

RESEARCH ARTICLE

Genome-wide identification of phospho-regulators of Wnt signaling in *Drosophila*

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ABSTRACT

Evolutionarily conserved intercellular signaling pathways regulate embryonic development and adult tissue homeostasis in metazoans. The precise control of the state and amplitude of signaling pathways is achieved in part through the kinase- and phosphatase-mediated reversible phosphorylation of proteins. In this study, we performed a genome-wide *in vivo* RNAi screen for kinases and phosphatases that regulate the Wnt pathway under physiological conditions in the *Drosophila* wing disc. Our analyses have identified 54 high-confidence kinases and phosphatases capable of modulating the Wnt pathway, including 22 novel regulators. These candidates were also assayed for a role in the Notch pathway, and numerous phospho-regulators were identified. Additionally, each regulator of the Wnt pathway was evaluated in the wing disc for its ability to affect the mechanistically similar Hedgehog pathway. We identified 29 dual regulators that have the same effect on the Wnt and Hedgehog pathways. As proof of principle, we established that Cdc37 and Gilgamesh/CK1 γ inhibit and promote signaling, respectively, by functioning at analogous levels of these pathways in both *Drosophila* and mammalian cells. The Wnt and Hedgehog pathways function in tandem in multiple developmental contexts, and the identification of several shared phospho-regulators serve as potential nodes of control under conditions of aberrant signaling and disease.

KEY WORDS: Wnt, Wingless, *In vivo* RNAi screen, Hedgehog, Notch

INTRODUCTION

The canonical Wnt signaling pathway is evolutionarily conserved and regulates essential biological processes such as cell fate specification, proliferation and migration during metazoan development. As a consequence, aberrant Wnt signaling can result in diverse human developmental disorders and cancers (MacDonald et al., 2009; Wodarz and Nusse, 1998). Cells use the reversible phosphorylation of proteins to control the state and amplitude of signaling pathways (Cohen, 1992; Hunter, 1995; Salazar and Höfer, 2009). In the silent state of the Wnt pathway, the transcriptional effector β -catenin is phosphorylated within a cytosolic Axin (Axn) complex by Casein Kinase 1 α (CK1 α) and Glycogen Synthase Kinase 3 β (GSK3 β). This modification facilitates its poly-ubiquitination and degradation (Aberle et al., 1997; Amit et al., 2002; Kitagawa et al., 1999; Liu et al., 1999, 2002). In the absence of stabilized β -catenin, the DNA-binding protein TCF represses target gene expression (Cavallo et al., 1998;

Roose et al., 1998). Binding of secreted Wnt ligand to its transmembrane Frizzled (Fz) receptor and LRP co-receptor initiates pathway activity through the recruitment of the Axn complex to the cell surface (Cliffe et al., 2003; Tamai et al., 2004; Umbhauer et al., 2000; Wong et al., 2003; Yang-Snyder et al., 1996). This induces the phosphorylation of LRP by CK1 α , GSK3 β and the plasma membrane-associated CK1 γ (Davidson et al., 2005; Tamai et al., 2004; Zeng et al., 2005), which then reciprocally facilitates the disassembly of the Axn complex to prevent β -catenin degradation (Cselenyi et al., 2008; Kim et al., 2013; Mi et al., 2006; Willert et al., 1999). Stabilized β -catenin translocates to the nucleus to form a transcriptional complex with TCF to direct expression of target genes (Behrens et al., 1996; Molenaar et al., 1996; van de Wetering et al., 1997). Preceding these events in signal-receiving cells, the secretion, diffusion and reception of Wnt itself is phospho-regulated in ligand-producing cells and in the extracellular environment (Buechling and Boutros, 2011).

Although several kinases and phosphatases are known to regulate Wnt signaling (Gao et al., 2014; Verheyen and Gottardi, 2010), our current knowledge of these enzymes remains incomplete. In fact, differential phospho-proteome analyses have identified both novel phospho-regulators and novel phosphorylation sites on known regulators of the pathway (Bodenmiller et al., 2007; Tang et al., 2007). It is unlikely that the reversible phosphorylation of all of these newly identified phospho-epitopes is catalyzed by the current subset of known kinases and phosphatases of Wnt signaling, thereby suggesting that there are additional unidentified phospho-regulators of the pathway.

Several large-scale *in vitro* screening analyses have been performed in various *Drosophila* and mammalian cell lines to identify regulators of the Wnt pathway. These high-throughput loss-and gain-of-function studies quantitated the response of exogenous Wnt pathway reporters under conditions of elevated signaling (Anton et al., 2011; Buechling et al., 2011; Caspi and Rosin-Arbesfeld, 2008; DasGupta et al., 2005; Firestein et al., 2008; Groenendyk and Michalak, 2011; Jacob et al., 2011; James et al., 2009; Kategaya et al., 2009; Major et al., 2008; Miller et al., 2009; Port et al., 2011; Tang et al., 2008). In this study, we have performed a comprehensive genome-wide *in vivo* RNAi screen for kinases and phosphatases in *Drosophila* to build a phospho-regulatory network of the Wnt pathway. *Drosophila* has significantly contributed to our understanding of the molecular mechanism of the Wnt pathway (Bejsovec, 2006), and has low functional redundancy but high functional conservation of the genes with humans (Fortini et al., 2000; Reiter et al., 2001). The strength of our approach is that we assayed the effects of putative phospho-regulators of the Wnt pathway on endogenous targets under physiological levels of signaling in an intact tissue. Wing discs from third instar larvae were immunostained against direct high- and low-threshold targets of the pathway, allowing us to evaluate the entire ligand-induced gradient of signaling (Barolo, 2006). As the wing disc comprises distinct

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cells that either produce or respond to the ligand, we could identify phospho-regulators at discrete positions in the pathway. Our analyses of the Wnt pathway have yielded a large subset of high-confidence kinases and phosphatases, including 22 previously unidentified regulators of signaling. Among candidate Wnt regulators, we determined which acted indirectly through the Notch pathway to modulate *wg* transcription. Furthermore, we established those shared between the Wnt and Hh pathways. Last, we validated two dual regulators of the Wnt and Hh pathways using biochemical assays to demonstrate that their roles are evolutionarily conserved from *Drosophila* to mammalian cells.

RESULTS

Design of an *in vivo* screen to identify phospho-regulators of the Wnt pathway

In the wing imaginal disc, Wingless (*Wg*) (*Drosophila* Wnt) produced from cells at the dorsal/ventral (D/V) compartment boundary induces the nested expression of pathway target genes *senseless* (*sens*) (high-threshold) and *Distal-less* (*Dll*) (low-threshold) in adjacent non-boundary cells that receive the ligand (Neumann and Cohen, 1997; Zecca et al., 1996) (Fig. 1). We compiled a list of all protein kinases and phosphatases, non-protein kinases and phosphatases, as well as factors that associate with these enzymes, such as cyclins and regulatory subunits, which we will collectively refer to as the kinase and phosphatome (supplementary material Figs S1, S2). Using transgenic RNAi libraries of UAS-driven inverted repeats (IRs) and tissue-specific Gal4 drivers, we knocked down in a spatially restricted fashion the expression of each of 385 and 205 genes present in the *Drosophila* kinase and phosphatome, respectively, to assay their effect on Wnt signaling (Fig. 1). At least two non-overlapping IRs per gene were independently tested to minimize positive and negative false discovery.

The primary screen was performed using a combination of *decapentaplegic* (*dpp*)-*Gal4* and *hedgehog* (*hh*)-*Gal4* that are expressed along the anterior/posterior compartment boundary and in the posterior compartment of the wing disc, respectively. These *Gal4* drivers were used to knock down gene function in both the ligand-producing and ligand-receiving cells (Fig. 1). A *UAS-dicer-2* transgene was used in combination with the *Gal4* drivers for all screening analyses to enhance the efficiency of RNAi-mediated gene knockdown (Dietzl et al., 2007). Wing discs from every genotype were immunostained against *Sens*, *Dll* and *Wg* to assay for pathway activity and status of the ligand. A subset of known regulators of the Wnt pathway was knocked down in a pilot screen to validate our experimental design (supplementary material Fig. S3). We consistently observed reproducible and expected defects in pathway targets (and *Wg* protein in certain cases) with known components of signaling, although some effects were subtle, yet highly penetrant and reproducible (e.g. supplementary material Fig. S3).

The wing disc is specified by Wnt and other signaling pathways (Couso et al., 1994; Rulifson et al., 1996), and disruption of this process may induce compensatory mechanisms that form a relatively normal adult wing (Herrera et al., 2013; Ryoo et al., 2004; Wells et al., 2006). Discs were therefore immunostained from every genotype irrespective of the presence of an adult wing or other phenotype. A gene that modified the levels of *Sens* and/or *Dll* when knocked down independently with at least two non-overlapping IRs using any combination of *Gal4* drivers was classified as a candidate of the Wnt pathway (examples of modifiers are shown in Fig. 2). We identified 90 candidates from the primary screen (Fig. 3). Scoring of

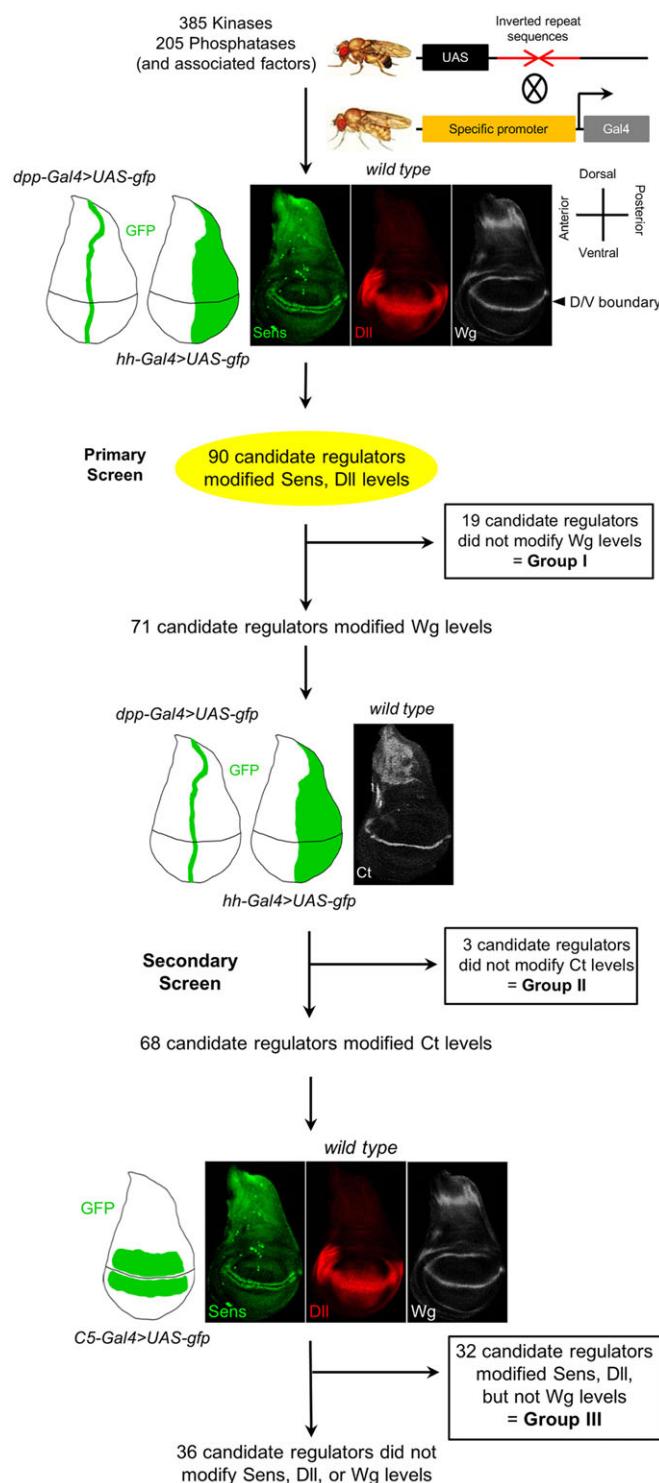


Fig. 1. Design of Wnt pathway screen in the *Drosophila* wing disc. Schematic illustration of *in vivo* Wnt pathway screen. Five-hundred and ninety kinases, phosphatases and associated factors were knocked down in a spatially restricted fashion in the wing disc through the use of *Gal4* drivers/*UAS*-IRs. Secondary screens were performed to further functionally classify the regulators, as described in the text. Ninety primary screen candidates were classified into 54 high-confidence Group I, Group II and Group III regulators based on secondary screens.

