

Selective expression of presenilin 1 in neural progenitor cells rescues the cerebral hemorrhages and cortical lamination defects in presenilin 1-null mutant mice

Paul H. Wen^{1,2}, Rita De Gasperi^{1,2}, Miguel A. Gama Sosa^{1,2}, Anne B. Rocher³, Victor L. Friedrich, Jr³, Patrick R. Hof^{3,4} and Gregory A. Elder^{1,2,*}

¹Department of Psychiatry, Mount Sinai School of Medicine, New York, NY 10029, USA

²Psychiatry Service, Bronx VA Medical Center, Bronx, NY 10468, USA

³Department of Neuroscience, Mount Sinai School of Medicine, New York, NY 10029, USA

⁴Department of Geriatrics and Adult Development, Mount Sinai School of Medicine, New York, NY 10029, USA

*Author for correspondence (e-mail: gregory.elder@mssm.edu)

Accepted 17 June 2005

Development 132, 3873–3883

Published by The Company of Biologists 2005

doi:10.1242/dev.01946

Summary

Mice with a null mutation of the presenilin 1 gene (*Psen1*^{−/−}) die during late intrauterine life or shortly after birth and exhibit multiple CNS and non-CNS abnormalities, including cerebral hemorrhages and altered cortical development. The cellular and molecular basis for the developmental effects of *Psen1* remain incompletely understood. *Psen1* is expressed in neural progenitors in developing brain, as well as in postmitotic neurons. We crossed transgenic mice with either neuron-specific or neural progenitor-specific expression of *Psen1* onto the *Psen1*^{−/−} background. We show that neither neuron-specific nor neural progenitor-specific expression of *Psen1* can rescue the embryonic lethality of the *Psen1*^{−/−} embryo. Indeed neuron-specific expression rescued none of the

abnormalities in *Psen1*^{−/−} mice. However, *Psen1* expression in neural progenitors rescued the cortical lamination defects, as well as the cerebral hemorrhages, and restored a normal vascular pattern in *Psen1*^{−/−} embryos. Collectively, these studies demonstrate that *Psen1* expression in neural progenitor cells is crucial for cortical development and reveal a novel role for neuroectodermal expression of *Psen1* in development of the brain vasculature.

Key words: Cortical development, CNS hemorrhages, Familial Alzheimer's disease, Neural progenitor cells, Presenilin 1 (PS1, *Psen1*), Transgenic mice, Vascular development

Introduction

Alzheimer's disease (AD) is a degenerative disorder of the central nervous system (CNS). Although most cases occur sporadically, in some families the disease is inherited in an autosomal dominant fashion, referred to as familial AD (FAD). Mutations in the presenilin 1 (*PSEN1*) gene are the most commonly recognized cause of early onset FAD and more than 100 mutations in the *PSEN1* gene have been linked to FAD (Lleo et al., 2004). Mutations in a related gene, presenilin 2, cause FAD in a more limited number of cases (Cruts et al., 1998).

In adult brain, *Psen1* is expressed primarily in neurons (Elder et al., 1996), although neural progenitor cells in adult hippocampus express *Psen1* (Wen et al., 2002a) and its expression can be induced in reactive astrocytes (Cribbs et al., 1996), including those surrounding senile plaques (Weggen et al., 1998). Developmentally, *Psen1* expression is found as early as the preimplantation embryo (Jeong et al., 2000) and *Psen1* is prominently expressed in neural progenitor cells in the ventricular zone of embryonic rodents (Moreno-Flores et al., 1999) and humans (Kostyszyn et al., 2001). Mice with a null mutation of the *Psen1* gene (*Psen1*^{−/−}) die during late intrauterine life or shortly after birth, and exhibit multiple CNS

and non-CNS abnormalities, including cerebral hemorrhages and altered cortical development (Hartmann et al., 1999; Shen et al., 1997; Wong et al., 1997).

The cellular and molecular basis for the developmental effects of *Psen1* remain incompletely understood. In particular, the role that *Psen1* plays in different cell types remains unknown. Recently, we generated transgenic mice with either neuron-specific (Wen et al., 2002b) or neural progenitor-specific expression of *Psen1* (this paper). We show by crossing these transgenes onto the *Psen1*^{−/−} background that neither neuron-specific nor neural progenitor-specific expression of *Psen1* rescues the embryonic lethality of the *Psen1*^{−/−} embryo. Indeed, neuron-specific expression ameliorates none of the abnormalities in the *Psen1*^{−/−} embryo. However, *Psen1* expression in neural progenitors rescues cortical development in *Psen1*^{−/−} embryos, including normalizing cortical lamination patterns and eliminating the cerebral hemorrhages.

Materials and methods

Transgenic and *Psen1*^{+/−} mice

Transgenic mice expressing human *Psen1* under the control of the neuron specific enolase (NSE) promoter/enhancer regions have been

previously described (Wen et al., 2002b; Wen et al., 2004b). A transgene allowing expression of human *Psen1* in neural progenitor cells (NesPsen1) was constructed in the vector gITKlacZ (Zimmerman et al., 1994), which contains the nestin intron 2 neural specific enhancer upstream of a 160 bp thymidine kinase (tk) minimal promoter followed by an artificial intron, cloning site and SV40 polyadenylation signal. The *lacZ* gene was replaced with a human *Psen1* wild-type cDNA (Elder et al., 1996). A Nestin/Cre recombinase transgene (NesCrenls) was prepared by cloning the nestin-tk promoter/enhancer from gITKlacZ into the plasmid pOG231 (O'Gorman et al., 1997), which places the nestin-tk promoter/enhancer upstream of an 0.2 kb synthetic intron followed by a Cre-coding sequence containing a nuclear localization sequence and a polyadenylation signal. A Cre reporter transgene was generated by replacing the *lacZ* sequences in the plasmid pcAct-XstopXnZ (obtained from Drs Eric Mercer and David Anderson, Howard Hughes Medical Institute, Caltech, USA) with an enhanced green fluorescent protein (EGFP) cDNA (Clontech, Palo Alto, CA, USA). This transgene (cActXstopXEGFP) includes the 2.1 kb chicken β -actin promoter along with an additional 1 kb containing the β -actin exon 1, intron 1 and 5' untranslated sequence from exon 2, while downstream of exon 2 it contains a translation 'stop' cassette sequence (Lakso et al., 1992) flanked by 34 bp *loxP* sites and the EGFP cDNA.

Transgenic mice were produced by pronuclear injection using C57Bl/6J \times C3H (B₆C₃) as a source of fertilized eggs. Genotypes were determined by PCR on DNA isolated from tail biopsies or from regions of the embryos or yolk sac. The NesPsen1 transgene was identified with primers homologous to the tk promoter (5'CACGCAGATGCAGTCGGG3') and the human *Psen1* cDNA (5'GTGTTCTCTCCAGGCCAAG3') that yield a 287 bp product. Primers to the Cre cDNA (5'GTCGAGCGATGGATTTCGGTCT3' and 5'GCTTGATGATCTCCGGTATT3') were used to identify a 274 bp product from the NesCrenls transgene. cActXstopXEGFP transgenic mice were identified with the primers 5'CGTAAACGGC-CACAAGTTCAG3' and 5'ATGCCGTTCTTCTGCTTGTCG3' that amplify a 420 bp product from the EGFP cDNA. Lines were maintained by breeding transgenic animals to C57Bl/6 wild-type mice.

The Z/EG transgenic line (Novak et al., 2000) was obtained from Jackson laboratories (Bar Harbor, MA, USA; stock name Tg(ACTB-Bgeo/GFP); stock number 003920). *Psen1*^{-/-} mice were obtained from Dr Huntington Potter (University of South Florida, Tampa FL, USA) and are those generated by Shen et al. (Shen et al., 1997). These animals were provided on a mixed genetic background and have been maintained by breeding to C57Bl/6 wild-type mice. Genotypes were determined as described by Shen et al. (Shen et al., 1997). Owing to the mixed genetic background of both the *Psen1*^{-/-} and the *Psen1* transgenic animals, non-transgenic littermates were used as controls in all studies.

Animals were housed on 12-hour light/dark cycles and received food and water ad libitum. All procedures were approved by the Mount Sinai School of Medicine Institutional Animal Care and Use Committee and were in conformance with the National Institutes of Health 'Guide for the Care and Use of Laboratory Animals'.

Western blotting

Western blotting was performed as previously described (Wen et al., 2002b).

BrdU injections

5'-bromo-2'-deoxyuridine (BrdU; Sigma, St Louis, MO, USA) was dissolved in 0.9% NaCl at a concentration of 10 mg/ml and sterilized through an 0.45 μ m filter. Mice received one intraperitoneal injection (50 μ g/gm of body weight).

Histology and immunohistochemistry

Timed pregnant female mice were euthanized with carbon dioxide.

The day of vaginal plug was designated as E0.5. Embryos were fixed in 4% paraformaldehyde overnight and embedded in paraffin wax. Brains were cut into 6–8 μ m coronal or horizontal sections. Adult animals were perfused transcardially and the tissues processed as previously described (Wen et al., 2002b).

Histological sections were stained with Cresyl Violet and a standard set of at least five sections per embryo was examined. For horizontally sectioned embryos, a series centered through the lateral ventricle above the level of the caudate/putamen was used. A series centered at the level of the medial septal nucleus was examined in coronally sectioned embryos.

