

## RESEARCH REPORT

# Conserved regulation of Nodal-mediated left-right patterning in zebrafish and mouse

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## ABSTRACT

Nodal is the major effector of left-right axis development. In mice, Nodal forms heterodimers with Gdf1 and is inhibited by Cerl2/Dand5 at the node, and by Lefty1 in the lateral plate mesoderm (LPM). Studies in zebrafish have suggested some parallels, but also differences, between left-right patterning in mouse and zebrafish. To address these discrepancies, we generated single and double zebrafish mutants for *southpaw* (*spaw*, the *Nodal* ortholog), *dand5* and *lefty1*, and performed biochemical and activity assays with Spaw and Vg1/Gdf3 (the Gdf1 ortholog). Contrary to previous findings, *spaw* mutants failed to initiate *spaw* expression in the LPM, and asymmetric heart looping was absent, similar to mouse *Nodal* mutants. In blastoderm assays, Vg1 and Spaw were interdependent for target gene induction, and contrary to previous results, formed heterodimers. Loss of *Dand5* or *Lefty1* caused bilateral *spaw* expression, similar to mouse mutants, and *Lefty1* was replaceable with a uniform Nodal signaling inhibitor. Collectively, these results indicate that *Dand5* activity biases Spaw-Vg1 heterodimer activity to the left, Spaw around Kupffer's vesicle induces the expression of *spaw* in the LPM and global Nodal inhibition maintains the left bias of Spaw activity, demonstrating conservation between zebrafish and mouse mechanisms of left-right patterning.

**KEY WORDS:** Left-right patterning, Nodal, Southpaw, Cerl2, Dand5, Lefty

## INTRODUCTION

On its surface, the vertebrate body plan appears bilaterally symmetric, but the morphology and positioning of internal organs reveal an underlying left-right asymmetry: the heart jogs to the left, the liver is positioned on the right and the gut undergoes asymmetric rotation. Nodal, a ligand in the TGFβ protein family, is a major regulator of left-right axis establishment (Blum and Ott, 2018; Blum et al., 2014; Grimes and Burdine, 2017). After its role in mesendoderm patterning, *Nodal* is expressed in the embryonic node and in the left lateral plate mesoderm (LPM) in mouse (Collignon et al., 1996; Lowe et al., 1996; Zhou et al., 1993), zebrafish (Long et al., 2003), chick (Levin et al., 1995; Pagán-

Westphal and Tabin, 1998) and *Xenopus* (Lowe et al., 1996), and it is expressed asymmetrically in *Amphioxus* (Li et al., 2017), snails (Grande and Patel, 2009; Kuroda et al., 2009), echinoderms (Duboc et al., 2005) and ascidians (Morokuma et al., 2002).

The mechanisms underlying left-right patterning are well understood in mice, but less established in zebrafish, owing to a reliance on morpholino knockdown phenotypes and an absence of null mutants. Knockdown of *southpaw* (*spaw*, a zebrafish *Nodal* ortholog) results in an absence of *spaw* in the LPM, and subsequent heart jogging and looping phenotypes (Long et al., 2003), similar to *Nodal* mutant mice (Brennan et al., 2002; Kumar et al., 2008; Saijoh et al., 2003). In contrast, *spaw* mutants with a C-terminal point mutation still initiate (but do not propagate) *spaw* expression in the LPM, and then develop heart jogging, but not looping, phenotypes (Noël et al., 2013). This has led to the conclusion that heart chirality in zebrafish is controlled by a Nodal-independent mechanism (Noël et al., 2013). However, it is unclear whether the existing *spaw* allele is a null mutant. Thus, a loss-of-function mutation in *spaw* is needed to directly test its function in zebrafish left-right patterning, and to determine whether Nodal function is conserved between fish and mouse.

A second TGFβ family member, *Gdf1*, is also expressed in the node and LPM (Rankin et al., 2000). Nodal-Gdf1 heterodimers pattern the mouse left-right axis (Tanaka et al., 2007). Consistent with *Gdf1* mutant phenotypes, knockdown of zebrafish *vg1/gdf3* causes a reduction or absence of *spaw* in the LPM and heart looping phenotypes (Peterson et al., 2013), and rescue experiments of maternal-zygotic *vg1* mutants suggest that Spaw requires Vg1 to pattern the left-right axis (Pelliccia et al., 2017). However, in contrast to mouse Nodal-Gdf1, Spaw-Vg1 heterodimers have not been detected in zebrafish (Peterson et al., 2013). Thus, it remains unclear whether the role of Vg1/Gdf1 in left-right patterning is conserved between fish and mouse.

Asymmetric *Nodal* expression in the mouse LPM is initiated and maintained through two means of inhibition. First, Nodal is inhibited on the right side of the node by Cerl2/Dand5, a member of the Cerberus/Dan family of secreted TGFβ antagonists. Like *Cerl2/Dand5* mouse mutants (Marques et al., 2004), *dand5/charon* zebrafish morphants develop bilateral *spaw* expression, and heart jogging and looping defects (Hashimoto et al., 2004). Lowering *dand5* levels by morpholino knockdown causes premature induction of *spaw* expression in the LPM, but no change in the rate of propagation was observed (Wang and Yost, 2008). *Dand5* mutants have not been reported in zebrafish.

Second, *Lefty1* at the midline is thought to restrict Nodal activity to the left. Zebrafish *lefty1* morphants (Wang and Yost, 2008) and mutants (Rogers et al., 2017) have defects in heart jogging. *lefty1* morphants also develop bilateral *spaw* expression (Wang and Yost, 2008), and similar to mouse *lefty1* mutants (Meno et al., 1998), left LPM *spaw* expression precedes the right (Wang and Yost, 2008). These data have led to the suggestion that *Lefty1* functions as a midline barrier during left-right patterning (Lenhart et al., 2011;

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Meno et al., 1998; Wang and Yost, 2008), but Lefty1 has also been proposed to act through a self-enhancement and lateral-inhibition (SELI) feedback system (Nakamura et al., 2006). Surprisingly, during mesendoderm patterning in zebrafish, Lefty1 and Lefty2 can be replaced with uniform inhibition via a Nodal inhibitor drug (Rogers et al., 2017). This raises the question of whether (1) *lefty1* localization, (2) the timing of *lefty1* expression and (3) dynamic feedback between Nodal and *lefty1* are required for left-right patterning.