all crosses with multiple RNAi lines is provided in supplementary material Figs S4, S5. No gene, when knocked down, modified the levels of only *Sens* or *Dll*, although in some cases one pathway

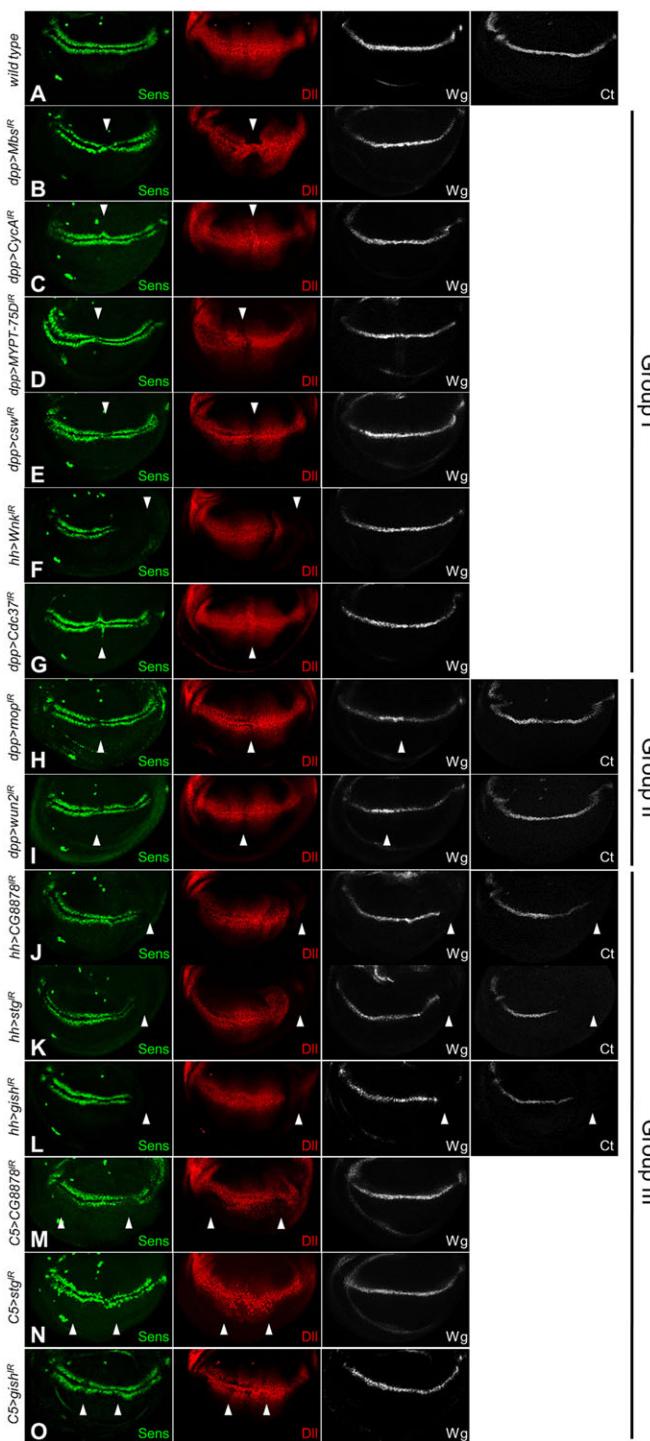


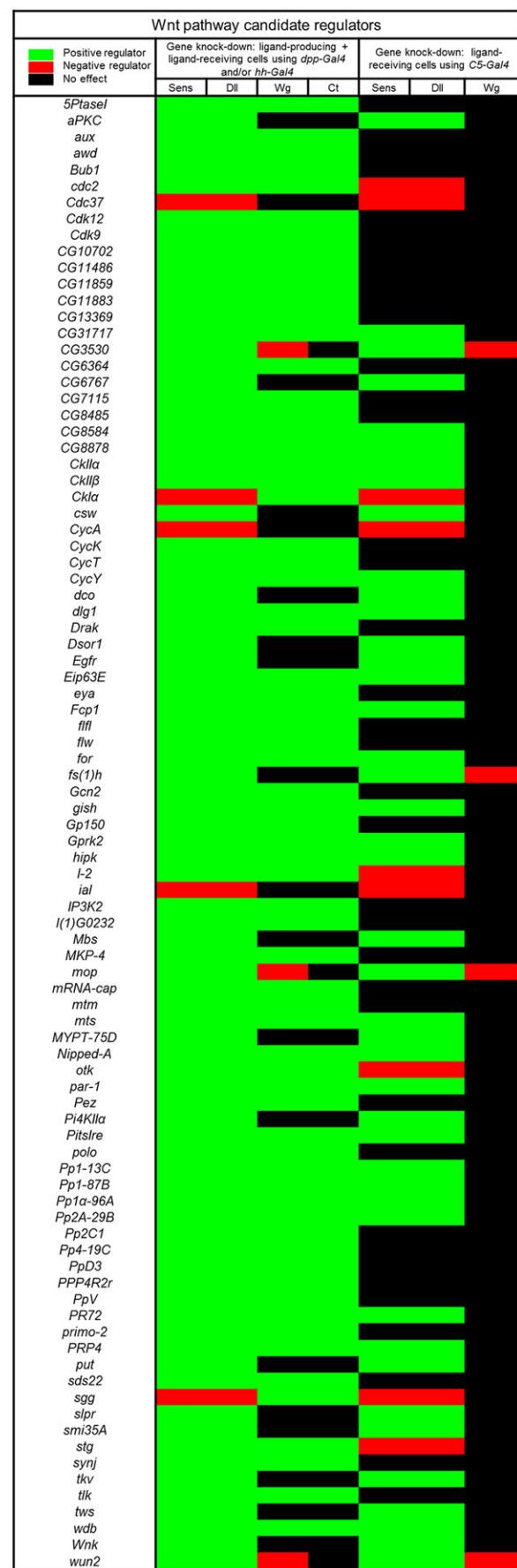
Fig. 2. High-confidence regulators of the Wnt pathway identified in the *Drosophila* wing disc. (A) A wild-type wing disc that displays the levels of Sens, Dll, Wg and Ct. (B–G) Group I regulators knocked down using *dpp-Gal4* or *hh-Gal4* modified the levels of Sens and Dll, but not Wg. Knockdown of *Mbs*, *MYPT-75D*, *csw* and *Wnk* decreased the levels of Sens and Dll (arrowheads), while knockdown of *CycA* and *Cdc37* increased the levels of Sens and Dll (arrowheads). (H,I) Group II regulators modified the levels of Sens, Dll and Wg, but not of Ct. Knockdown of *mop* and *wun2* decreased the levels of Sens and Dll (arrowheads), and increased Wg levels (arrowheads). (J–O) Group III regulators, such as *CG8878*, *stg* and *gish*, modified the levels of Sens, Dll, Wg and Ct (arrowheads). (M–O) Knockdown of these regulators in only the ligand-receiving cells with *C5-Gal4* modified the levels of Sens and Dll (arrowheads), but not Wg.

target was affected to a greater extent than the other (Figs 2, 3). Of note, most candidates had mild to moderate effects on targets, consistent with results obtained in our pilot screen with known Wnt pathway components, suggesting that under these assay conditions relatively subtle effects are valid, as confirmed by further analyses.

Nineteen of the 90 candidates had no observable effect on Wg levels or distribution, and thus do not function upstream of or at the level of the ligand-receptor interaction to affect the secretion, diffusion or reception of Wg. These candidates were classified as Group I high-confidence regulators of the Wnt pathway that function downstream of the ligand-receptor interaction in the ligand-receiving cells (Figs 2, 3). Group I includes known regulators of signaling such as *Wnk* (Serysheva et al., 2013) and novel regulators such as *Myosin binding subunit (Mbs)*, *Cyclin A (CycA)*, *MYPT-75D*, *corkscrew (csw)* and *Cdc37* (Fig. 2A–G).

Seventy-one of the 90 candidates modified the levels of Wg (Fig. 2; supplementary material Figs S4, S5). The Notch pathway signals from non-boundary cells to induce the expression of *wg* in cells at the D/V boundary of the wing disc (Diaz-Benjumea and Cohen, 1995; Neumann and Cohen, 1996; Rulifson and Blair, 1995; supplementary material Fig. S6). To test whether the candidates that affect Wg levels do so as a result of their regulation of the ligand after translation or by affecting expression of the ligand via the Notch pathway, they were re-analyzed in a secondary screen. To address transcriptional regulation by Notch signaling, we determined whether they also modified the levels of another Notch pathway target, Cut (Ct) (Micchelli et al., 1997; Fig. 1). Only three of the 71 candidates when knocked down with either *dpp-Gal4* or *hh-Gal4* had no effect on Ct levels. We inferred that these three candidates have no effect on the Notch pathway and *wg* expression, but rather modify the secretion, diffusion or reception of Wg, which was detected as a change in its levels. These candidates were classified as Group II high-confidence regulators of the Wnt pathway that function upstream of or at the level of the ligand-receptor interaction in the ligand-producing cells or extracellular environment, respectively (Figs 2, 3). Group II includes a known regulator of signaling, *myopic (mop)* (Miura et al., 2008; Pradhan-Sundd and Verheyen, 2014), and a novel regulator, *wunen2 (wun2)* (Fig. 2H–I). As further evidence of the functional role of Group II candidates, we and others have confirmed that two of the three Group II candidates (CG3530, Mop) have no effect on *wg-lacZ* but do affect the Wg protein (Silhankova et al., 2010; Pradhan-Sundd and Verheyen, 2014).

The remaining 68 of the 71 candidates modified Ct levels and were inferred to regulate *wg* expression as a result of their effect on multiple Notch targets (Fig. 3). These candidates were further tested to determine whether they also affected the Wnt pathway independently of their regulation of *wg* expression. The majority of regulators of developmental pathways function in the ligand-receiving cells and not the ligand-producing cells. Therefore, if a gene independently affects the Wnt and Notch pathways in the wing disc, it would likely do so in the ligand-receiving cells of both pathways, regulating Wnt signaling in the non-boundary cells and Notch signaling in the boundary cells. We knocked down the 68 candidates that we identified as regulators of the Notch pathway using the *C5-Gal4* driver, which is expressed in only non-boundary cells adjacent to the D/V compartment boundary (Fig. 1). By knocking down gene function in only the ligand-receiving cells of the Wnt pathway, which correspond to the ligand-producing cells of the Notch pathway (supplementary material Fig. S6), we could distinguish candidates that independently regulate both Wnt and Notch signaling from those that regulate only Notch signaling in the

**Fig. 3. Results of Wnt pathway screen in the *Drosophila* wing disc.**

Graphical summary that displays whether knockdown of candidates decreased (green), increased (red) or had no effect (black) on the levels of Sens, Dll, Wg and Ct. Summary reflects results obtained with at least two unique RNAi lines (see supplementary material Figs S4 and S5 for all data) knocked down in signal-producing and signal-receiving cells (using either *dpp-Gal4* or *hh-Gal4*) or in ligand-receiving cells only (using *C5-Gal4*) of the wing disc.

wing disc. When knocked down in the non-boundary cells, 32 of the 68 candidates modified the levels of the Wnt pathway targets Sens and Dll, but not Wg, and thus did not affect the Notch pathway in these cells to regulate *wg* expression at the D/V compartment boundary. These 32 candidates were classified as Group III high-confidence regulators of the Wnt pathway that function downstream of the ligand-receptor interaction in the signal-receiving cells and that affect the Notch pathway independently (Figs 2, 3). Group III includes known regulators of signaling, such as *string* (*stg*) (Davidson et al., 2009) and *gilgamesh* (*gish*) (Davidson et al., 2005; Zhang et al., 2006), and a novel regulator, *CG8878* (Fig. 2J-O). The remaining 36 candidates when knocked down in the non-boundary cells had no effect on the levels of Sens, Dll or Wg, and thus do not regulate the Wnt or Notch pathway in these cells (Fig. 3). Although these 36 candidates (supplementary material Fig. S7) regulate the Notch pathway and *wg* expression in the boundary cells, through our analyses we could not determine whether any of these candidates, albeit unlikely, also independently regulate Wg secretion from these cells to affect the Wnt pathway.

