

## Seasonality of the red blood cell stress response in rainbow trout (*Oncorhynchus mykiss*)

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### Summary

The  $\beta$ -adrenergic stress response in red blood cells (RBCs) of rainbow trout shows seasonal changes in expression. We have explored the mechanisms underpinning this response by following, over a period of 27 months, changes in  $\beta$ -adrenergic receptor ( $\beta$ -AR) binding characteristics,  $\beta$ -adrenergically stimulated RBC  $\text{Na}^+/\text{H}^+$  exchanger ( $\beta\text{NHE}$ ) activity, together with  $\beta$ -AR and  $\beta\text{NHE}$  mRNA levels and plasma steroid hormone and lactate levels. These parameters were measured at approximately monthly intervals in a single population of fish held under semi-natural conditions. Membrane-bound, high-affinity  $\beta$ -ARs were present in RBCs at all sampling times, varying from  $668 \pm 112$  receptors  $\text{cell}^{-1}$  to  $2654 \pm 882$  receptors  $\text{cell}^{-1}$  (mean  $\pm$  S.E.M.;  $N=8$ ).  $\beta\text{NHE}$  activity, however, was reduced by 57% and 34% in December 1999 and February 2001, respectively, compared with an otherwise sustained influx that averaged  $110.4 \pm 2.3$   $\text{mmol l}^{-1}$  RBCs  $\text{h}^{-1}$  ( $N=119$ ). Only one reduction coincided with a spawning period but both were preceded by transient increases in circulating testosterone.  $\beta\text{NHE}$  activity measured under standard conditions was not correlated with the number or affinity of  $\beta$ -ARs nor

with water temperature, but both  $\beta$ -AR numbers and  $\beta\text{NHE}$  activity were positively related to their respective mRNA levels ( $P=0.005$  and  $0.038$ , respectively). Pharmaceutical intervention in the transduction cascade linking the  $\beta$ -AR and  $\beta\text{NHE}$  failed to indicate any failure of the transduction elements in RBCs displaying low  $\beta\text{NHE}$  activity. Similarly, we failed to demonstrate any link between seasonal cortisol fluctuations and seasonally reduced  $\beta\text{NHE}$  activity. However, the  $\beta\text{NHE}$  activity of age-separated RBC fractions showed that younger RBCs had a significantly higher  $\beta\text{NHE}$  response than older RBCs, consistent with the seasonal reductions in  $\beta\text{NHE}$  being linked to turnover of RBCs and erythropoiesis. Testosterone is known to induce erythropoiesis and we conclude that seasonal reductions in  $\beta\text{NHE}$  are not caused by changes in  $\beta$ -AR numbers but may be linked to testosterone-induced erythropoiesis.

Key words: adrenergic receptor,  $\text{Na}^+/\text{H}^+$  exchanger, oestradiol, testosterone, cortisol, seasonal changes, rainbow trout, *Oncorhynchus mykiss*.

### Introduction

The red blood cell  $\beta$ -adrenergic response is of crucial importance for oxygen delivery to the tissues in numerous fish species during stressful events such as burst swimming or environmental hypoxia (Primmitt et al., 1986; Nikinmaa, 1992). In salmonid fish, the magnitude of the  $\beta$ -adrenergic response appears to vary seasonally. Thus, Cossins and Kilbey (1989) showed that *in vitro* the adrenergically stimulated  $\text{H}^+$  efflux in rainbow trout (*Oncorhynchus mykiss*) red blood cells (RBCs) was gradually reduced during winter and increased abruptly early in the following spring. Nikinmaa and Jensen (1986) found that neither exercise nor adrenaline injection of rainbow trout in winter affected RBC volume or the pH gradient across the RBC membrane, in contrast to summer fish. These authors suggested that the number or affinity of the  $\beta$ -adrenergic receptors ( $\beta$ -ARs) might be seasonally reduced or

that the activity of some other step in the cascade leading to  $\text{Na}^+/\text{H}^+$  exchanger ( $\beta\text{NHE}$ ) activation might be altered. Seasonality in magnitude and temperature sensitivity of the  $\beta$ -adrenergic response has also been shown *in vitro* in RBCs of Arctic charr (*Salvelinus alpinus*; Lecklin and Nikinmaa, 1999). By contrast, Tetens et al. (1988) found no difference in the magnitude and affinity of the  $\beta$ -adrenergic response of rainbow trout RBCs *in vitro* in winter and summer fish.

The seasonal loss of  $\beta\text{NHE}$  activity is likely to impair the normal respiratory response to stress and this might negatively affect stress tolerance. Understanding the underlying mechanisms of this seasonally reduced  $\beta$ -adrenergic responsiveness and its physiological significance requires an intimate understanding of how RBC biology varies over the seasonal timescale and how this relates to circulating

hormones. Thus, a reduction in rainbow trout RBC adrenergic responsiveness and flounder (*Platichthys flesus*)  $\text{Na}^+/\text{H}^+$  exchanger activity has been circumstantially linked to the reproductive cycle (Lecklin and Nikinmaa, 1999; Weaver et al., 1999, respectively). Furthermore, plasma cortisol levels also show seasonal variation in salmonid fish (Pickering and Christie, 1981), and Reid and Perry (1991) have shown an upregulation of RBC  $\beta$ -AR numbers in response to highly elevated plasma cortisol level. Also, Perry et al. (1996) observed an increased RBC  $\beta$ NHE responsiveness when cortisol was elevated through repeated physical stress.

Thus, in order to throw light on the occurrence, duration and magnitude of seasonal downregulation of the  $\beta$ -adrenergic response and to identify the underlying mechanisms, we have undertaken an extensive monthly monitoring programme over 27 months. We have sought to relate changes in RBC adrenergic responsiveness to the number and affinity of RBC  $\beta$ -ARs and also to link changes in both to a range of other physiological parameters, including reproductive condition and circulating levels of cortisol and reproductive hormones. In particular, we have sought measures of erythropoietic activity, since Lecklin et al. (2000) have shown that immature RBCs have a higher adrenergic responsiveness than mature RBCs. Thus, we have determined the cellular amounts of transcripts for both the  $\beta$ -AR and the  $\beta$ NHE through the seasons. Finally, we employed known pharmacological activators of the  $\beta$ NHE response to probe the effectiveness during winter and summer of the transduction pathway linking  $\beta$ -AR with adrenergically activated NHE activity.

## Materials and methods

### *Animals and sampling procedures*

Rainbow trout (*Oncorhynchus mykiss* Walbaum; Stirling strain; hatched March 1997; mean mass 728 g at the start of the study) were transferred to eight 1000 litre outdoor holding tanks in October 1999, 50 fish per tank, with a constant flow ( $35 \text{ l min}^{-1}$ ) of Windermere lake water. The fish were fed commercial feed three times per week according to the manufacturer's recommendations, and their mean mass increased to 1642 g and 1882 g in December 2000 and 2001, respectively. Water temperature was electronically logged at 6-h intervals. At approximately monthly intervals, for 27 months, eight female and eight male fish (until April 2000), two fish from each holding tank, were netted into a bucket containing anaesthetic (2-phenoxyethanol; 1:2000; v/v). Blood (~8–10 ml) was sampled from the caudal vessels into heparinised syringes, transferred to polypropylene tubes and kept on ice. Between May 2000 and May 2001, a second blood sample (~1 ml), for determination of  $\beta$ -AR and  $\beta$ NHE mRNA levels, was collected into sterile syringes and transferred to sterile micro-centrifuge tubes before being stored, unwashed, at  $-80^\circ\text{C}$ . The fish were then killed by a sharp blow to the head, length and weight measured, after which the fish were pithed and sexed. The gonads were weighed in male and unovulated female fish.

