

RESEARCH ARTICLE

Impact of elevated CO₂ background levels on the host-seeking behaviour of *Aedes aegypti*

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ABSTRACT

Mosquitoes rely on carbon dioxide (CO₂) to detect and orient towards their blood hosts. However, the variable and rapid fluctuations of atmospheric CO2 concentrations may have an impact on the hostseeking behaviour of mosquitoes. In this study, we analysed the effect of transient elevated background levels of CO₂ on the hostseeking behaviour and the physiological characteristics of the CO₂sensitive olfactory receptor neurones (ORNs) in female yellow fever mosquitoes, Aedes aegypti. We show that the take-off and source contact behaviour of A. aegypti is impeded at elevated background levels of CO₂ as a result of masking of the stimulus signal. The mechanism underlying this masking during take-off behaviour is one of sensory constraint. We show that the net response of the CO₂-ORNs regulates this CO₂-related behaviour. Since these neurones themselves are not habituated or fatigued by the transient elevation of background CO2, we propose that habituation of second-order neurones in response to the elevated CO₂-ORN activity could be one mechanism by which the net response is transduced by the olfactory system. The findings from this study may help to predict future shifts in mosquito-host interactions and consequently to predict vectorial capacity in the light of climate change.

KEY WORDS: Olfaction, Single sensillum recording, Wind tunnel, Climate change, Absolute detector

INTRODUCTION

Volatiles play an important role in the host-seeking behaviour of mosquitoes (Takken and Knols, 1999). Carbon dioxide (CO₂) is one of the most important volatiles, a key kairomone emitted by vertebrates, which has been shown to be a reliable cue for mosquitoes to detect and orient towards a host species (Mboera and Takken, 1997). Such a role for CO₂ in mosquito foraging was recognised by Rudolfs as early as 1922 (Rudolfs, 1922). However, diel atmospheric CO₂ concentrations vary between 350 parts per million (ppm) and 500 ppm (up to 1000 ppm in dense vegetation), and rapid fluctuations are features of natural CO₂ sources (Gillies, 1980; Guerenstein and Hildebrand, 2008). Consequently, variations in ambient CO₂ levels could affect the ability of mosquitoes to process CO₂ fluctuations (Grant et al., 1995) and modulate their host-seeking behaviour. To date, there have been no published studies that have dealt with the effects of elevated ambient CO₂ levels on mosquito behaviour. Analysis of CO₂ processing in the context of host-seeking behaviour could help to identify the mechanisms underlying these effects, to predict shifts in

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mosquito-host interactions and consequently to predict vectorial capacity.

Activation and source finding by host-seeking female mosquitoes occur when the fine-scale structure of the CO₂ plume is filamentous, i.e. when flying mosquitoes are exposed to intermittent increases in CO₂ concentration above background, as shown under controlled conditions (Geier et al., 1999; Dekker et al., 2001). Such a plume is encountered naturally by mosquitoes up to tens of metres from a host (Costantini et al., 1996; Zöllner et al., 2004). However, in a variable ambient CO₂ background, the effective range of attraction could decrease, due to limitations of the CO₂ chemosensory system.

Highly sensitive olfactory receptor neurones (ORNs), housed in the maxillary palp capitate peg sensilla of mosquitoes, detect CO_2 (Grant et al., 1995; Lu et al., 2007; Syed and Leal, 2007). Pulses of CO_2 elicit phasic—tonic responses in these ORNs, and the pulse duration is dependent on the concentration and length of the stimulus pulse (Grant et al., 1995). Grant et al. have shown that this response of the CO_2 -ORNs might be impeded in high ambient concentrations of CO_2 . However, we do not know how mosquitoes encode such an impediment and how it affects the behavioural response of mosquitoes to CO_2 .