In summary, the 590 phospho-regulators screened in the wing disc yielded 90 candidates that were classified into 54 high-confidence regulators of the Wnt pathway and 36 candidates that regulate *wg* expression to indirectly affect the Wnt pathway. The high-confidence regulators were further classified into three phenotypic categories, referred to as Groups I, II and III, each of which comprise known and novel regulators of signaling (Fig. 4). Of the high-confidence regulators, 33 are kinases (and associated factors) and 21 are phosphatases (and associated factors). Forty-five of the 54 high-confidence regulators promote (blue in Fig. 4A), while the remaining nine inhibit (yellow in Fig. 4A), signaling.

Remarkably, 32 of the 90 candidates are known regulators of the Wnt pathway (indicated by gray shading in Fig. 4). This large subset of known regulators is reflective of the robustness and low false-negative error rate of our screen design. Moreover, based on our unbiased genetic analyses, all 32 of these known regulators cluster together in the category of high-confidence Groups I, II and III regulators. This strongly suggests that at least some of the 22 novel high-confidence regulators identified are bona fide regulators of the Wnt pathway. We classified the high-confidence regulators of the Wnt pathway according to their respective kinase and phosphatase groups. The *Drosophila* kinase comprises 13 groups (defined in supplementary material Fig. S1), of which 12 are represented by the high-confidence regulators of the Wnt pathway (Fig. 4B; supplementary material Fig. S1). The *Drosophila* phosphatome comprises nine groups (defined in supplementary material Fig. S2), of which six are represented by the high-confidence phosphatases (and associated factors) of the Wnt pathway (Fig. 4C; supplementary material Fig. S2).

Hh pathway counterscreen identifies Cdc37 and Gish as dual regulators of signaling

The Hh pathway has diverse functions during metazoan development, such as the regulation of organogenesis and stem

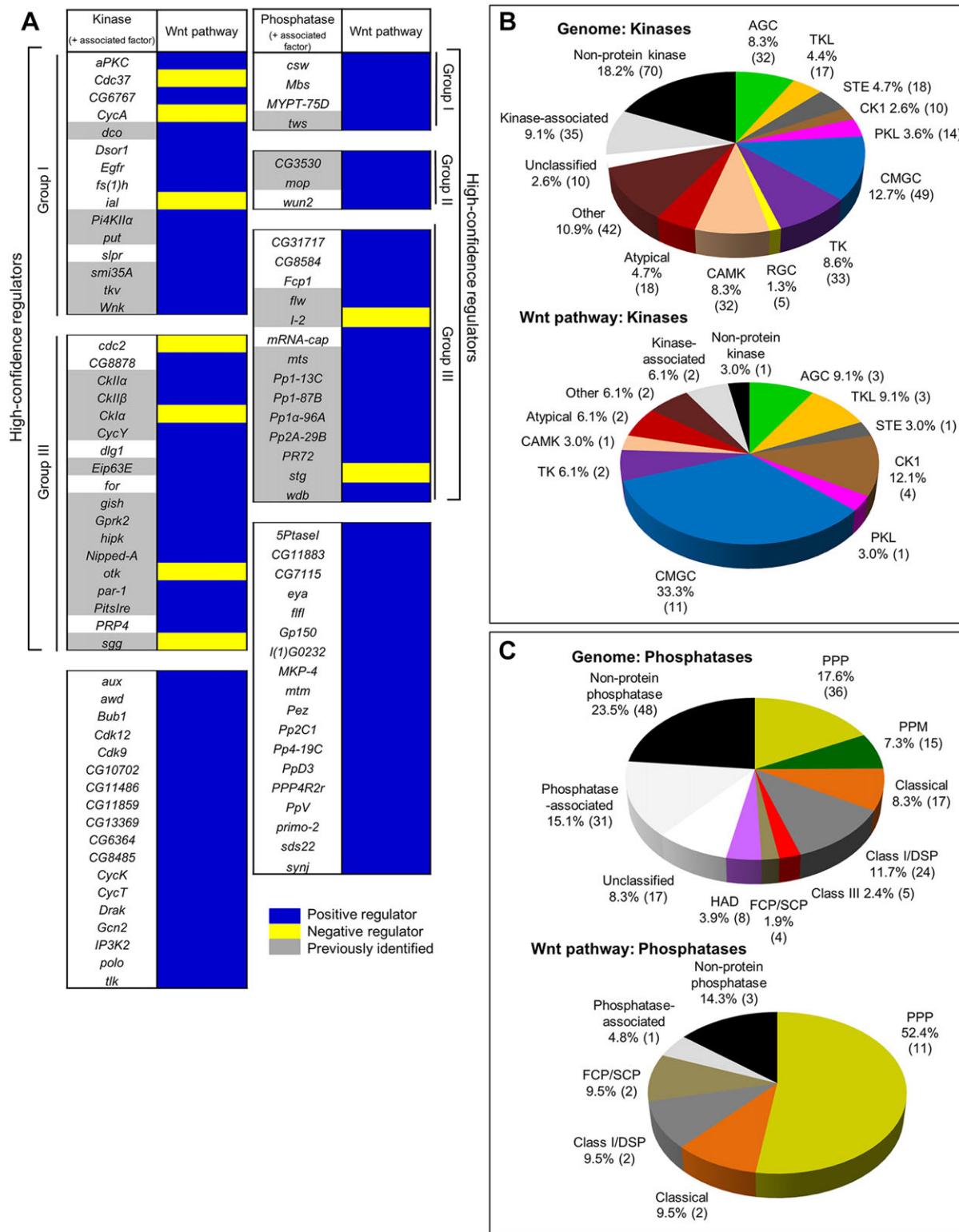


Fig. 4. Summary of phospho-regulators of the Wnt pathway identified in the *Drosophila* wing disc. (A) Ninety candidates of the Wnt pathway were refined into 54 high-confidence regulators (Group I, Group II and Group III). Forty-five high-confidence regulators promote (blue) and nine inhibit (yellow) the Wnt pathway. Three high-confidence regulators function upstream or at the level of the ligand-receptors interaction (Group II), whereas the remaining 51 function downstream of the ligand-receptor interaction (Group I, Group III). Thirty-two of the 90 candidates identified are previously validated regulators of the Wnt pathway (gray). (B,C) Graphical summary of the kinase (B) and phosphatase (C) groups in the *Drosophila* genome, and the subset of these that regulate the Wnt pathway.

cell homeostasis (Varjosalo and Taipale, 2008). Similar to the Wnt pathway, the Hh pathway is also subject to reversible phosphorylation in its silent and active states. In the absence of

signaling, the transcriptional effector GLI is phosphorylated by Protein Kinase A (PKA), GSK3 β and CK1 α within a cytosolic Kif7 complex (Chen et al., 1998; Jia et al., 2002, 2005; Price and

Kalderon, 1999, 2002; Zhang et al., 2005). The phosphorylation of GLI triggers its poly-ubiquitylation and partial degradation to yield a truncated form of the protein that represses target gene expression (Aza-Blanc et al., 1997; Jia et al., 2005; Jiang and Struhl, 1998; Méthot and Basler, 1999; Smelkinson and Kalderon, 2006; Smelkinson et al., 2007; Tempé et al., 2006). Binding of the Hh ligand to its receptor Patched (Ptc) recruits the Kif7 complex to the transmembrane signal transducer Smoothened (Smo) (Lum et al., 2003; Ogden et al., 2003; Ruel et al., 2003; Zheng et al., 2010). This induces Smo phosphorylation by PKA and CK1α, which promotes its accumulation at the cell surface (Apionishev et al., 2005; Denef et al., 2000; Jia et al., 2004; Zhang et al., 2004; Zhu et al., 2003). The interaction between Kif7 and phospho-Smo disassembles the complex to stabilize full-length GLI that directs target gene expression (Aza-Blanc et al., 1997; Jia et al., 2003; Liu et al., 2007; Ohlmeyer and Kalderon, 1998). Although the evolutionary relationship between Wnt and Hh signaling remains unclear, these pathways have a similar phospho-regulatory mechanism of signal transduction and comprise similar or identical regulators that exert the same effect on signaling by functioning at analogous levels of the relays (Kalderon, 2002).

We evaluated the ability of all 90 candidates of the Wnt pathway to regulate the Hh pathway *in vivo*. We performed this counter-screen to identify shared phospho-regulators that exert the same effect to either promote or inhibit signaling, and thereby potentially function at analogous levels of these pathways. Hh signaling stabilizes full-length Cubitus interruptus (Ci) (*Drosophila* GLI) to regulate expression of the target gene *ptc* along the anterior/posterior (A/P) boundary in the anterior compartment of the wing disc (Strigini and Cohen, 1997) (Fig. 5A). Each candidate was knocked down using *MS1096-Gal4* (with *UAS-dicer-2*) and wing discs were immunostained to detect Ci and Ptc. The *MS1096-Gal4* domain is in the center of the wing disc with stronger expression in the dorsal half relative to the ventral half (supplementary material Fig. S8A). The enhancer of *ptc*, unlike that of other Hh pathway targets, responds in a cooperative manner to both the levels and active state of Ci; thus, a change in the levels of Ci that modulates the amplitude of Hh signaling might not necessarily result in an effect on the expression of *ptc* (Parker et al., 2011). Any gene that modified the levels of Ci, but not necessarily Ptc, when independently knocked down with at least two non-overlapping IRs was classified as a regulator of the Hh pathway (Fig. 5B; supplementary material Fig. S8B). A limitation of this approach is that the counter-screen cannot identify regulators of the Hh pathway that do not affect the levels of Ci, but function downstream of its stabilization to modulate the expression of targets other than *ptc*. Sixty-six of the 90 candidates of the Wnt pathway modified the levels of Ci (and Ptc in most cases) to regulate the Hh pathway (supplementary material Figs S4, S5). The remaining 24 candidates of the Wnt pathway had no effect on the levels of Ci or Ptc (supplementary material Figs S4, S5). While the majority of Wnt regulators promoted signaling, 45 of 66 regulators of the Hh pathway inhibited signaling (supplementary material Fig. S8B). Nevertheless, 29 Wnt candidates exerted the same effect on the Hh pathway to either promote or inhibit signaling in the wing disc (Fig. 5B). Of these 29 dual regulators that we propose function at analogous levels of these pathways, 25 are high-confidence regulators and eight are novel regulators of the Wnt pathway. As proof of principle, we recovered *hipk* in our screen, which we have previously shown to be a dual regulator of Wnt and Hh acting on the E3 ubiquitin ligase Supernumerary limbs (Slimb) at analogous levels of the pathways (Swarup and Verheyen, 2011).

We identified *Cdc37* and *gish* as novel negative and positive regulators, respectively, of the Hh pathway. Knockdown of a negative control, *lacZ* (supplementary material Fig. S8A), displayed normal levels of Ci and Ptc, as seen in wild-type tissue (Fig. 5C). Knockdown of *Cdc37* resulted in the robust enhancement of Ci levels, both within and away from the signaling domain in the anterior compartment of the wing disc (Fig. 5D). Knockdown of *Cdc37* consistently led to the distortion of the morphology of the disc, which precluded an accurate evaluation of Ptc levels. Based on our data, *Cdc37* knockdown does not seem to appreciably alter Ptc levels, but we cannot definitively rule out this possibility. *Cdc37* knockdown phenocopies loss-of-function mutants of other negative regulators of the Hh pathway that strongly enhance the levels of Ci but have minimal or no effect on Ptc levels, such as *sgg* (*Drosophila* GSK3β) (Jia et al., 2002; Price and Kalderon, 2002) and *slimb* (Wang et al., 1999). Knockdown of *gish* resulted in the reduction of Ci and a reduction in the levels of Ptc, as indicated by the width of its expression domain in the central region of the wing disc (Fig. 5E).

