

## RESEARCH ARTICLE

# A subset of neurons controls the permeability of the peritrophic matrix and midgut structure in *Drosophila* adults

Hiroyuki Kenmoku<sup>1</sup>, Hiroki Ishikawa<sup>1,2</sup>, Manabu Ote<sup>1,3</sup>, Takayuki Kuraishi<sup>1,4,5,6</sup> and Shoichiro Kurata<sup>1,\*</sup>

## ABSTRACT

The metazoan gut performs multiple physiological functions, including digestion and absorption of nutrients, and also serves as a physical and chemical barrier against ingested pathogens and abrasive particles. Maintenance of these functions and structures is partly controlled by the nervous system, yet the precise roles and mechanisms of the neural control of gut integrity remain to be clarified in *Drosophila*. Here, we screened for GAL4 enhancer-trap strains and labeled a specific subsets of neurons, using Kir2.1 to inhibit their activity. We identified an NP3253 line that is susceptible to oral infection by Gram-negative bacteria. The subset of neurons driven by the NP3253 line includes some of the enteric neurons innervating the anterior midgut, and these flies have a disorganized proventricular structure with high permeability of the peritrophic matrix and epithelial barrier. The findings of the present study indicate that neural control is crucial for maintaining the barrier function of the gut, and provide a route for genetic dissection of the complex brain–gut axis in adults of the model organism *Drosophila*.

**KEY WORDS:** Peritrophic matrix, *Drosophila*, Enteric neurons

## INTRODUCTION

Maintenance of the proper structure and function of the gastrointestinal tract is central to host homeostasis in metazoan animals. Aside from its main role in digestion and nutrient absorption, the gut must also protect an animal from harmful substances and microorganisms, leading to a strong immune system and robust physical/structural barriers against invaders (Sansonet, 2004). The intestinal tract senses external cues, such as a nutrient availability, via the enteric endocrine or nervous system and sends systemic signals through hormonal or neuronal means to change both metabolism and behavior (Furness and Costa, 1987). These functions of the intestinal tract are also consistent for most insects, including the model organism *Drosophila melanogaster* (Kuraishi et al., 2013; Lemaître and Miguel-Aliaga, 2013).

Complex and highly organized tissue structures ensure the realization of these important tasks of the gut. Compartmentalization – the sequential organization of regions that vary histologically and

functionally – is an important feature of the intestinal tract (Karasov et al., 2011). In *Drosophila* adults, the gut is divided into three distinct domains based on developmental origin: the foregut, midgut and hindgut. The midgut, which is the main region responsible for intestinal functions, comprises a single layer of epithelium, surrounded by visceral muscles, nerves and tracheae, and is subdivided into six major anatomic regions with distinct functions (Buchon et al., 2013b).

The peritrophic matrix and septate junctions between epithelial cells have a central role as a physical barrier against external invaders (Hegedus et al., 2009; Tepass et al., 2001). The peritrophic matrix is an acellular structure that forms a layer of chitin polymers and glycoproteins, such as peritrophins, lining the insect midgut lumen (Lehane, 1997). The peritrophic matrix seems to be formed by either the midgut epithelium (type I) or the proventriculus (type II), which is a specialized structure located at the foregut/midgut junction that regulates passage of food to the midgut. In type I peritrophic matrix, delamination of successive concentric lamellae occurs along the length of the midgut. Diptera such as *Drosophila* have a type II peritrophic matrix that is continuously produced by specific cells of the proventriculus (King, 1988). The protective role of the peritrophic matrix against abrasive food particles and pathogens, as well as in sequestering ingested toxins, has been studied in many insects (Edwards and Jacobs-Lorena, 2000; Wang and Granados, 2000). In *Drosophila* adults, mutation of the *drosocrystallin* (*dcy*) gene, which encodes a structural component of the peritrophic matrix, results in its reduced thickness and higher permeability (Kuraishi et al., 2011). The *dcy* mutant flies show greater susceptibility to ingested entomopathogenic bacteria or pore-forming toxins. Septate junctions are functionally related to mammalian tight junctions and participate in epithelial barrier function, protecting the fly from oral infection by pathogenic bacteria. Bonnay et al. demonstrated that the *big bang* gene (*bbg*) encodes a PDZ domain-containing protein that is expressed in septate junctions (Bonnay et al., 2013). A mutation in *bbg* results in the loosening of septate junctions, and is associated with acute susceptibility to invasive enteric pathogens such as *Pseudomonas aeruginosa* and *Serratia marcescens*. The compartmentalization, peritrophic matrix and septate junctions of the gut are maintained throughout adult life by rapid turnover of the epithelium within 1 to 2 weeks under steady-state conditions (Buchon et al., 2013a,b). However, the cellular and molecular processes required to maintain these cellular and acellular structures of the intestinal tract are poorly understood.

A major function of the stomatogastric nervous system is to control peristalsis of the muscles surrounding the intestinal tract (Huizinga and Lammers, 2009) and to sense external conditions to regulate metabolism. A recent study in *Drosophila* adults revealed that enteric neurons also govern fluid homeostasis and sex peptide-induced changes in intestinal physiology, indicating an indispensable role for the brain–gut axis in maintaining host homeostasis (Cognigni et al., 2011; Talsma et al., 2012). We hypothesized that the enteric nervous system also has a role in maintaining the structural integrity of the gut, which is important for its barrier function. In this study, we identified

<sup>1</sup>Department of Molecular Genetics, Graduate School of Pharmaceutical Sciences, Tohoku University, Sendai 980-8578, Japan. <sup>2</sup>Immune Signal Unit, Okinawa Institute of Science and Technology Graduate University, Okinawa, 904-0495, Japan. <sup>3</sup>Division of Neurogenetics, Graduate School of Life Sciences, Tohoku University, Sendai 980-8577, Japan. <sup>4</sup>Department of Microbiology and Immunology, Keio University School of Medicine, Tokyo 160-8582, Japan. <sup>5</sup>Faculty of Pharmacy, Institute of Medical, Pharmaceutical and Health Sciences, Kanazawa University, Ishikawa 920-1192, Japan. <sup>6</sup>PRESTO, Japan Science and Technology Agency, Tokyo 102-0076, Japan.

\*Author for correspondence (kurata@mail.pharm.tohoku.ac.jp)

© S.K., 0000-0002-0301-872X

a subset of neurons required for maintaining gut impermeability against enteric pathogens, providing evidence for the neural control of gut integrity in *Drosophila* adults.

## MATERIALS AND METHODS

### Fly stocks

Oregon R flies were used as wild-type flies. The GAL4 lines screened in this study (Table S1) were obtained from the Bloomington Stock Center (Indiana University, Bloomington, IN, USA), *Drosophila* Genetic Resource Center (Kyoto Institute of Technology, Japan) and *384-GAL4*, *Bx-GAL4*, *elav-GAL4*, *tubP-GAL80<sup>ts</sup>*, *L<sup>1</sup>/CyO*; *UAS-DenMark*, *syt.eGFP* and *UAS-mCD8::GFP/CyO* were from Bloomington Stock Center. *UAS-dTrpA1* was a gift from P. Garrity (Hamada et al., 2008). *elav-GAL80* was a gift from Y. Jan (Yang et al., 2009). *UAS-lacZ* (Bloomington *Drosophila* Stock Center) or *w<sup>1118</sup>* was used as a control. *Drosophila* stocks and crosses were maintained at 18°C or 25°C in tubes containing standard cornmeal-agar medium. To inhibit or activate neural activity, 5- to 9-day-old flies of *NP3253-GAL4/tubP-GAL80<sup>ts</sup>*; *UAS-Kir2.1-EGFP* (Baines et al., 2001)/+ (*NP3253>Kir2.1* flies) or *NP3253-GAL4/UAS-dTrpA1* were maintained at 30°C for 2 days prior to use in all experiments. The NP3253 line was subjected to standard mitotic recombination over *yw* chromosomes to eliminate possible second-site mutations.