In this study, we analyse the sensory mechanism underlying the behavioural response of the yellow fever mosquito, *Aedes aegypti*, to CO_2 . We show that female *A. aegypti* encode pulsed CO_2 stimuli, dependent on ambient CO_2 levels, as a net sensory response. Furthermore, we show that the net sensory response dictates behavioural activation across ambient CO_2 levels. The effect of ambient CO_2 levels on the host-seeking behaviour and vectorial capacity of mosquitoes is discussed.

RESULTS

CO₂-ORN responses in different background levels of CO₂

Stimulation with increased concentrations of pulsed CO₂ elicited a significant increase in the A-cell response (background level 400 ppm: *F*=61.54, d.f.=3, *P*<0.001; background level 600 ppm: F=480.81, d.f.=3, P<0.001; background level 1200 ppm: F=379.54, d.f.=3, P<0.001), as well as in the net response when the stimulus concentration exceeded that of the background level (background level 400 ppm: F=75.36, d.f.=3, P<0.001; background level 600 ppm: *F*=425.76, d.f.=3, *P*<0.001; background level 1200 ppm: F=403.47, d.f.=3, P<0.001) (Fig. 1C; Fig. 2). Conversely, a significant decrease in the net response was observed when the stimulus concentration was lower than that of the background level (t=-17.82, d.f.=9, P<0.001, t-test), i.e. when stimulating with 600 ppm CO₂ in a 1200 ppm CO₂ background (Fig. 2). Hence, the overall net response of the CO₂-ORNs was significantly affected by the background level of CO_2 (F=4.0, d.f.= 2, P=0.03). The decrease in net response was contributed to by a significant increase in the interstimulus activity of the CO₂-ORNs at 600 and 1200 ppm CO₂ background levels, compared with that at 400 ppm CO_2 (F=46.47, d.f.= 2, P < 0.001) (Fig. 2).

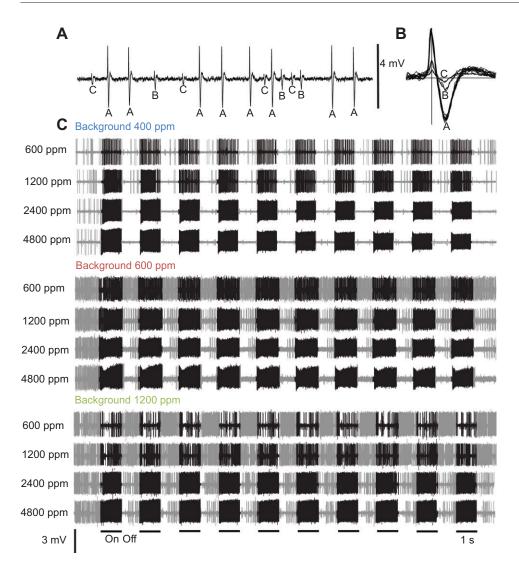


Fig. 1. The temporal characteristics of CO₂ chemosensation in female A. aegypti in different background levels of CO2 to four different concentrations of pulsed CO2 stimuli. (A) An extracellular recording from a capitate-peg sensillum showing the spontaneous activity of the large amplitude 'A', the intermediate 'B' and the small 'C' neurones, respectively. Vertical bar indicates the spike amplitude in millivolts (mV). (B) Waveform analysis of the extracellular recording in (A) shows the distinct waveforms of each neurone. (C) Detection of pulsed CO₂ by the CO₂-ORNs, in different CO₂ backgrounds, over ascending concentrations. Black spikes represent the response by CO2-ORNs to a pulsed train of 1 s CO₂ stimuli and the grey spikes indicate the interstimulus activity of the CO₂-ORNs. The vertical bar denotes the amplitude in millivolts. The horizontal bar represents the pulsed train of 1 s CO₂ stimuli