### *Red blood cell $\beta$ -adrenergic receptor determination*

Freshly drawn blood was washed three times in isotonic saline ( $145 \text{ mmol l}^{-1} \text{ NaCl}$ ,  $6 \text{ mmol l}^{-1} \text{ KCl}$ ,  $1 \text{ mmol l}^{-1} \text{ MgSO}_4$ ,  $5 \text{ mmol l}^{-1} \text{ CaCl}_2$ ,  $5 \text{ mmol l}^{-1} \text{ D-glucose}$  and  $10 \text{ mmol l}^{-1} \text{ Hepes}$ , pH 7.9 and an osmolality of  $319 \pm 1 \text{ mosmol kg}^{-1}$ ; mean  $\pm$  S.E.M.;  $N=4$ ) to remove the buffy coat. The red cells were re-suspended in fresh saline at a haematocrit (Hct) of ~30%. Plasma removed after the first spin was frozen ( $-20^\circ\text{C}$ ) in aliquots (200  $\mu\text{l}$ ) for subsequent hormone and metabolite assays. High-affinity  $\beta$ -ARs were measured using CGP 12177 as a  $\beta$ -adrenergic agonist according to Marttila and Nikinmaa (1988). Briefly, duplicate samples (50  $\mu\text{l}$ ) of the RBC suspension were added to a series of tubes containing 450  $\mu\text{l}$  saline with final concentrations of 0.3–4.3  $\text{nmol l}^{-1}$  of [ $^3\text{H}$ ]CGP 12177 {[(-)-4-(3-t-butylamino-2-hydroxypropoxy)-(5,7-[ $^3\text{H}$ ]benzimidazol-2-one)]}; specific activity 1.85 TBq  $\text{mmol}^{-1}$ ; Amersham, Hertfordshire, UK} either alone (total binding,  $B_t$ ) or in the presence of excess unlabelled CGP 12177 (gift from Novartis Pharma AG, Basel, Switzerland; non-specific binding,  $B_{ns}$ ). The samples were mixed and incubated for 6 h at  $4^\circ\text{C}$ , during which time they were regularly agitated. The RBCs were counted using a haemocytometer (Weber Scientific International Ltd, Teddington, UK). At the end of the incubation period, 400  $\mu\text{l}$  of each individual sample was filtered through glass microfibre filters (2.5 cm  $\varnothing$ , retention  $\geq 1.0 \mu\text{m}$ ; Whatman, Maidstone, UK) held on a vacuum filtration manifold (Model 1225; Millipore, Watford, UK) attached to a vacuum pump. The filters were washed three times with 2 ml ice-cold saline to remove unbound ligand. The filters were placed in vials containing 5 ml of scintillation cocktail (Eco-Safe; Meridian, Epsom, UK), shaken vigorously and stored in the dark at  $4^\circ\text{C}$  for 72 h before liquid scintillation counting (Tri-Carb 2100 TR; Packard, Perkin Elmer, Seer Green, UK). The activity present in the incubation media was determined by counting of the stock standard solution activities. Counts for  $B_t$  and  $B_{ns}$  were multiplied by 1.25 to account for the fact that only 80% of the total volume of the samples was counted. Values for specifically bound ligand ( $B_s$ ) were determined by subtraction of  $B_{ns}$  from  $B_t$  and subsequently converted to  $\text{fmol tube}^{-1}$  by division by the specific activity. Receptor density ( $B_{\text{max}}$ ) and affinity ( $K_d$ ) were determined by use of non-linear regression analysis (Sigmaplot 4; Jandel Scientific, Woking, UK) of  $B_s$  as a function of free [ $^3\text{H}$ ]CGP 12177 [total ([ $^3\text{H}$ ]-CGP)– $B_t$ ].  $B_{\text{max}}$  values were then converted from  $\text{fmol tube}^{-1}$  to receptors  $\text{cell}^{-1}$  by multiplication by Avogadro's number  $\text{fmol}^{-1} \text{ cells tube}^{-1}$ .

Cortisol may influence the number of RBC membrane-bound  $\beta$ -ARs upon exposure to a stressor (Reid and Perry, 1991). To establish whether seasonal fluctuations in plasma cortisol concentrations influence the number of  $\beta$ -ARs, we incubated sub-samples of RBC suspensions under anoxic conditions at  $5^\circ\text{C}$  for 1 h prior to incubation with labelled and unlabelled ligand (July 2000–March 2001). The subsequent procedures were as described above.

*Red blood cell βNHE activity measurements*

Samples of 1.2 ml washed RBC suspension were placed in rotating Eschweiler tonometers maintained thermostatically at 15°C and equilibrated with humidified N<sub>2</sub> (anoxic condition) for 60 min. After 50 min of equilibration, ouabain (5×10<sup>-4</sup> mol l<sup>-1</sup> final concentration) was added and duplicate samples of the suspension were taken for Hct determination. The RBC suspension was transferred from the tonometer flask into polypropylene test tubes containing saline with ouabain (5×10<sup>-4</sup> mol l<sup>-1</sup> final concentration; 0.5% DMSO v/v vehicle) and <sup>22</sup>Na (~0.05 MBq ml<sup>-1</sup>) equilibrated to the same conditions as the blood cells, resulting in a 10-fold RBC dilution (this was defined as time zero). The <sup>22</sup>Na influx was measured in unstimulated cells (basal flux) and in cells stimulated by 10<sup>-5</sup> mol l<sup>-1</sup> (final concentration) isoproterenol added at 5 min.

Triplicate samples (300 μl) from control and treated RBC suspensions were taken at 5 min and 10 min. The samples were centrifuged (Model 5410; Eppendorf, Cambridge, UK) and the supernatant removed, whereupon the RBCs were washed three times in ice-cold isotonic Hepes-containing MgCl<sub>2</sub> solution (adjusted to pH 7.9). The remaining RBC pellet was lysed in 0.5 ml 0.05% Triton-X solution and deproteinised by subsequent addition of 0.5 ml 5% trichloroacetic acid. The samples were centrifuged for 2 min and 0.5 ml of the supernatant was counted (Tri-Carb 2100 TR, Packard) in 5 ml scintillation cocktail (Eco-Safe, Meridian). In addition, triplicate samples of 200 μl extracellular medium were counted for each experiment. The Na<sup>+</sup> influx (mmol Na l<sup>-1</sup> RBC h<sup>-1</sup>) was calculated as:

$$\langle \{[(X \times A_2) - (X \times A_1)] Y^{-1} \times 10^3\} (t_2 - t_1)^{-1} \rangle 60,$$

where *X* denotes μmol Na<sup>+</sup> equivalent to 1 c.p.m.; *A*<sub>1</sub> and *A*<sub>2</sub> denote <sup>22</sup>Na (c.p.m.) in triplicate samples at time 1 and 2, respectively; *Y* denotes μl packed RBCs (determined from the Hct value) and *t*<sub>1</sub> and *t*<sub>2</sub> denote time in minutes.

*Plasma cortisol, sex steroids and lactate*

Plasma cortisol, testosterone and oestradiol-17β concentrations were measured by previously validated radioimmunoassays (cortisol: Pickering et al., 1987; testosterone: Pottinger and Pickering, 1985; oestradiol-17β: Pottinger and Pickering, 1990). Plasma lactate concentration was determined enzymatically using lactate oxidase and peroxidase followed by spectrophotometric analysis (Roche, Basel, Switzerland).