In this study, we assessed to what extent the mechanisms driving left-right patterning are conserved between mouse and zebrafish. We genetically tested the role of the secreted factors *spaw*, *dand5* and *lefty1* in zebrafish by generating single and double null mutants. Our results reveal that Spaw is required to transfer *spaw* expression to the LPM, Dand5 and Lefty1 restrict Spaw to the left side, and Spaw and Vg1 form heterodimers. Additionally, we show that Dand5 and Lefty1 control the timing and speed of Spaw propagation, and that the localization and timing of *lefty1* expression are not always crucial for left-right patterning. Collectively, these results clarify and unify the regulatory mechanisms of left-right patterning.

## RESULTS AND DISCUSSION

### Spaw, Dand5 and Lefty1 are required for left-right asymmetry

To test whether the mechanisms of left-right axis establishment are conserved between mouse and zebrafish, we analyzed single and double mutants for *spaw*, *dand5* and *lefty1* (Fig. 1A,B). We used CRISPR-Cas9 to generate frameshifting mutants for *dand5* and *spaw*, and used previously generated *lefty1* mutants (Rogers et al., 2017) (Table S1). All six mutant combinations were homozygous viable and lacked gross phenotypes (Fig. 1C) but displayed randomized or symmetric heart looping and jogging (Fig. 1D,E). In particular, *spaw* mutants displayed randomization of heart jogging, and failed to produce a normal heart loop, contrary to *spaw* C-terminal point mutants (Noël et al., 2013) but similar to *spaw* morphants (Long et al., 2003) and mouse *Nodal* mutants (Brennan et al., 2002; Kumar et al., 2008; Saijoh et al., 2003).

To understand the effect of the mutations on gene expression, we analyzed *lefty1*, *lefty2* and *spaw* expression during somitogenesis (Fig. 1F). In *spaw*, *dand5;spaw* and *lefty1;spaw* mutants, *spaw* expression was reduced around Kupffer's vesicle (the zebrafish equivalent to the mouse node in left-right patterning), and *lefty1* and *lefty2* expression was lost. In *dand5*, *lefty1* and *dand5;lefty1* mutants, *spaw* was expressed in both the left and right LPM, *lefty1* expression was higher in the midline, and *lefty2* was expressed in both heart fields (Fig. 1F). In addition, in *lefty1* and *dand5;lefty1* mutants, *spaw* was expressed in the midline, a phenotype not observed in zebrafish morphants (Wang and Yost, 2008) and only weakly visible in mouse mutants (Meno et al., 1998). However, *spaw* midline expression is consistent with the endogenous expression of the Nodal co-receptor *one-eyed pinhead* (*oep*)/*tdgf1* and the Nodal pathway transcription factor *foxh1* in the midline (Pogoda et al., 2000; Sirotkin et al., 2000; Zhang et al., 1998). Thus, Spaw is an upstream activator of *spaw*, *lefty1* and *lefty2*, and Dand5 and Lefty1 restrict the localization of *spaw*. Collectively, these results reveal that the roles of Nodal, Dand5 and Lefty1 are conserved from mouse to zebrafish.

### Spaw and Vg1 form heterodimers

Vg1 is not detectably processed or secreted on its own, but upon co-expression with its heterodimeric Nodal partners *cyclops* or *squint*, it is cleaved to its mature form and secreted (Montague and Schier, 2017). To determine whether Spaw and Vg1 also form functional heterodimers like Cyclops-Vg1, Squint-Vg1 and mouse Nodal-Gdf1, we generated superfolderGFP (sfGFP) and epitope-tagged versions of

Spaw and Vg1, and tested the activity, processing, interaction and localization of Spaw and Vg1. Expression of *vg1-sfGFP* in the presence and absence of *spaw* revealed that, while Spaw-sfGFP alone was cleaved to its mature form and secreted, Vg1-sfGFP was cleaved and secreted only in the presence of Spaw (Fig. 2A-C).

If Vg1 and Spaw form heterodimers, a biochemical interaction should be detectable. We performed co-immunoprecipitation experiments by co-expressing *vg1-Flag* with *spaw-HA*, or *squint-HA* as a positive control (Montague and Schier, 2017). In contrast to previous studies (Peterson et al., 2013), an interaction was detected between Vg1 and Spaw (Fig. 2D), indicating that Vg1 and Spaw form heterodimers.

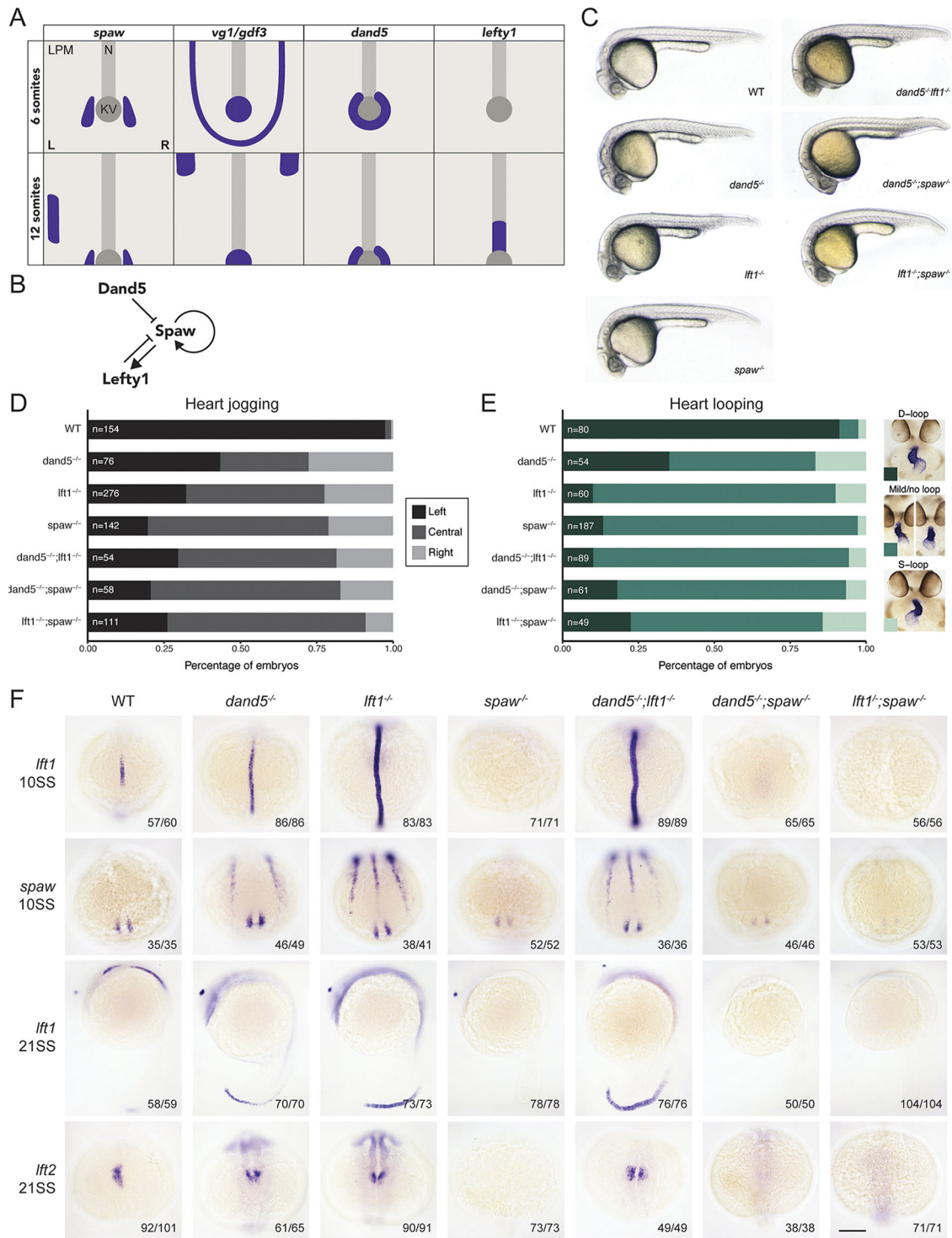
To test whether Spaw activity is dependent on Vg1, we expressed Spaw in the presence or absence of Vg1. Expression of 0.25-5 pg of *spaw* mRNA in blastula embryos induced ectopic Nodal target gene expression in wild-type embryos, but failed to induce ectopic gene expression in maternal *vg1* (*Mvg1*) mutants (Fig. 2E), indicating that Spaw requires Vg1 for full activity. Nodal target gene expression was induced in embryos injected with 20 pg or more of *spaw* mRNA in the absence of *vg1*, suggesting that, at high concentrations, Spaw might be able to form active homodimers. Collectively, these results suggest that, after Spaw is translated, it heterodimerizes with inactive, unprocessed Vg1. Vg1-Spaw dimerization facilitates the cleavage and secretion of Vg1, producing a mature active heterodimer.