In our screening analyses, all 90 candidates of the Wnt pathway were evaluated against Notch (as part of the secondary screen) and Hh (as part of the counter-screen) pathways in the wing disc (Fig. 6A). This allowed us to distinguish between candidates that are specific to the Wnt pathway from those that are shared between the Wnt and other signaling pathways (Fig. 6B; Tabata and Takei, 2004). These analyses also allowed us to evaluate whether a candidate of the Wnt pathway does so only indirectly as a result of its effect on cell death, cell proliferation or non-specific gene transcription. For example, if knockdown of a candidate decreases Wnt signaling but does not affect or increases Notch and/or Hh signaling, we inferred that this candidate does not affect cell death or non-specific gene transcription. However, if knockdown of a candidate decreases Wnt, Notch and Hh signaling, it is possible that it indirectly regulates the Wnt pathway due to an effect on cell death or non-specific gene transcription. Twelve of the 90 candidates of the Wnt pathway when knocked down had the same effect (i.e. positive or negative regulator of both pathways) on both the Notch and Hh pathways (Fig. 6C). This does not necessarily imply that these 12 candidates indirectly regulate the Wnt pathway, but that further analysis of these candidates is required. Coincidentally, 11 of these 12 candidates have been previously described to regulate the Wnt pathway (Fig. 6C).

Cdc37 inhibits the Wnt and Hh pathways by destabilizing their transcriptional effectors

As proof of concept for dual regulators that operate at analogous levels of the Wnt and Hh pathways, we further characterized the functions of *cdc37* and *gish*. *Cdc37* is a chaperone that functions to regulate the folding and biogenesis of diverse kinases (Caplan et al., 2007). We identified *cdc37* as a novel negative regulator whose knockdown resulted in increased pathway activity within the signaling domain and ectopic pathway activity outside the signaling domain (Fig. 2G). The *Drosophila* eye has frequently been used to screen for regulators of the Wnt pathway using a *sevenless* (*sev*)>*wg* (or equivalent) gain-of-function genetic background in which eye development is impaired (Fig. 7A; Greaves et al., 1999; Port et al., 2011). Heterozygosity for a loss-of-function allele of *cdc37* caused an enhancement of the *sev*>*wg* phenotype (Fig. 7B), suggesting that reduction of *cdc37* enhances the aberrant eye phenotype caused by *Wg* signaling. We examined embryos homozygous for the *cdc37^{ED4}* allele. *Wg* signaling is required for specification of regions of naked cuticle

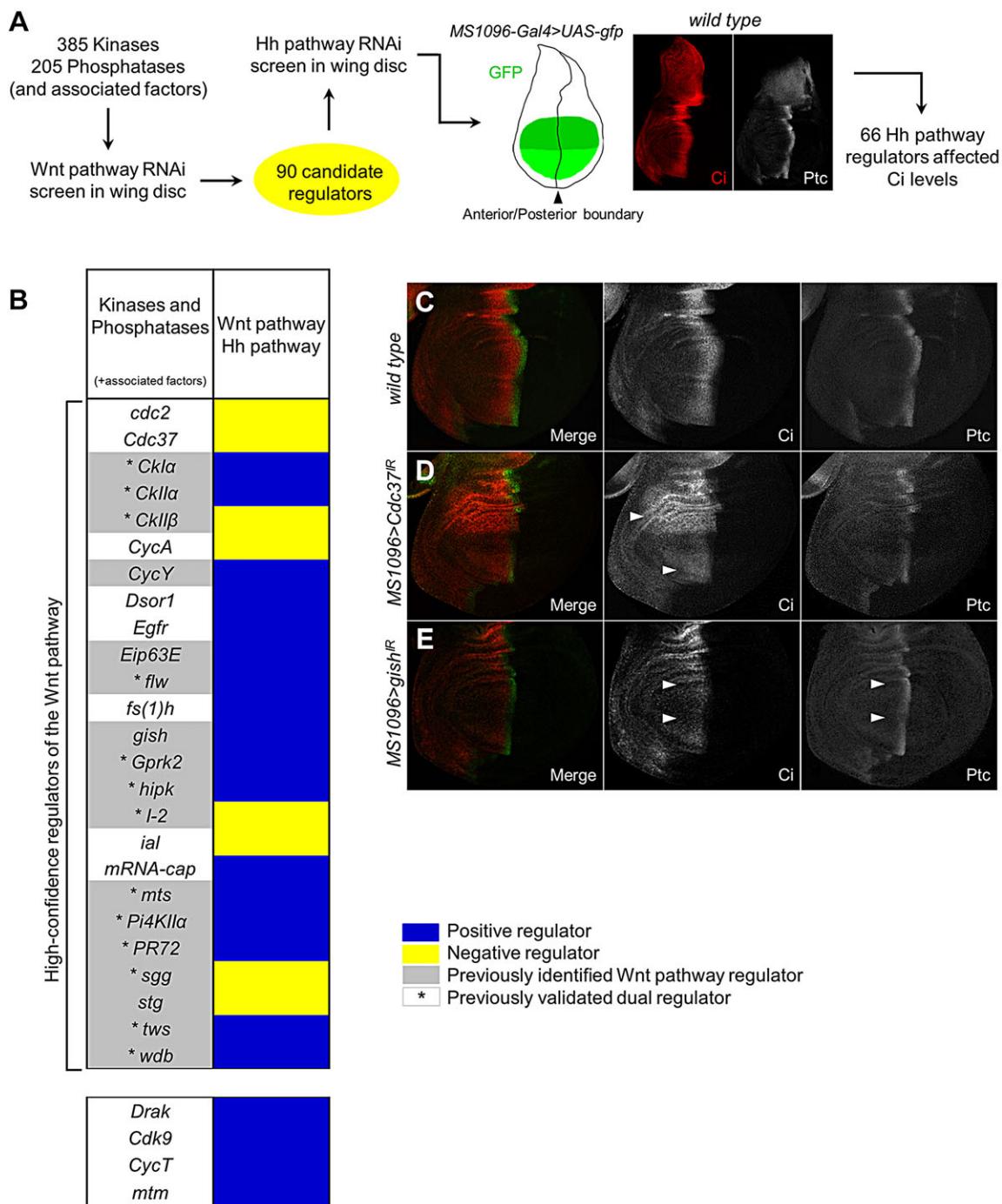


Fig. 5. Hh pathway counter-screen in the *Drosophila* wing disc. (A) Ninety candidates of the Wnt pathway were knocked down throughout the wing pouch using MS1096-Gal4 and wing discs were immunostained against Ci and Ptc. Sixty-six candidates modified Ci levels to regulate the Hh pathway. (B) Twenty-nine candidates of the Wnt pathway were identified to have the same effect on the Hh pathway to either promote (blue) or inhibit (yellow) signaling. All but four of these 29 candidates are high-confidence regulators and 17 are known regulators (gray) of the Wnt pathway. Thirteen of the 29 candidates have been previously validated as dual regulators (asterisks) of the Wnt and Hh pathways. (C) The levels of Ci and Ptc along the anterior/posterior compartment boundary of a wild-type wing disc. (D) Knockdown of *cdc37* increased the levels of Ci (arrowheads), but did not affect Ptc. (E) Knockdown of *gish* decreased the levels of Ci and Ptc (arrowheads).

between denticle belts in the cuticle (Fig. 7C). Enhanced Wg signaling caused loss of denticle belts, such as was seen in *cdc37* mutant embryos, suggesting loss of *cdc37* promotes Wg signaling (Fig. 7D,E). Somatic loss-of-function clones of *cdc37* do not survive in the wing disc due to its requirement for cell viability (Lange et al., 2002). We therefore generated MARCM loss-of-function clones of *cdc37* (positively marked with nuclear GFP) that overexpress the

apoptosis inhibitor *p35*. These MARCM clones, although small in size, displayed a cell-autonomous upregulation of Wnt and Hh signaling, as indicated by an increase in Dll and Ci levels, respectively. Similar to the effect seen with RNAi (Figs 2G, 5D), ectopic Dll (Fig. 7F) and Ci (Fig. 7G) were observed in MARCM clones away from the signaling domains of these pathways. This result suggests that Cdc37 may normally destabilize the effectors to function

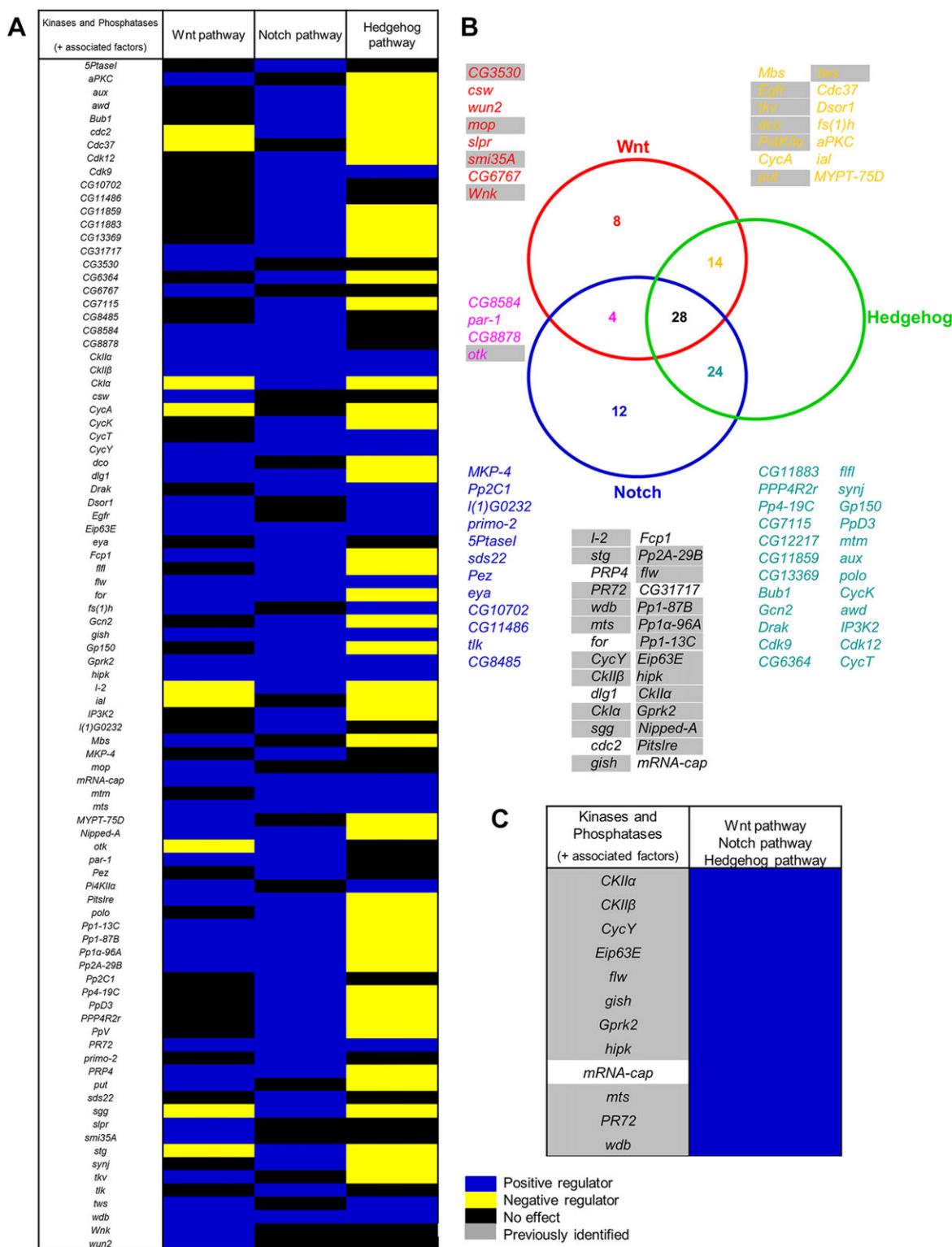


Fig. 6. Specific and shared phospho-regulators of the Wnt, Notch and Hh pathways in *Drosophila*. (A) Graphical summary that displays whether a regulator promotes (blue), inhibits (yellow) or has no effect (black) on the Wnt, Notch and Hh pathways in the wing disc. (B) Venn diagram showing specific or shared regulators of the Wnt, Notch and Hh pathways. Previously known regulators of the Wnt pathway are indicated with gray shading. (C) Twelve of the 90 candidates of the Wnt pathway identified also had the same effect on the Notch and Hh pathways in the wing disc.