Consistency of pulsed stimuli of CO₂

To investigate the correlation between CO₂-ORN and behavioural responses, a similar stimulus protocol was adopted in all experiments. To ensure that the train of pulsed stimuli were neither molecularly nor turbulently diffused in the air stream of the wind tunnel, we designed a protocol to produce pulse trains consistent in shape and amplitude throughout the wind tunnel. To verify this consistency, a train of distinct pulsed stimuli of CO₂ was visualised by using acetone as tracer gas using a mini-photo ionisation detector (mini-PID) to track the ascending flow of known concentrations of acetone at different positions (centre and lateral sides) and distances (release chamber, halfway and source contact). This explicitly showed that the pulsed stimuli had a consistent shape and were clearly separated from one another throughout the wind tunnel (Fig. 3A, left inset, N=10). The amplitude of each discrete pulse was shown by linear regression to be consistent in all positions in the wind tunnel assayed for each flow rate (Fig. 3A, right inset, N=10).

Behavioural responses to CO₂ in different CO₂ backgrounds

The host-seeking behaviour of female *A. aegypti* was significantly affected by both the stimulus concentration and the background level of CO₂ (Fig. 3B). Time to take-off was significantly decreased when stimulus concentrations exceeded 1200 ppm CO₂, at background levels of 400 and 600 ppm CO₂ (Fig. 3B). At a

background level of 1200 ppm CO₂, however, time to take-off was not affected by stimulus concentration (*F*=3.6, d.f.=29, *P*=0.3) (Fig. 3B). In addition, time to take-off at this background level was significantly increased compared with that observed at background levels of 400 and 600 ppm CO₂ (Fig. 3B). Once the mosquitoes had taken off, the time to reach the halfway mark in the wind tunnel was not significantly affected by either stimulus concentration or background level of CO₂ (data not shown). In contrast, the time to source contact was affected by both stimulus concentration and the background level of CO₂ (Fig. 3B). At a background level of 400 ppm CO₂, the time to source contact was significantly decreased as stimulus concentration increased (Fig. 3B). However, at background levels of 600 and 1200 ppm CO₂, mosquitoes did not take a significantly shorter time to reach the source at any stimulus concentration (Fig. 3B).

Correlation between behaviour and physiology

The net sensory response over increasing stimulus concentrations among the different background levels was compared, following normalisation (Fig. 4). The slopes of the net response curves in 600 and 1200 ppm $\rm CO_2$ background levels were significantly different from that of the 400 ppm $\rm CO_2$ level (F=6.52, d.f., d.f., d=76, P=0.013), but not from each other (F=0.48, d.f.,=1, d.f.,d=76, P=0.49). A net response threshold of \geq 100 spikes s⁻¹ was found

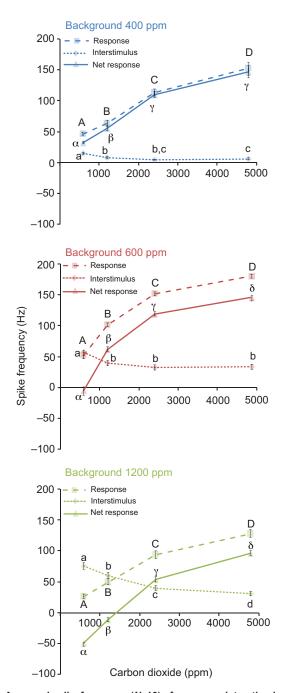


Fig. 2. Averaged spike frequency (*N*=10) of response, interstimulus activity and net response of the CO₂-ORNs in different CO₂ backgrounds. Letters and symbols denote significant differences within the response, interstimulus activity and net response over increasing concentrations. Error bars represent the standard error of the mean. The spike frequency is represented in hertz (Hz).

(Fig. 4), which correlated with a significant decrease in time to takeoff flight (Fig. 3B) regardless of background CO₂ level (Figs 4, 5). At no tested CO₂ concentration, in a background level of 1200 ppm, did the mosquitoes decrease their time to take-off (Fig. 4), which correlated with the maximum net response generated of 92.5±4.1 spikes s⁻¹ (Figs 4, 5).

