarity in suites of genes at play is of note. In the frog POA, cocaine- and amphetamine-regulated transcript (cart) is upregulated in parental males while corticotropin-releasing hormone binding protein (crhbp) is downregulated and in the medial pallium (MP), corticotropin-releasing factor receptor 2 (crhr2) as well as cart are upregulated while pro-thyrotropin-releasing hormone and thyrotropin-releasing hormone (trh)

receptor are downregulated (Fischer et al., 2019). In *A. burtoni*, homologs of these ligands and receptors (*cartl, trh, crhbpl* and *crh*) are strongly correlated in expression with increased expression in B02 and R02 surrounding the transition from mouth-brooding care to defense of fry. In mice, *Crhbp* inhibits endogenous *Crh* to stimulate maternal aggression and deletion of the gene inhibits maternal aggression (Gammie et al., 2008), suggesting it may play a similar role in the *A. burtoni* maternal transition.

Fasting and metabolic regulation

Under normal conditions, the arcuate nucleus contains two specific populations of neurons, one producing orexigenic NPY and AgRP to stimulate feeding behavior, and the other producing POMC and CART to inhibit feeding, which are directly responsive to the peripheral signals of satiety – leptin (Cone, 2005) and ghrelin (Cowley et al., 2003) - and indicate energy reserves and body condition. However, in brooding females, the reduction in body mass (Grone et al., 2012; Renn et al., 2009) that should signal to increase orexigenic signals does not result in feeding behavior during the mouthbrooding time points. Among the many hormone/ neuropeptide genes related to feeding behavior, we found several to be among those trending toward differential expression across the reproductive time points, including relaxin-3-like (rln3l), gal, cartl, cholecystokinin-like (cckl), pro-opiomelanocortin B (pomcb) and peptide YY (pvv) (Fig. 3). While many of these showed large variance within time points, they appeared to be responsive to the resumption of feeding post-release.

The anorexigenic genes *pomcb* and *cart* showed highest expression after resumption of feeding, at R02, and were low during brooding, again reflecting energetic state. These genes are regulated by the peripheral signals leptin and insulin (Lau and Herzog, 2014; Valen et al., 2011). The orexigenic genes *gal* and *rln3l* clustered with *nts* (Fig. 3) and showed increased expression during mouthbrooding when females are not eating. As these genes are expected to promote food intake (Ganella et al., 2013; Volkoff and Peter, 2001), their upregulation during brooding suggests that the brains may be receiving the typical signals of hunger from the periphery, although the females suppress or do not respond to the signals with increased feeding. Alternatively, these genes may be playing a role directly promoting maternal mouthbrooding care.

While anorexigenic neuropeptide Y (npy) did not show differential expression across time points in the current study, a related peptide, pyy, was elevated in mouthbrooding stages (unadjusted P=0.01). In the digestive system, pyy expression increases with food intake to signal satiety (Karra et al., 2009); however, in the brain, its response to fasting is variable across species (Volkoff, 2016). In a study of the mouse enteroendocrine system, pyy was shown to be co-expressed and co-secreted with both nts and glucagon-like peptide-1 (glp1) (Grunddal et al., 2016). The genes pyy and glp1 were each found to work synergistically with nts in these mice to inhibit gastric emptying and decrease food intake. In A. burtoni maternal brains, pvy expression correlates with nts expression (Fig. 3) across samples through a clustering of candidate differentially expressed neurohormones and neuropeptides. Its role in this system and potential interaction with nts in the regulation of fasting behavior warrants further study.

In addition to the potential role of *nts* in parental behavior, this neuropeptide has a pleiotropic role and also can regulate fasting. In a number of studies in mice and rats, researchers have shown that increases in *nts* in the brain suppressed feeding behavior through interaction with its associated receptor, *ntsr1* (Kim et al., 2008; Schroeder and Leinninger, 2018). In *A. burtoni*, we saw the highest

expression of *nts* in mouthbrooding fish and a sizable drop in non-brooders but no change in low-expressed *ntsr1* (Fig. 4). This pattern is consistent with the hypothesis that *nts* plays a role in fasting behavior in mouthbrooding females by eliciting an anorectic effect despite unchanged expression of *ntsr1*. It is possible that mouthbrooding *A. burtoni* may actually be suppressing signals that would normally increase hunger and feeding behavior. Such suppression has been suggested by studies addressing the expression of feeding behavior-associated hormones, neuropeptides and their receptors in *A. burtoni* and *Oreochromis niloticus* brains (Das et al., 2019; Grone et al., 2012).

Many of our DE genes point to compensatory metabolic mechanisms to allow fish to withstand long periods of fasting. For example, the pyruvate dehydrogenase kinases pdk2 (FDR=9.71E-5) and pdk4 (FDR=0.024) show significantly higher expression in the brain in the two mouthbrooding time points relative to the two non-brooding time points. These genes are associated with food deprivation in fish, mammals and other taxa, and have been shown to increase during periods of starvation then decrease after resumption of feeding (Wu et al., 2000; Yang et al., 2019). This energy intensive pathway mediates the interaction of glycolysis and the tricarboxylic acid cycle (Sugden and Holness, 2003). The functional conservation of pyruvate dehydrogenase kinase activity across taxa may have been a necessary mechanism allowing for the evolution of mouthbrooding behavior in cichlids and other independent fish lineages as a result of the associated fasting behavior.