For immunohistochemical staining sections were blocked with PBS/0.1% Triton X-100/5% goat serum (TBS-TGS) for 30 minutes and primary antibodies were applied in TBS-TGS at room temperature overnight. The primary antibodies used were Rat 401, a mouse monoclonal anti-rat nestin (1:500, Chemicon, Temecula, CA, USA); CS56, a mouse monoclonal anti-chondroitin sulfate proteoglycan (1:400, Sigma); G10, a mouse monoclonal anti-reelin (1:500, Chemicon); a rabbit polyclonal anti-EGFP (1:1500, Molecular Probes, Eugene, OR, USA); a rabbit polyclonal anti-von Willebrand factor (1:200, Sigma); and a rabbit polyclonal anti-fibronectin (1:400, Sigma). Immunofluorescence staining was detected with species-specific Alexa Fluor secondary antibody conjugates (1:400, Molecular Probes) applied for 2 hours; sections were mounted using Gel/Mount (Biomedex, Foster City, CA, USA). BrdU immunohistochemistry was performed as previously described (Wen et al., 2002b).

Vascular staining with Griffonia simplicifolia isolectin B4 was performed using methods similar to those described in Ashwell (Ashwell, 1991). Deparaffinized sections were incubated for 20 minutes at 90°C in 10 mM Na citrate buffer (pH 8.6) and allowed to cool to room temperature. After blocking with TBS containing 0.02% Triton X-100, 1 mM MgCl₂, 1 mM CaCl₂ and 1 mM MnCl₂ for 1 hour at room temperature, biotinylated Isolectin B4 (Sigma, 10 μ g/ml in the above buffer) was added and sections incubated overnight at 4°C. To detect lectin binding, sections were incubated with Streptavidin-Alexa 488 (Molecular Probes, 1:300 diluted in TBS) for 2 hours at room temperature. When Isolectin B4 staining was combined with immunohistochemistry, lectin staining was performed after the immunostaining.

Images were collected using a Zeiss Axiophot microscope (Zeiss, Thornwood, NY, USA) or a Nikon Eclipse E400 connected to a DXC-390 CCD camera (Nikon, Melville, NY, USA). Images were color balanced and merged using Adobe Photoshop 7.0 (Adobe Systems, San Jose, CA, USA).

When spontaneous EGFP fluorescence was imaged and combined with aquaporin immunohistochemistry, Vibratome sections (200 μ m) were immunostained with a polyclonal anti-aquaporin 4 antibody (1:100, Chemicon) followed by Texas Red-conjugated secondary antibodies (Vector Laboratories, Burlingame, CA, USA). Spontaneous EGFP fluorescence and Texas Red immunofluorescence were imaged with a Radiance 2000 MP system (Bio-Rad, Hercules, CA, USA) equipped with a Mira 900F Ti: sapphire laser (tuned to 860 nm) and a Verdi 10-W pump laser (Coherent, Santa Clara, CA, USA).

In situ hybridization

E12.5–E13.5 embryos were immersion fixed for 2 hours in 4% paraformaldehyde dissolved in diethylpyrocarbonate (DEPC)-treated PBS, equilibrated sequentially in 10%, 20% and 30% sucrose and embedded in OCT compound (Tissue Tek, Elkhart, IN, USA). Cryosections (15 μ m) were cut, air dried for 20 minutes and fixed in 4% paraformaldehyde in PBS-DEPC. Sections were treated with 1 μ g/ml proteinase K for 5 minutes at room temperature and fixed in paraformaldehyde for an additional 20 minutes. After several washes with PBS-DEPC, sections were acetylated with acetic anhydride in the presence of triethanolamine. The sections were then prehybridized for 2 hours in 50% formamide, 5 \times SSC, 5 \times Denhardt's solution, 250 μ g/ml yeast RNA, 500 μ g/ml herring sperm DNA and hybridized at

55°C for 16 hours with 400 ng/ml of a heat denatured (80°C) digoxigenin-labeled antisense RNA probe complementary to a 260 bp sequence in the human *PSEN1* 3' untranslated region (nucleotides 1854-2114 in GenBank Accession Number BC011729). Probes were labeled by random incorporation of digoxigenin-labeled deoxyuridine triphosphate using a commercially available kit (Roche, Indianapolis, IN, USA). Slides were washed for 1 hour in 0.2× SSC at 70°C and subsequently with 50 mM Tris-HCl (pH 8.0), 0.15 M NaCl (TBS) at room temperature. After blocking with 10% heat-inactivated goat serum in TBS at room temperature, sections were incubated overnight with a 1:250 dilution of anti-digoxigenin antibodies at 4°C (Roche). Following several washes with TBS, slides were equilibrated in alkaline phosphatase buffer [0.1 M Tris-HCl (pH 9.5), 0.1 M NaCl, 50 mM MgCl₂, 0.01% Tween-20, 0.25 mg/ml levamisole] for 30 minutes followed by staining with 0.4 mg/ml nitro tetrazolium blue chloride, 0.19 mg/ml 5-bromo-4-chloro-3-indolyl-phosphate in the same solution for 72 hours at 4°C. E16.5 embryos were hybridized in an identical manner except that the brains were dissected and frozen directly in OCT compound without prior fixation. Additionally, the proteinase K digestion step was omitted and the hybridization was performed at 60°C.

Results

Generation of transgenic mice expressing human presenilin 1 in neural progenitor cells under the control of the rat nestin intron 2 enhancer

Nestin is an intermediate filament that is expressed in neural progenitor cells of the developing CNS (Frisen et al., 1998; Lendahl et al., 1990; McKay, 1997). To express *Psen1* selectively in neural progenitor cells, we used the 257 bp enhancer from the second intron of the rat nestin gene that has been used to drive expression of heterologous genes in neural progenitor cells in transgenic mice in both embryonic and adult CNS (Aoki et al., 2000; Josephson et al., 1998; Zimmerman et al., 1994). Like endogenous nestin, expression of these transgenes ceases as progenitor cells differentiate into neurons or glial cells.

In the present studies, two lines (9 and 14) with similar patterns of transgene expression were used. Western blotting of E12.5 brain with a human specific anti-*Psen1* antibody showed that in both lines the human *Psen1* protein could be detected in transgenic brain while no signal was found in non-transgenic controls (Fig. 1). In addition, western blotting with a *Psen1* antibody that reacts with both the mouse and human proteins gave an approximately threefold higher signal in the transgenic embryos (Fig. 1). To establish the cell type specificity of transgene expression, we performed in situ hybridization on E13.5 embryos using a human *Psen1*-specific cRNA probe. In nestin-*Psen1* line 9 there was prominent human *Psen1* expression in the ventricular zone of the telencephalon as expected for a nestin-driven transgene (see Fig. S1 in the supplementary material; see Fig. 8).

Neither neural progenitor nor neuron specific expression of *Psen1* rescues the embryonic lethality of *Psen1*^{-/-} mice

Hemizygous nestin-*Psen1* transgenic mice were indistinguishable from wild-type littermates with regard to behavior and physical appearance. To determine whether selective expression of *Psen1* in neural progenitor cells could rescue the embryonic lethality of *Psen1*^{-/-} animals, we bred the

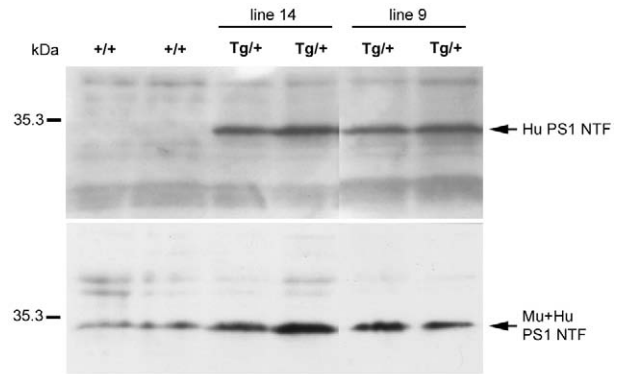


Fig. 1. Expression of human *PSEN1* protein in transgenic mice under the control of the nestin intron 2 enhancer. Western blotting was performed on brain extracts from E12.5 nestin-*Psen1* transgenic (Tg/+) embryos from line 9 and line 14, as well as non-transgenic controls (+/+). Blots were probed with NT.1 (upper panel) an antibody that recognizes the 30 kDa N-terminal fragment (NTF) of the transgenic human (Hu) *PSEN1* but not the endogenous mouse (Mu) *Psen1* (Wen et al., 2002b). The two wild-type (+/+) embryos are immunonegative. The lower panel was probed with a rabbit polyclonal antibody 222 that recognizes both the mouse and human NTFs (Wen et al., 2002b). Antibody 222 was raised against amino acids 2-12 of the human NTF, a region that contains one mismatch between the mouse and human proteins (Wen et al., 2002b). Thus, the apparent elevation in total *Psen1* (Mu + Hu) in the transgenic (Tg/+) embryos may in part reflect a higher affinity of this antibody for the human protein.

nestin-*Psen1* transgene onto the *Psen1*^{-/-} background. For this purpose, hemizygous transgenic/heterozygous *Psen1*^{+/-} animals were created and crossed with *Psen1*^{+/-} mice. From such matings one out of eight offspring should be *Psen1*^{-/-} if rescue is possible. We were not able to rescue live born *Psen1*^{-/-} animals from matings with either nestin-*Psen1* line 9 or 14 in over 40 pups generated from these matings ($P=0.004$ that no *Psen1*^{-/-} would be born).