### Dand5 and Lefty1 regulate the timing and speed of spaw propagation

Analysis of *spaw* expression in the single and double mutants revealed that *spaw* propagated through the LPM of *dand5* and *lefty1* mutants before it had initiated LPM expression in wild-type embryos (Fig. 1F). To understand whether this was due to a difference in the onset of *spaw* expression, the rate of *spaw* propagation, or both, we analyzed *spaw* expression (co-stained with the somite marker *myoD1*) across multiple stages of somitogenesis in the single and double mutants. *spaw* expression never initiated in the LPM of *spaw*, *dand5;spaw* or *lefty1;spaw* mutants (Fig. 3A,B), contrary to previous results (Noël et al., 2013), indicating that Spaw activity is required to initiate *spaw* expression in the LPM. *spaw* expression initiated prematurely in the LPM of *dand5* and *lefty1* mutants, consistent with previous results using morpholino knockdown (Wang and Yost, 2008), and it initiated earliest in *dand5;lefty1* mutants (Fig. 3A,B). To understand whether the premature LPM *spaw* expression was caused by earlier initiation of *spaw* around Kupffer's vesicle, we analyzed the onset of *spaw* expression in wild type, *dand5*, *lefty1* and *dand5;lefty1* mutants. Indeed, *spaw* expression was already present around Kupffer's vesicle two to four somite stages (1-2 h) earlier in *dand5*, *lefty1* and *dand5;lefty1* mutants than in wild-type embryos (Fig. S1). Thus, Dand5 and Lefty1 both spatially and temporally regulate Spaw activity.

To test whether the rate of *spaw* propagation is affected by the loss of its inhibitors, we measured the distance of *spaw* expression from the tailbud as a function of somite number (as a proxy for time) in the *dand5*, *lefty1* and *dand5;lefty1* mutants. *spaw* propagated at an approximate rate of 108  $\mu\text{m}$  per somite generation in wild-type embryos and  $\sim 112$   $\mu\text{m}$ /somite in *dand5* mutants. Contrary to previous morpholino studies (Wang and Yost, 2008), the rate of *spaw* propagation was slightly higher in *lefty1* mutants ( $\sim 140$   $\mu\text{m}$ /somite) and *dand5;lefty1* double mutants ( $\sim 125$   $\mu\text{m}$ /somite) (Fig. 3B, Fig. S2). Thus, Lefty1 may reduce the speed of Spaw propagation during normal left-right patterning.

Collectively, these results suggest that peri-nodal Spaw induces the expression of *spaw* in the LPM. Dand5 and Lefty1 inhibit Spaw activity both temporally, which delays the onset of *spaw* expression,