as a negative regulator of signaling in the silent and active states of the Wnt and Hh pathways. Indeed, when we assayed for levels of stabilized Armadillo (Arm) (*Drosophila* β -catenin), we found more in wing discs reduced for *cdc37* function compared with wild type

(Fig. 7I). Although *cdc37* RNAi did not significantly affect *ptc* expression, it did result in increased expression of another target gene of the Hh pathway, *dpp-lacZ* (supplementary material Fig. S9A). Although loss of *cdc37* clearly stabilizes Ci, studies suggest that there

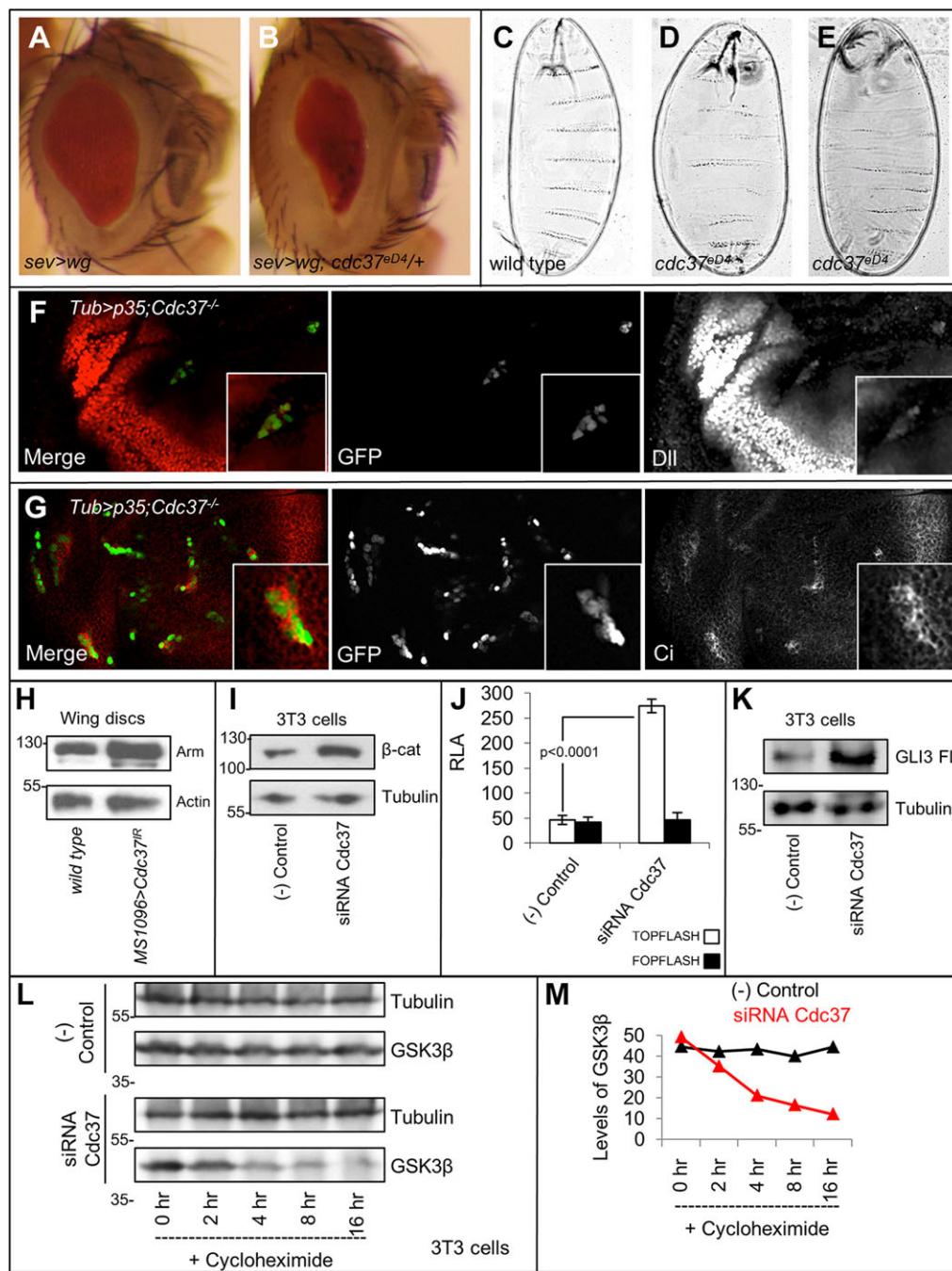


Fig. 7. *Cdc37* destabilizes the effectors of the Wnt and Hh pathways to inhibit signaling. (A,B) Expression of *wg* from the *sev* promoter (*sev>wg*) results in a small rough eye, which is enhanced upon heterozygosity for the *cdc37^{eD4}* allele. (C-E) Homozygous mutant *cdc37^{eD4}* embryos display cuticle defects reminiscent of *wg* gain-of-function defects, including loss of denticle belts. (F,G) MARCM clones (with *p35* transgene) of *cdc37^{eD4}* (marked by nuclear GFP) in the wing disc resulted in an increase in DII (F) and Ci (G) levels. (H) Protein lysate from wing discs reduced for *Cdc37* function had elevated levels of stabilized Arm compared with wild-type discs. (I) Compared with the mock-transfected control, siRNA-mediated knockdown of *Cdc37* in 3T3 cells enhanced the levels of stabilized β -catenin. (J) Increased Wnt pathway activity is observed following siRNA-mediated knockdown of *Cdc37*, as measured from the TOPFLASH reporter but not FOPFLASH (negative control) (RLA=Relative Luciferase Activity). Student's *t*-test was performed and s.d. was calculated. (K) siRNA-mediated knockdown of *Cdc37* in 3T3 cells enhanced the levels of full-length GLI3 compared with the mock-transfected control. (L,M) siRNA-mediated knockdown of *Cdc37* in cycloheximide-treated 3T3 cells displayed progressively lower levels of GSK3 β over the course of 16 h, compared with the mock-transfected control, as seen in western blot (L) and following densitometry (M). The levels of GSK3 β were normalized to the loading control. Molecular weights are indicated in kDa next to each blot.

is an additional step that regulates Ci import into nucleus. We propose that only a small amount of Ci enters the nucleus in *cdc37* clones, which may explain why there are no apparent changes in *ptc* (which requires high-level Hh for expression), although we do observe a change in *dpp*, which is a low level target. Knockdown of *cdc37* caused increased cell proliferation in discs, as indicated by increased levels of phospho-Histone H3 (PH3; supplementary material Fig. S9B), but this proliferation cannot solely explain the expression of targets in regions outside the normal domain of signaling. Thus, these results indicate that loss of *cdc37* enhances both Wg and Hh signaling outputs.

To examine whether the function of *Cdc37* is evolutionarily conserved, we carried out biochemical studies in mammalian cells. Knockdown of *Cdc37* with siRNA in 3T3 cells strongly enhanced

levels of stabilized β -catenin (Fig. 7J). Furthermore, knockdown of *Cdc37* also induced signaling in unstimulated 3T3 cells, as measured using the Wnt pathway-specific TOPFLASH reporter, compared with mock-transfected 3T3 cells (Fig. 7K). Knockdown of *Cdc37* also enhanced the levels of full-length GLI3 (Fig. 7L). We were unable to detect the truncated form of GLI3 in our assay. These effects of *Cdc37* in mammalian cells mimic those of negative regulators of the Wnt and Hh pathways, such as GSK3 β and CK1 α . As *Cdc37* is a kinase-associated chaperone, we propose that it functions to promote the stability of GSK3 β and/or CK1 α , which both constitutively destabilize the effectors of the Wnt and Hh pathways. Accordingly, cycloheximide-treated 3T3 cells reduced for *Cdc37* displayed progressively lower levels of GSK3 β due to its shorter half-life, compared with mock-transfected cells (Fig. 7M,N). We did not

evaluate whether Cdc37 regulates the stability of CK1 α in our assay. Thus, our analyses suggest that Cdc37 has a novel evolutionarily conserved function from *Drosophila* to mammalian cells to promote the stability of GSK3 β and inhibit both the Wnt and Hh pathways.

Gish/CK1 γ promotes the Hh pathway by phosphorylating Smo

Gish (*Drosophila* CK1 γ) is a plasma membrane-associated kinase that has been described to promote the Wnt pathway by phosphorylating the co-receptor LRP (Davidson et al., 2005; Zhang et al., 2006). Consistently, it was recovered as a high-confidence Wnt regulator (Fig. 2L,O). Thus far, no role has been ascribed to this kinase in the regulation of the Hh pathway. Although knockdown of *gish* decreased the expression of targets of the Wnt, Notch and Hh pathways in the wing disc (Fig. 6A), the levels of Delta (DI) and cleaved-Caspase 3 (Casp 3) were unaffected (supplementary material Fig. S9C). Thus, *gish* does not have a non-specific effect on gene transcription or cell death. Compared with wild type, a *gish* mutant wing disc displayed lower levels of Ci (Fig. 8A). Hh signaling stabilizes Ci to regulate expression of *dpp-lacZ* within the morphogenetic furrow (MF) of the eye disc (Fig. 8B). A somatic clone of *gish* in the eye disc had decreased levels of Ci and correspondingly *dpp-lacZ* expression (Fig. 8C), thereby confirming its role as a positive regulator of Hh signaling across multiple tissues.

We propose that Gish/CK1 γ regulates the phosphorylation of the transmembrane protein Smo to promote the Hh pathway, analogous to its role in regulating the phosphorylation of LRP in the Wnt pathway. We tested this hypothesis using protein lysates from wing discs in a gel

mobility shift assay (Fig. 8D). Knockdown of *gish* throughout the wing blade using 71B-Gal4 reduced the phosphorylation of Smo in disc protein lysates (as detected by a faster migrating band). As Smo phosphorylation is a prerequisite to its accumulation, knockdown of *gish* also decreased Smo levels. Conversely, overexpression of the Hh ligand, *hh-N*, expectedly enhanced the phosphorylation (as detected by a slower migrating band in lane 3) and levels of Smo (Fig. 8D). The simultaneous knockdown of *gish* in the presence of ectopic *hh-N* reduced the phosphorylation (migration) and levels of Smo, compared with ectopic *hh-N* alone (determined by densitometry), thus indicating that Gish acts downstream of the Hh ligand.

To determine whether the effect of Gish on the Hh pathway is evolutionarily conserved, the effect of CK1 γ on signaling was assessed in mammalian cells. 3T3 cells transfected with CK1 γ displayed increased levels of full-length GLI3, compared with mock-transfected cells (Fig. 8E). When exogenous CK1 γ and Smo were co-transfected into 3T3 cells, they were detected in a complex in a co-immunoprecipitation assay (Fig. 8F). Furthermore, CK1 γ robustly phosphorylated Smo at one or more serine and/or threonine residues (as detected by phospho-specific antibodies) in the presence of ATP in an *in vitro* kinase assay (Fig. 8G). Thus, Gish/CK1 γ regulates the phosphorylation of Smo to promote the Hh pathway in *Drosophila* and mammalian cells.