*Effects of repeated disturbance stress on adrenergic responsiveness*

Eighty rainbow trout from a stock population were divided evenly between four 1000 litre outdoor holding tanks (conditions as above) and acclimated for two weeks. At the onset of the experiment, the water level in two of the tanks was dropped to ~10 cm for ~5 min once or twice daily for a two-week period. The control tanks were left undisturbed. Upon sampling, three fish from each tank (mean mass ± S.E.M.,

1773±92 g; *N*=12) were anaesthetised and blood was collected and prepared for Na<sup>+</sup> influx measurements exactly as described above. The abundance of RBC β-ARs was determined for RBC suspensions incubated under normoxic and anoxic conditions as described above.

*The effects of isoproterenol, forskolin, dibutyryl cAMP and calyculin A on RBC Na<sup>+</sup> influx*

Six rainbow trout from the stock population (mean mass ± S.E.M., 1847±83 g) were netted into anaesthetic and blood was sampled, washed and stored overnight for Na<sup>+</sup> influx measurements as described above. The Na<sup>+</sup> influx was measured as described above 5 min after addition of (1) isoproterenol (10<sup>-5</sup> mol l<sup>-1</sup>; non-selective β-AR agonist), (2) forskolin (1.5×10<sup>-4</sup> mol l<sup>-1</sup>; adenylate cyclase activator), (3) dibutyryl cAMP (10<sup>-3</sup> mol l<sup>-1</sup>; membrane-permeable cAMP analogue) or (4) calyculin A (10<sup>-7</sup> mol l<sup>-1</sup>; phosphatase inhibitor). In addition, the Na<sup>+</sup> influx was measured in unstimulated RBCs.

*βNHE activity in RBCs separated according to density (age)*

Blood was sampled, washed and stored as described above from rainbow trout (Shasta strain; mean mass ± S.E.M., 937±68 g; *N*=6) during October–November 2001. This treatment assured that the RBC membrane transporters and cell volume were in an unstimulated steady-state condition (Bourne and Cossins, 1982). After overnight storage, the RBCs were washed once and re-suspended in saline containing 1% bovine serum albumin (BSA) at an Hct of approximately 80%. The RBCs were separated into age fractions by fixed-angle (30°) centrifugation (10 000 g, 4°C, 15 min) in narrow tubes (diameter 4 mm, length 45 mm, volume 0.5 ml; Speckner et al., 1989; optimized for trout by Phillips et al., 2000). The youngest, least dense, cells are located in the top layer, whereas the older, more dense cells are located in the middle and bottom of the tube. The tubes were cut into a top (24±2%), middle (60±3%) and bottom (15±1%) fraction, containing RBCs of increasing age, which were washed in isotonic saline three times to remove BSA. The mean cellular haemoglobin concentration (MCHC), determined as [Hb]/Hct, was used to verify that the cells were separated according to age. MCHC is lower in younger cells than in older cells (Speckner et al., 1989; Lund et al., 2000). Adrenergically stimulated Na<sup>+</sup> influx was measured in each of the different cell fractions and also in the original unseparated population of RBCs.

*RBC βNHE and β3b receptor mRNA determinations*

Total RNA was isolated from frozen tissue by homogenisation in guanidinium thiocyanate (Chomczynski and Sacchi, 1987) using Trizol Reagent (Invitrogen, Ontario, Canada). After treatment with DNase I (5 units per μg RNA; Invitrogen) to remove any remaining genomic DNA, the quality of the RNA was assessed by gel electrophoresis. cDNA was synthesised from 1–2 μg RNA using random hexamer primers and Superscript II reverse transcriptase (Invitrogen). Previous studies (Nickerson, 2003) have demonstrated that the

trout RBC  $\beta$ -AR most closely resembles the  $\beta_3$ -AR of mammals or the  $\beta_{4c}$ -AR of turkey RBCs (Chen et al., 1994). Because the trout RBC  $\beta$ -AR is found exclusively in the blood, it has been termed  $\beta_{3b}$ -AR (Nickerson et al., 2003).  $\beta_{3b}$ -AR or  $\beta$ NHE mRNA levels were assessed by Q-PCR on duplicate samples of cDNA (1  $\mu$ l) using a Hot StarTaq Master Mix kit (Qiagen, Ontario, Canada) and a Stratagene (West Cedar Creek, TX, USA) MX-4000 multiplex quantitative PCR system. CYBR green (Molecular Probes Inc., Boulder, CO, USA) and ROX (Stratagene) were used as DNA and reference dyes, respectively. The PCR conditions (final reaction volume=20  $\mu$ l) were as follows: cDNA template=1  $\mu$ l; forward and reverse [primer]=150 pmol l<sup>-1</sup>; [Mg<sup>2+</sup>]=2.0 mmol l<sup>-1</sup>; CYBR green=1:50 000 final dilution; ROX=1:30 000 final dilution; dNTP=200  $\mu$ mol l<sup>-1</sup>. The annealing and extension temperatures over 40 cycles were 58°C (45 s) and 72°C (60 s), respectively. The following primer pairs were designed using Primer3 software ([http://www-genome.wi.mit.edu/cgi-bin/primer/primer3\\_www.cgi](http://www-genome.wi.mit.edu/cgi-bin/primer/primer3_www.cgi)):

actin forward 5'-CAC CAT GAA GAT CAA GAT CAT YGC-3'

actin reverse 5'-ATT TRC GGT GGA CGA TGG AG-3'

$\beta_{3b}$  forward 5'-CTT GGG CTA TGG TGG CAG TA-3'

$\beta_{3b}$  reverse 5'-CCA TGA TAA TGC CCA AGG TC-3'

$\beta$ NHE forward 5'-GGG TAA TGC GTC AGA CAA CC-3'

$\beta$ NHE reverse 5'-CCA TGA TAA TGC CCA AGG TC-3'

The specificity of the primers was verified by cloning (TOPO TA cloning kit; Invitrogen) and sequencing of the amplified products. To ensure that CYBR green was not being incorporated into primer dimers or non-specific amplicons during the Q-PCR runs, PCR products were analysed by gel electrophoresis in initial experiments; single bands of expected size were obtained in all instances. Furthermore, the

construction of CYBR green dissociation curves after completion of 40 PCR cycles revealed the presence of single amplicons for each primer pair. Relative expression of mRNA levels was determined (using actin as an endogenous standard) by a modification of the delta-delta Ct method (Pfaffl, 2001). Amplification efficiencies were determined from standard curves generated by serial dilution of plasmid DNA.

#### Data presentation and statistics

Throughout, the data are presented as means  $\pm$  S.E.M. The data sets were analysed using analysis of variance (ANOVA; Genstat 5, Lawes Agricultural Trust, Harpenden, UK) with, for the seasonal study, individual fish, tank and time as factors; for the implant studies, fish, tank and treatment as factors; and for the separated cell study, fish and cell fraction as factors. Significant differences between times, treatment groups or fractions were determined using the estimated standard error of the differences between means. Where mean and variance were found to be interdependent, the data were log-transformed prior to analysis. Sigma Plot 4.0 was used to assess statistical significance of fitted linear and non-linear regressions.

#### Results

Only data for female fish are presented. Because of differential mortality rates during the previous spawning season (male rainbow trout tend to mature one year earlier than females and are more vulnerable to fungal infections when sexually mature), sufficient numbers of male fish were available only for the initial five samples. Over the five-month period, which included one sample with a reduced  $\beta$ -adrenergic responsiveness (December 1999; Fig. 2), we observed no statistically significant sex-related differences in  $\beta$ -AR characteristics or  $\beta$ NHE activity. Maximum androgen and estrogen levels occurred at the same time in males as in female fish (data not shown). These limited data suggest that our observations of female fish are also valid for male fish.