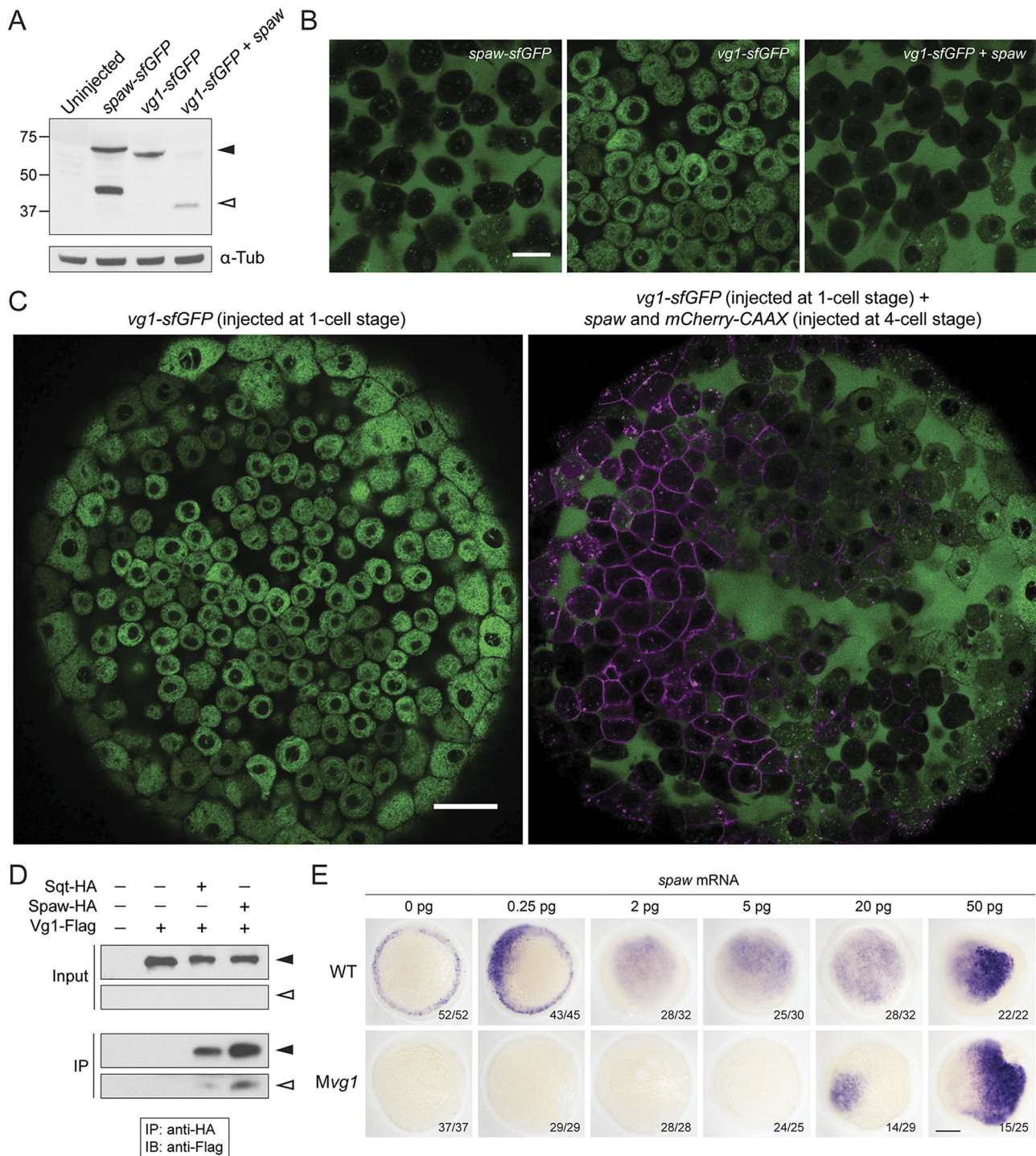


**Fig. 1. Spaw, Dand5 and Lefty1 are required for left-right asymmetry.** (A) Expression patterns of *spaw*, *vg1/gdf3*, *dand5* and *lefty1* in wild-type zebrafish embryos. LPM, lateral plate mesoderm; N, notochord; KV, Kupffer's vesicle. (B) Spaw induces itself and its inhibitor Lefty1. Spaw is also inhibited by Dand5. (C) Wild-type (WT), *dand5*, *lefty1* (*lft1*), *spaw*, *dand5;lft1*, *dand5;spaw* and *lft1;spaw* embryos at 28 h post-fertilization (hpf). See Table S1 for information about the mutant alleles. (D) Quantification of heart jogging at 28 hpf in wild-type and mutant embryos. (E) Quantification of heart looping at 2 days post-fertilization (dpf) in wild-type and mutant embryos, detected by expression of *myl7/cmlc2* by *in situ* hybridization. D-loop, dextral loop; S-loop, sinistral loop. A large proportion of hearts failed to loop correctly in either direction, forming no loop or a very mild loop. We designated these embryos as 'mild/no loop'. (F) Expression of *lefty1*, *spaw* and *lefty2* (*lft2*) at the 10-somite stage (10SS) and 21SS in wild-type and mutant embryos. Each image is representative of three to five independent experiments (embryo number is in bottom right corner of each panel). Scale bar: 150  $\mu$ m.

and spatially, which restricts *spaw* expression to the left LPM. In addition, Lefty1 may reduce the speed at which *spaw* propagates through the LPM. Although the timing of *Nodal* propagation in the

presence and absence of inhibitors has not been assessed in mouse, the spatial restriction of *Nodal* by its inhibitors is conserved from fish to mouse.