A correlation was observed between net sensory response of the CO₂-ORNs and the behavioural response to pulsed stimuli of CO₂ in elevated background of CO₂ levels (Fig. 5). The time to take-off

flight was significantly decreased as the stimulus concentration of CO₂ exceeded 1200 ppm at the CO₂ background of 400 and 600 ppm, which was significantly correlated with the net response threshold of \geq 100 spikes s⁻¹ (400 ppm at r=0.9764, 95% confidence interval: 0.2489 to 0.9995, P=0.0236; 600 ppm at r=0.9830, 95% confidence interval: 0.3970 to 0.9997, P=0.0170; Figs 4, 5). In addition, the upwind flight towards source contact at CO₂ background of 400 ppm was correlated with the net response (r=0.9644, 95% confidence interval: 0.04438 to 0.9993, P=0.0356; Fig. 5). However, at the higher background levels of CO₂, the stimulus concentration had no effect on upwind flight (Fig. 5).

DISCUSSION

In this study, we have analysed the sensory mechanism that is involved in constraining the behavioural response of A. aegypti to CO_2 at elevated background CO_2 levels. Knowledge gained through this study sheds new light on the effects of varying CO_2 environments on the interactions between insects and hosts, in general, and between disease-vector mosquitoes and their blood hosts specifically. The study also improves our ability to predict shifts in vectorial capacity and other community interactions in future environments.

Impact of elevated CO₂ levels on mosquito behaviour

Carbon dioxide activates and modulates the host-seeking behaviour of insects, including mosquitoes, in a concentration-dependent manner (Dekker et al., 2001; Guerenstein and Hildebrand, 2008; Dekker and Cardé, 2011). We have shown that a transient elevation of the background level of CO₂ significantly affects the behavioural response of the mosquitoes. Specifically, our behavioural data suggest that an elevation of the background level of CO₂ adds a masking effect that reduces the detection of the CO₂ stimulus, affecting both activation and source finding. A similar mechanism has been reported to affect oviposition by the pyralid moth *Cactoblastis cactorum*, and its peripheral reception (see below) (Stange, 1997). This mechanism is closely analogous to the attraction of male moths towards the pheromone of calling females, which decreases with an increase in the background concentration of pheromone (Sanders, 1982; Sanders and Lucuik, 1996; Schofield et al., 2003).

Our behavioural data suggest that mosquitoes are able to cope with the present natural diurnal and seasonal changes in atmospheric CO_2 concentrations, i.e. 350–500 ppm (up to 1000 ppm in dense vegetation) (Gillies, 1980; Guerenstein and Hildebrand, 2008). However, it is unclear if they will be able to do so following the predicted ongoing increase in atmospheric CO_2 , 550–1000 ppm by the turn of the next century (Guerenstein and Hildebrand, 2008). This ongoing increase in ambient CO_2 level may reduce the vectorial capacity of mosquitoes by limiting the effective range of attraction to their blood hosts (Zöllner et al., 2004). Additional studies, however, are required to investigate the ability of mosquitoes to adapt to ongoing increases in atmospheric CO_2 levels.

Constraint of the CO₂-ORNs

Limitations of the CO₂-induced behavioural response are dependent on the physiological response of their CO₂-ORNs. In *A. aegypti*, CO₂-ORNs respond in a concentration-dependent manner at all background levels of CO₂. A dampening of the signal resolution occurs between 2400 and 4800 ppm, which corresponds to a previously observed upper behavioural response threshold (Costantini et al., 1996). A plausible mechanism that the olfactory system of mosquitoes uses to minimise this loss of signal resolution is to subtract the spontaneous (interstimulus) activity of the ORNs