GSEA results also corroborate this change in mitochondrial function, with GO sets related to mitochondrial ATP production and electron transport showing upregulation in B02 fish relative to R14 fish (Fig. 2; Table S3). These GO terms signal a shift in energy metabolic processes at the start of a long period of prolonged fasting, and include gene sets such as 'ATP biosynthetic process', 'mitochondrial fusion', 'mitochondrial electron transport, NADH to ubiquinone', 'proton-transporting ATP synthase activity, rotational mechanism' and 'mitochondrial proton-transporting ATP synthase complex'. Metabolic downshifts are common among starving or hibernating animals and are often characterized by a reduction in mitochondrial function in liver and skeletal muscle tissue for energy conservation (Barger et al., 2003). That we see these changes anticipated in the periphery occurring in brain samples may reflect the high metabolic demand of neural tissue.

Hypoxic signaling and neuroprotection

Late-stage (post-hatching) mouthbrooding cichlids show reduced tolerance to hypoxia (*P. multicolor*: Corrie et al., 2008) possibly as a result of the costs of maternal behavior including buccal churning to aerate the eggs. However, we found a dramatic reduction in gene expression for several globin genes, which could also contribute to the observed sensitivity to hypoxia. Four genes reached, and one approached, a statistically significant reduction in expression at the late-brooding B14 and R02 time points (Fig. 5). These contribute to GO term enrichment (Table 2) and GSEA results (Fig. 2) for oxygen transport-related terms. These are among the most dramatic gene expression changes observed, likely reflecting consistent regulation across multiple brain regions. The reduction in globin expression may be a response to nutrient state as seen under starvation in seabream (Ntantali et al., 2020). The sustained expression could reflect 'carryover', the persistence of acquired neurogenomic states into subsequent stages (Burkhari et al., 2019). However, we also found hypoxia-inducible factor 1-alpha-like (FDR=0.12, unadjusted P=7.72E-4) elevated in both brooding time points

along with two target genes, *pdk4*, known to be also induced by starvation (Lee et al., 2012), and *round spermatid basic protein 1-like (rsbn1l)*, a less understood gene known to be expressed in testes as well as breast cancer tissue (Abu-Jamous et al., 2017). The correlated expression of *rsbn1l* and *pdk4* supports the hypothesis that mouthbrooding cichlids are responding to hypoxia-related signals possibly due to reduced globin gene expression.

The reduction in nutrient intake and apparent reduction in neural oxygen transport capacity confer a large metabolic cost that could be mitigated by neuroprotective pathways as suggested by GSEA (Figs 2 and 4). We saw varied and dynamic patterns among the neuroprotection-related genes, some being upregulated in late brooding and release such as two butyrophilin-related DE genes (butyrophilin subfamily 2 member A1-like and V-set domaincontaining T-cell activation inhibitor 1-like) that co-localize on a genome scaffold and are known immune system regulators that may have neuroprotective function in response to hypoxia, reduced nutrient intake or the action of other DE genes associated with foreign DNA/RNA (Abeler-Dörner et al., 2012). Conversely, other genes involved in neuroprotection were downregulated at late brooding time points and upregulated at early brooding time points. Downregulation of ECM/collagen-related DE genes including matrix metalloproteinases such as matrix metalloproteinase 9 (mmp9) could impact recovery via delayed promotion of neuroblast cell migration (Chen et al., 2009), while regulation of platelet-derived growth factor receptor-like (pdgfrl), similar to other PDGF proteins seen in cerebral hypoxia of stroke victims (Krupinski et al., 1997), could aid in neural regeneration in response to neuronal damage.

Nutrient and oxygen deficit impact the regulation of ion gradients and ion transport pathways in the brain that require energy and aerobic ATP synthesis. Both ion transport and cell junction genes are upregulated in whole-brain samples under starvation in seabream (Ntantali et al., 2020) and hypoxia-tolerant species are known to have evolved means to reduce ion leakage in the CNS (Boutilier, 2001), thus reducing energy demands in such conditions. Similarly, increased myelination mitigates the loss of action potentials under hypoxia (Waxman et al., 1990). These facts are consistent with the observed upregulation at B14 and R02 time points of cell adhesion molecule 4 (cadm4) (FDR=2.21E-14 and 3.2-fold change), which regulates myelination of the CNS (Elazar et al., 2019; Golan et al., 2013) and could contribute to the maintenance of action potentials under starvation and hypoxic conditions during mouthbrooding. These and other DE genes, including those coding for myosin, actin and troponin complex proteins, contribute both to molecular function GO terms, such as ion transport-related and action potential gradient-related GO sets. and biological process GO terms related to the structure and function of skeletal muscle and heart tissue (Fig. 2; Table S3), supporting the hypothesis of added demand to regulate ion gradients for neural function during mouthbrooding. Here again, the patterns of regulation among genes contributing to ion transport-related GO terms is highly variable as a result of the individual genes being negative versus positive regulators of the same or related processes.