Previously, we generated transgenic mice that express human wild-type *Psen1* in a neuron specific fashion under control of the neuron specific enolase (NSE) promoter/enhancer regions (Wen et al., 2002b; Wen et al., 2004b). The *Psen1* transgene in these lines was widely expressed in neurons in adult brain (Wen et al., 2002b). We also determined the expression pattern in developing brain by in situ hybridization. As shown in Fig. 2, in E16.5 brain the transgenic human *Psen1* in NSE-*Psen1* line 30 is widely expressed in neurons in the developing cortex but not in the ventricular zone, suggesting that transgene expression begins in the earliest stages of neuronal differentiation but is not present in neural progenitor cells. Thus, the NSE driven transgene seemed optimal to determine the effect of *Psen1* expression in the earliest developing postmitotic neurons in the absence of neural progenitor expression.

To determine if neuron-specific expression of *Psen1* could rescue the embryonic lethality of *Psen1*^{-/-} mice we bred NSE-*Psen1* line 30 onto the *Psen1*^{-/-} background. Again, we were unable to rescue live born *Psen1*^{-/-} mice in over 20 pups born from these matings ($P=0.06$). Thus, neither neuron-specific nor neural progenitor-specific expression of *Psen1* can rescue the embryonic lethality of the *Psen1*^{-/-} mouse.

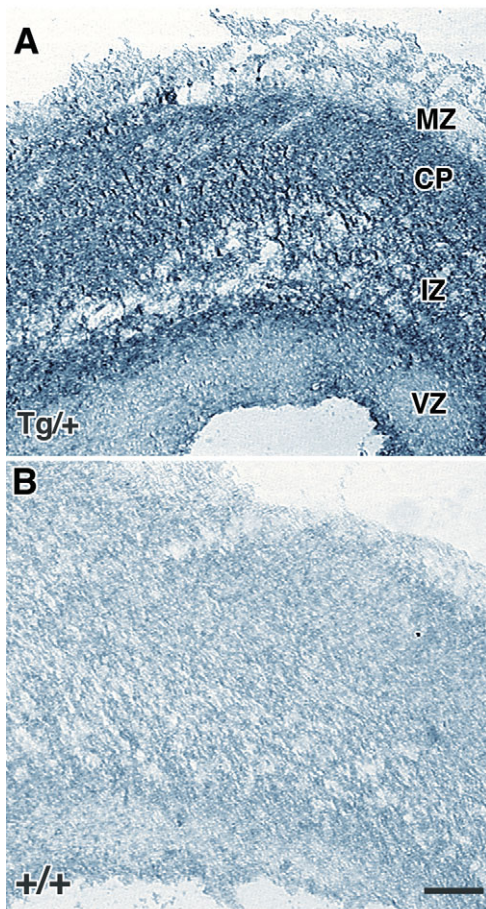


Fig. 2. Expression pattern of an NSE-Psen1 transgene. (A,B) Horizontal sections through the lateral telencephalon of an E16.5 NSE-Psen1 transgenic (A) or non-transgenic littermate control (B) embryos hybridized with a human-specific PSEN1 probe. In the transgenic embryo there is prominent hybridization in the intermediate zone (IZ), cortical plate (CP) and marginal zone (MZ), and an absence of signal in the ventricular zone (VZ). Scale bar: 50 μ m.

Selective expression of *Psen1* in neural progenitor cells rescues the cerebral hemorrhages and normalizes the appearance of the developing neocortex in *Psen1*^{-/-} embryos

As we could not rescue live born *Psen1*^{-/-} animals with either nestin- or NSE-driven transgenes we next examined the status of the embryos. By E12.5 *Psen1*^{-/-} embryos possess a generally hypomorphic caudal body with a kinked tail and by E18.5 a disorganized cortical laminar structure and almost invariably

Fig. 3. A nestin-Psen1 transgene rescues the cerebral hemorrhages but not the caudal defects in the *Psen1*^{-/-} embryo. (A-C) E18.5 wild-type (A), *Psen1*^{-/-} (B) embryos and an embryo with the nestin-Psen1 transgene on the *Psen1*^{-/-} background (C). The *Psen1*^{-/-} embryo has an abnormal tail region (arrow) and a cerebral hemorrhage (arrowhead). By contrast, the embryo with the nestin-Psen1 transgene on the *Psen1*^{-/-} background still exhibits the caudal defects (arrow) but no cerebral hemorrhages.



gross cerebral hemorrhages (Shen et al., 1997; Wong et al., 1997). Therefore, we examined E12.5 and E18.5 embryos from matings of hemizygous transgenic/heterozygous *Psen1*^{+/-} with *Psen1*^{+/-} animals. As shown in Fig. 3, E18.5 embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background still possessed a hypomorphic caudal body with a kinked tail, suggesting that the nestin driven transgene does not rescue the defects in somite segmentation in *Psen1*^{-/-} animals (Wong et al., 1997). However, none of the embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background evidenced gross cerebral hemorrhages that are nearly always present in pure *Psen1*^{-/-} embryos at E18.5 (Fig. 3).

Because defects in cortical development occur in *Psen1*^{-/-} mice (Hartmann et al., 1999; Shen et al., 1997), we examined the histology of the telencephalon in crosses of the nestin-Psen1 transgene onto the *Psen1*^{-/-} background. Cresyl Violet-stained sections of E12.5 and E18.5 embryos are shown in Figs 4 and 5. At E12.5 (Fig. 4), the telencephalon consists of a ventricular zone (VZ) and preplate (PP), and there were no obvious differences between *Psen1*^{+/+}, *Psen1*^{+/-}, *Psen1*^{-/-} mice or animals with the nestin-Psen1 transgene on the *Psen1*^{+/+}, *Psen1*^{+/-} or *Psen1*^{-/-} backgrounds. At E18.5, wild-type telencephalon shows a recognizable laminar structure (Fig. 5). By contrast, *Psen1*^{-/-} animals show hemorrhages and a general disorganization of the laminar structure of the cortex, even in areas that are free of hemorrhage. In many cortical areas, *Psen1*^{-/-} embryos also exhibited cellular ectopias in the marginal zone (Fig. 5) as well as the leptomenigeal gaps and thickening first described by Hartmann et al. (Hartmann et al., 1999).

None of six embryos with the nestin-Psen1 transgene on *Psen1*^{-/-} background derived from four separate litters showed microscopic evidence of hemorrhage and the cortical lamination pattern appeared identical to wild-type embryos (Fig. 5). Rescued embryos also showed no heterotopias in the marginal zone and the leptomeninges showed neither the gaps nor fibrosis seen in *Psen1*^{-/-} embryos (see Fig. S2 in the supplementary material). Likewise, the previously described loss of chondroitin sulfate proteoglycan staining in the marginal zone (Hartmann et al., 1999) was normalized (see Fig. S3 in the supplementary material), and reelin immunostaining (see Fig. S4 in the supplementary material) showed that Cajal-Retzius cells were as easily identified in the marginal zone of rescued animals as in wild-type embryos, showing no evidence for the loss of these cells that occurs in latter stage *Psen1*^{-/-} embryos (Hartmann et al., 1999; Kilb et al., 2004; Wines-Samuelson et al., 2005). Indeed, except for pure *Psen1*^{-/-} embryos, there were no differences between wild-type embryos and any of the genotypes that were generated in these crosses (i.e. *Psen1*^{+/-} or nestin-Psen1

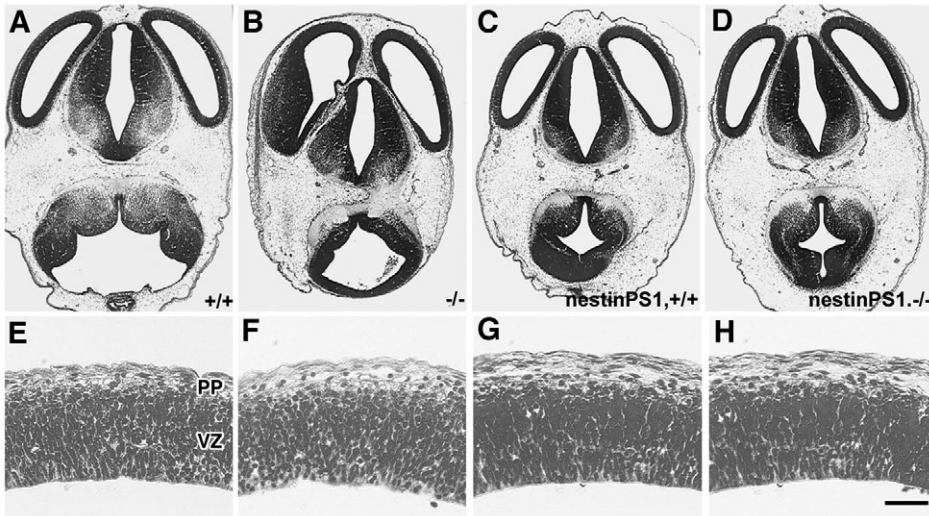


Fig. 4. Normal appearance of the developing cortical plate in E12.5 embryos with or without *Psen1*. Cresyl Violet-stained horizontal sections from E12.5 wild-type (A,E) and *Psen1*^{-/-} (B,F) embryos, as well embryos with the nestin-*Psen1* transgene on the *Psen1*^{+/-} (C,G) and *Psen1*^{-/-} backgrounds (D,H). (E-H) Higher power views through the lateral wall of the telencephalon. The ventricular zone (VZ) and preplate (PP) are indicated. No differences were apparent between any of the genotypes. Scale bar: 200 μ m for A-D; 25 μ m for E-H.

transgene on mouse *Psen1*^{+/-}, *Psen1*^{+/-} or *Psen1*^{-/-} backgrounds) Thus, neural progenitor specific expression of *Psen1* is sufficient to normalize cortical lamination patterns as well as prevent cerebral hemorrhages in *Psen1*^{-/-} embryos.