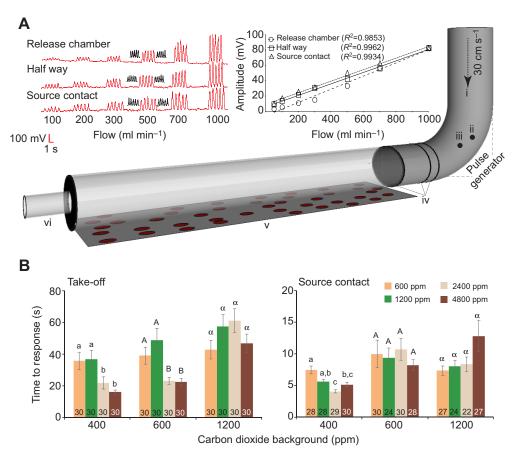


Fig. 3. Behavioural responses of female A. aegypti in different background levels of CO_2 . (A) Behaviour was assessed in a wind tunnel assay, as shown in the schematic diagram in the bottom panel: (i) charcoal-filtered and humidified air, (ii) pressurised air inlet, (iii) stimulus inlet into which CO_2 was injected, (iv) stainless-steel mesh plume diffusers, (v) glass flight tunnel and (vi) release chamber. The upper panels demonstrate that the pulsed stimuli (here shown as five cycles of 1 s on and 1 s off) maintain their amplitude and shape throughout the wind tunnel and at all tested flow rates. The upper left panel shows the consistent and distinct pulsed stimuli at ascending flow rates of known concentration of acetone in the wind tunnel. Discrete pulsed stimuli were measured in the centre (in red) and at the lateral sides (in black) of the release chamber, at halfway and at the source. The upper right panel presents a graphical representation of the distinct pulsed stimuli, which shows the average amplitude of each of the five distinct pulses (N=10) at different positions and the regression correlation coefficients (R=10) that demonstrate the consistency of the stimulus amplitude at the different positions within the wind tunnel with increasing flow rates. (B) Female mosquito attraction towards pulsed stimuli of four concentrations of CO_2 in the wind tunnel at different CO_2 background levels is shown (N=30 for each background level). Numbers inside the bars represent the number of tested individuals that took off (left panel) or made source contact (right panel). Letters indicate the significant differences within the background levels over increasing concentrations.

at a given CO₂ background level from the stimulus response. This generates a net response that remains linear throughout the ecologically relevant range of CO2 stimuli. This strategy, while maintaining signal resolution to elevated CO₂ stimuli, requires stronger stimuli in increased CO₂ background levels to produce an equivalent signal to that generated in a lower CO₂ background. The requirement for a stronger stimulus in the elevated backgrounds is likely to be mitigated by the reduction of the membrane potential needed to achieve the threshold for firing an action potential. This is a result of the increased baseline membrane potential predicted during the interstimulus periods in elevated background CO₂. Thus mosquitoes are able to accurately detect the level of CO₂ stimulus above background CO₂ levels over a broad range of concentrations. This sensitivity ensures that a mosquito is able to respond to a plume of host-emitted CO2 in which distance from the source becomes the limiting factor in elevated backgrounds of CO₂.

Carbon dioxide-ORNs have been described as absolute concentration detectors, i.e. at or below the background CO₂ level, the sensory response to CO₂ remains linear (Syed and Leal, 2007), and background CO₂ concentration has little to no effect on the

stimulus response (Grant et al., 1995). Our electrophysiological analyses revealed that the CO₂-ORNs meet the first criterion, but not the second. Mosquito CO₂-ORNs exhibit compressive non-linearity (Stevens, 1971). The response to CO₂, as a function of the stimulus (spikes s⁻¹ ppm⁻¹), was significantly reduced when the background level increased. Similar observations have been made for CO₂ detection in the moth *C. cactorum* (Stange and Wong, 1993; Stange, 1997), for 1-octen-3-ol detection in the fly *Musca domestica* (Kelling et al., 2002), and for pheromone detection in a wide variety of moths (Willis and Baker, 1984; Mafra-Neto and Baker, 1996; Evenden et al., 2000). The compressive non-linearity suggests that insects experiencing elevated background concentrations of stimuli either are unable to perceive the stimulus or are habituated, which consequently affects their behavioural performance.