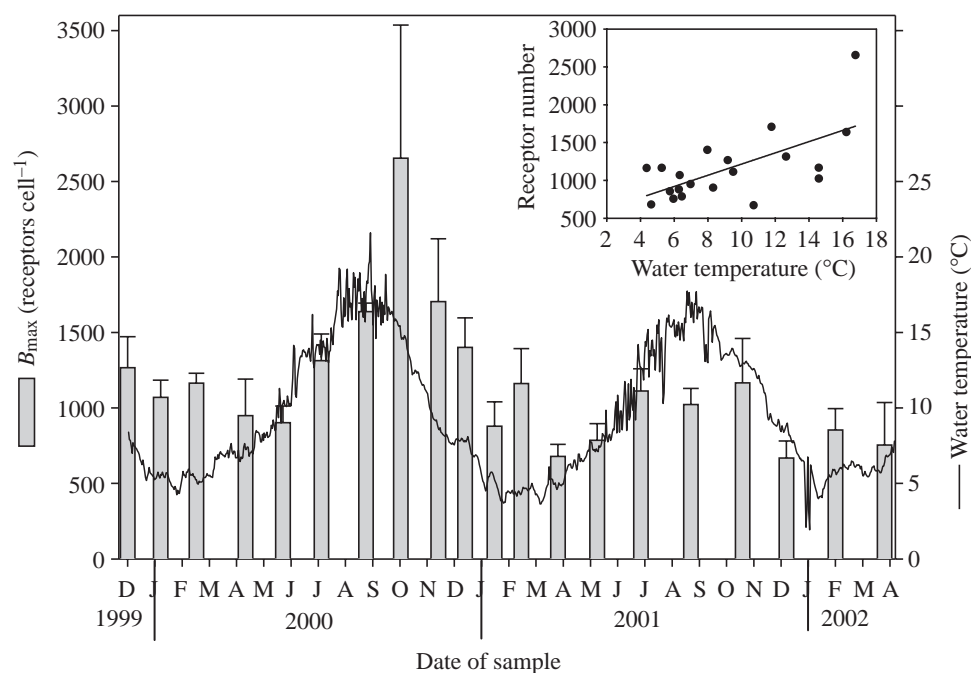


Fig. 1. Seasonal variation in the number of  $\beta$ -adrenergic receptors ( $\beta$ -ARs; grey bars) and daily ambient water temperature (black line) throughout the study. Values are means  $\pm$  S.E.M. ( $N=8$ ). The inset shows the linear regression curve for  $\beta$ -AR number vs mean water temperature ( $P=0.002$ ,  $f(x)=474.7+74.1x$ ,  $r^2=0.417$ ). Water temperature in the inset was calculated from the temperatures measured over the full period since the previous sampling.

### Seasonal variations in RBC $\beta$ -AR characteristics and $\beta$ NHE activity

Fig. 1 presents the time course over 27 months of  $\beta$ -AR numbers together with the measured water temperatures. The number of  $\beta$ -ARs varied seasonally from a minimum of  $668 \pm 112$  receptors cell<sup>-1</sup> to a maximum of  $2654 \pm 882$  receptors cell<sup>-1</sup> ( $N=8$ ; Fig. 1). During late summer 2000, the  $\beta$ -AR number increased, remained high until late autumn and then decreased into the winter. A similar, though less pronounced, rhythm was evident in 2001 (Fig. 1). Throughout this period, the  $\beta$ -AR number was positively related in a linear fashion to the average ambient water temperature ( $P=0.002$ ,  $r^2=0.417$ ; Fig. 1 inset). The equilibrium dissociation constant ( $K_D$ ) for CGP binding to the RBC  $\beta$ -ARs varied between  $0.31 \pm 0.08$  nmol l<sup>-1</sup> and  $1.35 \pm 0.26$  nmol l<sup>-1</sup> ( $N=8$ ; not shown) but did not correlate with the ambient water temperature (linear regression:  $P=0.77$ ).

Reid and Perry (1991) showed that cortisol applied through an osmotic pump can pre-adapt the RBCs to cope with a stressor by elevating the number of membrane-bound  $\beta$ -ARs, and this potentially increases adrenergic responsiveness. During our routine procedures for measuring  $\beta$ -adrenergic responsiveness we also stressed the RBCs by incubating cells throughout under anoxic conditions. We therefore examined whether seasonal fluctuations in cortisol, with elevated levels during the spawning periods (Fig. 2), influenced the number or affinity of functional  $\beta$ -ARs when incubated under oxygenated or anoxic conditions. There was, however, no statistical difference in  $\beta$ -AR characteristics (number and affinity) between oxygenated and anoxically treated RBC suspensions during the entire period between July 2000 and March 2001 (data not shown).

The mean isoproterenol-stimulated Na<sup>+</sup> influx was  $110.4 \pm 2.3$  mmol l<sup>-1</sup> RBCs h<sup>-1</sup> ( $N=128$ ) during the months where the  $\beta$ -adrenergic response was not reduced (Fig. 2). A downregulated response was arbitrarily defined as any flux lower than  $85$  mmol l<sup>-1</sup> RBCs h<sup>-1</sup>. In December 1999 and February 2001, the  $\beta$ NHE activity was reduced by 57% and 34%, respectively, compared with the average value, whereas during the third winter of the study there was no statistically significant reduction in the  $\beta$ NHE activity (Fig. 2). Surprisingly, there was no obvious dependence of the magnitude of the Na<sup>+</sup> influx on the number (linear regression:

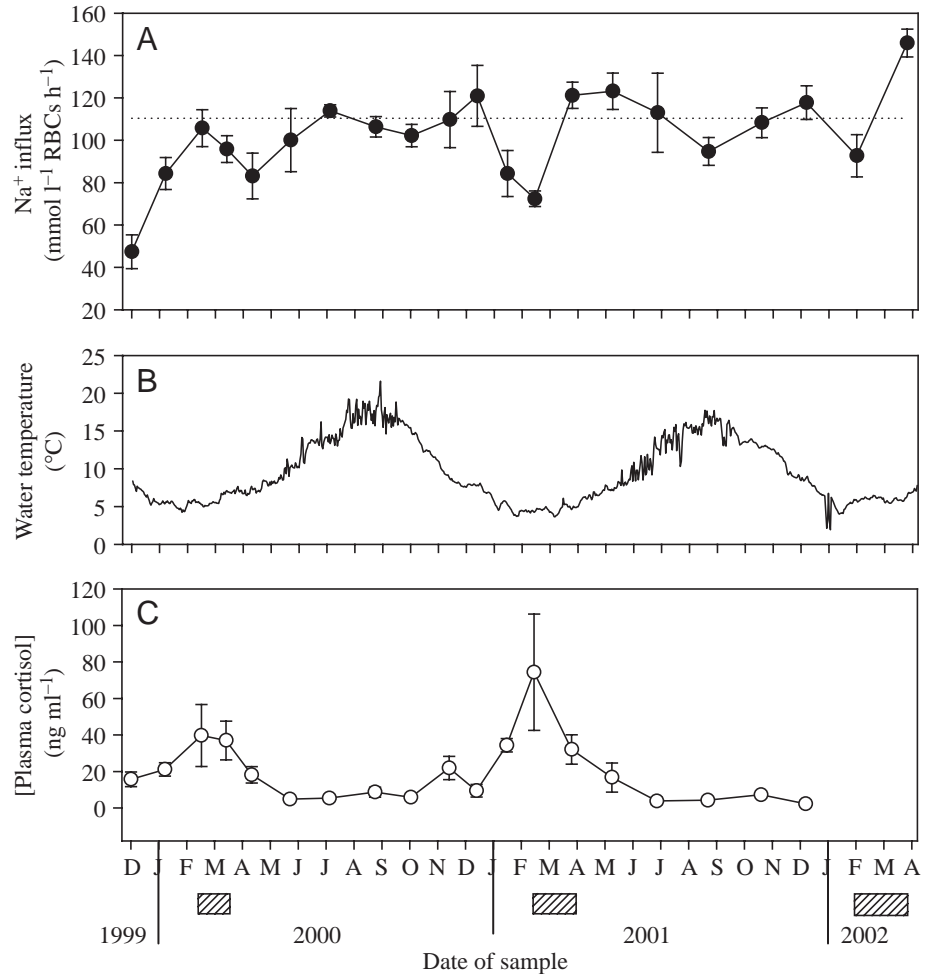


Fig. 2. Seasonality in the  $\beta$ -adrenergically stimulated red blood cell (RBC) Na<sup>+</sup> influx (A), daily ambient water temperature (B) and plasma cortisol levels (C). The dotted line in A illustrates the mean Na influx at the times of the year when the  $\beta$ -NHE activity is not downregulated. The crosshatched boxes on the x-axis indicate the spawning periods, defined by high proportions of ovulated females. Values are means  $\pm$  S.E.M. ( $N=8$ ).

$P=0.83$ ;  $N=163$ ) or affinity (linear regression:  $P=0.68$ ;  $N=163$ ) of the  $\beta$ -ARs. These data suggest that the seasonal changes in  $\beta$ NHE activity do not reflect changes in the binding characteristics of the hormone to the receptors. Also, the Na<sup>+</sup> influx did not correlate with ambient water temperature (linear regression:  $P=0.73$ ; non-linear regression:  $P=0.72$ ), i.e. the temperature at which the fish were acclimated, which agrees with the results obtained by Cossins and Kilbey (1989).

### Effects of repeated disturbance stress on $\beta$ -adrenergic responsiveness

RBC  $\beta$ -AR density has been shown to decrease in response to chronically elevated plasma catecholamine levels (Gilmour et al., 1994). We exposed fish to repeated daily stress over a 2-week period in January 2001, when the  $\beta$ NHE responsiveness was normal, to elevate plasma catecholamine levels and test if unidentified stressors in the holding facilities would affect the  $\beta$ -adrenergic responsiveness. The  $\beta$ NHE

Table 1.  $\beta$ -adrenergically stimulated  $\text{Na}^+$  influx in RBCs with normal or seasonally reduced  $\beta\text{NHE}$  activity

Compound	Normal $\beta\text{NHE}$ activity	Reduced $\beta\text{NHE}$ activity
Isoproterenol	100	100
Forskolin	97 $\pm$ 15	82 $\pm$ 6
Dibutyryl cAMP	52 $\pm$ 11	45 $\pm$ 19
Calyculin A	61 $\pm$ 11	45 $\pm$ 15

Values are  $\text{Na}^+$  influxes relative to the  $\text{Na}^+$  influx in isoproterenol-stimulated red blood cells (RBCs) with normal or seasonally reduced  $\text{Na}^+/\text{H}^+$ -exchanger ( $\beta\text{NHE}$ ) activity.  $N=3$  for each experiment.

activity was not affected by repeated physical stress, with  $\text{Na}^+$  influx values of 129.7 $\pm$ 4.7 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> and 128.8 $\pm$ 7.4 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> in control and disturbed fish, respectively.

#### Comparison of the effects of isoproterenol, forskolin, dibutyryl cAMP and calyculin A

Having established that the seasonal reduction in  $\beta\text{NHE}$  activity was not linked to seasonal changes in  $\beta$ -AR number or affinity, we looked for other causal factors. Binding of catecholamine to the membrane-bound  $\beta$ -AR triggers an adenylate cyclase-catalysed synthesis of cAMP (Mahe et al., 1985), which in turn stimulates protein kinase A to activate the  $\beta\text{NHE}$  (Guizouarn et al., 1993). Accordingly, we tested whether the cause of the seasonally reduced adrenergic responsiveness lay in the transduction pathway linking the receptor with the transporting effector using chemical

compounds known to stimulate the  $\beta\text{NHE}$  by intervening at different points on a possibly long transduction pathway. None of the compounds stimulated the  $\text{Na}^+$  influx to a greater extent than isoproterenol (data not shown). More significantly in the present context, the extent to which these compounds stimulated the  $\beta\text{NHE}$  relative to stimulation by isoproterenol was similar in RBCs with a seasonally reduced response compared with RBCs with a normal response (Table 1). Thus, the reduced isoproterenol-induced  $\beta\text{NHE}$  activity could not be rescued by any of the transduction activators, indicating that the seasonal diminution of  $\beta\text{NHE}$  activity was not linked to any of the steps lying between the  $\beta$ -AR and the  $\beta\text{NHE}$ . This together with the absence of any changes in the binding affinity and number of  $\beta$ -ARs, suggests that the winter loss of  $\beta\text{NHE}$  activity was linked to changes in the  $\beta\text{NHE}$  itself.

#### Red blood cell $\beta\text{NHE}$ and $\beta 3b$ mRNA levels

The seasonal variation in  $\beta 3b$  mRNA levels is shown in Fig. 3. The relative quantity of  $\beta 3b$  mRNA was more or less constant between May and October, decreased markedly by ~90% between October and December and remained at low levels between December and May. The data showed that the level of  $\beta 3b$  mRNA, like the receptor number per cell, was correlated in a linear fashion with the ambient water temperature ( $P=0.01$ ,  $r^2=0.632$ ) and also that the number of receptors depended in a linear fashion ( $P=0.005$ ,  $r^2=0.755$ ) on the level of  $\beta 3b$  mRNA present in the RBCs (Fig. 3 inset).

Fig. 4 illustrates the corresponding changes in  $\beta\text{NHE}$  mRNA. The relative quantity increased between May and November, decreased markedly between November and December and increased again progressively between December and May (Fig. 4). Examination of the levels of  $\beta\text{NHE}$  mRNA at different times of the year showed that the  $\beta\text{NHE}$  activity had a one-sampling time delayed hyperbolic dependency on the mRNA level ( $P=0.038$ ;  $r^2=0.538$ ) present in the cell (Fig. 4 inset).