In addition to oxygen transport and ion regulation, our GSEA analysis detected the inflammatory pathway as a large family of 16 GO sets under the umbrella term 'interleukin-1 beta production' (Table S3) that showed decreased expression at the early brooding B02 time point and increased expression at the late brooding B14 time point. While IL1B, a cytokine secreted by microglia in the brain in response to hypoxia or injury to the brain (Hewett et al., 2012), is not itself differentially expressed in our dataset, GSEA

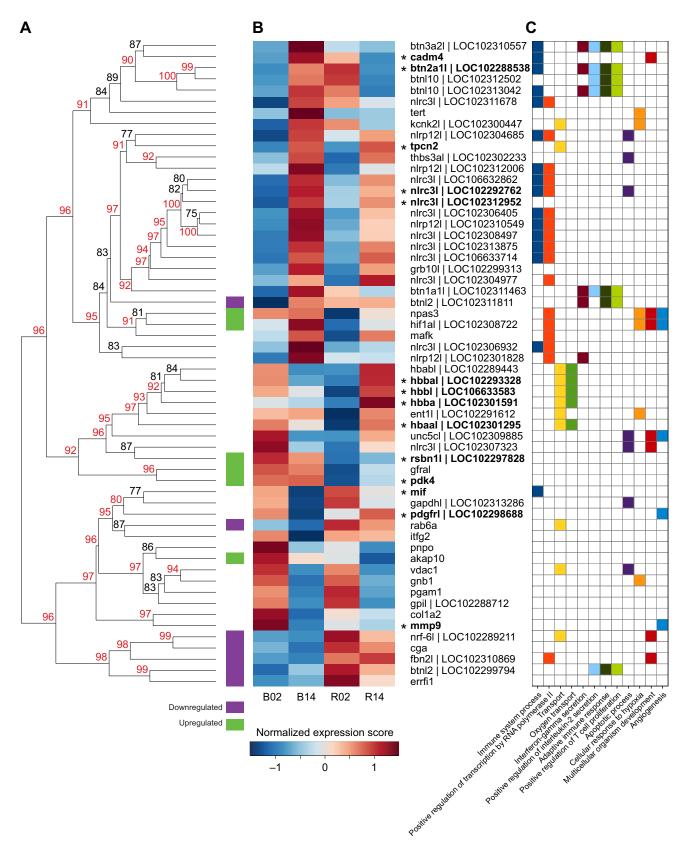


Fig. 5. See next page for legend.

identifies genes involved in IL1B suppression via (NF-κB) signaling (e.g. protein nlrc3-like, NACHT, LRR and PYD domains-containing protein 12-like) (Allen, 2014). The

upregulation of these pathways and static expression of cytokines may be adaptations that help avoid negative physiological consequences of prolonged inflammation associated with hypoxic

Fig. 5. Hierarchical clustering of gene expression patterns of candidate genes involved in the response to hypoxia and neuroprotection. Genes in this figure were selected from the list of genes trending toward differential expression for having functional GO annotations related to oxygen transport, neuroprotection, neuroinflammation or response to hypoxia, plus many genes from a list of hif1a-interacting genes from the Harmonizome database (https://maayanlab.cloud/Harmonizome/; accessed June 2020). (A) Gene hierarchical clustering was performed using the average linkage method with a correlation metric distance. Bootstrap values were generated in the PVclust R package, with red values highlighting nodes conserved in ≥90% of iterations. (B) Heatmap values represent mean VST expression of samples within groups scaled by row for visualization in the made4 R package. Gene abbreviations were manually assigned based on reference gene names for those that include A. burtoni accessions. Asterisks and bold indicate genes that are differentially expressed based on LRT analysis at FDR<0.1, while all other genes show near significance at an unadjusted P<0.05. Genes marked with colored bars on the left are either upregulated (green) or downregulated (purple) in pooled mouthbrooding time points relative to pooled release time points based on DESeq2 binomial Wald results at an unadjusted P<0.05. (C) The top 12 biological process GO terms, selected by the abundance of represented genes within this set, are displayed in a binary heatplot with columns clustered by the average linkage method and Euclidean distance. For B02 and R02 time points, n=5; for B14 and R14 time points, n=6.

conditions and starvation experienced during each repeated reproductive cycle (Martin and Król, 2017; Schäfer et al., 2021). To the best of our knowledge, these immune function pathways have not been implicated in other whole-brain studies of fish under starvation (e.g. Ntantali et al., 2020; Yang et al., 2019) but have been associated with paternal care in stickleback (Burkhari et al., 2019) suggesting their regulation may represent an adaptation in cichlids that reduce the costs of mouthbrooding.

Conclusions

Using female A. burtoni, we carried out a comprehensive analysis of whole-brain gene expression and how it changes across distinct brooding stages. The gene expression patterns reveal how the female brain may resolve trade-offs between metabolic demands and parental care. Similar conclusions can be drawn from lists of DE genes, GO analysis and the GSEA results that rely on differing levels of statistical significance for individual genes. In addition to identifying *itnp*, classically associated with parental care, our data support the involvement of nts, a gene that has more recently emerged as a candidate in the regulation of social phenotypes. These, and other identified DE genes, could be subjects for exciting functional analysis with gene editing techniques that are being made available in A. burtoni and other emerging model organisms (Juntti, 2019). Both GO enrichment and GSEA analysis further support the involvement of receptors and downstream activity of this pathway. Furthermore, correlation of gene expression ties these pathways to important neuropeptides involved in the regulation of feeding behavior and metabolism, highlighting the known tradeoffs between care and feeding (O'Rourke and Renn, 2015).

Interestingly, we found both orexigenic and anorexigenic genes that showed increased expression during mouthbrooding stages as well as anorexigenic genes that showed decreased expression during mouthbrooding, which underscores the complexity of feeding regulation under voluntary starvation. Future transcriptomic studies should include food-deprived individuals as an additional control group for comparison with mouthbrooding fish. Such a comparison would help elucidate the degree to which our observed increase in neuroprotective pathways and increased expression of negative regulators of inflammation represent evolved mechanisms to counter the hypoxic and metabolic demands of mouthbrooding.