We found no indication of sensory neurone habituation or fatigue during the electrophysiological experiments (<5 min exposure to the elevated CO₂ background), as is demonstrated by there being no difference in CO₂-ORN response to stimulus or interstimulus during the first, fifth or tenth stimulation. The duration of the exposure of the mosquitoes to the background CO₂ during the behavioural

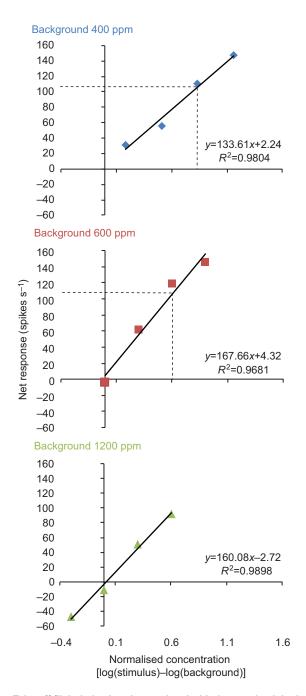


Fig. 4. Take-off flight behaviour is correlated with the net physiological response of the CO_2 -ORNs to normalised CO_2 concentrations in A. aegypti females. The dotted lines indicate the behavioural threshold in net physiological response, i.e. at which time to take-off flight was significantly reduced.

assays (<3 min) was less than that used for the physiological assays. Yet, with no evidence of ORN habituation or fatigue, the modulation behaviour is still evident within this time period. We argue that this is due to the sensory information included both in the stimulus response and the interstimulus response: what we have termed the net response. Although we have not investigated the response profile of second-order neurones in the antennal lobe in this study, we propose that second-order neurones may become habituated to the background firing of the CO₂-ORNs. In fact, this may be one mechanism by which the difference between the background firing

of the ORN could be subtracted from the stimulus, resulting in the transfer of the net response to the higher brain centres. In this model, the absolute sensory constraint on the activation behaviour is the difference between the background and maximum firing rate of the CO₂-ORNs. If the difference is less than the physiological threshold then no decrease in time to activation behaviour can occur. This system provides signal resolution in a sea of noise, which would otherwise result in the activation of host-seeking behaviour in elevated background CO₂ concentrations in the absence of a host.

Future perspectives

Considering the reliance on CO₂ sources, the CO₂-sensory system of mosquitoes is an ideal target to disrupt their host-seeking behaviour. In this study, we showed that an elevation of ambient CO₂, of up to three times that of normal concentration, adds a masking effect that significantly reduces the detection of this key host kairomone. In line with our finding, Turner et al. (Turner et al., 2011) recently showed that host-seeking mosquitoes become disoriented by artificially prolonged activation of CO₂-ORNs, which masks the ability of the mosquitoes to detect changes in the concentration of CO₂ in the environment. We believe that our study will provide a better understanding of the natural mechanisms involved in this masking process.

MATERIALS AND METHODS

Insects

Aedes aegypti (Rockefeller strain) were kept at 27°C, 65±5% relative humidity (RH) and at a 12:12 h light:dark period, as previously described (Cook et al., 2011). The ambient concentration of CO₂ during rearing and experiments was 400±5 ppm sugar-fed, 4- to 7-days post-emergence, female mosquitoes were used in this study.

Single sensillum recordings

Capitate peg sensilla are found on the fourth segment of the maxillary palps of female *A. aegypti* (McIver, 1972). Each sensillum houses three ORNs, distinguishable by spike amplitude (Fig. 1A,B) (Cook et al., 2011). The ORN with the largest amplitude is, by convention, referred to as the A-cell and has previously been shown to respond to CO₂ (Grant et al., 1995). Single sensillum recordings from this cell were performed as previously described (Cook et al., 2011). A single recording was taken from each of 10 preparations at each background concentration. In total, recordings were made from 30 mosquitoes.