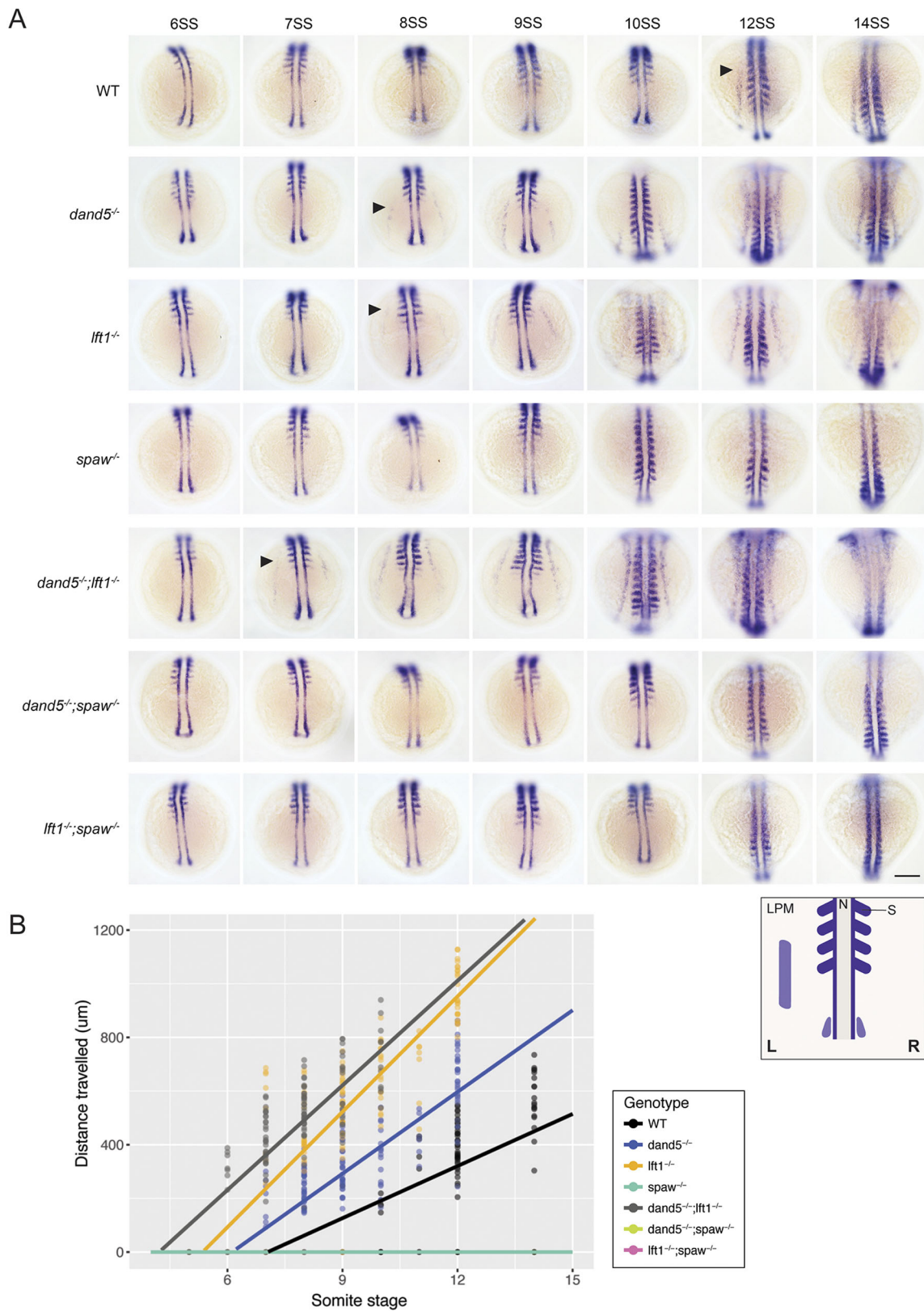


**Fig. 2. Spaw and Vg1/Gdf3 form heterodimers.** (A) Anti-GFP reducing immunoblot of wild-type embryos injected with 25 pg of *spaw-sfGFP*, *vg1-sfGFP* or *vg1-sfGFP* and *spaw* mRNA. Black arrowhead indicates the position of full-length protein; open arrowhead represents cleaved protein. Lower panel: anti- $\alpha$ -Tubulin loading control. (B) Animal cap of sphere-stage live embryos injected at the one-cell stage with 50 pg of *spaw-sfGFP*, 50 pg of *vg1-sfGFP* mRNA or 50 pg of *vg1-sfGFP* and 50 pg of *spaw* mRNA. Scale bar: 20  $\mu$ m. (C) Animal cap of a sphere-stage live embryo injected with 50 pg of *vg1-sfGFP* mRNA at the one-cell stage (left) or 50 pg of *vg1-sfGFP* mRNA at the one-cell stage and 25 pg of a *spaw* mRNA/*mCherry-CAAX* mRNA mix at the four-cell stage (right). Scale bar: 40  $\mu$ m. (D) Anti-Flag reducing immunoblot (IB) of anti-HA immunoprecipitates (IPs) from lysates of wild-type embryos injected with 50 pg of *squint-HA* (*sqt-HA*), 50 pg of *spaw-HA* and/or 50 pg of *vg1-Flag* mRNA. Black arrowheads indicate the position of full-length protein; open arrowheads represent cleaved protein. The input and IP blots were exposed for different lengths of time. (E) Expression of *lefty1* in wild-type and maternal *vg1* (*Mvg1*) mutant embryos injected with 0.25-50 pg of *spaw* mRNA. Scale bar: 150  $\mu$ m.

### Uniform Nodal inhibition rescues *lefty1* mutants

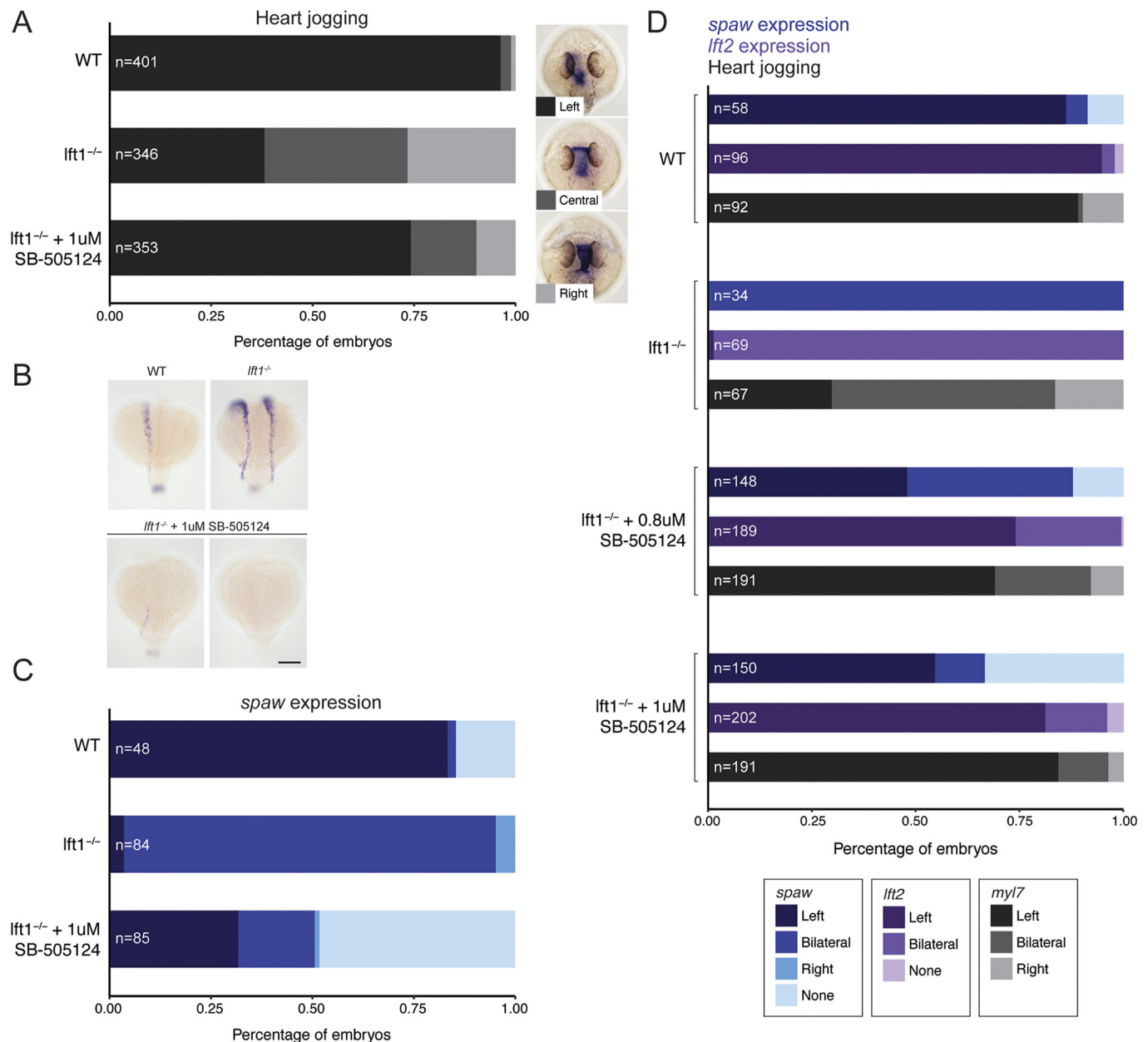
Our results indicate that Lefty1 is required to restrict and/or maintain Spaw activity in the left LPM. To test whether the localization or

timing of *lefty1* expression is necessary for correct patterning, we replaced *lefty1* expression with uniform Nodal inhibition by soaking *lefty1* embryos in 1  $\mu$ M of a Nodal signaling inhibitor drug,