Our results underscore that cichlid maternal behavior and fasting are potentially regulated by pleiotropic activity of novel peptides such as neurotensin as well as the expected classical neurohormones and neuropeptides. It is noteworthy that these hormonal signals were dwarfed by dramatic changes in oxygen transport capacity, neuroprotection and ionic regulation pathways in the brain that mirror metabolic shifts from animals regularly subjected to starvation or hypoxic conditions. Here, additional time points through these transitions could be used to further test the degree to which gene expression changes persist to reflect previous neurogenomic states versus priming the brain for future neurogenomic states (Burkhari et al., 2019). While the observed changes in neurohormones are logically brain specific, many of the other gene expression changes are likely not unique to brain tissue and rather reflect systemic responses to the stress of starvation and energy demands.

Acknowledgements

Samples and preliminary data were collected by Kimberly Engeln with fish husbandry support from several Reed lab undergraduates.

Competing interests

The authors declare no competing or financial interests.

Author contributions

Conceptualization: S.C.P.R.; Methodology: J.J.F.-H.; Formal analysis: J.J.F.-H., S.C.P.R.; Data curation: J.J.F.-H.; Writing - original draft: J.J.F.-H., S.C.P.R.; Writing - review & editing: J.J.F.-H., S.C.P.R.; Supervision: S.C.P.R.

Funding

This work was supported by grants from the National Institutes of Health (NIGMS R15-DK116224-01) and National Science Foundation (IOS 1456486) to S.C.P.R. Deposited in PMC for release after 12 months.

Data availability

Raw transcriptome data and the processed read count table are available in NCBI's GEO repository under accession number GSE192958. R-code, input files and supplementary (as well as additional WGCNA analysis) files can be found in GitHub repository: github.com/jfaberha/cichlid_transcriptome/tree/main/lab_stock_ analysis

References

Abeler-Dörner, L., Swamy, M., Williams, G., Hayday, A. C. and Bas, A. (2012). Butyrophilins: an emerging family of immune regulators. *Trends Immunol.* 33, 34-41. doi:10.1016/j.it.2011.09.007

Abu-Jamous, B., Buffa, F. M., Harris, A. L. and Nandi, A. K. (2017). In vitro downregulated hypoxia transcriptome is associated with poor prognosis in breast cancer. *Mol. Cancer* 16, 105. doi:10.1186/s12943-017-0673-0

Allen, I. C. (2014). Non-inflammasome forming NLRs in inflammation and tumorigenesis. *Front. Immunol.* **5**, 169. doi:10.3389/fimmu.2014.00169

Balaban, M., Moshiri, N., Mai, U., Jia, X. and Mirarab, S. (2019). TreeCluster: clustering biological sequences using phylogenetic trees. *PLoS One* 14, e0221068. doi:10.1371/journal.pone.0221068

Barger, J. L., Brand, M. D., Barnes, B. M. and Boyer, B. B. (2003). Tissue-specific depression of mitochondrial proton leak and substrate oxidation in hibernating arctic ground squirrels. Am. J. Physiol.-Regul. Integr. Comp. Physiol. 284, R1306-R1313. doi:10.1152/ajpregu.00579.2002

Barlow, G. W. (2000). The Cichlid Fishes: Nature's Grand Experiment in Evolution. Cambridge, MA: Perseus Publ.

Boutilier, **R. G.** (2001). Mechanisms of cell survival in hypoxia and hypothermia. *J. Exp. Biol.* **204**, 3171-3181. doi:10.1242/jeb.204.18.3171

Brawand, D., Wagner, C. E., Li, Y. I., Malinsky, M., Keller, I., Fan, S., Simakov, O., Ng, A. Y., Lim, Z. W., Bezault, E. et al. (2014). The genomic substrate for adaptive radiation in African cichlid fish. *Nature* **513**, 375-381. doi:10.1038/nature13726

Bukhari, S. A., Saul, M. C., James, N., Bensky, M. K., Stein, L. R., Trap, R. and Bell, A. M. (2019). Neurogenomic insights into paternal care and its relation to territorial aggression. *Nat. Commun.* **10**, 4437. doi:10.1038/s41467-019-12212-7

Butler, J. M., Maruska, K. P. (2021). Noise during mouthbrooding impairs maternal care behaviors and juvenile development and alters brain transcriptomes in the African cichlid fish Astatotilapia burtoni. Genes Brain Behav. 20, e12692. doi:10. 1111/gbb.12692