CO₂ stimulation

A continuous humidified airstream containing a background of either ambient (400), 600 or 1200 ppm CO₂ was delivered at $21 \,\mathrm{min}^{-1}$ via a glass tube (7 mm i.d.). The elevated CO₂ backgrounds were obtained by diluting pure CO₂ (Strandmöllen, Ljungby, Sweden) directly into the airstream. The outlet of the tube was placed approximately 10 mm from the maxillary palps. An IDAC-4 (Syntech, Kirchzarten, Germany) was used to activate two-way Teflon solenoid valves (Teddington, Lanna, Sweden) that controlled the delivery of an embedded CO₂ stimulus into the glass tube through a separate CO₂ line. The stimulus was embedded into the airstream through a hole (2 mm i.d.) in the glass tube, 11 cm upstream of the maxillary palps, and the pulses were verified by using a CO₂ analyser (LI-820, LI-COR Biosciences, Lincoln, NE, USA). The solenoid valves were connected to separate gas cylinders containing metered amounts of CO₂ (600, 1200, 2400 and 4800 ppm) and oxygen (20%), balanced by nitrogen (Strandmöllen). A pulsed stimulus train of CO₂ was used, with stimulation for 1 s and an interstimulus interval of 1 s.

Wind tunnel bioassay

Behavioural experiments with pulsed CO₂ stimuli were performed in a glass wind tunnel (80×9.5 cm i.d.) (Fig. 3A), illuminated from above at 280 lx. A charcoal-filtered and humidified air stream (25±2°C, RH 65±2%) flowed

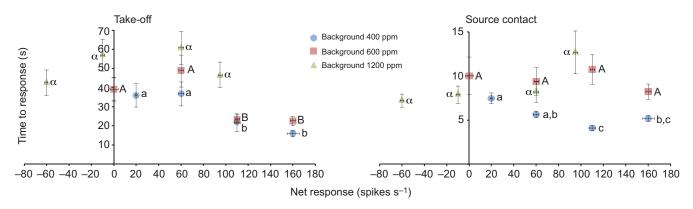


Fig. 5. The behavioural response of female *A. aegypti* is correlated with the net physiological response of the CO₂-ORNs. Letters and symbols denote significant differences among the behavioural responses to CO₂ stimuli within each CO₂ background. Vertical and horizontal error bars represent the standard error of the means of time to take-off and net response, respectively.

through the wind tunnel at 30 cm s⁻¹. The air was passed through a series of stainless-steel mesh screens to generate a laminar flow and a homogenous plume structure.

To investigate the direct correlation between sensory input and behaviour, we created distinct pulsed stimuli of 1 s on and 1 s off, embedded on the CO₂ background of 400, 600 and 1200 ppm, in a wind tunnel (Fig. 3A, inset). The transient elevated backgrounds were obtained by diluting pure CO₂ in the main airstream. The pulsatile plume structure was designed to be consistent in amplitude and structure throughout the length and breadth of the wind tunnel. For this, homogenous discrete pulse stimuli were created by pushing pure CO2-laden air into a pulse generator placed behind the stainless-steel mesh screens through a stimulus controller (SEC-2/b, Syntech) (Fig. 3A). Desired concentrations (600, 1200, 2400 and 4800 ppm) of pulsed CO₂ were obtained by regulating the CO₂ flow to the stimulus controller. To ensure distinct pulses of stimuli, pressurised air at 4.5 l min⁻¹ was introduced into the pulse generator just downstream of the point where the CO2 was introduced (Fig. 3A). Various flow rates of known amounts of acetone as a tracer gas (99.9%, Chromasolv, Sigma-Aldrich, Stockholm, Sweden) were used to investigate the consistency of distinct pulse stimuli throughout the wind tunnel and measured at different positions (centre and lateral sides) and distances (40 cm 'source contact', 80 cm 'halfway' and 120 cm 'release chamber') from the pulse generator (Fig. 3A, inset). Five cycles of 1 s on and 1 s off were tested at each flow rate. The consistency in amplitude and the structure of the pulsed stimuli was visualised using a mini-PID (Aurora Scientific, Aurora, Ontario, Canada). The corresponding concentrations of CO₂ were calculated backwards from mini-PID responses to the known concentration of acetone with different flow rates (Fig. 3A, inset). The concentration of CO2 was measured at the downwind and upwind end of the wind tunnel via the CO2 analyser.