**Fig. 3. Dand5 and Lefty1 regulate the timing and speed of spaw propagation.** (A) *spaw* and *myoD1* expression in six-somite stage (6SS) to 14SS wild-type and mutant embryos, staged by *myoD1* somite expression. Black arrowheads represent the earliest stage *spaw* expression appears in the lateral plate mesoderm. The diagram indicates *myoD1* expression (dark purple) and *spaw* expression (light purple). N, notochord, S, somite, LPM, lateral plate mesoderm. Scale bar: 150 µm. (B) Scatterplot of distance *spaw* has traveled through the LPM in wild-type and mutant embryos. The *spaw*, *dand5*;*spaw* and *lft1*;*spaw* graphs overlap at zero. *n*=160 (wild type), 183 (*dand5*), 148 (*lft1*), 149 (*spaw*), 115 (*dand5*;*lft1*), 126 (*dand5*;*spaw*) and 90 (*lft1*;*spaw*). These values represent the sum of three to six independent experiments. To calculate the rate of *spaw* propagation, a linear regression was fitted to the data after *spaw* had initiated in the LPM, see Fig. S2.





**Fig. 4. Uniform Nodal inhibition rescues *lefty1* mutants.** (A) Quantification of heart jogging in wild-type, *lefty1* embryos and *lefty1* embryos soaked in 1 μM of the Nodal inhibitor SB-505124. (B) Examples of left, bilateral and no *spaw* expression in wild-type, *lefty1* and SB-505124-treated *lefty1* mutant embryos. Scale bar: 150 μm. (C) Quantification of *spaw* expression in wild-type, *lefty1* and SB-505124-treated *lefty1* mutant embryos. (D) Quantification of *spaw* expression, *lft2* expression and heart jogging in wild-type, *lefty1*, and 0.8 μM and 1 μM SB-505124-treated *lefty1* embryos.

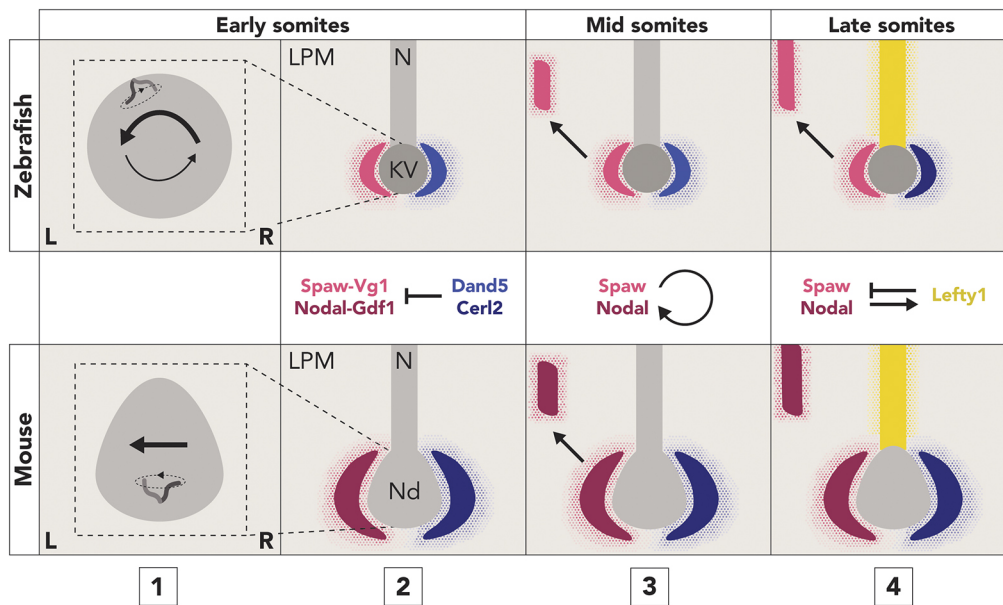
SB-505124, from 90% epiboly onwards (prior to the onset of *lefty1* expression) (Rogers et al., 2017). Surprisingly, leftward heart jogging was rescued in the majority of drug-treated *lefty1* embryos (Fig. 4A,D). Analysis of *spaw* expression in drug-treated or untreated *lefty1* mutants revealed that treatment with SB-505124 was also able to rescue left LPM *spaw* expression in a subset of embryos (Fig. 4B-D). To dissect why there was a difference between the proportion of embryos with left *spaw* expression versus left-biased hearts, we analyzed *lefty2* expression in the heart field at an intermediate stage between *spaw* LPM expression and heart positioning at two different concentrations of Nodal inhibitor. *lefty2* expression and leftward heart jogging were rescued in a similar proportion of drug-treated *lefty1* mutant embryos (Fig. 4D). This suggests that *in situ* hybridization is not sensitive enough to detect

low levels of *spaw* expression that can lead to asymmetric *lefty2* expression, or that leftward heart jogging is not entirely dependent on the presence of an asymmetric Nodal signal.

These results reveal that left-right patterning can occur in the absence of timely expression and midline localization of *lefty1*. In addition, although Lefty1 and Spaw normally function in a feedback system (Fig. 1B), Spaw-Lefty1 feedback can be removed for normal left-right patterning. Thus, *lefty1*-mediated inhibition may primarily function to dampen Nodal signaling to prevent its amplification and extension to the right LPM.

### Conclusions

Our mutant analyses clarify four roles of Spaw and its inhibitors in zebrafish left-right patterning. First, *spaw*-null mutants show



**Fig. 5. Left-right patterning mechanisms are conserved from zebrafish to mouse.** (1) Rotation of motile cilia in the left-right organizer (Kupffer's vesicle, zebrafish; node, mouse) causes the leftward flow of fluid that precedes asymmetric gene expression. (2) Spaw-Vg1 and Nodal-Gdf1 heterodimers are restricted to the left by Dand5/Cerl2 activity at the node. (3) Spaw/Nodal induces expression of *spaw/Nodal* in the left LPM, and (4) Spaw activity is maintained on the left by Lefty1-mediated inhibition. LPM, lateral plate mesoderm; N, notochord; KV, Kupffer's vesicle; Nd, node.

disruptions to both heart jogging and looping, indicating that Spaw is required for proper heart asymmetry. In addition, *spaw*, *spaw*; *dand5* and *spaw*; *lefty1* mutants fail to initiate *spaw* expression in the LPM, revealing that Spaw around Kupffer's vesicle is required to transfer *spaw* expression to the LPM. This is consistent with mouse data: mouse mutants that lack Nodal activity in the node fail to express *Nodal* in the LPM, and later develop left-right patterning defects, including heart jogging and looping phenotypes (Brennan et al., 2002; Kumar et al., 2008; Saijoh et al., 2003). Thus, these results reveal conservation between the roles of zebrafish Spaw and mouse Nodal in left-right patterning.