#### $\beta\text{NHE}$ activity of age-separated RBCs

After separation, the top layer contained 23.9 $\pm$ 2.9%, the middle layer 61.5 $\pm$ 3.0% and the bottom layer 14.7 $\pm$ 1.1% of the cells ( $N=6$ ). The values for MCHC are presented in Table 2. MCHC increased from the top to the bottom fractions of RBCs, showing that the cells had been separated according to age. Based on measurements for the individual fractions, MCHC of the unseparated RBCs suspension was calculated to be 4.19 mmol l<sup>-1</sup>, which is within the 0.05 significance limits of the measured value. The activity of the

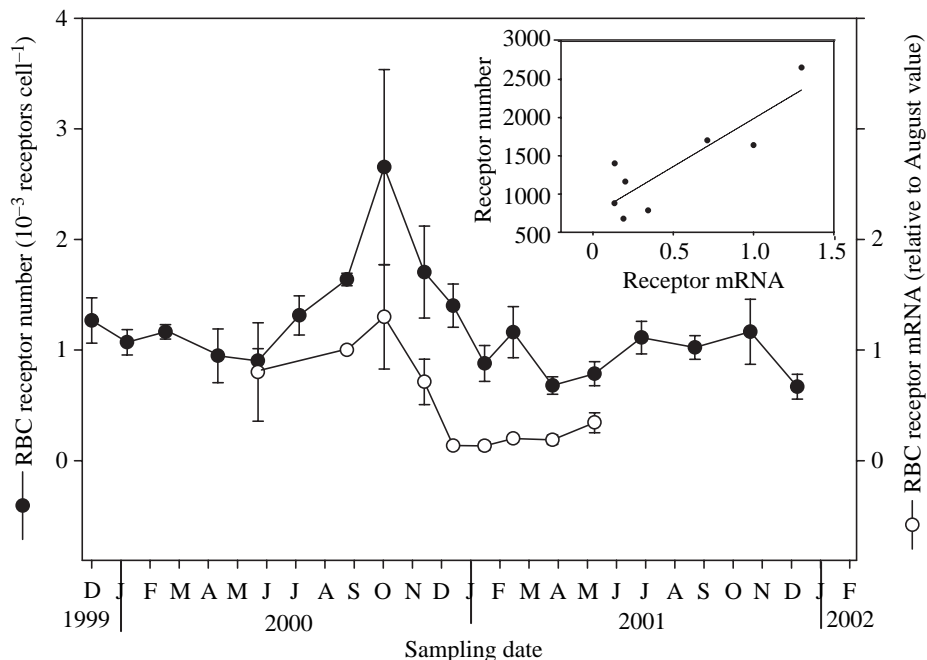


Fig. 3. The seasonal variations in  $\beta$ -adrenergic receptor ( $\beta$ -AR) numbers (filled circles) and  $\beta 3b$ -AR mRNA levels (relative to the August value; open circles). Values are means  $\pm$  S.E.M. ( $N=8$  for  $\beta$ -AR numbers and  $N=3-8$  for  $\beta 3b$ -AR mRNA values). The inset illustrates the linear regression curve for receptor number vs  $\beta 3b$ -AR mRNA levels ( $P=0.005$ ,  $f(x)=734.5+1252.8x$ ,  $r^2=0.755$ ).

βNHE of the top fraction RBCs was 23.6 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> and 35.5 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> higher than that of the middle and bottom fractions, respectively (*P*<0.01 and *P*<0.05, respectively; Table 2).

*Seasonal variations in plasma oestradiol, testosterone and lactate*

The seasonal fluctuations in plasma sex steroids in female fish within the experimental population are illustrated together with the βNHE activity in Fig. 5. Plasma testosterone levels in the female fish increased steeply just prior to or concomitantly with the observed major changes in the RBC βNHE activity (Fig. 5). It is noticeable that the maximal levels of plasma testosterone observed in successive years decreased gradually and in a statistically significant manner; i.e. in the third year of the study, the plasma testosterone level was reduced compared with the level in the first (*P*<0.01) and second (*P*<0.001) year of the study. Plasma oestradiol levels varied in a very predictable manner, i.e. the maximal levels and the timing of the increases and decreases were similar each year (Fig. 5).

Plasma lactate varied seasonally throughout the study, with minimal values of 1.13±0.11 mmol l<sup>-1</sup> (*N*=16) in February and maximal values of 2.54±0.24 mmol l<sup>-1</sup> (*N*=16) in August. The response to β-adrenergic stimulation, which lowers the risk of anoxic metabolism and lactic acid production, was also lowest in the winter (see above). The lactate data, therefore, indicate that, despite the reduction in β-adrenergic responsiveness, the oxygen uptake from the water was entirely sufficient to prevent activation of anaerobic metabolism under conditions of routine activity in the well-aerated holding facilities.

**Discussion**

*Seasonal variations in βNHE activity*

In agreement with previous studies (Cossins and Kilbey, 1989), the magnitude of the RBC β-adrenergic response, as indicated by βNHE activity measured under standard conditions, varied seasonally. Responses declined significantly below normal levels during some winter months and recovered during the early spring. As before, the spring recovery occurred prior to any increase in ambient temperature (Cossins and Kilbey, 1989). However, the timing of the downregulation was not consistent over the life of the experimental trout population; during the first winter the response was minimal in December, 2–3 months prior to spawning, whereas during the following winter the maximal reduction coincided exactly

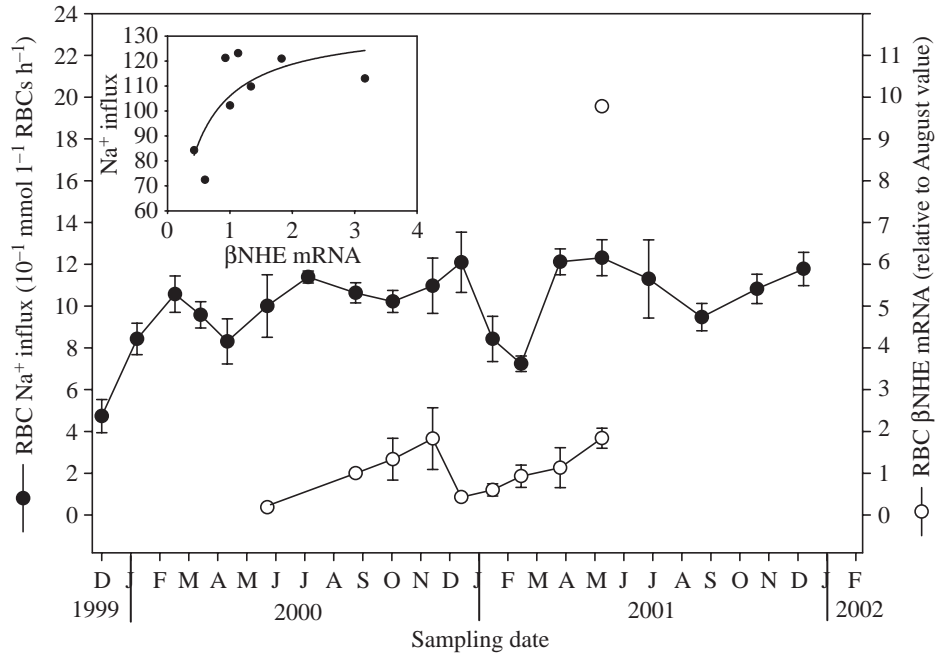


Fig. 4. Seasonal changes in the β-adrenergically stimulated RBC Na<sup>+</sup>/H<sup>+</sup> exchanger (βNHE) activity (filled circles) and βNHE mRNA levels (relative to August value; open circles). Note one outlying βNHE mRNA value is plotted separately. Values are means ± S.E.M. (*N*=8 for βNHE activity values and *N*=3–8 for βNHE mRNA values). The inset shows the linear regression curve for βNHE activity vs βNHE mRNA values (*P*=0.038, *f*(*x*)=135.0*x*/(0.274+*x*), *r*<sup>2</sup>=0.538).

with the spawning period in February. The inconsistency in timing of the reduced βNHE activity was somewhat unexpected but is in accordance with the timing of peaks in plasma testosterone levels (see below). One of the factors that varied during our study was the age of the fish. To our knowledge, no studies have previously looked into possible age-related variations in timing of parameters involved in the reproductive cycle or the stress response. Also, the magnitude of reduction varied during the study; thus, it was larger (57% of the mean βNHE activity) during the first winter than during the following winter (34% of the mean βNHE activity) whilst there was no significant reduction in the third winter. In part, this may be explained by our sampling point not coinciding

Table 2. βNHE activity and mean cellular haemoglobin concentration (MCHC) measured in density-separated red blood cells (RBCs) and in the original RBC suspension

Cell fraction	Na <sup>+</sup> influx (mmol l <sup>-1</sup> RBCs h <sup>-1</sup> )	MCHC (mmol l <sup>-1</sup> )
Top (young)	125.6±9.5	3.82±0.16
Middle (older)	90.1±8.5**	4.24±0.14*
Bottom (oldest)	102.0±7.7*	4.57±0.10***
Original RBC suspension	91.0±4.9**	4.10±0.18*

Statistically significant differences from values for the top fraction are indicated by \**P*<0.05, \*\**P*<0.01 and \*\*\**P*<0.001. *N*=6 for each experiment.

exactly with time of the maximal reduction but might represent a trend that is associated with increasing age or size.