- Butler, J. M., Herath, E. M., Rimal, A., Whitlow, S. M. and Maruska, K. P. (2020).
 Galanin neuron activation in feeding, parental care, and infanticide in a mouthbrooding African cichlid fish. Horm. Behav. 126, 104870. doi:10.1016/j. yhbeh.2020.104870
- Chen, W., Hartman, R., Ayer, R., Marcantonio, S., Kamper, J., Tang, J. and Zhang, J. H. (2009). Matrix metalloproteinases inhibition provides neuroprotection against hypoxia-ischemia in the developing brain. *J. Neurochem.* 111, 726-736. doi:10.1111/j.1471-4159.2009.06362.x
- Clauss, J. A., Avery, S. N., Benningfield, M. M. and Blackford, J. U. (2019). Social anxiety is associated with BNST response to unpredictability. *Depress. Anxiety* 36, 666-675. doi:10.1002/da.22891
- Cone, R. D. (2005). Anatomy and regulation of the central melanocortin system. *Nat. Neurosci.* 8, 571-578. doi:10.1038/nn1455
- Corrie, L. W.-C., Chapman, L. J. and Reardon, E. E. (2008). Brood protection at a cost: mouth brooding under hypoxia in an African cichlid. *Environ. Biol. Fishes* 82, 41-49. doi:10.1007/s10641-007-9251-4
- Cowley, M. A., Smith, R. G., Diano, S., Tschöp, M., Pronchuk, N., Grove, K. L., Strasburger, C. J., Bidlingmaier, M., Esterman, M., Heiman, M. L. et al. (2003). The distribution and mechanism of action of Ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* 37, 649-661. doi:10.1016/S0896-6273(03)00063-1
- Crossin, G. T., Trathan, P. N., Phillips, R. A., Gorman, K. B., Dawson, A., Sakamoto, K. Q. and Williams, T. D. (2012). Corticosterone predicts foraging behavior and parental care in macaroni penguins. *Am. Nat.* 180, E31-E41. doi:10. 1086/666001
- Culhane, A. C., Thioulouse, J., Perriere, G. and Higgins, D. G. (2005). MADE4: an R package for multivariate analysis of gene expression data. *Bioinformatics* 21, 2789-2790. doi:10.1093/bioinformatics/bti394
- Das, K., Ogawa, S., Kitahashi, T. and Parhar, I. S. (2019). Expression of neuropeptide Y and gonadotropin-releasing hormone gene types in the brain of female Nile tilapia (*Oreochromis niloticus*) during mouthbrooding and food restriction. *Peptides* 112, 67-77. doi:10.1016/j.peptides.2018.10.009
- Deangelis, R., Gogola, J., Dodd, L. and Rhodes, J. S. (2017). Opposite effects of nonapeptide antagonists on paternal behavior in the teleost fish *Amphiprion* ocellaris. Horm. Behav. 90, 113-119. doi:10.1016/j.yhbeh.2017.02.013
- Deng, K., Yang, L., Xie, J., Tang, H., Wu, G.-S. and Luo, H.-R. (2019). Whole-brain mapping of projection from mice lateral septal nucleus. *Biol. Open* 8, bio043554. doi:10.1242/bio.043554
- De Vos, L. and Seegers, L. (1998). Seven new Orthochromis species (Teleostei: Cichlidae) from the Malagarasi, Luiche and Rugufu basins (Lake Tanganyika drainage), with notes on their reproductive biology. *Ichthyol. Explor Freshw.* 9, 371-420
- Dobin, A., Davis, C. A., Schlesinger, F., Drenkow, J., Zaleski, C., Jha, S., Batut, P., Chaisson, M. and Gingeras, T. R. (2013). STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* 29, 15-21. doi:10.1093/bioinformatics/bts635
- Driessen, T. M., Zhao, C., Whittlinger, A., Williams, H. and Gammie, S. C. (2014).
 Endogenous CNS expression of neurotensin and neurotensin receptors is altered during the postpartum period in outbred mice. *PLoS One* 9, e83098. doi:10.1371/journal.pone.0083098
- Elazar, N., Vainshtein, A., Golan, N., Vijayaragavan, B., Schaeren-Wiemers, N., Eshed-Eisenbach, Y. and Peles, E. (2019). Axoglial adhesion by Cadm4 regulates CNS myelination. *Neuron* 101, 224-231.e5. doi:10.1016/j.neuron.2018. 11.032
- Faber-Hammond, J. J., Coyle, K. P., Bacheller, S. K., Roberts, C. G., Mellies, J. L., Roberts, R. B. and Renn, S. C. P. (2019). The intestinal environment as an evolutionary adaptation to mouthbrooding in the Astatotilapia burtoni cichlid. FEMS Microbiol. Ecol. 95, fiz016. doi:10.1093/femsec/fiz016
- Fischer, E. K. and O'Connell, L. A. (2017). Modification of feeding circuits in the evolution of social behavior. *J. Exp. Biol.* **220**, 92-102. doi:10.1242/jeb.143859
- Fischer, E. K., Roland, A. B., Moskowitz, N. A., Tapia, E. E., Summers, K., Coloma, L. A. and O'connell, L. A. (2019). The neural basis of tadpole transport in poison frogs. *Proc. R. Soc. B Biol. Sci.* 286, 20191084. doi:10.1098/rspb.2019. 1084
- Fryer, G. and Iles, T. D. (1972). The Cichlid Fishes of the Great Lakes of Africa: Their Biology and Evolution. Edinburgh: Oliver and Boyd.
- Gammie, S. C., Seasholtz, A. F. and Stevenson, S. A. (2008). Deletion of corticotropin-releasing factor binding protein selectively impairs maternal, but not intermale aggression. *Neuroscience* 157, 502-512. doi:10.1016/j.neuroscience. 2008.09.026
- Gammie, S. C., D'anna, K. L., Gerstein, H. and Stevenson, S. A. (2009). Neurotensin inversely modulates maternal aggression. *Neuroscience* 158, 1215-1223. doi:10.1016/j.neuroscience.2008.11.045
- Ganella, D. E., Ma, S. and Gundlach, A. L. (2013). Relaxin-3/RXFP3 signaling and neuroendocrine function – a perspective on extrinsic hypothalamic control. *Front. Endocrinol.* 4, 128. doi:10.3389/fendo.2013.00128
- Golan, N., Kartvelishvily, E., Spiegel, I., Salomon, D., Sabanay, H., Rechav, K., Vainshtein, A., Frechter, S., Maik-Rachline, G., Eshed-Eisenbach, Y. et al. (2013). Genetic deletion of Cadm4 results in myelin abnormalities resembling charcot-marie-tooth neuropathy. *J. Neurosci.* 33, 10950-10961. doi:10.1523/JNEUROSCI.0571-13.2013