An NSE driven transgene rescues neither the cerebral hemorrhages nor the cortical lamination defects found in *Psen1*^{-/-} mice

We also examined brains from embryos with the NSE-*Psen1*

transgene on the *Psen1*^{-/-} background. As with the nestin-*Psen1* transgene, the NSE driven transgene failed to rescue the hypomorphic caudal body and kinked tail of *Psen1*^{-/-} animals (data not shown). In addition, E18.5 embryos with the NSE-*Psen1* transgene on the *Psen1*^{-/-} background had gross cerebral hemorrhages that were evident microscopically to the same degree found in pure *Psen1*^{-/-} embryos (Fig. 6), as well as cellular ectopias in the marginal zone and cortical lamination defects that were identical to those found in *Psen1*^{-/-} mice (Fig.

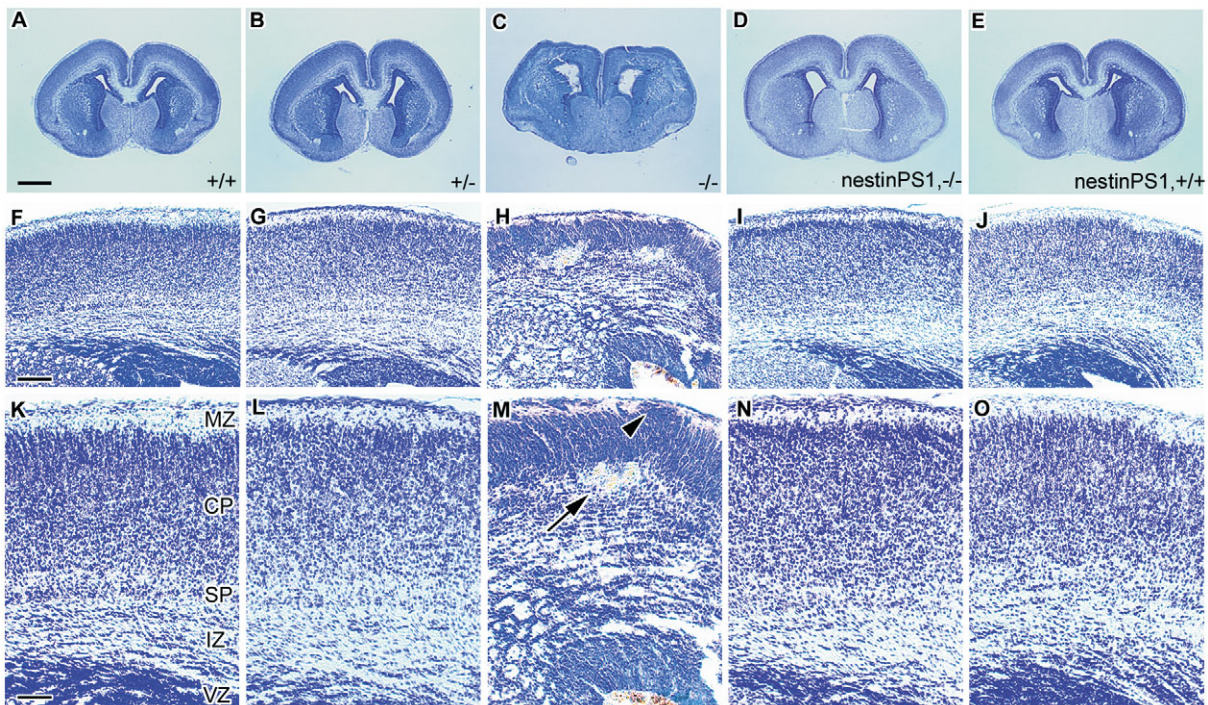


Fig. 5. Rescue of cerebral hemorrhages and cortical lamination defects in *Psen1*^{-/-} embryos with a nestin-*Psen1* transgene. Cresyl Violet-stained coronal sections of E18.5 embryos from wild-type (A,F,K), *Psen1*^{+/-} (B,G,L), *Psen1*^{-/-} (C,H,M), nestin-*Psen1* transgene on *Psen1*^{-/-} background (D,I,N) and nestin-*Psen1* transgene on *Psen1*^{+/-} background (E,J,O). (F-O) Higher power views through the lateral telencephalon. In the E18.5 embryos, the wild-type embryo shows a clear lamination pattern with a recognizable ventricular zone (VZ), intermediate zone (IZ), subplate (SP), cortical plate (CP) and marginal zone (MZ). The *Psen1*^{-/-} animal has multiple areas of hemorrhage (one indicated by arrow in M) and lacks distinct cortical layers. A cortical ectopia in the *Psen1*^{-/-} embryo is indicated by an arrowhead (M). By contrast, the embryo with the nestin-*Psen1* transgene on the *Psen1*^{-/-} background, as well as the *Psen1*^{+/-} and the embryo with the nestin-*Psen1* transgene on the *Psen1*^{+/-} background, are normal in appearance. Scale bars: 500 μ m in A-E; 100 μ m in F-J; 50 μ m in K-O.

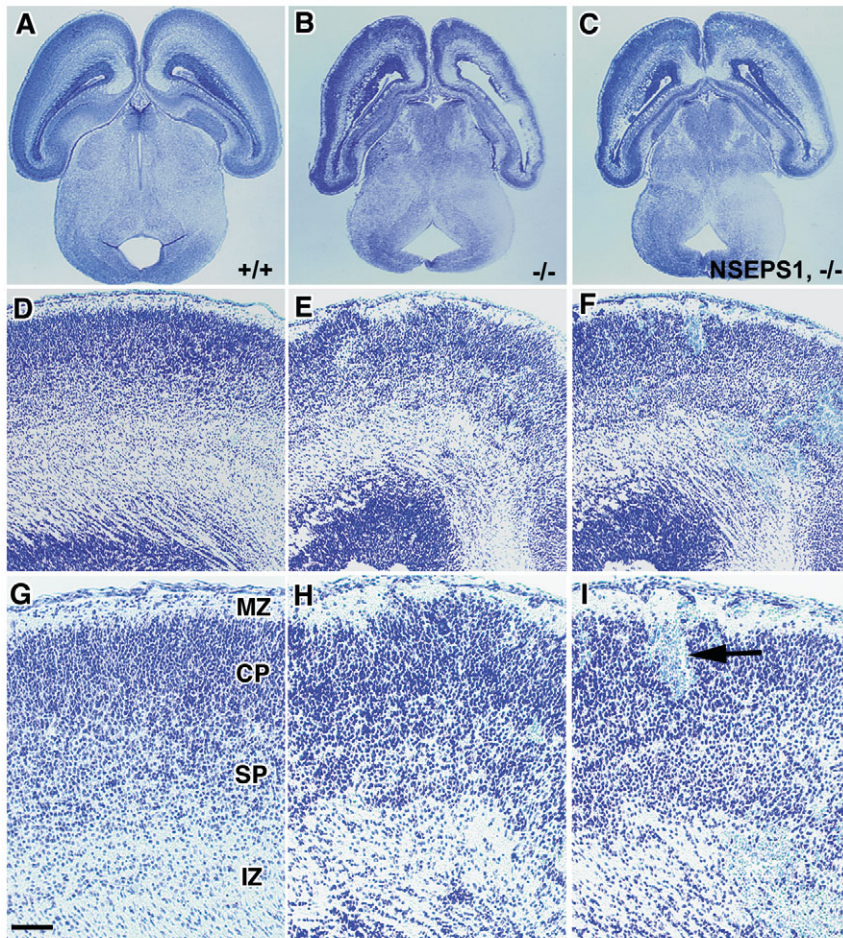


Fig. 6. Failure of an NSE-Psen1 transgene to rescue the brain pathology in *Psen1*^{-/-} mice. Shown are Cresyl Violet-stained horizontal sections of E18.5 embryos that are wild type (A,D,G), *Psen1*^{-/-} (B,E,H) or have the NSE-Psen1 transgene on *Psen1*^{-/-} background (C,F,I). (D-I) Higher power views of the lateral cerebral wall. The embryo with the NSE-Psen1 transgene on the *Psen1*^{-/-} background exhibits multiple areas of hemorrhage (one indicated by arrow), as well as disrupted cortical lamination in a pattern that is indistinguishable from that of the *Psen1*^{-/-} embryo. Cortical layers (MZ, CP, SP, IZ) are indicated as in Fig. 5. Scale bar: 500 μm in A-C; 100 μm in D-F; 50 μm in G-I.

patterns 2 hours after injections into E12.5 embryos. There were no discernable differences in labeling patterns between the various genotypes (see Fig. S5 in the supplementary material), indicating that at E12.5 the progenitor population in the ventricular zone is still grossly intact in the *Psen1*^{-/-} embryo and that this pattern is not affected by overexpression of Psen1 in neural progenitor cells.