DISCUSSION

Divergent disease states have been attributed to be a cause or consequence of aberrant protein phosphorylation (Reiter et al.,

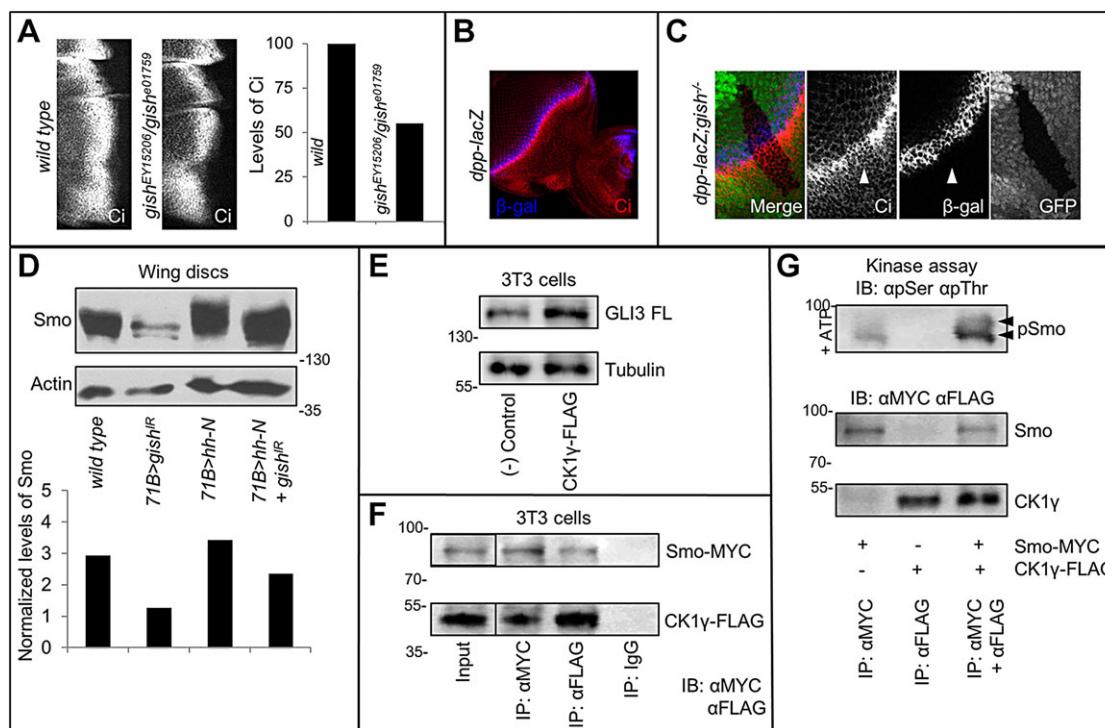


Fig. 8. Gish/CK1 γ promotes the Hh pathway by phosphorylating Smo. (A) Ci protein levels are reduced in a *gish* mutant wing disc compared with wild type. (B) The Hh pathway stabilizes Ci to promote *dpp-lacZ* expression within the MF of the eye disc. (C) A *gish* somatic clone (GFP negative) in the eye disc had decreased levels of Ci and *dpp-lacZ* (detected by anti- β -gal; arrowheads). (D) Knockdown of *gish* using 71B-Gal4 in wing discs reduced the phosphorylation (detected by mobility shift) and levels of Smo, compared with wild type. Expression of *hh-N* with 71B-Gal4 enhanced the phosphorylation (as detected by mobility shift) and levels of Smo. The simultaneous knockdown of *gish* in the presence of ectopic *hh-N* reduced the phosphorylation (as detected by mobility shift) and levels of Smo. (E) Overexpression of CK1 γ in 3T3 cells resulted in increased levels of full-length GLI3 compared with the mock-transfected control. (F) CK1 γ co-precipitated with Smo when transfected into 3T3 cells. Epitope-tagged exogenous proteins were precipitated with corresponding antibodies or with IgG (negative control). (G) Smo was phosphorylated by CK1 γ in an *in vitro* kinase assay, as detected by antibodies against phosphorylated serine and threonine residues. The loading control had equal amounts of immunoprecipitated proteins. Molecular weights are indicated in kDa next to each blot.

2001). Wnt signaling is phosphor-regulated both in its silent and active states, but thus far our understanding of kinases, phosphatases and associated factors of the pathway has been limited. In this study, we performed the first genome-wide *in vivo* screen under physiological conditions in the *Drosophila* wing disc for phospho-regulators of the Wnt pathway. We identified 54 high-confidence regulators, 22 of which are novel. The results of our analyses do not indicate whether a high-confidence regulator has a direct or indirect effect on signaling. However, as ~60% of the high-confidence regulators identified have been previously validated to have a direct effect on Wnt signaling, we predict that at least some of the novel high-confidence regulators identified would also have a direct effect on the pathway. Indeed, subsequent analyses of Myopic revealed a novel role in regulating Wg secretion (Pradhan-Sundd and Verheyen, 2014). Although the mechanism and components of the Wnt pathway are for the most part conserved between *Drosophila* and humans, there are possibly vertebrate-specific phospho-regulators of signaling that would not have been identified in our analyses. Our dataset represents the largest list of putative phospho-regulators of the Wnt pathway identified to date, almost all of which have identified human orthologs (supplementary material Figs S1, S2) and are therefore likely to be functionally conserved.

As part of this study, we also established previously unknown relationships between the Wnt and Hh pathways *in vivo* by identifying 12 novel dual regulators that we propose function at analogous levels of signaling (Fig. 5B). As proof of concept, we biochemically characterized the roles of Cdc37 and Gish/CK1 γ to demonstrate that their functions are conserved from *Drosophila* to mammalian cells. We also describe an initial analysis of candidate regulators of Notch signaling during wing disc development. Although these findings are preliminary, they highlight an emerging theme of phospho-regulation of Notch that likely hold parallels in vertebrate biology. The comparison of signaling pathways *in vivo* and the identification of specific versus shared phospho-regulators facilitate our understanding of human development and disease states.

MATERIALS AND METHODS

Drosophila genetics

The following *Drosophila* strains were used: *w¹¹¹⁸* (wild type), *dpp-Gal4/TM6B, C5-Gal4, MS1096-Gal4, omb-Gal4, 71B-Gal4, UAS-flp, UAS-dicer-2, UAS-p35, dpp-lacZ/CyO, Cdc37^{eD4} FRT79/TM6B, FRT82, GFP/TM6B, MARCM79, hs-flp;FRT82, GFP/TM6B* and *gish^{EY15206}* (Bloomington *Drosophila* Stock Center); *hh-Gal4/TM6B* and *eyFlp;ey-Gal4, GMR-Gal4; sev>y+>wg* (Port et al., 2011); *UAS-hhN* (Su et al., 2011); and *FRT82 gish^{e01759}/TM6B* (Gault et al., 2012). The transgenic RNAi strains used for the screens were obtained from the Vienna *Drosophila* RNAi Center (Dietzl et al., 2007), National Institute of Genetics and Harvard Transgenic RNAi Project (supplementary material Fig. S4, Fig. 5). The percentage of inverted repeats with predicted sequence-dependent off-target effects are as follows: 65% (0), 27% (1-2) and 8% (>2). All genes (at least two independent RNAi lines per gene) were first tested with the *dpp-Gal4* driver. If a phenotype was only observed with one RNAi line for *dpp-Gal4* or with two lines targeting the same region of the mRNA, the gene was re-tested with *hh-Gal4*. The one or more RNAi lines that were re-tested with *hh-Gal4* were either the same lines that were used with *dpp-Gal4* or additional lines (if new ones were available). If a RNAi line displayed a phenotype with both the *dpp-Gal4* and *hh-Gal4* drivers in the primary screen, the *hh-Gal4* driver was used in the secondary screen. Wing discs from 20 larvae of each genotype were immunostained (30 larvae for genotypes that were non-homozygous with a balancer chromosome). Knockdown of no gene precluded analysis due to lethality before the third instar stage. Twenty flies of each genotype were scored for adult phenotypes. The penetrance of all phenotypes indicated is

between 80 and 100%. False-positive results due to cell death, cell proliferation or non-specific gene transcription were evaluated through testing the ability of candidate regulators to affect different signaling pathways in the wing disc. *cde37* MARCM clones were generated by crossing the *UAS-p35; Cdc37^{eD4} FRT79/TM6B* and *MARCM79* strains, and progeny were heat-shocked 48 h after egg laying (AEL) for 2 h at 38°C. *gish* somatic clones in the wing disc were generated by crossing the *UAS-flp; FRT82 gish^{e01759}/TM6B* and *omb-Gal4; FRT82, GFP/TM6B* strains. *gish* somatic clones in the eye disc were generated by crossing the *hs-flp; FRT82, GFP/TM6B* and *FRT82 gish^{e01759}/TM6B* strains, and progeny were heat-shocked 48 h AEL for 2 h at 38°C.

Immunostaining of wing discs

Drosophila wing and eye discs from third instar larvae were dissected, immunostained and mounted according to standard procedures (Swarup and Verheyen, 2011). The following primary antibodies were used: anti-Ci 2A1 (1:50), anti-Ptc (1:50), anti-Wg 4D4 (1:100), anti-Ct 2B10 (1:75), anti-Delta C594.9B (1:50) (Developmental Studies Hybridoma Bank), anti-Phospho Histone H3 (1:100), anti- β -galactosidase (1:1500), anti-Cleaved Caspase 3 (1:100) (Cell Signaling Technology), anti-Sens (1:1000) (Nolo et al., 2000) and anti-Dll (1:400) (Duncan et al., 1998; Panganiban et al., 1995). Fluorescent secondary antibodies (1:400) were from Jackson Immunolabs. Wing disc images were obtained on a Nikon A1R laser scanning confocal microscope and all images are derived from stacked z-series. A fixed-size box (width=width of stabilized Ci domain in wild type, height=bottom of disc to top of wing pouch) was used to compare the integrated density of wild type and *gish* mutant. For quantification of immunostained discs, integrated density (pixel area multiplied by baseline subtracted intensity) was calculated.

Cell culture

NIH-3T3 cells (American Type Culture Collection) were cultured at 37°C in DMEM supplemented with 10% FBS (Invitrogen). 3T3 cells were transfected with CK1 γ -FLAG (Davidson et al., 2005), Smo-MYC (Chen et al., 2011), TOPFLASH, FOPFLASH, or Renilla luciferase (Ishitani et al., 2003) using Polyfect Reagent (Qiagen) and harvested 36–48 h post-transfection. Knockdown of *Cdc37* was performed with siRNA and Lipofectamine RNAiMAX Reagent (Invitrogen). As a negative control, cells were mock-transfected with empty vector. Cells were treated with cycloheximide (25 μ g/ml) (Sigma) 24 h after transfection with *Cdc37* siRNA [a mixture of two distinct siRNAs targeted to different parts of the genes from Ambion (136227 and 136228)] and harvested at the indicated time points.

Biochemical assays

Protein lysates were prepared from 3T3 cells and wing discs. Proteins of interest were immunoprecipitated from lysates using antibodies and Protein-G Sepharose beads (Sigma). The kinase assay with non-radioactive ATP/kinase assay buffer (Cell Signaling Technology) was performed with immunoprecipitated proteins at 30°C for 30 min. The de-phosphorylation assay was performed on protein lysate using lambda protein phosphatase at 30°C for 1 h (New England Biolabs).

Western blot analyses

Western blot analyses were performed using the following primary antibodies: anti-Tubulin (1:1000), anti-Myc (1:1000), anti-FLAG (1:1000) (Sigma), anti-GSK3 β (1:500) (Abcam), anti-GLI3 N19 (1:200) (Santa Cruz Biotechnology), anti-Phosphoserine (1:500), anti-Phosphothreonine (1:500) (Sigma), anti-Smo 20C6 (1:10) and anti-Armadillo N27A1 (1:200) (Developmental Studies Hybridoma Bank). For quantification of western blot bands, integrated density (pixel area multiplied by baseline subtracted intensity) of each band was calculated within an appropriate exposure range and then normalized to the loading control.

Transcriptional assays

Transcriptional assays were performed using TOPFLASH, FOPFLASH and Renilla (control) luciferase reporter plasmids. The experiment was performed using the Dual Luciferase Reporter Assay System (Promega).