- Goodson, J. L. (2005). The vertebrate social behavior network: evolutionary themes and variations. *Horm. Behav.* **48**, 11-22. doi:10.1016/j.yhbeh.2005.02.
- Goodson, J. L. and Kingsbury, M. A. (2013). What's in a name? Considerations of homologies and nomenclature for vertebrate social behavior networks. *Horm. Behav.* 64, 103-112. doi:10.1016/j.yhbeh.2013.05.006
- Goodwin, N. B., Balshine-Earn, S. and Reynolds, J. D. (1998). Evolutionary transitions in parental care in cichlid fish. *Proc. R. Soc. Lond. B Biol. Sci.* 265, 2265-2272. doi:10.1098/rspb.1998.0569
- Grone, B. P., Carpenter, R. E., Lee, M., Maruska, K. P. and Fernald, R. D. (2012). Food deprivation explains effects of mouthbrooding on ovaries and steroid hormones, but not brain neuropeptide and receptor mRNAs, in an African cichlid fish. *Horm. Behav.* **62**, 18-26. doi:10.1016/j.yhbeh.2012.04.012
- Grunddal, K. V., Ratner, C. F., Svendsen, B., Sommer, F., Engelstoft, M. S., Madsen, A. N., Pedersen, J., Nøhr, M. K., Egerod, K. L., Nawrocki, A. R. et al. (2016). Neurotensin is coexpressed, coreleased, and acts together with GLP-1 and PYY in enteroendocrine control of metabolism. *Endocrinology* 157, 176-194. doi:10.1210/en.2015-1600
- Gu, Z., Eils, R. and Schlesner, M. (2016). Complex heatmaps reveal patterns and correlations in multidimensional genomic data. *Bioinformatics* 32, 2847-2849. doi:10.1093/bioinformatics/btw313
- Hewett, S. J., Jackman, N. A. and Claycomb, R. J. (2012). Interleukin-1β in central nervous system injury and repair. *Eur. J. Neurodegener. Dis.* 1, 195-211.
- Huerta-Cepas, J., Szklarczyk, D., Heller, D., Hernández-Plaza, A., Forslund, S. K., Cook, H., Mende, D. R., Letunic, I., Rattei, T., Jensen, L. J. et al. (2019). eggNOG 5.0: a hierarchical, functionally and phylogenetically annotated orthology resource based on 5090 organisms and 2502 viruses. Nucleic Acids Res. 47, D309-D314. doi:10.1093/nar/gky1085
- Iwao, H., Okuda, N. and Ito, S. (2004). Mate availability and somatic condition affect filial cannibalism in a paternal brooding goby. *Behaviour* 141, 279-296. doi:10. 1163/156853904322981851
- Juntti, S. (2019). The future of gene-guided neuroscience research in non-traditional model organisms. *Brain Behav. Evol.* 93, 108-121. doi:10.1159/000500072
- Karra, E., Chandarana, K. and Batterham, R. L. (2009). The role of peptide YY in appetite regulation and obesity: peptide YY in appetite regulation and obesity. J. Physiol. 587, 19-25. doi:10.1113/jphysiol.2008.164269
- Kasumyan, A. (2004). The vestibular system and sense of equilibrium in fish. *J. lchthyol.* **44**, S224.
- Keverne, E. B. and Kendrick, K. M. (1992). Oxytocin Facilitation of Maternal Behavior in Sheepa. *Ann. N. Y. Acad. Sci.* **652**, 83-101. doi:10.1111/j.1749-6632. 1992 tb34348 x
- Kim, E. R., Leckstrom, A. and Mizuno, T. M. (2008). Impaired anorectic effect of leptin in neurotensin receptor 1-deficient mice. *Behav. Brain Res.* 194, 66-71. doi:10.1016/j.bbr.2008.06.024
- Krupinski, J., İssa, R., Bujny, T., Slevin, M., Kumar, P., Kumar, S. and Kaluza, J. (1997). A putative role for platelet-derived growth factor in angiogenesis and neuroprotection after ischemic stroke in humans. *Stroke* 28, 564-573. doi:10. 1161/01.STR.28.3.564
- Kulczykowska, E. and Kleszczyńska, A. (2014). Brain arginine vasotocin and isotocin in breeding female three-spined sticklebacks (Gasterosteus aculeatus): the presence of male and egg deposition. Gen. Comp. Endocrinol. 204, 8-12. doi:10.1016/j.ygcen.2014.04.039
- **Kuwamura, T.** (1997). The evolution of parental care and mating systems among Tanganyikan cichlids. In *Fish Communities in Lake Tanganyika*, pp. 57-86. Kyoto: Kyoto University Press.
- Langfelder, P. and Horvath, S. (2008). WGCNA: an R package for weighted correlation network analysis. BMC Bioinform. 9, 559. doi:10.1186/1471-2105-9-550
- Lau, J. and Herzog, H. (2014). CART in the regulation of appetite and energy homeostasis. Front. Neurosci. 8, 313. doi:10.3389/fnins.2014.00313
- Lee, J. H., Kim, E.-J., Kim, D.-K., Lee, J.-M., Park, S. B., Lee, I.-K., Harris, R. A., Lee, M.-O. and Choi, H.-S. (2012). Hypoxia induces PDK4 gene expression through induction of the orphan nuclear receptor ERRγ. PLoS One 7, e46324. doi:10.1371/journal.pone.0046324
- Lopes, P. C. and De Bruijn, R. (2021). Neurotranscriptomic changes associated with chick-directed parental care in adult non-reproductive Japanese quail. Sci. Rep. 11, 15481. doi:10.1038/s41598-021-94927-6
- Love, M. I., Huber, W. and Anders, S. (2014). Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biol. 15, 550. doi:10. 1186/s13059-014-0550-8
- Maere, S., Heymans, K. and Kuiper, M. (2005). BiNGO: a Cytoscape plugin to assess overrepresentation of Gene Ontology categories in Biological Networks. *Bioinformatics* 21, 3448-3449. doi:10.1093/bioinformatics/bti551
- Martin, S. A. M. and Król, E. (2017). Nutrigenomics and immune function in fish: new insights from omics technologies. *Dev. Comp. Immunol.* 75, 86-98. doi:10. 1016/i.dci.2017.02.024
- Martinez, V. G. and O'driscoll, L. (2015). Neuromedin U: a multifunctional neuropeptide with pleiotropic roles. Clin. Chem. 61, 471-482. doi:10.1373/ clinchem.2014.231753