Next, we injected BrdU at E12.5 and examined embryos at E18.5. As shown in Fig. 7, the majority of BrdU-labeled cells in a *Psen1*^{+/+} embryo were concentrated in the deeper layers of the cortical plate, a pattern expected for a wild-type animal. In *Psen1*^{-/-} embryos BrdU labeled cells were spread diffusely throughout the presumptive cortical

plate, similar to the cell migration defects observed by others (Handler et al., 2000; Yuasa et al., 2002). By contrast, BrdU labeled cells in embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background were concentrated in the deeper layers of the cortical plate as in wild-type animals. Embryos with the nestin-Psen1 transgene on the *Psen1*^{+/+} background or *Psen1*^{+/-} mice also had BrdU labeling patterns that were similar to wild-type embryos. These effects have been observed in at least three embryos for each genotype, in each case the embryos coming from more than one litter, and have been observed in embryos from crosses of both the nestin-Psen1 line 9 and line 14 onto the *Psen1*^{-/-} background.

6). Thus, expression of Psen1 in postmitotic neurons using an NSE promoter rescues none of the defects found in *Psen1*^{-/-} embryos.

Neural progenitor specific expression of Psen1 normalizes cell migration patterns in the telencephalon in *Psen1*^{-/-} embryos

Neural progenitor cells migrate prematurely into the developing neocortex of *Psen1*^{-/-} embryos (Handler et al., 2000; Yuasa et al., 2002). We therefore determined whether neural progenitor specific expression of Psen1 could rescue cell migration defects. Initially, we examined the BrdU labeling

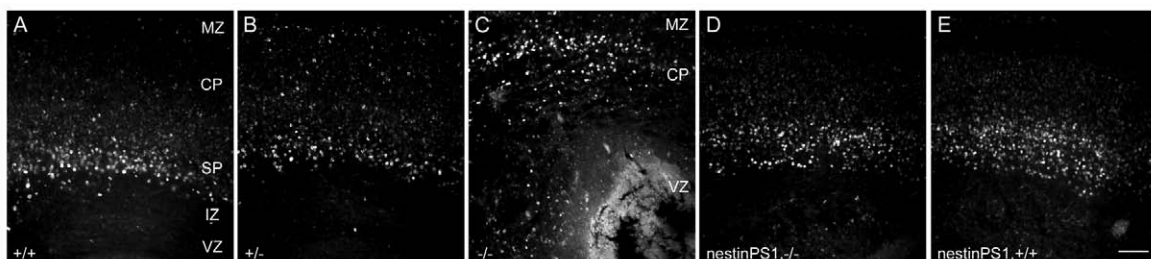


Fig. 7. Rescue of cell migration defects in *Psen1*^{-/-} embryos with a nestin-Psen1 transgene. Pregnant females were given one dose of BrdU at E12.5 and the embryos were collected at E18.5 and immunostained with anti-BrdU antibodies. Shown are BrdU-immunostained sections through the lateral cerebral wall of E18.5 embryos from wild-type (A), *Psen1*^{+/-} (B), *Psen1*^{-/-} (C), nestin-Psen1 transgene on *Psen1*^{-/-} background (D) and nestin-Psen1 transgene on *Psen1*^{+/+} background (E). BrdU labeling is dispersed in the *Psen1*^{-/-} embryo and normalized by the nestin-Psen1 transgene. Cortical layers (MZ, CP, SP, IZ, VZ) are indicated as in Fig. 5. Scale bar: 100 μm.

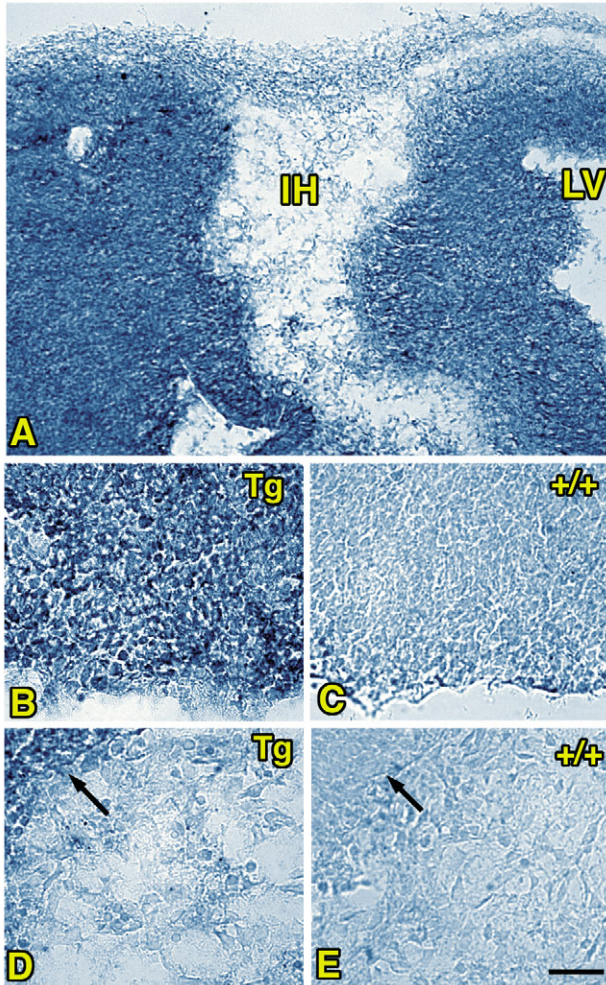


Fig. 8. Lack of detectable transgene expression in cephalic mesenchyme by in situ hybridization with a human-specific *Psen1* cRNA. Shown are horizontal sections through the telencephalon of an E12.5 nestin-*Psen1* transgenic embryo from line 9 (A,B,D) or a non-transgenic littermate control (C,E) embryo hybridized with a human specific *Psen1* probe. (A) A section through the anterior telencephalon. The lateral ventricle (LV) and interhemispheric fissure (IH) are indicated. There is prominent hybridization in the brain parenchyma. Hybridization in the ventricular zone from a transgenic (Tg) and non-transgenic (+/+) embryo is shown in B and C. The transgenic embryo is strongly hybridized. (D) Hybridization at the juncture between the brain parenchyma and the overlying cephalic mesenchyme in the interhemispheric fissure from a transgenic embryo. (E) A comparable region from a non-transgenic embryo (+/+) is shown to indicate the level of background staining. An arrow indicates the border between the brain parenchyma (showing hybridization product in D but not in E) and the cephalic mesenchyme. Scale bar: 20 μ m for A; 50 μ m for B-E.

The nestin intron 2 enhancer is not active in vascular endothelial cells or vascular progenitor cells

The brain vascular system develops from mesodermally derived angioblasts that invade the brain from overlying mesenchymal tissue (Harrigan, 2003). Additional blood vessels form via sprouting or splitting of initially established

vessels (Harrigan, 2003). Thus, rescue of the cerebral hemorrhages is somewhat surprising given that the nestin intron 2 enhancer is not regarded as active in mesoderm (Zimmerman et al., 1994).

As leakage of the transgene into mesodermal- or possibly neural crest-derived cephalic mesenchymal cells would provide a trivial explanation for the rescue of the cerebral hemorrhages, we re-examined transgene expression pattern in E12.5 brain by in situ hybridization, with particular attention as to whether the transgenic human *PSEN1* was expressed in the overlying cephalic mesenchyme. As expected, strong hybridization was seen within the brain parenchyma of the transgenic specimen, with the signal being especially prominent in cells surrounding the ventricular zone (Fig. 8). Owing to the resolution of the digoxigenin detection system, it was difficult to determine unequivocally whether hybridization product was present in endothelial cells within brain parenchyma. However, no hybridization was apparent in any region of the overlying cephalic mesenchyme, including blood vessels and perivascular cells within this structure (Fig. 8) indicating that the transgene is not expressed in mesoderm or in neural crest-derived elements that migrate into this region (Etchevers et al., 1999).

We have also generated transgenic mice expressing Cre recombinase under the control of the nestin intron 2 enhancer/tk promoter (NesCrenls mice). Cre expression in these lines has been verified by both western blotting and by immunohistochemistry on E13.5 mouse brain (data not shown). Because low-level transgene expression could be missed by in situ hybridization, as an additional test of nestin intron 2 enhancer activity, we crossed NesCrenls transgenic mice to two lines of Cre reporter mice, a technique that should be sensitive to even low levels of nestin intron 2 enhancer activity. NesCrenls line 27 mice were firstly crossed with a Cre/loxP reporter line cActXstopXEGFP44. This line serves as an indicator of Cre expression in that Cre removes a *loxP* flanked stop cassette sequence and activates EGFP expression. Spontaneous EGFP fluorescence was imaged and combined with immunohistochemistry for aquaporin 4, which is expressed in astrocytic foot processes that surround blood vessels and thus serves as an effective vascular marker (Verkman, 2002). EGFP expression from the hippocampal CA1 region of an adult double transgenic mouse is shown in Fig. 9, where prominent expression of EGFP is seen in CA1 pyramidal cells while no EGFP expression is seen in the aquaporin outlined vessels.