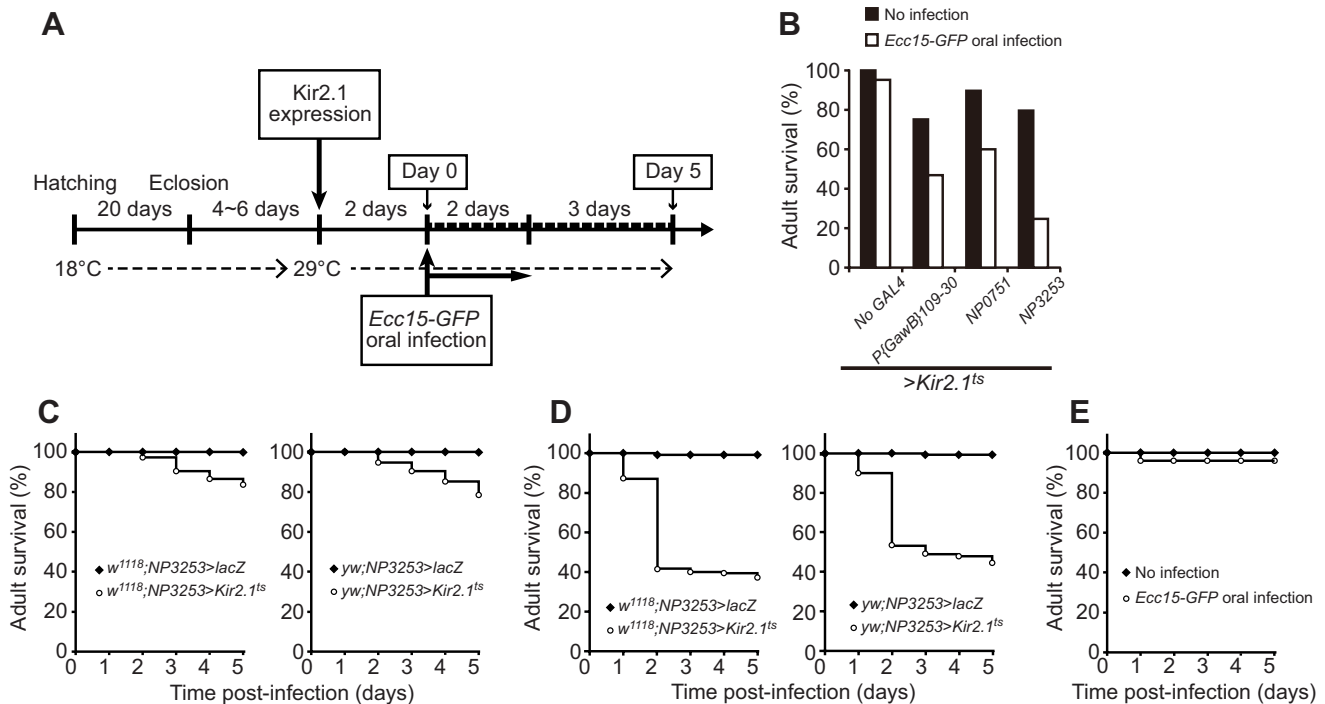
### Microbial infection

The *Ecc15-GFP* strain was described previously (Basset et al., 2000) and was grown in Luria Bertani broth for all experiments. Flies were grown at 29–30°C and allowed to reach the stationary phase. Cells

were then concentrated at OD<sub>600</sub>=200 with 2.5% sucrose solution. For oral infection, flies were starved for 2 h at 30°C and then placed in a fly vial with food solution. The food solution was made by mixing a pellet of bacteria, added to a filter disk that completely covered the surface of standard fly medium. Flies were maintained at 30°C and survival was monitored at different time points.

### Immunohistochemistry

*Drosophila* adults were dissected into cold phosphate-buffered saline (PBS) and the guts or brains were immediately fixed with 4% paraformaldehyde in PBS for 30 min at room temperature. The samples were rinsed in 0.5% Triton X-100 in PBS and then incubated with primary antibodies: mouse anti-GFP (1:500, Invitrogen, cat. no. A-6455), rabbit anti-RFP (1:500, Clontech, cat. no. 632496), rabbit anti-PH3 (1:500, Cell Signaling, cat. no. 9701), mouse anti-Discs-large 4F3 (1:100, Developmental Studies Hybridoma Bank), Alexa Fluor 647-conjugated goat anti-HRP antibody (1:500, Jackson ImmunoResearch, cat. no. 123-605-021) in 0.5% Triton X-100 in PBS at 4°C overnight. The samples were washed twice with 0.2% Triton X-100 in PBS and primary antibodies were labeled with Alexa Fluor 488-, 546- or 647-coupled secondary antibodies (Invitrogen). Actin filaments were stained with Rhodamine-Phalloidin (1:100, Sigma) and nuclei were stained with 4',6-diamidino-2-phenylindole (DAPI; 3 µg ml<sup>-1</sup>, Dojindo, Japan). The samples were then washed with 0.2% Triton X-100 in PBS, incubated with 50% glycerol (Wako, Japan) in PBS, and mounted in 80% glycerol in PBS or in Vectashield. For anti-PH3 antibody staining, the samples were fixed with 3.7% formaldehyde in PBS, permeabilized with 99.5% pre-chilled ethanol at -30°C for 5 min. The samples were visualized



**Fig. 1. NP3253>Kir2.1 flies are susceptible to oral infection with Ecc15-GFP.** (A) Timetable for neuronal inhibition by Kir2.1. Kir2.1 was expressed with temperature-sensitive GAL80, and the expressed flies were raised at 18°C until adulthood. They were then moved to 29°C and kept for 2 days before oral infection with *Ecc15-GFP*. The flies were maintained in vials with *Ecc15-GFP* for 2 days, and then moved to standard medium. (B) Survival analysis of Kir2.1-expressing flies by several GAL4 driver lines upon oral infection with *Ecc15-GFP*. Graph shows the survival rate of ~30 flies 3 days after infection. (C) Survival analysis of NP3253>lacZ or NP3253>Kir2.1 flies upon sucrose feeding. NP3253 is *w<sup>1118</sup>* background (left) or *yw* background (right). (D) Survival analysis of NP3253>lacZ or NP3253>Kir2.1 flies orally infected with *Ecc15-GFP*. NP3253 is *w<sup>1118</sup>* background (left) or *yw* background (right);  $P < 0.0001$  (log-rank test, left and right, comparing NP3253>lacZ with NP3253>Kir2.1). (E) Survival analysis of NP3253>dTrpA1 flies orally infected with *Ecc15-GFP*. No infection indicates sucrose feeding after starvation. Each survival curve corresponds to at least 2 independent experiments of 3 tubes of 30 flies each.