Second, our results show that in blastoderm assays, Vg1 is required for Spaw activity, Spaw allows the secretion and processing of Vg1, and Spaw and Vg1 interact. These results are consistent with genetic and biochemical data showing Gdf1 is a heterodimeric partner of Nodal during mouse left-right patterning (Tanaka et al., 2007).

Third, mutations in the Nodal inhibitors *dand5* and *lefty1* result in bilateral *spaw* expression, demonstrating functional conservation with the mouse orthologs *Cerl2/Dand5* and *Lefty1*, the absence of which leads to bilateral *Nodal* expression and organ asymmetry defects (Marques et al., 2004; Meno et al., 1998). In addition, our results reveal that the loss of Dand5 and Lefty1 leads to precocious *spaw* expression, and the loss of Lefty1 leads to accelerated *spaw* progression, suggesting these inhibitors normally control the timing, speed and localization of *Nodal* expression.

Fourth, application of a Nodal inhibitor drug to *lefty1* mutant embryos is sufficient to rescue normal heart jogging and left-sided expression of *spaw* in most animals, suggesting that the localization of Lefty1 activity is not always crucial. Rather, Lefty1 dampens Nodal activity to prevent its amplification and extension to the right side of the embryo.

Collectively, our results show that the mechanisms of left-right patterning are highly conserved from mouse to zebrafish and suggest a refined model of zebrafish left-right patterning: (1) rotation of cilia in Kupffer's vesicle causes leftward flow of fluid; (2) Dand5 creates an initial bias in Spaw-Vg1 heterodimer activity at the node; (3) Spaw-Vg1 induces the expression of *spaw* in the LPM; and (4) Lefty1-mediated inhibition maintains the left-right bias of Spaw activity during auto-induction (Fig. 5). Given that the basic functions of Nodal, Dand5, Lefty and cilia/the left-right organizer

are also conserved in *Xenopus* left-right patterning (Blum et al., 2014; Branford et al., 2000; Schweickert et al., 2010, 2007; Vonica and Brivanlou, 2007), these results suggest that the roles of the Nodal cascade and left-right organizer may be conserved across all vertebrates, with the exception of the sauropsida (Blum and Ott, 2018). These studies lay the foundation for future biochemical and biophysical analyses needed to determine how fluid flow is converted into asymmetric gene expression, and how Nodal pathway components interact to generate precise timing and localization during left-right patterning.

## MATERIALS AND METHODS

### Ethics statement

All vertebrate animal work was performed at the facilities of Harvard University, Faculty of Arts and Sciences (HU/FAS). The HU/FAS animal care and use program maintains full AAALAC accreditation, is assured with OLAW (A3593-01) and is currently registered with the USDA. This study was approved by the Harvard University/Faculty of Arts and Sciences Standing Committee on the Use of Animals in Research and Teaching under protocol number 25-08.

### Zebrafish husbandry and microinjection

Zebrafish embryos were grown at 28°C and staged according to Kimmel et al. (1995). For somite stages (except drug treatments), embryos were grown at 28°C until sphere stage, and then grown overnight at room temperature. Embryos were cultured in blue water (250 mg/l Instant Ocean salt, 1 mg/l Methylene Blue in reverse osmosis water adjusted to pH 7 with NaHCO<sub>3</sub>). Embryos for immunoblot and live imaging experiments were chemically dechorionated with 1 mg/ml pronase (protease type XIV from *Streptomyces griseus*, Sigma) prior to injection, and cultured in agarose-coated dishes. Embryos were injected at the one-cell stage unless otherwise stated.

### CRISPR-Cas9-mediated mutagenesis of *dand5* and *spaw*

sgRNAs targeting the *dand5/charon* and *spaw* genes were designed using CHOPCHOP (Labun et al., 2016; Montague et al., 2014) (Table S1) and synthesized as previously described (Gagnon et al., 2014). TLAB wild-type zebrafish embryos were injected at the one-cell stage with sgRNAs and ~0.5 nl of 50 μM Cas9 protein. Injected embryos were raised to adulthood and outcrossed to TLAB adults. The resulting clutches were genotyped to identify potential founders with germline mutations in *dand5* or *spaw* by MiSeq sequencing. Animals were recovered with a 4 bp deletion in the first exon of *dand5* (*dand5<sup>a204</sup>*), causing a frameshift and truncation of Dand5

from 243 amino acids to 54 amino acids, and a 49 bp deletion in the first exon of *spaw* (*spaw*<sup>a205</sup>), resulting in a frameshift and truncation of Spaw from 404 amino acids to 49 amino acids (Table S1). The offspring of confirmed founders were raised to adulthood and genotyped by fin-clipping to determine heterozygous individuals. Homozygous animals were generated by intercrossing heterozygous fish.

### Generation of double mutants

*dand5;lefty1*, *dand5;spaw* and *lefty1;spaw* mutants were generated by intercrossing homozygous *dand5*, *spaw* and *lefty1* (Rogers et al., 2017) mutants to generate double heterozygous mutants. Double heterozygous mutants were intercrossed and genotyped to identify double homozygous mutants.

### Genotyping of mutants

Genomic DNA was extracted from embryos and fin-clips using the HotSHOT method (Meeker et al., 2007), and PCR was performed using standard methods. *spaw* and *vg1/gdf3* mutants (*vg1*<sup>a165</sup>) (Montague and Schier, 2017) were genotyped by gel electrophoresis, *dand5* mutants were genotyped by Sanger sequencing, and *lefty1* mutants (*lefty1*<sup>a145</sup>) were genotyped by *PshAI* digestion (Rogers et al., 2017). Primer sequences were as follows: *dand5*\_genotype\_F, TCACAACAATTGGCGCTTTC; *dand5*\_genotype\_R, GCCAAGAACGCGGGAAAC; *lefty1*\_genotype\_F, CGTG-GCTTTCATGTATCACCTTC; *lefty1*\_genotype\_R, GGATGCCGCCA-AACTG; *spaw*\_genotype\_F, CGCGTTATTTGTTTACGCGTTG; *spaw*\_genotype\_R, TGCTTACCTGTGCAATCAAGC; *vg1*\_genotype\_F, CT-TGCAGATGTGGATTCTGGCC; and *vg1*\_genotype\_R, CATGATGCG-ATGGTTTGGGTCG.