As an independent test of nestin intron 2/tk promoter activity, we also crossed NesCrenls lines 27 and 13 mice with the Z/EG reporter line (Novak et al., 2000). In this line, Cre-mediated recombination removes a *lacZ* gene and activates expression of an EGFP reporter under the control of CMV/chicken β -actin promoter. These mice have been used in several recent studies to follow Cre-mediated excision in both early embryonic and adult tissues (Guo et al., 2002; Malatesta et al., 2003; Novak et al., 2000). Crosses with both NesCrenls lines gave similar results in showing widespread EGFP activation in adult brain but no activation in or around brain vessels. Immunohistochemical staining for EGFP combined with staining for von Willebrand factor as an endothelial cell marker (Costa et al., 2001) is shown in Fig. 9, where EGFP labeled cells can be seen to the left and right of a large

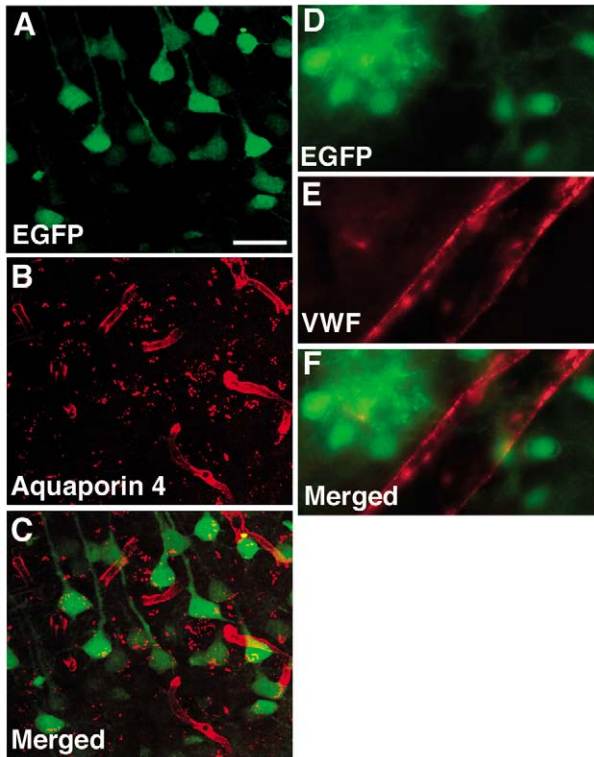


Fig. 9. Lack of the nestin intron 2 enhancer activity in vascular progenitor cells or endothelial cells. NesCrenls line 27 was crossed with a Cre/loxP reporter line cActXstopXEGFP44. The hippocampal CA1 region of an adult double transgenic mouse is shown in A-C. Spontaneous EGFP fluorescence (A) was imaged and combined with immunohistochemistry for aquaporin 4 (B). EGFP is prominently expressed in the pyramidal cells but not in the aquaporin 4 outlined vessels (merged image is shown in C). (D-F) Combined immunohistochemical staining is shown for EGFP (D), and von Willebrand factor (E) on an adult brain from a NesCrenls line 27 mouse crossed to the Z/EG reporter line (merged image is shown in F). Immunostaining in D-F was performed on sections cut from tissue perfusion fixed with paraformaldehyde. A large penetrating vessel is visible at the cortical surface. There is no EGFP labeling of the von Willebrand factor-stained vessel. Scale bar: 50 μm for A-C; 10 μm for D-F.

penetrating vessel at the cortical surface, while no EGFP expression is apparent in the von Willebrand factor stained vessel. In these same sections, we have also been unable to detect EGFP activation in either aquaporin 4- or fibronectin- (Peters and Hynes, 1996) labeled vessels (data not shown). Thus, collectively, these studies show that the nestin intron 2 enhancer is not active in vascular endothelial cells or in vascular progenitor cells at any stage of development.

Selective expression of *Psen1* in neural progenitor cells restores a normal vasculature in *Psen1*^{-/-} embryos

Cerebral hemorrhages can occur in *Psen1*^{-/-} embryos as early as E12.5 but are unusual at this age, while at E18.5 intracerebral as well as intraventricular hemorrhages are invariably present. To investigate how *Psen1* expression in neural progenitor cells might influence vascular development, we labeled vascular endothelial cells in *Psen1*^{-/-} and wild-type

embryos with Griffonia simplicifolia isolectin B4 (Ashwell, 1991). In E18.5 *Psen1*^{-/-} brains, lectin-stained vessels in the cerebral hemispheres appeared distorted and more lobular than vessels in wild-type brains (Fig. 10). Such changes were evident even in areas without macroscopic or microscopic hemorrhage, and were also present in vessels in the ganglionic eminence (data not shown). In E12.5 embryos, differences between vessels in *Psen1*^{-/-} and wild-type control brains were more subtle. However, vessels in both the cortical plate (Fig. 10) and the ganglionic eminence (data not shown) of *Psen1*^{-/-} brain appeared enlarged compared with wild type. These changes at both E12.5 and E18.5 were, however, completely normalized in embryos with the nestin-*Psen1* transgene on the *Psen1*^{-/-} background (Fig. 10).

During cerebral angiogenesis, endothelial cells form close contacts with perivascular mural cells or pericytes (Beck and D'Amore, 1997), and once within the neuroectoderm, endothelial/pericyte complexes make close contacts with brain cells, including radial glia (Bass et al., 1992). To address whether defective interactions between neuroepithelial cells and developing vessels might be occurring in *Psen1*^{-/-} embryos, we used nestin immunostaining as a marker of neuroepithelial cells and isolectin B4 staining as a vascular marker. Nestin staining in E12.5 *Psen1*^{-/-} telencephalon appeared similar to wild-type embryos, with many radially distributed processes extending throughout the telencephalic wall (data not shown). At E18.5, nestin staining in wild-type controls continued to show many radially distributed processes extending throughout the full thickness of the cerebral hemispheres (Fig. 10) with some nestin-stained processes traversing near cerebral vessels. By contrast, in E18.5 *Psen1*^{-/-} animals, nestin staining was largely absent in the cerebral hemispheres with at most sparse fibers that failed to extend through the full thickness of the cerebral wall, a pattern that was evident even in areas that lacked cerebral hemorrhages. Interestingly, the nestin staining that was present in *Psen1*^{-/-} brains was concentrated around and possibly within distorted cerebral vessels. Similar patterns were observed in both the cerebral hemispheres (Fig. 10) and the developing caudate/putamen (data not shown) of *Psen1*^{-/-} embryos. However, in embryos with the nestin-*Psen1* transgene on the *Psen1*^{-/-} background, many radially directed processes were nestin stained, and these processes associated with brain vessels in a pattern that was identical to wild-type controls (Fig. 10).

Discussion

Psen1-null mutant mice die during late intrauterine life or shortly after birth (Shen et al., 1997; Wong et al., 1997). The embryos are small and exhibit a generally hypomorphic caudal body with a kinked tail, as well as skeletal malformations. Abnormalities in the tail region can be seen as early as E9.5, with the somites being indistinct and largely absent in the most caudal region (Wong et al., 1997). By E15.5, the vertebral column is shortened and fails to undergo proper segmentation, resulting in fused vertebrae. Some embryos have midline defects in the body wall and cranial vault (Hartmann et al., 1999). The telencephalon appears relatively normal at E12.5. However, by E14.5 to E15.5 homozygous mutants exhibit abnormalities in cortical and subcortical development and, by

E18.5, invariably display CNS hemorrhages (Handler et al., 2000; Louvi et al., 2004; Shen et al., 1997). The cellular basis for these defects remains largely unexplained. A prior study (Nakajima et al., 2003), reported abnormal vascular dilation in *Psen1*^{-/-} embryos, similar to that observed here, as well as reduced capillary sprouting and abnormal vascular morphologies, including endothelial cell multiplication, apoptosis and necrosis. Although one study has suggested that neural progenitor cells differentiate prematurely in *Psen1*^{-/-} mice (Handler et al., 2000), a more recent study has cast doubt on the generality of this conclusion (Wen et al., 2004a).

We show that selective expression of Psen1 in neural progenitors fails to rescue the embryonic lethality of *Psen1*^{-/-} mice, as well as failing to rescue the hypomorphic caudal body malformations. However within brain, the nestin-Psen1 transgene rescued the cortical lamination defects found in *Psen1*^{-/-} mice, normalized cell migration patterns (Handler et al., 2000; Yuasa et al., 2002) and eliminated the CNS hemorrhages. By contrast neuron-specific expression rescued neither the embryonic lethality nor any of the histological abnormalities in the brain. The ability of a nestin-driven transgene to rescue the cortical abnormalities in *Psen1*^{-/-} mice documents a central role for Psen1 expression in neural progenitor cells. More surprising is the ability of the transgene to rescue the cerebral hemorrhages, because the brain vascular system develops from mesodermally derived angioblasts that invade brain from overlying leptomeninges (Marin-Padilla, 1985).

Several studies have suggested that vascular endothelial cells, including embryonic blood vessels in brain as well as pericytes and periendothelial cells may express nestin based on immunostaining with antibodies such as Rat 401 (Alliot et al., 1999; Mokry and Nemecek, 1998; Mokry and Nemecek, 1999; Tohyama et al., 1992). However, we found no evidence in *in situ* hybridization of transgene expression in the overlying cephalic mesenchyme, including blood vessels contained within this region and there was also no evidence for activity of the nestin intron 2 enhancer in endothelial cells or vascular progenitor cells when this enhancer was used to drive Cre recombinase and crossed to Cre reporter lines. Thus, leakage of the transgene into cephalic mesenchymal elements, whether mesodermal or neural crest derived does not appear to be the basis for the rescue of the vascular abnormalities. The failure of the nestin-Psen1 transgene to rescue the caudal defects in the *Psen1*^{-/-} mouse also argues against mesodermal expression. Several other studies have also failed to detect vascular activity of the nestin intron 2 enhancer in transgenic mice when it was used to drive EGFP directly (Filippov et al., 2003; Kawaguchi et al., 2001; Mignone et al., 2004; Yamaguchi et al., 2000).