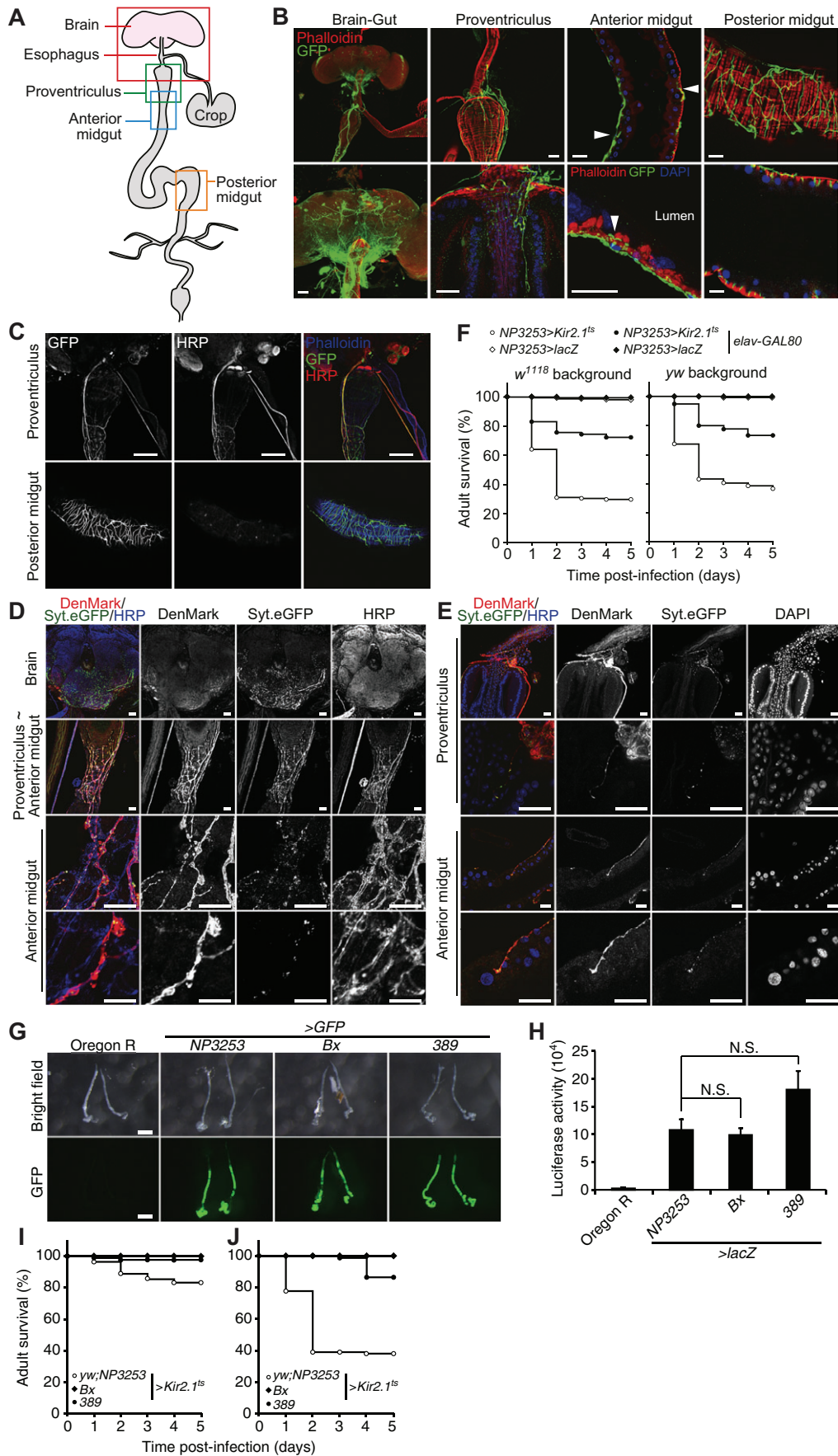


Fig. 2. See next page for legend.



**Fig. 2. Some NP3253-positive cells are neurons responsible for the survival phenotype.** (A) A schematic representation of the adult midgut. (B) Fluorescent confocal imaging of *NP3253>mCD8::GFP* flies. Green indicates NP3253-positive cells (anti-GFP). Blue indicates nuclei (DAPI). Red indicates visceral muscles (Phalloidin). Arrowheads indicate NP3253-positive cells that appear to innervate epithelial cells. Scale bars, 20  $\mu$ m. (C) Fluorescent confocal imaging of the proventriculus or posterior midgut of *NP3253>mCD8::GFP* flies. Green indicates NP3253-positive cells (anti-GFP). Blue indicates visceral muscles (Phalloidin). Red indicates neuronal marker (anti-HRP). Scale bars, 50  $\mu$ m. (D) Characterization of NP3253-positive cells by neuronal markers. Fluorescent confocal imaging of *NP3253>DenMark, syt.eGFP* flies. Green indicates synaptic vesicles (anti-GFP). Blue indicates neurons (anti-HRP). Red indicates DenMark (anti-RFP). Scale bars, 20  $\mu$ m (top 2 rows) and 10  $\mu$ m (bottom 2 rows). (E) Fluorescent confocal imaging of *NP3253>DenMark, syt.eGFP* flies. Green indicates synaptic vesicle (anti-GFP). Blue indicates nuclei (DAPI). Red indicates DenMark (anti-RFP). Scale bars, 20  $\mu$ m. (F) Survival analysis of flies orally infected with *Ecc15-GFP*. *lacZ* or *Kir2.1* expression is driven by NP3253-GAL4, together with (filled symbols) or without (open symbols) *elav-GAL80*. NP3253 is *w<sup>1118</sup>* background (left) or *yw* background (right).  $P < 0.0001$  (log-rank test, left and right, comparing *NP3253>Kir2.1* with *elav-GAL80; NP3253>Kir2.1*). Each survival curve corresponds to at least 2 independent experiments of 3 tubes of 30 flies each. (G) Survival analysis with salivary gland GAL4 drivers. Whole salivary gland imaging of flies expressing GFP by several GAL4 drivers. Salivary glands were observed under a light microscope (top) or fluorescence microscope (bottom). Green indicates GFP signal. Scale bars, 200  $\mu$ m. (H) Measurement of  $\beta$ -gal protein activity by  $\beta$ -glo assay. Five flies were examined and the graph shows a representative result of two independent experiments (N.S., not significant). (I) Survival analysis of *Kir2.1*-expressing flies by Bx-GAL4 or 389-GAL4 lines without infection. (J) Survival analysis of *Kir2.1*-expressing flies by Bx-GAL4 and 389-GAL4 lines upon oral infection with *Ecc15-GFP*. No infection indicates sucrose feeding after starvation. Each survival curve corresponds to at least 2 independent experiments of 3 tubes of 30 flies each.  $P < 0.01$  (NP3253 versus Bx, and NP3253 versus 389), log-rank test.

with a Leica TCS-SPE confocal microscope, and images were reconstructed using Photoshop (Adobe) and ImageJ (NIH).

PH3-positive cells in the whole midgut of 10 to 12 female flies were counted under a confocal microscope. Confocal images that showed the maximum measured area were calculated by ImageJ in the luminal region of the proventriculus or in the anterior midgut (from the top of the proventriculus to the 200  $\mu$ m point).