### Cloning of expression and *in situ* probe constructs

The *spaw* CDS sequence was PCR amplified from a somite-stage cDNA library and cloned into the pCS2(+) vector using Gibson cloning (Gibson et al., 2009) to generate pCS2(+)-*spaw*. *spaw-sfGFP* was generated by inserting the superfolder GFP (sfGFP) sequence (Pédélec et al., 2006) downstream of the *spaw* cleavage site (RHKR) in pCS2(+)-*spaw* with Gibson cloning. *mCherry-CAAX* was generated by inserting a farnesylation sequence at the 3' end of the *mCherry* CDS in a pCS2(+) vector. *vg1*, *vg1-sfGFP*, *vg1-Flag* and *sqt-HA* have been previously reported (Montague and Schier, 2017). *spaw-HA* was generated by inserting the HA tag (YPYDVPDYA) sequence downstream of the cleavage site by site-directed mutagenesis of pCS2(+)-*spaw*. For all fusion constructs, the epitope or sfGFP tag was flanked by a GSTGTT linker at the 5' end of the tag and by a GS linker at the 3' end of the tag. *In situ* probes targeting *lefty1*, *spaw*, *lefty2*, *myl7* and *myoD* were generated by inserting the full CDS, or up to 1000 bp of the CDS of the gene into the pSC vector using Strataclone (Agilent).

### mRNA and probe synthesis

Expression constructs were linearized by digestion with *NotI*, followed by mRNA synthesis using the SP6 mMessage Machine Kit (ThermoFisher). *In situ* antisense probes were synthesized using a DIG Probe Synthesis Kit (Roche).

### *In situ* hybridization

Embryos were fixed overnight at 4°C in 4% formaldehyde, followed by whole-mount *in situ* hybridization according to standard protocols (Thisse and Thisse, 2008). NBT/BCIP/alkaline phosphatase-stained embryos were imaged using a Zeiss Axio Imager.Z1 after dehydration with methanol and clearing with benzyl benzoate:benzyl alcohol (BBBA).

### Morphological analysis of mutant phenotypes and live imaging

Embryos at 24 h post-fertilization (hpf) were anesthetized using Tricaine (Sigma) and imaged in 2% methylcellulose. *spaw-sfGFP*-injected embryos were mounted at sphere stage in 1% low gelling temperature agarose (Sigma) and live imaged using a Zeiss LSM 700 confocal microscope.

### Immunoblotting and co-immunoprecipitation

Embryos were injected with 50 pg of mRNA (unless otherwise stated), and grown until 50% epiboly, then flash frozen (eight embryos per sample) in

liquid nitrogen after manual deoylking using forceps. The samples were boiled for 5 min at 95°C with 2× SDS loading buffer and DTT, and then loaded onto Any kD protein gels (Bio-Rad). The samples were transferred to polyvinylidene fluoride (PVDF) membranes (GE Healthcare) before blocking in 5% non-fat milk (Bio-Rad) in TBST at 4°C overnight with primary antibodies (rabbit anti-GFP, 1:5000, ThermoFisher A11122; mouse anti- $\alpha$ -Tubulin, 1:5000, ICN Biomedicals 691251; rabbit anti-Flag, 1:2000, Sigma F7425). HRP-coupled secondary antibody was applied to the membranes (goat anti-rabbit, 1:15,000, Jackson ImmunoResearch Labs 111-035-144) and Amersham ECL reagent (GE Healthcare) was used for chemiluminescence detection. For co-immunoprecipitation experiments, 80 embryos per sample were homogenized in 400  $\mu$ l of ice-cold lysis buffer (50 mM Tris at pH 7.5, 150 mM NaCl, 1 mM EDTA, 10% glycerol, 1% Triton X-100 and protease inhibitors, Sigma 11836170001) and vortexed every 5 min during a 30-min incubation on ice. Samples were spun at 4°C for 30 min, and then the supernatant was transferred to a tube with 50  $\mu$ l of anti-HA affinity matrix (Roche 11815016001). Samples were rocked at 4°C overnight. The samples were spun for 2 min and washed in ice-cold wash buffer (50 mM Tris at pH 7.5, 150 mM NaCl, 1% Triton X-100 and protease inhibitors) five times before adding 2×SDS loading buffer and DTT. Immunoblots were performed as above.

### Image adjustments

Images were processed in ImageJ/FIJI (Schindelin et al., 2012). Brightness, contrast and color balance were applied uniformly to images.

### Drug treatments

Embryos were grown at 28°C until 75% epiboly, then dechorionated using pronase. At 90% epiboly, the embryos were placed in agarose-coated dishes containing pre-warmed Nodal inhibitor SB-505124 (S4696, Sigma) diluted in blue water. After development to 18 somites, 22 somites or 28 hpf at 28°C, the embryos were fixed for *in situ* hybridization in 4% formaldehyde. SB-505124 was stored at 4°C in a 10 mM stock.

### *spaw* LPM measurements

Embryos were co-stained using a mix of the *spaw* and *myoD* probes. To calculate *spaw* propagation in the LPM, a measurement was made from the lowest point of *myoD* staining (base of the notochord) to the highest point of *spaw* staining on the left side of the embryo. For embryos in which *spaw* had propagated a substantial distance up the LPM, the embryo was imaged from multiple orientations, and the distances summed. Embryos were also imaged from the side to ensure that the curved surface of the embryo did not affect the final measurement. For *spaw* propagation calculations, a linear regression line was fit to the scatter plot of *spaw* distance at time points after initiation of expression in the LPM. This ensured that the timing of *spaw* propagation did not affect the slope of *spaw* propagation.

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### Competing interests

The authors declare no competing or financial interests.

### Author contributions

Conceptualization: T.G.M., A.F.S.; Methodology: T.G.M.; Validation: T.G.M.; Formal analysis: T.G.M.; Investigation: T.G.M.; Resources: J.A.G.; Writing - original draft: T.G.M.; Writing - review & editing: T.G.M., J.A.G., A.F.S.; Visualization: T.G.M.; Supervision: A.F.S.; Project administration: A.F.S.; Funding acquisition: A.F.S.

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### Supplementary information

Supplementary information available online at <http://dev.biologists.org/lookup/doi/10.1242/dev.171090.supplemental>



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