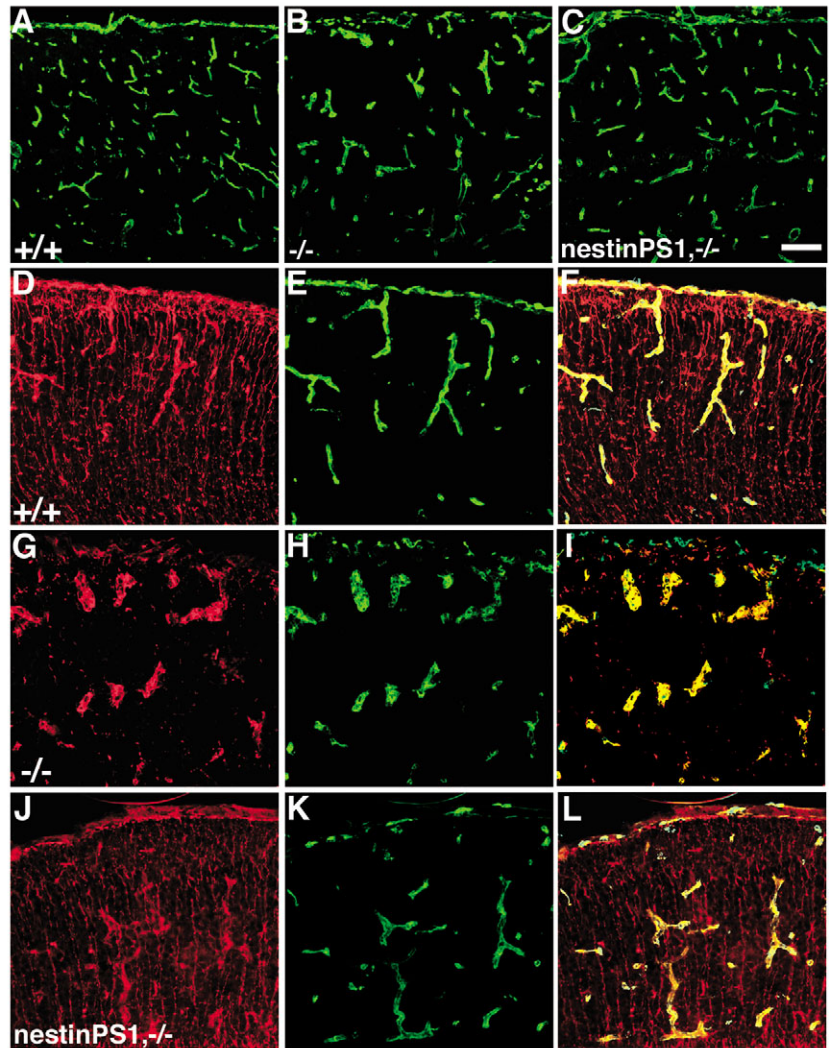


Fig. 10. Rescue of brain vascular abnormalities in embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background. (A-C) Staining with isolectin B4 was performed on paraffin-embedded E12.5 embryos from wild-type (A) and *Psen1*^{-/-} (B) embryos, and embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background (C). Sections through the lateral telencephalon are shown. Vessels in the *Psen1*^{-/-} embryo are enlarged. (D-L) Isolectin staining (E,H,K) was combined with nestin immunostaining (D,G,J) on paraffin-embedded E18.5 embryos from wild-type (D-F) and *Psen1*^{-/-} (G-I) embryos, and embryos with the nestin-Psen1 transgene on the *Psen1*^{-/-} background (J-L). Merged images are shown in F,I,L. Regions along the cortical surface of the lateral telencephalon are illustrated. The lectin-stained vessels in the *Psen1*^{-/-} brain are distorted and lobular appearance, and this pattern is normalized by the nestin-Psen1 transgene. The pattern of nestin staining is altered in the *Psen1*^{-/-} embryo. Scale bar: 100 μ m for A-C; 50 μ m for D-L.

In addition, another study (Wines-Samuelson et al., 2005) recently reported the production of a conditional knockout of Psen1 in neural progenitor cells by crossing a nestin-Cre transgene onto a floxed *Psen1* background. Although the cortical phenotype in these mice was milder than a pure *Psen1* null mutant, the conditional knockout mice exhibited widespread intracranial hemorrhages that were thought to be a proximal cause of postnatal death in these animals. This study is thus consistent with our results suggesting that neural progenitor specific expression of Psen1 is necessary for the development of a competent vasculature.

Collectively, these results argue that *Psen1* is essential for neural-derived signals that are necessary for formation of a normal vasculature. The disturbed nestin staining pattern in *Psen1*^{-/-} embryos points to an abnormal association of neuroepithelial cells with brain blood vessels. In this context, two recent studies have also reported abnormal radial glial development in *Psen1*^{-/-} embryos (Louvi et al., 2004; Wines-Samuelson et al., 2005). At the molecular level, it remains unclear how *Psen1* expression in neural progenitor might mediate this effect. *Psen1* influences a variety of pathways known to be important developmentally (Koo and Kopan, 2004). Yet whatever the mechanisms of action of *Psen1*, the studies described here show that *Psen1* expression in neural progenitor cells is sufficient to rescue completely the brain abnormalities found in *Psen1*^{-/-} mice, including the unexpected rescue of the vascular defects. As such, the combination of nestin-*Psen1* transgenic mice and the *Psen1*-null mutant provide a model for dissecting the specific molecular pathways that *Psen1* influences in neural progenitor cells.

This work was supported by grants from the National Institutes of Health (AG20139, AG21305, MH70603, AG23599, AG02219 and AG05138). Transgenic mice were generated through the Mount Sinai School of Medicine Mouse Genetics Shared Resource Facility. We thank Dr Paul Matthews for supplying the NT.1 antibody and Dr Nikolaos Robakis for antibody 222. We also thank Drs Ron McKay, Stephen O'Gorman, Eric Mercer and David Anderson for providing plasmids, and Dr Huntington Potter for making the *Psen1*^{-/-} mice available. P.R.H. is the Regenstreif Professor of Neuroscience. R.D.G. is a recipient of a Young Investigator Award from the National Alliance for Research in Schizophrenia and Affective Disorders (NARSAD).

Supplementary material

Supplementary material for this article is available at <http://dev.biologists.org/cgi/content/full/132/17/3873/DC1>