Female mosquitoes were kept individually in $7\times2.6\,\mathrm{cm}$ i.d. glass release chambers in the wind tunnel room for $24\,\mathrm{h}$ before the experiments. Mosquitoes were provided with water through a moistened filter paper placed against the stainless-steel mesh, which covered one end of the chamber. The release chambers were placed in the centre of the downwind end of the wind tunnel. Thereafter, the following behavioural steps were observed: time to take-off flight, halfway and source contact, for a maximum of $120\,\mathrm{s}$. Equal numbers of experimental and control flights were performed each day.

Statistical analysis

Physiological activity was analysed (AUTOSPIKE, Syntech) as the number of spikes during interstimulus interval (1 s) subtracted from the number of spikes during the stimulus response (1 s), resulting in the net response (spikes $\rm s^{-1}$; 10 replicates per elevated $\rm CO_2$ background level). The response, interstimulus activity and net response resulting from the first, fifth and tenth stimulus pulses were first assessed for normal distribution using the D'Agostino omnibus $\rm \it K^2$ test (Prism version 5.01, GraphPad Software, La Jolla, CA, USA) and then compared by 2-way repeated measures analysis of variance (ANOVA) followed by Bonferroni *post hoc* test (Prism version 5.01).

Data resulting from the fifth stimulus pulse was then used for all subsequent analyses, as there were no significant differences between factors among the stimulus pulses. Power analysis was performed on net physiological response to determine the sample size. Furthermore, we determined the significant effect of stimulus concentration on each variable by a general linear model (GLM) 2-way ANOVA followed by Tukeys' *post hoc* test (Minitab version 16.1.0). A one-sample *t*-test was performed to analyse the significance level of response to the stimulus of 600 ppm CO₂ in the background of 1200 ppm CO₂ compared with zero (Minitab version 16.1.0).

To define the relationship between CO_2 -ORN response and CO_2 background level, a regression analysis was performed separately for each insect tested. The slopes were used to analyse the statistical variation, with respect to background level of CO_2 , for net response (Prism version 5.01). Prior to linear regression analyses, we normalised the concentration regime by subtracting the logarithm of the stimulus concentration from the logarithm of the background level.

To describe the correlation between CO_2 -ORN net response (10 replicates per elevated CO_2 background level) and behavioural observation (30 replicates per elevated CO_2 background level), Pearson's correlation was used. Based on the prior behavioural (Figs 3, 5) and physiological (Figs 2, 5) analyses, thresholds for a significant reduction in time to take off (\leq 30 s; \geq 100 spikes s⁻¹) and source contact (<6 s; \geq 65 spikes s⁻¹) were identified. The number of observations that matched these criteria for each dose in each background condition were counted and recorded as a ratio of the total number of observations. Pearson's correlation coefficients were calculated comparing the ratios of significant behavioural and physiological responses for each background condition.

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Competing interests

The authors declare no competing financial interests.

Author contributions

All authors had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Study concept and design: R.I., S.R.H. and S.M. Acquisition of data: S.M. Analysis and interpretation of data: R.I., S.R.H. and S.M. Drafting of the article: R.I., S.R.H. and S.M. Critical revision of the article for important intellectual content: R.I. and S.R.H. Obtained funding: R.I.

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