### Feeding assay with FITC-labeled beads

Flies were starved for 2 h at 30°C, fed with FITC-labeled beads (50 nm diameter, Polysciences, Warrington, PA, USA) to monitor the permeability of the peritrophic matrix, as described previously (Kuraishi et al., 2011). Images were captured with a Zeiss conventional fluorescence microscope or a Leica confocal microscope with a 1.5 AU pinhole. For quantification, the guts were dissected out 10 min after feeding and observed under a conventional fluorescence microscope using 20 to 30 female flies.

### $\beta$ -glo assay

Five pairs of salivary glands were dissected out from adult flies in 50  $\mu$ l of PBS, homogenized with a pestle and mortar in 350  $\mu$ l of PBS and 50  $\mu$ l of PBS containing 5% Triton X-100. Samples were then incubated for 10 min at room temperature, and diluted 100 $\times$  with PBS. Fifty microliters of diluted sample was then mixed with 10 $\times$  diluted  $\beta$ -glo reagent (Promega), incubated for 30 min and emission at 570 nm measured using a luminometer. Assays were performed on triplicate samples.

### Bromophenol Blue feeding assay

Assays were performed largely based using a published method (Cognigni et al., 2011). For quantification of feeding, three female

flies were starved for 3 h at 30°C, fed with 0.5% Bromophenol Blue (BPB) sodium salt/cornmeal-agar for 1 or 2 h, and then each fly was placed into 50  $\mu$ l MilliQ water, homogenized with a pestle and centrifuged twice (16,000 g, 2 min) to remove debris. Absorption at 594 nm was measured using a NANODROP2000 (Thermo Scientific, Waltham, MA). For quantification of the excretion rate, five female flies were starved for 3 h at 30°C, fed with 0.5% BPB sodium salt/cornmeal-agar for 1 h at 30°C, moved to a new vial containing normal food and maintained there for several hours. Each fly was placed into 80  $\mu$ l MilliQ water, homogenized with pestle and centrifuged twice to remove debris. Absorption at 594 nm was measured using a Nanodrop2000 spectrophotometer.

### Lifespan analysis

All flies were raised at 18°C for 5 to 6 days after eclosion. Three vials (each containing 30 flies) were moved to 30°C. After 2 days, lifespan analysis was started (this day was set to day 0) at 30°C. Live flies were counted every day and transferred to new vials every 2 days.

### Statistical analysis

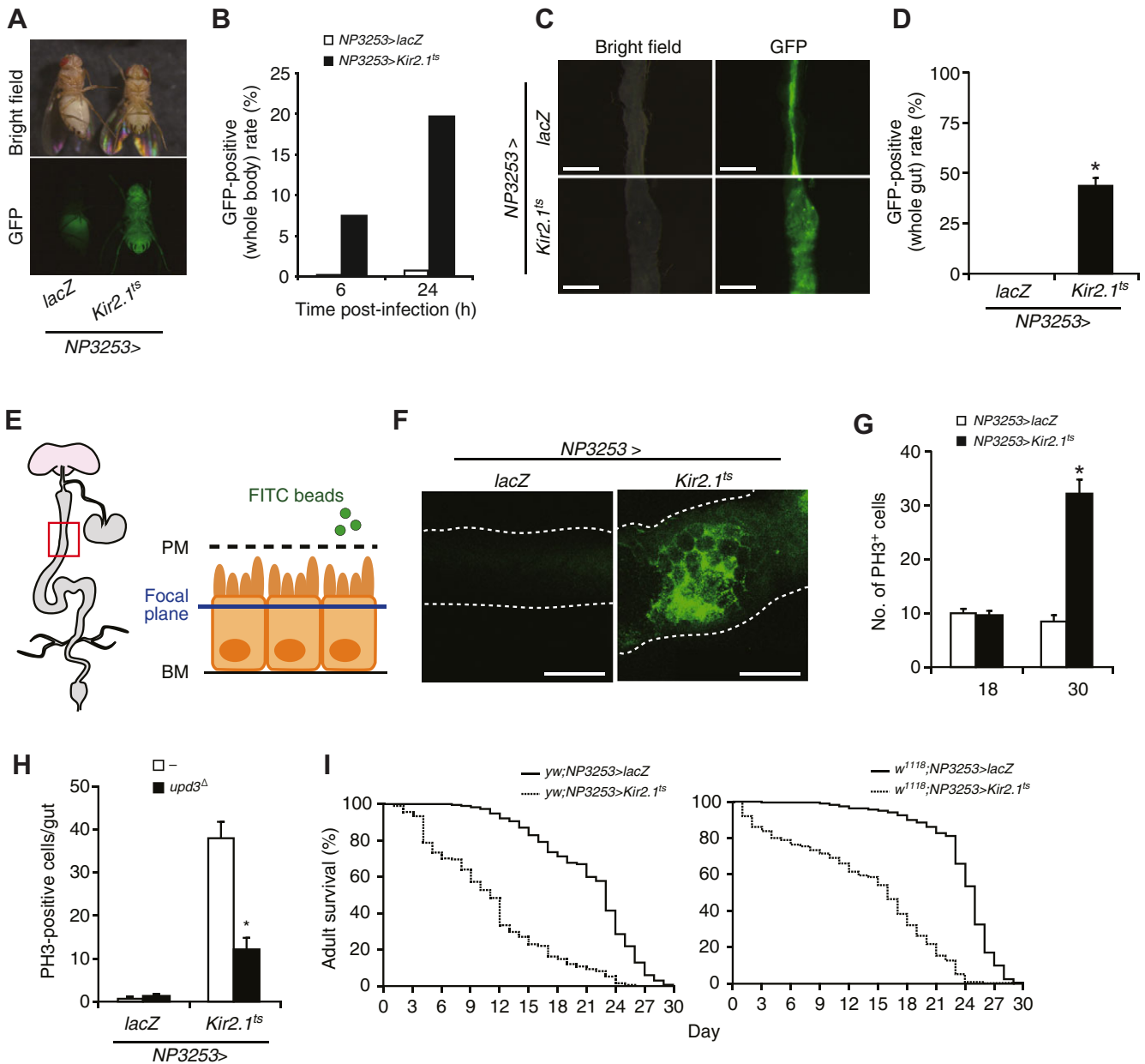
Statistical analyses were performed using Student's *t*-test or the log-rank test, and *P* values less than 0.05 were considered significant.