References

- Alliot, F., Rutin, J., Leenen, P. J. and Pessac, B. (1999). Pericytes and periendothelial cells of brain parenchyma vessels co-express aminopeptidase N, aminopeptidase A, and nestin. *J. Neurosci. Res.* **58**, 367-378.
- Aoki, Y., Huang, Z., Thomas, S. S., Bhide, P. G., Huang, I., Moskowitz, M. A. and Reeves, S. A. (2000). Increased susceptibility to ischemia-induced brain damage in transgenic mice overexpressing a dominant negative form of SHP2. *FASEB J.* **14**, 1965-1973.
- Ashwell, K. (1991). The distribution of microglia and cell death in the fetal rat forebrain. *Dev. Brain Res.* **58**, 1-12.
- Bass, T., Singer, G., Slusser, J. and Liuzzi, F. J. (1992). Radial glial interaction with cerebral germinal matrix capillaries in the fetal baboon. *Exp. Neurol.* **118**, 126-132.
- Beck, L., Jr and D'Amore, P. A. (1997). Vascular development: cellular and molecular regulation. *FASEB J.* **11**, 365-373.
- Costa, C., Harding, B. and Copp, A. J. (2001). Neuronal migration defects in the Dreher (*Lmx1a*) mutant mouse: role of disorders of the glial limiting membrane. *Cereb. Cortex* **11**, 498-505.
- Cribbs, D. H., Chen, L. S., Cotman, C. W. and LaFerla, F. M. (1996). Injury induces presenilin-1 gene expression in mouse brain. *NeuroReport* **7**, 1773-1776.
- Cruts, M., van Duijn, C. M., Backhovens, H., Van den Broeck, M., Wehnert, A., Serneels, S., Sherrington, R., Hutton, M., Hardy, J., St George-Hyslop, P. H. et al. (1998). Estimation of the genetic contribution of presenilin-1 and -2 mutations in a population-based study of presenile Alzheimer disease. *Hum. Mol. Genet.* **7**, 43-51.
- Elder, G. A., Tezapsidis, N., Carter, J., Shioi, J., Bouras, C., Li, H.-C., Johnston, J. M., Efthimiopoulos, S., Friedrich, V. L. and Robakis, N. K. (1996). Identification and neuron specific expression of the S182/presenilin 1 protein in human and rodent brains. *J. Neurosci. Res.* **45**, 308-320.
- Etchevers, H. C., Couly, G., Vincent, C. and Le Douarin, N. M. (1999). Anterior cephalic neural crest is required for forebrain viability. *Development* **126**, 3533-3543.
- Filippov, V., Kronenberg, G., Pivneva, T., Reuter, K., Steiner, B., Wang, L. P., Yamaguchi, M., Kettenmann, H. and Kempermann, G. (2003). Subpopulation of nestin-expressing progenitor cells in the adult murine hippocampus shows electrophysiological and morphological characteristics of astrocytes. *Mol. Cell. Neurosci.* **23**, 373-382.
- Frisen, J., Johansson, C. B., Lothian, C. and Lendahl, U. (1998). Central nervous system stem cells in the embryo and adult. *Cell Mol. Life Sci.* **54**, 935-945.
- Guo, C., Yang, W. and Lobe, C. G. (2002). A cre recombinase transgene with mosaic, widespread tamoxifen-inducible action. *Genesis* **32**, 8-18.
- Handler, M., Yang, X. and Shen, J. (2000). Presenilin-1 regulates neuronal differentiation during neurogenesis. *Development* **127**, 2593-2606.
- Harrigan, M. R. (2003). Angiogenic factors in the central nervous system. *Neurosurgery* **53**, 639-660.
- Hartmann, D., De Strooper, B. and Saftig, P. (1999). Presenilin-1 deficiency leads to loss of Cajal-Retzius neurons and cortical dysplasia similar to human type 2 lissencephaly. *Curr. Biol.* **9**, 719-727.
- Jeong, S. J., Kim, H. S., Chang, K. A., Geum, D. H., Park, C. H., Seo, J. H., Rah, J. C., Lee, J. H., Choi, S. H., Lee, S. G. et al. (2000). Subcellular localization of presenilins during mouse preimplantation development. *FASEB J.* **14**, 2171-2176.
- Josephson, R., Muller, T., Pickel, J., Okabe, S., Reynolds, K., Turner, P. A., Zimmer, A. and McKay, R. D. (1998). POU transcription factors control expression of CNS stem cell-specific genes. *Development* **125**, 3087-3100.
- Kawaguchi, A., Miyata, T., Sawamoto, K., Takashita, N., Murayama, A., Akamatsu, D., Ogawa, M., Okabe, M., Tano, Y., Goldman, S. A. et al. (2001). Nestin-EGFP transgenic mice: visualization of the self-renewal and multipotency of CNS stem cells. *Mol. Cell. Neurosci.* **17**, 259-273.
- Kilb, W., Hartmann, D., Saftig, P. and Luhmann, H. J. (2004). Altered morphological and electrophysiological properties of Cajal-Retzius cells in cerebral cortex of embryonic presenilin-1 knockout mice. *Eur. J. Neurosci.* **20**, 2749-2756.
- Koo, E. H. and Kopan, R. (2004). Potential role of presenilin-regulated signaling pathways in sporadic neurodegeneration. *Nat. Med.* **10**, S26-S33.
- Kostyszyn, B., Cowburn, R. F., Seiger, A., Kij, A. A. and Sundstrom, E. (2001). Expression of presenilin-1 and notch-1 receptor in human embryonic CNS. *Neuroscience* **103**, 885-898.
- Lakso, M., Sauer, B., Mosinger, B., Jr, Lee, E. J., Manning, R. W., Yu, S. H., Mulder, K. L. and Westphal, H. (1992). Targeted oncogene activation by site-specific recombination in transgenic mice. *Proc. Natl. Acad. Sci. USA* **89**, 6232-6236.
- Lendahl, U., Zimmerman, L. B. and McKay, R. D. (1990). CNS stem cells express a new class of intermediate filament protein. *Cell* **60**, 585-595.
- Lleo, A., Berezovska, O., Growdon, J. H. and Hyman, B. T. (2004). Clinical, pathological, and biochemical spectrum of Alzheimer disease associated with PS-1 mutations. *Am. J. Geriatr. Psychiat.* **12**, 146-156.
- Louvi, A., Sisodia, S. S. and Grove, E. A. (2004). Presenilin 1 in migration and morphogenesis in the central nervous system. *Development* **131**, 3093-3105.
- Malatesta, P., Hack, M. A., Hartfuss, E., Kettenmann, H., Klinkert, W., Kirchhoff, F. and Gotz, M. (2003). Neuronal or glial progeny: regional differences in radial glia fate. *Neuron* **37**, 751-764.
- Marin-Padilla, M. (1985). Early vascularization of the embryonic cerebral cortex: Golgi and electron microscopic studies. *J. Comp. Neurol.* **241**, 237-249.
- McKay, R. (1997). Stem cells in the central nervous system. *Science* **276**, 66-71.
- Mignone, J. L., Kukekov, V., Chiang, A. S., Steindler, D. and Enikolopov, G. (2004). Neural stem and progenitor cells in nestin-GFP transgenic mice. *J. Comp. Neurol.* **469**, 311-324.
- Mokry, J. and Nemecek, S. (1998). Angiogenesis of extra- and intraembryonic blood vessels is associated with expression of nestin in endothelial cells. *Folia Biol. (Praha)* **44**, 155-161.
- Mokry, J. and Nemecek, S. (1999). Cerebral angiogenesis shows nestin expression in endothelial cells. *Gen. Physiol. Biophys.* **18**, 25-29.
- Moreno-Flores, M. T., Medina, M. and Wandosell, F. (1999). Expression of presenilin 1 in nervous system during rat development. *J. Comp. Neurol.* **410**, 556-570.

- Nakajima, M., Yuasa, S., Ueno, M., Takakura, N., Koseki, H. and Shirasawa, T. (2003). Abnormal blood vessel development in mice lacking presenilin-1. *Mech. Dev.* **120**, 657-667.
- Novak, A., Guo, C., Yang, W., Nagy, A. and Lobe, C. G. (2000). Z/EG, a double reporter mouse line that expresses enhanced green fluorescent protein upon cre-mediated excision. *Genesis* **28**, 147-155.
- O'Gorman, S., Dagenais, N. A., Qian, M. and Marchuk, Y. (1997). Protamine-Cre recombinase transgenes efficiently recombine target sequences in the male germline of mice, but not in embryonic stem cells. *Proc. Natl. Acad. Sci. USA* **94**, 14602-14607.
- Peters, J. H. and Hynes, R. O. (1996). Fibronectin isoform distribution in the mouse. I. The alternatively spliced EIIIB, EIIIA, and V segments show widespread codistribution in the developing mouse embryo. *Cell Adhes. Commun.* **4**, 103-125.
- Shen, J., Bronson, R. T., Chen, D. F., Xia, W., Selkoe, D. J. and Tonegawa, S. (1997). Skeletal and CNS defects in presenilin-1-deficient mice. *Cell* **89**, 629-639.
- Tohyama, T., Lee, V. M., Rorke, L. B., Marvin, M., McKay, R. D. and Trojanowski, J. Q. (1992). Nestin expression in embryonic human neuroepithelium and in human neuroepithelial tumor cells. *Lab. Invest.* **66**, 303-313.
- Verkman, A. S. (2002). Aquaporin water channels and endothelial cell function. *J. Anat.* **200**, 617-627.
- Weggen, S., Diehlmann, A., Buslei, R., Beyreuther, K. and Bayer, T. A. (1998). Prominent expression of presenilin-1 in senile plaques and reactive astrocytes in Alzheimer's disease brain. *NeuroReport* **9**, 3279-3283.
- Wen, P. H., Friedrich, V. L., Jr, Shioi, J., Robakis, N. K. and Elder, G. A. (2002a). Presenilin-1 is expressed in neural progenitor cells in the hippocampus of adult mice. *Neurosci. Lett.* **318**, 53-56.
- Wen, P. H., Shao, X., Shao, Z., Hof, P. R., Wisniewski, T., Kelley, K., Friedrich, V. L., Jr, Ho, L., Pasinetti, G. M., Shioi, J. et al. (2002b). Overexpression of wild type but not an FAD mutant presenilin-1 promotes neurogenesis in the hippocampus of adult mice. *Neurobiol. Dis.* **10**, 8-19.
- Wen, P. H., De Gasperi, R., Gama Sosa, M. A. and Elder, G. A. (2004a). Neural progenitor cells do not differentiate prematurely in presenilin-1 null mutant mice. *Neurosci. Lett.* **371**, 249-254.
- Wen, P. H., Hof, P. R., Chen, X., Gluck, K., Austin, G., Younkin, S. G., Younkin, L. H., DeGasperi, R., Gama Sosa, M. A., Robakis, N. K. et al. (2004b). The presenilin-1 familial Alzheimer disease mutant P117L impairs neurogenesis in the hippocampus of adult mice. *Exp. Neurol.* **188**, 224-237.
- Wines-Samuelson, M., Handler, M. and Shen, J. (2005). Role of presenilin-1 in cortical lamination and survival of Cajal-Retzius neurons. *Dev. Biol.* **277**, 332-346.
- Wong, P. C., Zheng, H., Chen, H., Becher, M. W., Sirinathsinghji, D. J., Trumbauer, M. E., Chen, H. Y., Price, D. L., Van der Ploeg, L. H. and Sisodia, S. S. (1997). Presenilin 1 is required for notch1 and DII1 expression in the paraxial mesoderm. *Nature* **387**, 288-292.
- Yamaguchi, M., Saito, H., Suzuki, M. and Mori, K. (2000). Visualization of neurogenesis in the central nervous system using nestin promoter-GFP transgenic mice. *NeuroReport* **11**, 1991-1996.
- Yuasa, S., Nakajima, M., Aizawa, H., Sahara, N., Koizumi, K., Sakai, T., Usami, M., Kobayashi, S., Kuroyanagi, H., Mori, H. et al. (2002). Impaired cell cycle control of neuronal precursor cells in the neocortical primordium of presenilin-1-deficient mice. *J. Neurosci. Res.* **70**, 501-513.
- Zimmerman, L., Parr, B., Lendahl, U., Cunningham, M., McKay, R., Gavin, B., Mann, J., Vassileva, G. and McMahon, A. (1994). Independent regulatory elements in the nestin gene direct transgene expression to neural stem cells or muscle precursors. *Neuron* **12**, 11-24.