- Maruska, K. P. and Fernald, R. D. (2013). Social regulation of male reproductive plasticity in an African cichlid fish. *Integr. Comp. Biol.* 53, 938-950. doi:10.1093/ icb/ict017
- Maruska, K. P., Butler, J. M., Field, K. E., Forester, C. and Augustus, A. (2020a).
 Neural activation patterns associated with maternal mouthbrooding and energetic state in an African Cichlid fish. *Neuroscience* 446, 199-212. doi:10.1016/j. neuroscience.2020.07.025
- Maruska, K. P., Butler, J. M., Anselmo, C. and Tandukar, G. (2020b). Distribution of aromatase in the brain of the African cichlid fish *Astatotilapia burtoni*: aromatase expression, but not estrogen receptors, varies with female reproductive—state. *J. Comp. Neurol.* **528**, 2499-2522. doi:10.1002/cne.24908
- Mrowka, W. (1984). Brood care motivation and hunger in the mouthbrooding cichlid Pseudocrenilabrusm multicolor. Behav. Process. 9, 181-190. doi:10.1016/0376-6357(84)90039-1
- Numan, M. and Woodside, B. (2010). Maternity: neural mechanisms, motivational processes, and physiological adaptations. *Behav. Neurosci.* 124, 715-741. doi:10.1037/a0021548
- Ntantali, O., Malandrakis, E. E., Abbink, W., Golomazou, E., Karapanagiotidis, I. T., Miliou, H. and Panagiotaki, P. (2020). Whole brain transcriptomics of intermittently fed individuals of the marine teleost Sparus aurata. Comp. Biochem. Physiol. D Genomics Proteomics. 36, 100737. doi:10.1016/j.cbd.2020.100737
- O'Connell, L. A. and Hofmann, H. A. (2011). The vertebrate mesolimbic reward system and social behavior network: a comparative synthesis. *J. Comp. Neurol.* 519, 3599-3639. doi:10.1002/cne.22735
- O'Connell, L. A., Matthews, B. J. and Hofmann, H. A. (2012). Isotocin regulates paternal care in a monogamous cichlid fish. *Horm. Behav.* **61**, 725-733. doi:10. 1016/j.yhbeh.2012.03.009
- Olazábal, D. E. and Young, L. J. (2006). Oxytocin receptors in the nucleus accumbens facilitate "spontaneous" maternal behavior in adult female prairie voles. *Neuroscience* **141**, 559-568. doi:10.1016/j.neuroscience.2006.04.017
- Olszewski, P. K., Klockars, A., Schiöth, H. B. and Levine, A. S. (2010). Oxytocin as feeding inhibitor: maintaining homeostasis in consummatory behavior. *Pharmacol. Biochem. Behav.* **97**, 47-54. doi:10.1016/j.pbb.2010.05.026
- O'Rourke, C. F. and Renn, S. C. (2015). Integrating adaptive trade-offs between parental care and feeding regulation. *Curr. Opin. Behav. Sci.* **6**, 160-167. doi:10. 1016/j.cobeha.2015.11.010
- Osborne, P. G. and Hashimoto, M. (2008). Mammalian cerebral metabolism and amino acid neurotransmission during hibernation. *J. Neurochem.* **106**, 1888-1899. doi:10.1111/j.1471-4159.2008.05543.x
- Pedersen, C. A., Ascher, J. A., Monroe, Y. L. and Prange, A. J. (1982). Oxytocin induces maternal behavior in virgin female rats. *Science* 216, 648-650. doi:10. 1126/science.7071605
- Pertea, M., Pertea, G. M., Antonescu, C. M., Chang, T.-C., Mendell, J. T. and Salzberg, S. L. (2015). StringTie enables improved reconstruction of a transcriptome from RNA-seq reads. *Nat. Biotechnol.* 33, 290-295. doi:10.1038/ nbt.3122
- Renn, S. C. P., Carleton, J. B., Magee, H., Nguyen, M. L. T. and Tanner, A. C. W. (2009). Maternal care and altered social phenotype in a recently collected stock of Astatotilapia burtoni cichlid fish. *Integr. Comp. Biol.* 49, 660-673. doi:10.1093/icb/icp085
- Rosa Salva, O., Sovrano, V. A. and Vallortigara, G. (2014). What can fish brains tell us about visual perception? *Front. Neural Circuits* 8, 119. doi:10.3389/fncir. 2014.00119
- Rüber, L., Britz, R., Tan, H. H., Ng, P. K. L. and Zardoya, R. (2004). Evolution of mouthbrooding and life-history correlates in the fighting fish genus Betta. *Evolution* **58**, 799-813. doi:10.1111/j.0014-3820.2004.tb00413.x
- Salzburger, W., Mack, T., Verheyen, E. and Meyer, A. (2005). Out of Tanganyika: genesis, explosive speciation, key-innovations and phylogeography of the haplochromine cichlid fishes. BMC Evol. Biol. 5, 17. doi:10.1186/1471-2148-5-17
- Schäfer, N., Matoušek, J., Rebl, A., Stejskal, V., Brunner, R. M., Goldammer, T., Verleih, M. and Korytář, T. (2021). Effects of chronic hypoxia on the immune status of Pikeperch (Sander lucioperca Linnaeus, 1758). *Biology* 10, 649. doi:10. 3390/biology10070649