## RESULTS

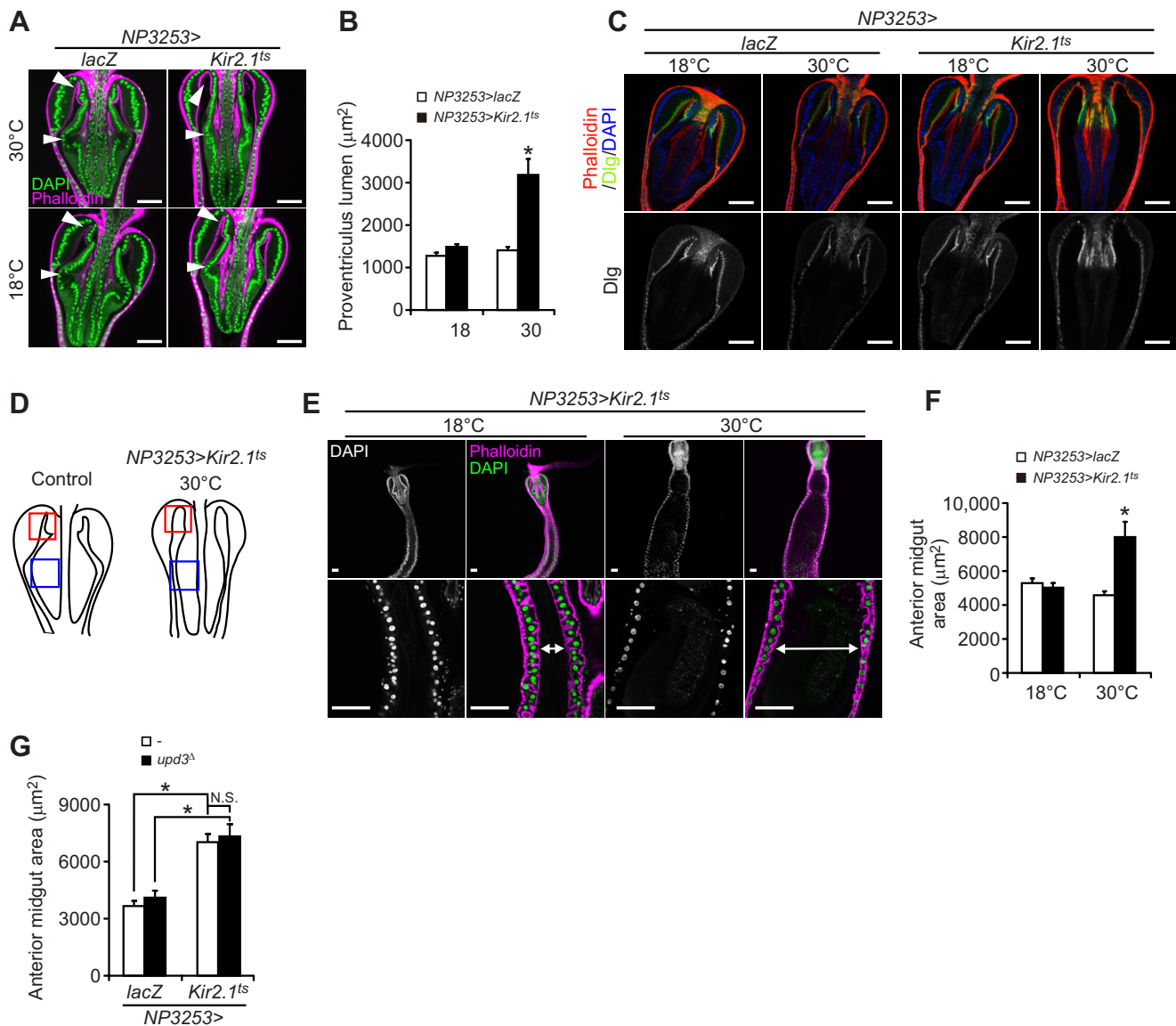
### NP3253-positive cells are required for defense against bacterial oral infection

To identify neurons important for gut integrity, we screened GAL4 enhancer trap lines with *Kir2.1*, a mammalian inwardly rectifying K<sup>+</sup> channel, to block neural activity (Baines et al., 2001), and examined their susceptibility to oral bacterial challenge as a measure of gut integrity. Sensitivity to bacterial infection is a complex phenomenon (Ayres and Schneider, 2012; Lemaitre and Hoffmann, 2007) because not only resistance mechanisms, such as the expression of antimicrobial peptides, but also tolerance mechanisms, such as permeability of the epithelial barrier, feeding behavior, excretion of ingested materials and damage repair after infection, are required for normal survival upon infection (Buchon et al., 2013a; Kuraishi et al., 2013). Therefore, if some lines are susceptible to oral infection, the underlying mutations are expected to be involved in some aspect of gut function, including structural integrity.

We selected 350 GAL4 enhancer trap lines (Table S1) known to induce expression in neurons based on the FLYBRAIN and Flytrap databases (Kelso et al., 2004; Shinomiya et al., 2011). *Kir2.1* expression was repressed by temperature-sensitive GAL80 (GAL80<sup>ts</sup>) until adulthood and then induced by shifting the flies to a restrictive temperature for 2 days (Fig. 1A). Several enhancer trap lines that expressed *Kir2.1* were susceptible to *Ecc15* oral infection (Fig. 1B). Of those, a fly line expressing *Kir2.1* using NP3253-GAL4, designated *NP3253>Kir2.1*, exhibited strong susceptibility to *Ecc15* oral infection, but not to normal fly foods (Fig. 1C,D). In contrast, flies with hyperactive NP3253-positive cells upon expression of dTrpA1 (Rosenzweig et al., 2005) were not sensitive to *Ecc15* oral infection (Fig. 1E). To exclude the possibility that these phenotypes resulted from the genetic background, the NP3253 line was backcrossed with the *yw* strain and *Kir2.1*-induced susceptibility was tested upon oral infection with *Ecc15*. The findings demonstrated that these flies are also susceptible to infection (Fig. 1C,D), indicating that the activity of NP3253-positive cells is specifically required for gut defense upon bacterial infection.



**Fig. 3. Increased permeability of gut barriers in *NP3253>Kir2.1* flies.** (A) Whole-body imaging of *NP3253>Kir2.1* flies after ingesting *Ecc15-GFP*. Flies were observed under a light microscope (top) or fluorescence microscope (bottom). Green indicates GFP signal. (B) Percentage of flies that show GFP signals in the whole body 6 h or 24 h after ingestion. Approximately 100 flies were examined and the graph shows representative results of two independent experiments. (C) Adult *NP3253>Kir2.1* flies were fed FITC-labeled 50 nm latex beads. Guts were dissected and examined under a conventional fluorescence microscope. The image shows the anterior part of the midgut. FITC signals are retained in the lumen if the beads cannot pass through the peritrophic matrix. Note that FITC signals were diffuse in *NP3253>Kir2.1* flies. Scale bars, 200  $\mu$ m. (D) Percentage of flies with FITC signal in the whole anterior midgut 10 min after feeding; 20–30 flies were examined and the graph shows a representative result of three independent experiments ( $*P<0.05$ ). (E) Schematic representation of the bead-feeding assay with a confocal microscope. The left panel shows the adult midgut and the red square indicates the examined part in F. The right panel presents the cross-section of the adult midgut and the focal plane that was scanned by a confocal microscope in F. PM, peritrophic matrix; BM, basement membrane. (F) Fluorescent confocal imaging of *NP3253>mCD8::GFP* flies and *NP3253>Kir2.1* flies fed FITC-labeled latex beads. Green indicates FITC signal. Dashed lines delineate the gut. Bars, 50  $\mu$ m. (G) The number of PH3-positive cells per one adult midgut in *NP3253>Kir2.1* flies. The flies were kept at 30°C for 2 days, and their guts were dissected and stained with anti-PH3 antibody; 10–12 flies were examined and the graph shows the average of three independent experiments ( $*P<0.05$ ). (H) PH3-positive cells in *upd3* mutant background flies; 10–12 flies were examined and the graph shows representative results of two independent experiments ( $*P<0.05$ ). (I) Survival analysis of *yw;NP3253>lacZ* or *NP3253>Kir2.1* flies at 30°C (left), or *w<sup>1118</sup>;NP3253>lacZ* or *NP3253>Kir2.1* flies at 30°C (right). Each survival curve corresponds to at least 3 independent experiments of 3 tubes of 30 flies each.  $P<0.0001$  (*yw*; *NP3253* and *w*; *NP3253*, log-rank test).