The value for each data point is the average of three individual experiments. The s.d. was calculated and Student's *t*-test was performed for statistical significance.

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Competing interests

The authors declare no competing or financial interests.

Author contributions

S.S., E.M.V. designed the experiments. S.S., T.P.-S. performed the experiments. S.S., T.P.-S., E.M.V. analyzed the data. S.S., E.M.V. wrote the manuscript.

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Supplementary material

Supplementary material available online at
<http://dev.biologists.org/lookup/suppl/doi:10.1242/dev.116715/-DC1>

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FIGURE LEGENDS

Fig. S1 Kinases and kinase-associated factors assayed in screen

All genes (sorted by CG number) encoding either kinases or associated factors for which UAS-controlled inverted repeat strains were tested. Kinase group and family is indicated, as well as human orthologs.

Fig. S2 Phosphatases and Phosphatase-associated factors assayed in screen

All genes (sorted by CG number) encoding either phosphatase or associated factors for which UAS-controlled inverted repeat strains were tested. Phosphatase group and family is indicated, as well as human orthologs.

Fig. S3 Wnt pathway pilot screen in the Drosophila wing disc.

Graphical summary of the Wnt pathway pilot screen in the wing disc that displays whether knock-down of pathway components decreased (green), increased (red), or had no effect (black) on the levels of the ligand and downstream targets. A subset of known components of the Wnt pathway were knocked-down using *dpp-Gal4* and *hh-Gal4* in the wing disc to evaluate their effects on the levels of Sens, Dll, and Wg. Knock-down of *lacZ* (negative control) did not affect the levels of Sens, Dll, or Wg. Knock-down of the positive regulator *porcupine (por)* resulted in a decrease of Sens and Dll levels (arrowheads) in the ligand-receiving cells due to the impaired secretion of Wg (arrow) (detected as an increase in levels) from the ligand-producing cells. Knock-down of the negative regulator *Axn* resulted in increased levels of Sens and Dll (arrowheads) in the ligand-receiving cells, but had no effect on Wg levels. Knock-down of the ligand *wg* resulted in the loss of Wg (arrow) in the ligand-producing cells and a corresponding loss of Sens and Dll (arrowheads) in the ligand-receiving cells.

Fig. S4 Results of primary and secondary screening of kinome

Results obtained with indicated RNAi lines in crosses indicated in top line of table, under primary screen and secondary screen headings. Blank infill indicates that there was no change in targets in primary screen. Black infill means there was no change observed in in that particular assay although there was an effect either with another readout or another RNAi line corresponding to the same gene. Coloured infill represents changes to readouts as indicated in legend. Genes that were tested in secondary screen are highlighted by lines above and below the gene. Adult phenotypes (if observed) are indicated. Any indicated lethality occurred after third instar, thus allowing larval discs to be examined. Transgenic RNAi Project strains are in blue font, while National Institute of Genetics (NIG) strains have the letter R in the stock number. All other strains are from Vienna Drosophila RNAi Center (VDRC).

Fig. S5 Results of primary and secondary screening of phosphatome

Results obtained with indicated RNAi lines in crosses indicated in top line of table, under primary screen and secondary screen headings. Blank infill indicates that there was no change in targets in primary screen. Black infill means there was no change observed in in that particular assay although there was an effect either with another readout or another RNAi line corresponding to the same gene. Coloured infill represents changes to readouts as indicated in legend. Genes that were tested in secondary screen are highlighted by lines above and below the gene. Adult phenotypes (if observed) are indicated. Any indicated lethality occurred after third instar, thus allowing larval discs to be examined. Transgenic RNAi Project strains are in blue font, while National Institute of Genetics (NIG) strains have the letter R in the stock number. All other strains are from Vienna Drosophila RNAi Center (VDRC).

Fig. S6 Signaling at the dorsal/ventral compartment boundary of the *Drosophila* wing disc.

The Notch pathway signals from non-boundary cells to induce expression of the target genes *wg* (red) and *ct* (orange) in cells at the dorsal/ventral compartment boundary of the wing disc. Wg that is subsequently produced from cells at the dorsal/ventral compartment boundary induces the nested expression of the target genes *sens* (blue) and *Dll* (grey) in non-boundary cells that receive the ligand. The expression domains of *dpp-Gal4*, *hh-Gal4*, and *C5-Gal4* in the wing disc relative to boundary and non-boundary cells are shown in green.

Fig. S7 Identification of phospho-regulators of the Notch pathway.

(A) 36 kinases (and associated factors) [left column] and 32 phosphatases (and associated factors) [right column] of the 590 phospho-regulators screened in the wing disc were shown to regulate the Notch pathway and the levels of Wg and Ct. (B) All 68 phospho-regulators identified positively (blue) regulate the Notch pathway. (C) The kinase and phosphatase groups are displayed in a graphical summary, and family names are as defined in Figs. S1 and S2.

Fig.S8 Identification of phospho-regulators of the Hh pathway.

(A) *MS1096-Gal4* is expressed throughout the central region of the wing disc, but displays stronger expression in the dorsal half. Knock-down of *lacZ* (negative control) using *MS109-Gal4* did not affect Ci or Ptc levels. (B) 66 of the 90 candidates of the Wnt pathway also regulated the Hh pathway to either promote (21; blue) or inhibit (45; yellow) signaling in the

wing disc. 24 candidates of the Wnt pathway did not affect (black) the Hh pathway in the wing disc.

Fig.S9 Effects of *cdc37* and *gish* in the wing disc

(A) Relative to the knock-down of *lacZ* (negative control), knock-down of *Cdc37* with *MS1096-Gal4* in the wing disc resulted in increased levels of expression of the Hh pathway target *dpp-lacZ* (arrowheads) as detected by anti- β -gal. (B) The *dpp-Gal4* driver is expressed along the anterior/posterior compartment boundary of the wing disc (green). Knock-down of *cdc37* with *dpp-Gal4* increases the levels of PH3 (white). (C) The *hh-Gal4* driver is expressed in the posterior compartment of the wing disc (green). Knock-down of *gish* with *hh-Gal4* did not affect Dl (red) or Casp 3 levels (white).

Fig S1

Classifications of kinases and associated factors

AGC	Named after Protein Kinase A, G, and C families
Tyrosine Kinase-Like	
STE	Homologs of the yeast STE7, STE11 and STE20 genes
Casein Kinase 1	
Protein Kinase-Like	
CMGC	Named after a set of families (CDK, MAPK, GSK3 and CLK)
Tyrosine Kinase	
Receptor Guanylate Cyclase	
Calcium & Calmodulin-regulated Kinase	
Atypical	
Other	
Unclassified	Diverse group of kinases and candidate kinases
Kinase-associated	Co-factors such as cyclins
Non-protein Kinase	Such as lipid and sugar kinases

Kinase Gene			Gene Group	Gene Family	Human ortholog(s)
Annotation Symbol	Gene Symbol	Gene Name			
CG10023	Fak56D	Focal adhesion kinase	TK	FAK	PTK2, PTK2B
CG10033	for	foraging	AGC	PKG	PRKG1, PRKG2
CG10079	Egfr	Epidermal growth factor receptor	TK	EGFR	ERBB2, ERBB4, EGFR, ERBB3
CG10082			Non-Protein Kinase	Lipid Kinase	
CG10155	Spred	Sprouty-related protein with EVH-1 domain	Kinase-associated		SPRED3, SPRED2, SPRED1
CG10177			CAMK	DCAMKL	
CG10244	Cad96Ca	Cadherin 96Ca	TK		
CG10260			PKL		PI4KA
CG10268			Unclassified		PMVK
CG10295	Pak	PAK-kinase	STE	STE20	PAK3, PAK2, PAK1
CG10308	CycJ	Cyclin J	CMGC	CDK	
CG10498	cdc2c	cdc2c	CMGC	CDK	CDK1, CDK2, CDK3
CG10504	Ilk	Integrin linked kinase	TKL	MLK	ILK
CG10522	sticky		AGC	DMPK	CIT
CG10539	S6k	RPS6-p70-protein kinase	AGC	RSK	RPS6KB1
CG10572	Cdk8	Cyclin-dependent kinase 8	CMGC	CDK	CDK8, CDK19
CG10579	Eip63E	Ecdysone-induced protein 63E	CMGC	CDK	CDK16, CDK14, CDK18
CG10637	Nak	Numb-associated kinase	Other	NAK	BMP2KL, BMP2K, AAK1
CG10673			Other	Bud32	TP53RK
CG10702			Kinase-associated		
CG10738			RGC	RGC	NPR2, NPR1
CG10776	wit	wishful thinking	TKL	STKR	BMPR2, ACVR2A, TGFBR2, ACVR2B
CG10895	lok	loki	CAMK	RAD53	CHEK2
CG10933			Kinase-associated		
CG10951	niki	nimA-like kinase	Other	NEK	
CG10967	Atg1	Autophagy-specific gene 1	Other	ULK	

Kinases and Associated factors

CG1098	Madm	MLF1-adaptor molecule	Other	NRBP	NRBP1, NRBP2
CG1107	aux	auxillin	Other	NAK	DNAJC6, GAK
CG11221			Other	NKF1	
CG11228	hpo	hippo	STE	STE20	STK3, STK4
CG11249			Non-Protein Kinase	Carbohydrate Kinase	Pyruvate Kinase
CG11255			Non-Protein Kinase	Carbohydrate Kinase	ADK
CG11420	png	pan gu	Other		
CG11486			Kinase-associated		PAN3
CG11489	srpk79D	serine-arginine protein kinase at 79D	CMGC	SRPK	SRPK1, SRPK3, SRPK2
CG11525	CycG	Cyclin G	CMGC	CDK	
CG11533	Asator	Asator	CK1	TTBK	TTBK1, TTBK2
CG11594			Non-Protein Kinase	Carbohydrate Kinase	FGGY
CG11621	Pi3K68D	Phosphotidylinositol 3 kinase 68D	Non-Protein Kinase	Lipid Kinase	PIK3C2G, PIK3C2A, PIK3C2B
CG11660			Atypical	RIO	RIOK3, RIOK1
CG11811			Non-Protein Kinase	Nucleic Acid Kinase	GUK1
CG11859			Atypical	RIO	RIOK2
CG12019	Cdc37	Cdc37	Kinase-associated		CDC37
CG12066	Pka-C2	cAMP-dependent protein kinase 2	AGC	PKA	PRKACB
CG12069			AGC	PKA	
CG12072	warts	warts	AGC	NDR	LATS2, LATS1
CG1210	Pdk1	Phosphoinositide-dependent kinase 1	AGC	PDK1	PDPK1, PDPK2
CG12147			CK1	CK1	
CG1216	mri	mrityu	Kinase-associated		KCTD20, BTBD10
CG12229			Non-Protein Kinase	Carbohydrate Kinase	Pyruvate Kinase
CG12244	lic	licorne	STE	STE7	MAP2K4, MAP2K6, MAP2K3
CG1227			Other	NAK	STK16
CG12289			Non-Protein Kinase	Carbohydrate Kinase	KHK
CG12306	polo	polo	Other	PLK	PLK3, PLK2, PLK1
CG12559	rl	rolled	CMGC	MAPK	MAPK3, MAPK1
CG1271			Non-Protein Kinase	Lipid Kinase	GK5
CG13369			Non-Protein Kinase	Carbohydrate Kinase	RBKS
CG13388	Akap200	A kinase anchor protein 200	Kinase-associated		UNC79
CG1344			Other	SCY1	SCYL3
CG13591	Ssl	Suppressor of Stellate-like	CMGC		CSNK2B
CG1362	cdc2rk	cdc2-related-kinase	CMGC	CDK	CDC2L2
CG13688	lpk2	lpk2	PKL	IPK	IPMK
CG1389	tor	torso	Atypical	PIKK	
CG14026	tkv	thickveins	TKL	STKR	ACVR1, BMPR1A, ACVR1C, BMPR1B, ACVRL1, ACVR1B
CG14030	Bub1	Bub1 homologue	Other	BUB	BUB1
CG14217	Tao	Tao	STE	STE20	TAOK1, TAOK2, TAOK3
CG14305			CAMK	TSSK	TSSK6, TSSK2, TSSK4, TSSK3, TSSK1