In considering the magnitude of the winter reduction in  $\beta$ -adrenergic responsiveness, it is worth pointing out that the trout RBC  $\beta$ NHE activity is markedly temperature dependent, with a  $Q_{10}$  of 7.9 over the temperature range 0–19°C (Cossins and Kilbey, 1990). Thus, assay of  $\beta$ NHE activity *in vitro* at a standard temperature of 15°C irrespective of seasonally varying environmental temperature must, therefore, profoundly underestimate the seasonal fluctuation in the *in vivo* RBC  $\beta$ -adrenergic response. Thus, the flux of 72 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> measured *in vitro* in February 2001 at 15°C would be reduced *in vivo* to ~9 mmol l<sup>-1</sup> RBCs h<sup>-1</sup> at the prevailing water temperature at that time.

Repeated physical stress and confinement both elevate plasma catecholamine and cortisol levels, with opposite effects on the  $\beta$ -adrenergic response (Perry et al., 1996). Thus, exposure of rainbow trout to daily physical stress reduced the number of membrane-bound  $\beta$ -ARs determined both under

normoxic and hypoxic conditions but elevated adrenergic responsiveness due to increased  $\beta$ -AR affinity (Perry et al., 1996). We therefore considered the possibility that (unidentified) seasonal differences in the degree of stress to which the experimental fish were exposed could have influenced RBC adrenergic responsiveness and contributed to the seasonal variation in RBC function observed during this study. However, at a time of the year when the  $\beta$ -adrenergic response was normal, exposure of the fish to a regime of repeated daily disturbance stress did not change the  $\beta$ NHE responsiveness. The seasonal reductions in adrenergic responsiveness, therefore, seem to occur independently of increased stress levels.

An obvious factor that might underlie the seasonal variations in RBC  $\beta$ -adrenergic response is the effectiveness of the cellular  $\beta$ -AR system. Previous work has implicated altered  $\beta$ -AR numbers in the enhanced response to hypoxia (Reid and Perry, 1991; Marttila and Nikinmaa, 1988); however, the fluctuations in  $\beta$ -AR numbers or affinity in the

present study were not in any way related to the seasonal changes in  $\beta$ NHE activity. An alternative explanation is that there was some impairment during winter in the effectiveness of the transduction pathway linking the  $\beta$ -AR with the  $\beta$ NHE. We stimulated the  $\beta$ NHE in a number of receptor-independent ways, using forskolin and calyculin A, which activate the  $\beta$ NHE by effects on adenylate cyclase and protein phosphatase, respectively (Seamon et al., 1981; Guizouarn et al., 1995), and the membrane-permeable cAMP analogue dibutyryl cAMP. We compared the effect of these compounds on the  $\beta$ NHE activity of winter fish with reduced  $\beta$ -adrenergic responsiveness with those in summer fish with the normal high  $\beta$ -adrenergic responsiveness. If winter suppression were due to impairment at a specific step in this pathway we would expect a larger effect of pharmacological activation at a downstream step, relative to the effects of the  $\beta$ -adrenergic agonist isoproterenol acting alone. We found that all three compounds stimulated the  $\beta$ NHE to the same extent in RBCs with a reduction in  $\beta$ NHE activity compared with RBCs with a normal response. This indicates no rescue of  $\beta$ -adrenergic response in RBCs of winter fish by downstream activation; the red cells from animals collected at different times of the year behaved identically with respect to transduction manipulation when tested under common experimental conditions.

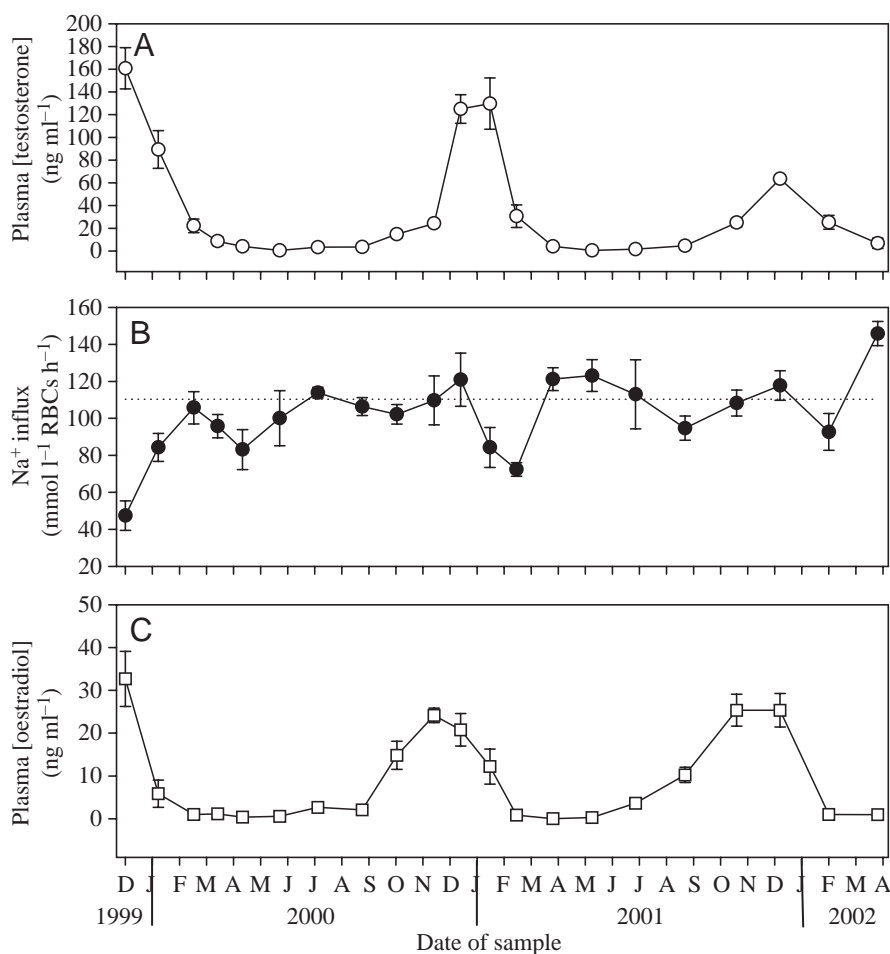


Fig. 5. Seasonal fluctuations in plasma testosterone (A) and plasma oestradiol-17 $\beta$  concentrations (C) in relation to fluctuations in red blood cell (RBC)  $\beta$ -adrenergically stimulated RBC Na<sup>+</sup>/H<sup>+</sup> exchanger ( $\beta$ NHE) activity (B). The dotted line illustrates the mean Na<sup>+</sup> influx at the times of the year when the  $\beta$ -NHE activity is not downregulated. Values are means  $\pm$  S.E.M. ( $N=8$ ).