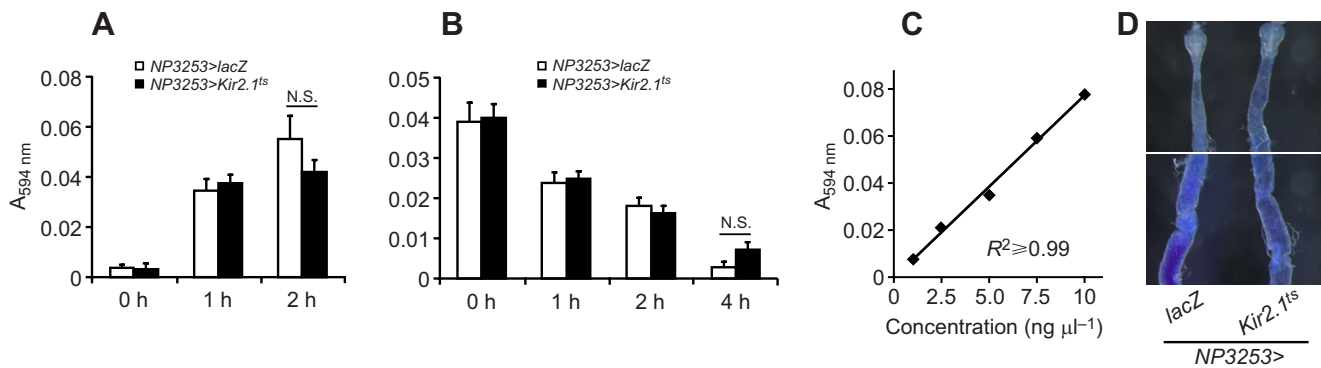
**Fig. 4. Aberrant gut structure of *NP3253>Kir2.1* flies.** (A) Fluorescent confocal imaging of the proventriculus of *NP3253>Kir2.1* flies. Green indicates nuclei (DAPI). Magenta indicates visceral muscles (Phalloidin). Arrowheads indicate areas of the proventriculus of *NP3253>Kir2.1* flies with an abnormal structure. Scale bars, 50 µm. (B) Area of the luminal region of the proventriculus was measured using ImageJ. 16–25 flies were examined and graph shows representative results of two independent experiments (\* $P < 0.05$ ). (C) Fluorescent confocal imaging of the proventriculus of *NP3253>Kir2.1* flies. Green indicates a marker of septate junctions (anti-Dlg). Blue indicates nuclei (DAPI). Red illustrates visceral muscles (Phalloidin). Bottom row shows anti-Dlg signal. Scale bars, 50 µm. (D) Schematic representation of results in A and C. Red squares indicate the location of arrowheads in A. Blue squares show the location of arrowheads in A. (E) Fluorescent confocal imaging of the anterior midgut of *NP3253>Kir2.1* flies. Magenta indicates visceral muscles (Phalloidin). Green or white indicates nuclei (DAPI). Arrows indicate the luminal width of the anterior midgut. Bottom panels are magnified views of top panels. Scale bars, 50 µm. (F,G) Area of anterior midgut (from the top of the proventriculus to the 200 µm point) measured using ImageJ at two different temperatures (F) or in *upd3* mutant or control (-) background flies (G); 16–25 flies were examined and each graph shows representative results of two independent experiments (\* $P < 0.05$ ; N.S., not significant).

### NP3253-positive neurons are responsible for the survival phenotype

We next examined expression patterns in the tissues of adult flies to evaluate whether NP3253-GAL4 could induce expression in enteric neurons. Many green fluorescent protein (GFP)-positive cells driven by NP3253-GAL4 were detected in the brain, proventriculus and anterior midgut, as well as in the posterior midgut (Fig. 2A,B), salivary glands, trachea, and reproductive organs (data not shown). A previous study (Tanaka et al., 2008) reported that NP3253 labels neurons in the mushroom body. To analyze whether the NP3253-positive cells in the gut are neurons, they were stained with horseradish peroxidase (HRP), a neural

marker protein, together with anti-GFP. The GFP-positive cells driven by NP3253-GAL4 in the proventriculus and anterior midgut were HRP positive, whereas those in posterior midgut were not (Fig. 2C). NP3253-positive cells in the proventriculus and anterior midgut were positive for the synaptic vesicle marker Syt.eGFP and the dendrite marker DenMark (Fig. 2D,E). These findings suggest that NP3253-positive cells in the proventriculus and anterior midgut are neurons, and indicate that not all NP3253-positive cells are neurons. This led us to examine whether NP3253-positive neurons are responsible for the survival phenotype upon *Ecc15* oral infection. We analyzed *NP3253>Kir2.1* flies in combination with *elav-GAL80* to inhibit GAL4 activity in all neurons (Rideout et al.,





**Fig. 5. Feeding assay with BPB-containing food.** (A) Measurement of feeding in starved *NP3253>Kir2.1* flies fed food containing 0.5% BPB for the indicated period. Three tubes of three flies each were examined and the graph shows a representative result of three independent experiments. (B) Measurement of the excretion rate in starved *NP3253>Kir2.1* flies after feeding with food containing 0.5% BPB for 1 h and then changing to normal food for the indicated period. Three tubes of five flies each were examined and the graph shows representative results of three independent experiments. (C) Calibration curve for BPB measurement. (D) Microscopic observation of the midgut of BPB-fed flies. Proventriculus anterior midguts (top) and middle-posterior (bottom) midguts were observed under a light microscope. Blue indicates BPB signal.

2010). Survival analysis revealed that susceptibility to *Ecc15* oral infection was partially rescued by co-expression with *elav-GAL80* (Fig. 2F). Both *NP3253-GAL4* and *elav-GAL80* drive gene expression in the salivary gland (Fig. 2G); therefore, to rule out the possibility that the salivary gland is responsible for the survival phenotype, we used *Bx-GAL4* and *384-GAL4* to drive *Kir2.1*. Both drivers induced reporter expression in the salivary gland as strong as *NP3253-GAL4* (Fig. 2G,H), but neither *Bx>Kir2.1* flies nor *384>Kir2.1* flies were susceptible to oral infection with *Ecc15* (Fig. 2I,J). Together, these results suggest that a subset of neurons driven by *NP3253-GAL4* is partly involved in the survival phenotype.