- Schneider, J. E. (2004). Energy balance and reproduction. *Physiol. Behav.* 81, 289-317. doi:10.1016/j.physbeh.2004.02.007
- Schroeder, L. E. and Leinninger, G. M. (2018). Role of central neurotensin in regulating feeding: implications for the development and treatment of body weight disorders. *Biochim. Biophys. Acta BBA Mol. Basis Dis.* 1864, 900-916. doi:10. 1016/j.bbadis.2017.12.036
- Secor, S. M. and Carey, H. V. (2016). Integrative physiology of fasting. Compr Physiol. 6, 773-825. doi:10.1002/cphy.c150013
- Shahjahan, M., Kitahashi, T. and Parhar, I. (2014). Central pathways integrating metabolism and reproduction in teleosts. *Front. Endocrinol.* 5, 36. doi:10.3389/ fendo.2014.00036
- Shannon, P., Markiel, A., Ozier, O., Baliga, N. S., Wang, J. T., Ramage, D., Amin, N., Schwikowski, B. and Ideker, T. (2003). Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res.* 13, 2498-2504. doi:10.1101/gr.1239303
- Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., Paulovich, A., Pomeroy, S. L., Golub, T. R., Lander, E. S. et al. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc. Natl. Acad. Sci. USA* 102, 15545-15550. doi:10.1073/pnas.0506580102
- Sugden, M. C. and Holness, M. J. (2003). Recent advances in mechanisms regulating glucose oxidation at the level of the pyruvate dehydrogenase complex by PDKs. Am. J. Physiol. Endocrinol. Metab. 284, E855-E862. doi:10.1152/ ajpendo.00526.2002
- Suzuki, R. and Shimodaira, H. (2006). Pvclust: an R package for assessing the uncertainty in hierarchical clustering. *Bioinformatics* 22, 1540-1542. doi:10.1093/ bioinformatics/btl117
- Taborsky, B. and Foerster, K. (2004). Female mouthbrooders adjust incubation duration to perceived risk of predation. *Anim. Behav.* 68, 1275-1281. doi:10.1016/ j.anbehav.2004.03.005
- The UniProt Consortium (2019). UniProt: a worldwide hub of protein knowledge. Nucleic Acids Res. 47, D506-D515. doi:10.1093/nar/gky1049
- Valen, R., Jordal, A.-E. O., Murashita, K. and Rønnestad, I. (2011). Postprandial effects on appetite-related neuropeptide expression in the brain of Atlantic salmon, Salmo salar. Gen. Comp. Endocrinol. 171, 359-366. doi:10.1016/j.ygcen. 2011.02.027
- Volkoff, H. (2016). The neuroendocrine regulation of food intake in fish: a review of current knowledge. Front. Neurosci. 10, 540. doi:10.3389/fnins.2016.00540
- Volkoff, H. and Peter, R. E. (2001). Interactions between orexin A, NPY and galanin in the control of food intake of the goldfish, *Carassius auratus*. Regul. Pept. 101, 59-72. doi:10.1016/S0167-0115(01)00261-0
- Ward, J. H. (1963). Hierarchical grouping to optimize an objective function. J. Am. Stat. Assoc. 58, 236-244. doi:10.1080/01621459.1963.10500845
- Waxman, S. G., Davis, P. K., Black, J. A. and Ransom, B. R. (1990). Anoxic injury of mammalian central white matter: decreased susceptibility in myelin-deficient optic nerve. Ann. Neurol. 28, 335-340. doi:10.1002/ana.410280306
- Wu, P., Blair, P. V., Sato, J., Jaskiewicz, J., Popov, K. M. and Harris, R. A. (2000). Starvation increases the amount of pyruvate dehydrogenase kinase in several mammalian tissues. *Arch. Biochem. Biophys.* 381, 1-7. doi:10.1006/abbi.2000. 1946
- Wu, Z., Autry, A. E., Bergan, J. F., Watabe-Uchida, M. and Dulac, C. G. (2014).
 Galanin neurons in the medial preoptic area govern parental behaviour. *Nature* 509, 325-330. doi:10.1038/nature13307
- Yanagisawa, Y., Ochi, H. and Rossiter, A. (1996). Intra-buccal feeding of young in an undescribed Tanganyikan cichlid *Microdontochromis* sp. *Environ. Biol. Fishes* 47, 191-201. doi:10.1007/BF00005042
- Yang, Y., Zhou, H., Shu, H., Zhong, D., Zhang, M. and Xia, J. H. (2019). Effect of fasting and subsequent refeeding on the transcriptional profiles of brain in juvenile Spinibarbus hollandi. PLoS One 14, e0214589. doi:10.1371/journal.pone. 0214589
- Yin, Q., Ge, H., Liao, C.-C., Liu, D., Zhang, S. and Pan, Y.-H. (2016). Antioxidant defenses in the brains of bats during hibernation. *PLoS One* 11, e0152135. doi:10. 1371/journal.pone.0152135