Whilst we cannot exclude the possibility that there is a critical step lying downstream of the calyculin A-sensitive protein phosphatase, we suggest that the limiting step during winter months in the  $\beta$ -adrenergic response was not upstream to the  $\beta$ NHE. Given that it was not due to changes in  $\beta$ -AR numbers, we therefore conclude that it was due to variations in the number or affinity of the exchanger itself or properties of its microenvironment.

The linkage between seasonal changes in  $\beta$ -adrenergic responsiveness and reduced expression of the  $\beta$ NHE is supported by measurements of  $\beta$ NHE transcript expression. We found that transcript amounts declined in the autumn and increased in the spring, the changes in the mRNA levels preceding by one sampling period the spring increase in  $\beta$ NHE activity. The changes in  $\beta$ NHE mRNA levels could be caused either by seasonal effects on the transcriptional activity in the circulating population of RBCs or by seasonality in erythropoietic activity coupled with age-dependent changes in RBC transcriptional activity. Age-dependent reductions in mRNA levels have been reported for carbonic anhydrase and the Band 3 anion exchanger in rainbow trout RBCs (Lund et al., 2000). At the physiological level, Lecklin et al. (2000) showed that the volume increase following  $\beta$ -adrenergic stimulation was considerably lower in mature than in immature RBCs, suggesting lower  $\beta$ NHE activity in the older cells. Consistent with this observation, age-dependent reductions in enzyme activity levels have been reported for citrate synthase, cytochrome oxidase, lactate dehydrogenase and pyruvate kinase in rainbow trout RBCs (Phillips et al., 2000). Age-dependent decreases in activity levels therefore seem to be widespread at both the transcriptional and functional levels. Erythropoietic activity and the release of newly synthesized RBCs into the circulation is reduced in rainbow trout during the winter (Lane, 1979), and in Baltic salmon (*Salmo salar*) the proportion of immature cells in circulation decreases during the winter (Härdig and Höglund, 1984). Seasonal changes in the age profile of the circulating RBCs make the age dependency of the RBC  $\beta$ -adrenergic volume response and enzyme activities of great interest in the interpretation of seasonal changes in RBC function. We show with density-separated RBCs that the  $\beta$ NHE activity of the top (youngest RBCs) of the separated fractions was significantly higher than that of the middle and bottom fractions, containing older RBCs, which is consistent with the changing age profile of circulating RBCs underpinning the seasonally reduced  $\beta$ -adrenergic responsiveness. In the present study, we do not have any measure of the erythropoietic activity. However, previous work in the same fish-holding facilities at CEH Windermere showed that the circulating RBC number decreased during autumn and winter in mature females of the closely related brown trout *Salmo trutta* (Pottinger and Pickering, 1987). The critical factor might be influence of low water temperatures during late autumn and winter upon erythropoietic activity (Hevesy et al., 1964), progressively reducing the proportion of young RBCs with a high  $\beta$ -adrenergic responsiveness in circulation.

#### *Linking circulating hormones and erythropoietic activity with the $\beta$ NHE activity*

Elevated levels of plasma oestradiol were observed prior to and concomitant with the seasonally reduced  $\beta$ NHE activity in December 1999 and January–February 2001. It remains to be investigated whether seasonal elevations of plasma oestradiol play any role in the  $\beta$ -adrenergic response of the circulating RBCs.

Testosterone is known to stimulate erythropoiesis in humans (Barcelo et al., 1999; Snyder et al., 2000) and birds (Jones and Johansen, 1972; Robinzon and Rogers, 1979; Thapliyal et al., 1982) and is suspected to have the same effect in fish (Pottinger and Pickering, 1987). Thus, we suggest that the increased testosterone levels observed during the spawning period in the present work may, by stimulating erythropoiesis, have promoted the rapid increase in adrenergic responsiveness during winter or early spring, the increasing proportion of immature RBCs with increased  $\beta$ NHE activity explaining the enhanced  $\beta$ -adrenergic responsiveness. The increases in  $\beta$ -adrenergic responsiveness were observed 1–2 sampling periods after the increases in plasma testosterone and decreases in  $\beta$ NHE activity. This delay is consistent with the lengthy lag time known to precede the entry of immature RBCs into the circulation (Lecklin et al., 2000).

#### *Seasonality in the $\beta$ -adrenergic receptor characteristics*

We provide the first evidence linking, in a linear fashion, the abundance of a receptor, i.e. the  $\beta$ -AR, and its cognate transcript with the ambient water temperature. The spring increase in water temperature was closely associated with an increase in  $\beta$ 3b-receptor mRNA and subsequently to an increase in the number of functional receptors. This seems appropriate, as oxygen consumption increases with a rise in water temperature, and stimulation of the  $\beta$ -AR optimizes oxygen transport (Nikinmaa, 1992). It is paradoxical, therefore, that the increase in receptor  $\beta$ -AR numbers did not elevate  $\beta$ NHE activity and, consequently, in functional terms, does not seem to improve oxygen transport. No other parameter monitored during the present study co-varied with the  $\beta$ 3b mRNA levels and  $\beta$ -AR numbers. However, we cannot exclude the idea that the influence of temperature on  $\beta$ 3b mRNA levels and  $\beta$ -AR numbers was indirect. How would the lower ambient temperatures routinely experienced by fish in Northern USA, Canada and Scandinavia (0–2°C) affect seasonal fluctuations in  $\beta$ -ARs? Extrapolation of the linear relationship between  $\beta$ -AR numbers and ambient temperature to 1°C predicts 549 receptors cell<sup>-1</sup>. This might be entirely sufficient to initiate events leading to  $\beta$ NHE activation. Indeed, the Windermere fish included specimens with very low  $\beta$ -AR numbers yet with powerfully expressed  $\beta$ NHEs, suggesting that exceptionally low winter temperatures would not necessarily incur greater reductions in  $\beta$ NHE activity.

Experimental elevation of plasma cortisol is able to increase  $\beta$ -AR density, probably by increasing transcription of the  $\beta$ 3b-receptor gene. This enlarges the pool of cytosolic, physiologically inactive  $\beta$ -receptors, which can be mobilised

to the plasma membrane on exposure to stress (Reid and Perry, 1991). Whilst we found considerable circannual fluctuations in plasma cortisol, with high levels during February–March, we failed to observe changes in membrane-bound  $\beta$ -ARs in anoxically treated RBCs compared with control RBCs. Cortisol therefore appears to play no role in the seasonal variation of RBC adrenergic responsiveness.

### Conclusions

We showed that the number of RBC  $\beta$ -ARs is linearly related both to the level of  $\beta$ -AR transcript and the ambient water temperature. Naturally occurring seasonal fluctuation in  $\beta$ -AR numbers was not related to the more inconsistent fluctuations in  $\beta$ NHE activity. On the other hand,  $\beta$ NHE activity was positively related to the  $\beta$ NHE transcript level and depended on the age of the RBC, showing reduced activity in older RBCs. The seasonally reduced  $\beta$ NHE activity could not be rescued by pharmacological intervention in the transduction cascade, suggesting that fluctuations in  $\beta$ NHE activity were a property of the transporter itself rather than any other factor. The inconsistency in timing of the reductions in  $\beta$ NHE activity was matched by corresponding fluctuations in plasma testosterone. Since testosterone stimulates erythropoietic activity in birds and humans, we suggest that the seasonally occurring fluctuations in  $\beta$ NHE activity of trout red cells are caused by changes in the age profile of circulating RBCs. We further hypothesise that this is due to the seasonal influences of testosterone.

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