#### The gut barriers of *NP3253>Kir2.1* flies are highly permeable

Next, we examined why the *NP3253>Kir2.1* flies exhibit sensitivity to bacterial oral infection. After feeding the flies GFP-labeled *Ecc15* (*Ecc15-GFP*), GFP signals were observed in the whole body of *NP3253>Kir2.1* flies, in contrast to wild-type flies, which expressed the GFP signal only in the abdomen (Fig. 3A). GFP signals were observed throughout the whole body in ~10% of the *NP3253>Kir2.1* flies at 6 h after *Ecc15-GFP* feeding and in up to 20% at 24 h (Fig. 3B). This observation indicated that the bacteria intruded into the hemolymph of *NP3253>Kir2.1* flies, suggesting that gut barrier function was compromised in these flies. The peritrophic matrix is an acellular layer that protects the gut epithelium and its permeability can be assessed by feeding adults fluorescein isothiocyanate (FITC)-labeled beads (Kuraishi et al., 2011). Conventional fluorescence microscopy revealed that the 50 nm FITC-labeled beads remained in the lumen of wild-type flies after feeding (Fig. 3C,D). In contrast, FITC signals were diffuse in the gut of *NP3253>Kir2.1* flies (Fig. 3C,D). Close examination using a confocal microscope with the focal plane on the epithelial cells (Fig. 3E) revealed FITC signals outside the peritrophic matrix in the *NP3253>Kir2.1* flies (Fig. 3F). Consistent with these observations, staining with the mitotic marker PH3 revealed that *upd3*-dependent stem cell proliferation, an indicator of gut damage, was increased in the midgut of *NP3253>Kir2.1* flies (Fig. 3G,H). Furthermore, *NP3253>Kir2.1* flies had a shorter lifespan, and began to die 1 to 2 weeks after emergence (Fig. 3I). Indeed, Rera et al. (2012) reported that increased gut permeability is a cause and predictor of imminent death. These findings indicate that the peritrophic matrix of *NP3253>Kir2.1* flies is more permeable or

damaged, providing an explanation for the susceptibility of *NP3253>Kir2.1* flies to oral infection.

#### Gut structure and function of *NP3253>Kir2.1* flies

We then performed histological analysis of the gut of *NP3253>Kir2.1* flies. The proventriculus, the organ responsible for secretion of the peritrophic matrix in *Drosophila* adults, was stained with Phalloidin and DAPI to visualize the actin filaments and nuclei, respectively. As shown in Fig. 4A,B, the proventriculus morphology in *NP3253>Kir2.1* flies differed from that in wild-type flies: the bulge formed by the inner cells (indicated by arrowheads) was lost in *NP3253>Kir2.1* flies, whereas the top of the inner part of the proventriculus was expanded (indicated by the arrows). This observation was supported by visualizing the tissue structure following staining with the marker for cell junctions, discs large (*Dlg*; Fig. 4C). These findings indicated that a part of the proventriculus of *NP3253>Kir2.1* flies was flattened (Fig. 4D). We also observed a morphological abnormality of the midgut of *NP3253>Kir2.1* flies. The diameter of the anterior part of the midgut, especially the R1 region of the midgut (Buchon et al., 2013b), was increased without a significant change in the number and shape of epithelial cells (Fig. 4E,F). This phenotype was also observed in starved *NP3253>Kir2.1* flies. Notably, the increased diameter of the anterior midgut was also observed in the *upd3* mutant background (Fig. 4G), suggesting that the increased diameter is not due to damage-induced stem cell proliferation. We further examined the feeding and excretion of the *NP3253>Kir2.1* flies. As shown in Fig. 5, neither the feeding nor the excretion rate of *NP3253>Kir2.1* flies, quantified by the amount of BPB food dye that flies ate, was compromised. These results suggest that the increased diameter was not due to defective excretion of the foods they had eaten, but rather to the homeostatic dysfunction of the *NP3253>Kir2.1* flies to maintain normal gut morphology.

#### DISCUSSION

The stomatogastric nervous system controls peristalsis, fluid homeostasis and sex peptide-induced changes in intestinal physiology in adult *Drosophila*. Here, we describe a role of the nervous system in maintaining the impermeable gut physical barrier and organized epithelial structure of the anterior midgut. Several questions remain, however, as discussed below.

## The type of defect of the peritrophic matrix and epithelial barrier

We demonstrated that the peritrophic matrix of *NP3253>Kir2.1* flies is more permeable than that of wild-type flies. This phenotype is much stronger than that of *dcy<sup>1</sup>* mutant flies. The peritrophic matrix of *dcy<sup>1</sup>* mutant flies is not permeable to FITC-labeled beads with a size >70 kDa (Kuraishi et al., 2011). The peritrophic matrix of the *NP3253>Kir2.1* flies, however, was permeable not only to latex beads, but also to bacteria, implying that the nature of the peritrophic matrix defects of *NP3253>Kir2.1* flies differs from that of the *dcy<sup>1</sup>* mutant.

What is the defect that occurs in the epithelial barrier? We observed that the epithelial structure of the proventriculus and anterior midgut was disorganized and expanded in *NP3253>Kir2.1* flies. We speculate that ingested bacteria augment the epithelial expansion and might affect the septate junctions between epithelial cells, resulting in a leaky epithelial barrier in the flies. This possibility should be examined in future studies.

## The nature of NP3253-positive neurons and mechanisms of control of structural integrity

NP3253-GAL4 drives expression in neuronal subsets in the brain and the anterior midgut in *Drosophila* adults. It is unclear which NP3253-positive neurons are involved in the observed phenotype and whether efferent or sensory neurons are responsible. We cannot rule out the possibility that NP3253-positive neurons only in the brain and not enteric neurons, are responsible for the observed phenotype. Further screening of enhancer trap lines is needed to identify drivers that have a similar phenotype as NP3253-GAL4.

Our study does not address the mechanisms of the NP3253-positive neurons that maintain the impermeability of the epithelial barrier and morphology. A possible mechanism by which NP3253-positive neurons control gut integrity is endocrine/paracrine regulation. Gut patterning is primarily achieved through interactions between the pan-midgut and region-specific transcription factors, together with spatial activities of morphogens (Buchon et al., 2013b). It is thus possible that secreted factors from NP3253-positive neurons affect morphogen expression or the activities of transcriptional factors in the anterior midgut. Although we showed that peristalsis is not severely compromised in the *NP3253>Kir2.1* flies, another possibility is that NP3253-positive neurons control the pumping action of the proventriculus. The secreted components of the peritrophic matrix from the cells of the proventriculus are squeezed to form the peritrophic matrix sleeve and conveyed throughout the midgut by pumping of the proventriculus (Lehane, 1997). Therefore, if the activity of NP3253-positive neurons is inhibited, the peritrophic matrix does not form correctly or smoothly, and thus may accumulate around the anterior midgut, leading to an enlarged and abnormal structure with compromised permeability of the gut barriers. Because constitutive activation of NP3253-positive cells does not induce susceptibility to *Ecc15* oral infection (Fig. 1D), the latter explanation is more plausible.

## Acknowledgements

We are grateful to Dr B. Lemaître for *Ecc15-GFP*. We thank Drs P. Garrity and Y. Jan at the Bloomington Stock Center (NIH P40OD018537), the *Drosophila* Genomics Resource Center at Indiana University, the *Drosophila* Genetic Resource Center at the Kyoto Institute of Technology, the *Drosophila* RNAi Screening Center, the Genetic Strain Research Center of National Institute of Genetics, and the Vienna *Drosophila* RNAi Center for fly stocks. We also thank the Developmental Studies Hybridoma Bank for antibodies; and FlyBase, FLYBRAIN, Flytrap and FlyAtlas for use of the databases.

## Competing interests

The authors declare no competing or financial interests.

## Author contributions

H.K. performed many of the experiments in this study with input from M.O., T.K. and S.K. H.I. performed screening for enhancer trap lines. M.O. took the picture of the brain-gut neurons. All authors analyzed the data. H.K. and T.K. wrote the draft, and all authors finalized the manuscript.

## Funding

This work was supported by grants from the Japan Science and Technology Agency (JST); the Ministry of Education, Culture, Sports, Science, and Technology of Japan (MEXT); the Kao Foundation for Arts and Sciences; the Uehara Memorial Foundation; and the Futaba Electronics Memorial Foundation.

## Supplementary information

Supplementary information available online at <http://jeb.biologists.org/lookup/doi/10.1242/jeb.122960.supplemental>

## References

- Ayres, J. S. and Schneider, D. S. (2012). Tolerance of infections. *Annu. Rev. Immunol.* **30**, 271-294.
- Baines, R. A., Uhler, J. P., Thompson, A., Sweeney, S. T. and Bate, M. (2001). Altered electrical properties in *Drosophila* neurons developing without synaptic transmission. *J. Neurosci.* **21**, 1523-1531.
- Basset, A., Khush, R. S., Braun, A., Gardan, L., Boccard, F., Hoffmann, J. A. and Lemaître, B. (2000). The phytopathogenic bacteria *Erwinia carotovora* infects *Drosophila* and activates an immune response. *Proc. Natl. Acad. Sci. USA* **97**, 3376-3381.
- Bonnay, F., Cohen-Berros, E., Hoffmann, M., Kim, S. Y., Boulianne, G. L., Hoffmann, J. A., Matt, N. and Reichhart, J. M. (2013). big bang gene modulates gut immune tolerance in *Drosophila*. *Proc. Natl. Acad. Sci. USA* **110**, 2957-2962.
- Buchon, N., Broderick, N. A. and Lemaître, B. (2013a). Gut homeostasis in a microbial world: insights from *Drosophila melanogaster*. *Nat. Rev. Microbiol.* **11**, 615-626.
- Buchon, N., Osman, D., David, F. P. A., Fang, H. Y., Boquete, J.-P., Deplancke, B. and Lemaître, B. (2013b). Morphological and molecular characterization of adult midgut compartmentalization in *Drosophila*. *Cell Rep.* **3**, 1725-1738.
- Cognigni, P., Bailey, A. P. and Miguel-Aliaga, I. (2011). Enteric neurons and systemic signals couple nutritional and reproductive status with intestinal homeostasis. *Cell Metab.* **13**, 92-104.
- Edwards, M. J. and Jacobs-Lorena, M. (2000). Permeability and disruption of the peritrophic matrix and caecal membrane from *Aedes aegypti* and *Anopheles gambiae* mosquito larvae. *J. Insect Physiol.* **46**, 1313-1320.
- Furness, J. B. and Costa, M. (1987). *The Enteric Nervous System*. London: Churchill-Livingstone.
- Hamada, F. N., Rosenzweig, M., Kang, K., Pulver, S. R., Ghezzi, A., Jegla, T. J. and Garrity, P. A. (2008). An internal thermal sensor controlling temperature preference in *Drosophila*. *Nature* **454**, 217-220.
- Hegedus, D., Erlandson, M., Gillott, C. and Toprak, U. (2009). New insights into peritrophic matrix synthesis, architecture, and function. *Annu. Rev. Entomol.* **54**, 285-302.
- Huizinga, J. D. and Lammers, W. J. E. P. (2009). Gut peristalsis is governed by a multitude of cooperating mechanisms. *Am. J. Physiol. Gastrointest. Liver Physiol.* **296**, G1-G8.
- Karasov, W. H., Martínez del Rio, C. and Caviedes-Vidal, E. (2011). Ecological physiology of diet and digestive systems. *Annu. Rev. Physiol.* **73**, 69-93.
- Kelso, R. J., Buszczak, M., Quiñones, A. T., Castiblanco, C., Mazzalupo, S. and Cooley, L. (2004). Flytrap, a database documenting a GFP protein-trap insertion screen in *Drosophila melanogaster*. *Nucleic Acids Res.* **32**, D418-D420.
- King, D. G. (1988). Cellular organization and peritrophic membrane formation in the cardia (proventriculus) of *Drosophila melanogaster*. *J. Morphol.* **196**, 253-282.
- Kuraishi, T., Binggeli, O., Opota, O., Buchon, N. and Lemaître, B. (2011). Genetic evidence for a protective role of the peritrophic matrix against intestinal bacterial infection in *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. USA* **108**, 15966-15971.
- Kuraishi, T., Hori, A. and Kurata, S. (2013). Host-microbe interactions in the gut of *Drosophila melanogaster*. *Front. Physiol.* **4**, 375.
- Lehane, M. J. (1997). Peritrophic matrix structure and function. *Annu. Rev. Entomol.* **42**, 525-550.
- Lemaître, B. and Hoffmann, J. (2007). The host defense of *Drosophila melanogaster*. *Annu. Rev. Immunol.* **25**, 697-743.
- Lemaître, B. and Miguel-Aliaga, I. (2013). The digestive tract of *Drosophila melanogaster*. *Annu. Rev. Genet.* **47**, 377-404.



- Rera, M., Clark, R. I. and Walker, D. W.** (2012). Intestinal barrier dysfunction links metabolic and inflammatory markers of aging to death in *Drosophila*. *Proc. Natl. Acad. Sci. USA* **109**, 21528-21533.
- Rideout, E. J., Dornan, A. J., Neville, M. C., Eadie, S. and Goodwin, S. F.** (2010). Control of sexual differentiation and behavior by the doublesex gene in *Drosophila melanogaster*. *Nat. Neurosci.* **13**, 458-466.
- Rosenzweig, M., Brennan, K. M., Tayler, T. D., Phelps, P. O., Patapoutian, A. and Garrity, P. A.** (2005). The *Drosophila* ortholog of vertebrate TRPA1 regulates thermotaxis. *Genes Dev.* **19**, 419-424.
- Sansonetti, P. J.** (2004). War and peace at mucosal surfaces. *Nat. Rev. Immunol.* **4**, 953-964.
- Shinomiya, K., Matsuda, K., Oishi, T., Otsuna, H. and Ito, K.** (2011). Flybrain neuron database: a comprehensive database system of the *Drosophila* brain neurons. *J. Comp. Neurol.* **519**, 807-833.
- Talsma, A. D., Christov, C. P., Terriente-Felix, A., Linneweber, G. A., Perea, D., Wayland, M., Shafer, O. T. and Miguel-Aliaga, I.** (2012). Remote control of renal physiology by the intestinal neuropeptide pigment-dispersing factor in *Drosophila*. *Proc. Natl. Acad. Sci. USA* **109**, 12177-12182.
- Tanaka, N. K., Tanimoto, H. and Ito, K.** (2008). Neuronal assemblies of the *Drosophila* mushroom body. *J. Comp. Neurol.* **508**, 711-755.
- Tepass, U., Tanentzapf, G., Ward, R. and Fehon, R.** (2001). Epithelial cell polarity and cell junctions in *Drosophila*. *Annu. Rev. Genet.* **35**, 747-784.
- Wang, P. and Granados, R. R.** (2000). Calcofluor disrupts the midgut defense system in insects. *Insect Biochem. Mol. Biol.* **30**, 135-143.
- Yang, C.-h., Rumpf, S., Xiang, Y., Gordon, M. D., Song, W., Jan, L. Y. and Jan, Y.-N.** (2009). Control of the postmating behavioral switch in *Drosophila* females by internal sensory neurons. *Neuron* **61**, 519-526.