Kinases and Associated factors

CG14396	Ret	Ret oncogene	TK	Ret	FGFR4, FGFR1, FGFR3, RET, FGFR2
CG14895	Pak3	Pak3	STE	STE20	PAK3, PAK1, PAK2
CG14939	CycY	Cyclin Y	CMGC	CDK	CCNYL2, CCNY, CCNYL1, CCNYL3
CG1495	CaMK1	Calcium/calmodulin-dependent protein kinase I	CAMK	CAMK1	PSKH2, CAMK4, CAMK1D, CAMK1D, PSKH1, PNCK, CAMK1
CG14992	Ack	Activated Cdc42 kinase	TK	Ack	TNK2, TNK1
CG1511	Eph	Eph receptor tyrosine kinase	TK	Eph	EPHB6, EPHB3, EPHA1, EPHA10, EPHA4, EPHB2, EPHA8, EPHA2
CG15218	CycK	Cyclin K	CMGC	CDK	CCNK
CG15224	CkII β	Casein kinase II β subunit	CMGC	CK2	CSNK2B
CG15547			Non-Protein Kinase	Nucleic Acid Kinase	
CG15793	Dsor1	Downstream of raf1	STE	STE7	MAP2K1, MAP2K2
CG15862	Pka-R2	cAMP-dependent protein kinase R2	AGC	PKA	PRKAR2A, PRKAR2B
CG1594	hop	hopscotch	TK	Jak	
CG1609	Gcn2	Gcn2	Other	PEK	EIFA2K4
CG16708	Cerk	Ceramide kinase	Non-Protein Kinase	Lipid Kinase	CERK
CG16903			Kinase-associated		CCNL1, CCNL2
CG16910	key	kenny	Kinase-associated		
CG16973	msn	misshapen	STE	STE20	MINK1, MAP4K4, TNK1
CG17010			Non-Protein Kinase	Carbohydrate Kinase	RBKS
CG17090	hipk	homeodomain interacting protein kinase	CMGC	DYRK	HIPK1, HIPK2, HIPK3
CG17146	Adk1	Adenylate kinase-1	Non-Protein Kinase	Nucleic Acid Kinase	AK1, AK5
CG17161	grp	grapes	CAMK	CAMKL	CHEK1
CG17216	KP78b	KP78b	CAMK	CAMKL	MARK2, MARK4, MARK3, MARK1
CG17245	plexB	plexin B	Kinase-associated		PLXNA3, PLXNB1, PLXNA1, PLXND1, PLXNA4, PLXNB3, PLXNB2, PLXNA2
CG1725	dlg1	discs large 1	Kinase-associated		DLG3, DLG2, DLG4, DLG1
CG17256	Nek2	Nek2	Other	NEK	NEK7, NEK2, NEK6
CG17299	SNF4Ay	SNF4/AMP-activated protein kinase gamma subunit	PKL	PIP	PRKAG1, PRKAG2, PRKAG3
CG17342	Lk6	Lk6	CAMK	MAPKAPK	MKNK2
CG17348	drl	derailed	TK	Ryk	RYK, TYRO, AXL, MERTK
CG1747	Sk1	Sphingosine kinase 1	Non-Protein Kinase	Lipid Kinase	SPHK1, SPHK2
CG17471			PKL	PIP	PIP4K2B, PIP4K2C, PIP4K2A
CG17520	CkII α	casein kinase II α	CMGC	CK2	CSNK2A1, CSNK2A2
CG17528			CAMK	DCAMKL	DCX, DCLK3, DCLK1, DCLK2
CG17559	dnt	doughnut on 2	TK	Ryk	RYK, TYRO3, AXL, MERTK
CG17596	S6kII	Ribosomal protein S6 kinase II	AGC	RSK	RPS6KA6, RPS6KA5, RPS6KA4, RPS6KA3, RPS6KA2, RPS6KA1
CG17603	Taf1	TBP-associated factor 1	Atypical	TAF1	TAF1L, TAF1
CG17698			Other	CAMKK	CAMKK1, CAMKK2
CG1772	dap	dacapo	Kinase-associated		CDKN1B
CG17998	Gprk2	G protein-coupled receptor kinase 2	AGC	GRK	GRK4, GRK5, GRK6
CG18069	CaMKII	Calcium/calmodulin-dependent protein kinase II	CAMK	CAMK2	CAMK2A, CAMK2D, CAMK2B, CAMK2G
CG18085	sev	sevenless	TK	Sev	ROS1

Kinases and Associated factors

CG18247	shark	SH2 ankyrin repeat kinase	TK	Syk	
CG18255	Strn-Mlck	Stretchin-Mick	CAMK	MLCK	MYLK4, MYLK2, MYLK, MYLK3
CG1830	PhKy	Phosphorylase kinase γ	CAMK	PHK	PHKG2
CG18374	Gyk	Glycerol kinase	Non-Protein Kinase	Lipid Kinase	GK, GK2
CG18402	InR	Insulin-like receptor	TK	InsR	IGF1R
CG1848	LIMK1	LIM-kinase1	TKL	LISK	LIMK1, LIMK2
CG18492	Tak1	TGF-β activated kinase 1	TKL	MLK	MAP3K7
CG1851	Ady43A	Ady43A	Non-Protein Kinase	Nucleic Acid Kinase	
CG18582	mbt	mushroom bodies tiny	STE	STE20	PAK6, PAK4, PAK7
CG18854			Non-Protein Kinase	Lipid Kinase	
CG1891	sax	saxophone	TKL	STKR	ACVR1, BMPR1A, ACVR1C, BMPR1B, ACVRL1
CG1915	sls	sallimus	Kinase-associated		TTN
CG1939	Dpck	Dephospho-CoA kinase	Unclassified		DCAKD
CG1951			Other	SCY1	SCYL2
CG1954	Pkc98E	Protein C kinase 98E	AGC	PKC	PRKCG, PRKCB, PRKCE, PRKCQ, PRKCA, PRKCH,
CG1973	yata	yata	Other	SCY1	SCYL1
CG2028	Ckla	Casein kinase Iα	CK1	CK1	CSNK1A1, CSNK1D, CSNK1A1L, CSNK1E
CG2048	dco	discs overgrown	CK1	CK1	CSNK1A1, CSNK1D, CSNK1A1L, CSNK1E
CG2049	Pkn	Protein kinase related to protein kinase N	AGC	PKN	PKN2, PKN3, PKN1
CG2087	PEK	pancreatic eIF-2α kinase	Other	PEK	EIF2AK3
CG2201			Non-Protein Kinase	Lipid Kinase	CHKB-CPT1B, CHKB
CG2210	awd	abnormal wing discs	Atypical	NDK	NME4, NME2, NME1, NME1-NME2, NME3
CG2246			Non-Protein Kinase	Carbohydrate Kinase	PRPSAP2, PRPSAP1
CG2252	fs(1)h	female sterile (1) homeotic	Atypical		BRD2, BRD3, BRDT, BRD4
CG2272	slpr	slipper	TKL	MLK	RP5-862P8.2, MLTK, MAP3K9, MAP3K10, MAP3K11
CG2577			CK1	CK1	CSNK1E, CSNK1A1, CSNK1A1L, CSNK1D
CG2615	ik2	IκB kinase-like 2	Other	IKK	IKBKE, TBK1
CG2621	sgg	shaggy	CMGC	GSK	GSK3B, GSK3A
CG2699	Pi3K21B	Pi3K21B	PKL	PIK	PIK3R3, PIK3R2, PIK3R1
CG2794			Non-Protein Kinase	Carbohydrate Kinase	
CG2845	pfl	pole hole	TKL	RAF	ARAF, BRAF, RAF1
CG2846			Non-Protein Kinase	Carbohydrate Kinase	RFK
CG2899	ksr	kinase suppressor of ras	TKL	RAF	KSR2, KSR1
CG2929	Pi4KIIα	Pi4KIIα	PKL	PIK	PI4K2A, PI4K2B
CG2964			Non-Protein Kinase	Carbohydrate Kinase	PKLR, PKM
CG3001	Hex-A	Hexokinase A	Non-Protein Kinase	Carbohydrate Kinase	HK3, HK2, HKDC1, HK1
CG30021	metro	menage a trois	Non-Protein Kinase	Nucleic Acid Kinase	MPP4, MPP5, MPP7, MPP2, MPP1, MPP3
CG3008			Atypical	RIO	RIOK3, RIOK1
CG30184			Kinase-associated		
CG30295	lpk1	lpk1	PKL		IPPK
CG3051	SNF1A	SNF1A/AMP-activated protein kinase	CAMK	CAMKL	PRKAA1, PRKAA2
CG3068	aur	aurora	Other	Aur	AURKA, AURKAB, AURKBC

Kinases and Associated factors

CG3000	dai	dai1d	Other	Aui	AURK, AURKA, AURKB
CG3086	MAPk-Ak2	MAP kinase activated protein-kinase-2	CAMK	MAPKAPK	MAPKAPK2, MAPKAPK5, MAPKAPK3
CG31003	gskt	gasket	CMGC	GSK	GSK3B, GSK3A
CG3105	Pask	PAS kinase	CAMK	CAMKL	PASK
CG31097			Unclassified		
CG31127	Wsck	Wsck	TK		
CG31140			Non-Protein Kinase	Lipid Kinase	DGKQ
CG31183			Kinase-associated		NPR2, NPR1
CG3127	Pgk	Phosphoglycerate kinase	Non-Protein Kinase	Lipid Kinase	PGK1, PGK2
CG31349	pyd	polychaetoid	Kinase-associated		TJP2, TJP1, TJP3
CG3140	Adk2	Adenylate kinase-2	Non-Protein Kinase	Nucleic Acid Kinase	AK4, AK2, AK3
CG31421	Takl1	Tak1-like 1	TKL		MAP3K7
CG31643			Atypical		FASTKD1, TBRG4, FASTKD3, FASTK, FASTKD3
CG3172	twf	twinfilin	TK		TWF2, TWF1
CG31751			Unclassified		AGPHD1
CG31873			Non-Protein Kinase	Lipid Kinase	FLJ10842
CG32019	bt	bent	CAMK		MYLK4, MYLK2, MYLK3, MYLK
CG32031	Argk	Arginine kinase	Unclassified		CKMT2, CKMT1A, CKMT1B,
CG32134	btl	breathless	TK		CKM, CKB, FGFR4, FGFR1, FGFR3, RET, FGFR2
CG3216			RGC	RGC	NPR2, NPR1
CG32417	Myt1	Myt1	AGC		PKMYT1
CG32484	Sk2	Sphingosine kinase 2	Non-Protein Kinase	Lipid Kinase	
CG32649			Atypical		ADCK4, ADCK3
CG32666	Drak	Death-associated protein kinase related	CAMK	MLCK	
CG32703			CMGC		MAPK15
CG32717	sdt	stardust	CAMK		CASK, MPP4, MPP5, MPP7, MPP2, MPP1, MPP3
CG32742	I(1)G0148	lethal (1) G0148	Other		CDC7
CG32743	nonC	no-on-and-no-off transient C	PKL		SMG1
CG3277			TK	VEGFR	
CG32849	Hex-t2	Hex-t2	Non-Protein Kinase	Carbohydrate Kinase	HK3, HK2, HKDC1, HK1
CG32944			CAMK		STK32C, STK32B, STK32A
CG33102	Hex-t1	Hex-t1	Non-Protein Kinase	Carbohydrate Kinase	
CG33114	Gyc32E	Guanyl cyclase at 32E	RGC		NPR2, NPR1
CG3319	Cdk7	Cyclin-dependent kinase 7	CMGC	CDK	CDK7, CDK20
CG3324	Pkg21D	cGMP-dependent protein kinase 21D	AGC	PKG	PRKG2, PRKG1
CG3338	p38c	p38c	CMGC	MAPK	MAPK12, MAPK13, MAPK14, MAPK11
CG33519	Unc-89	Unc-89	Unclassified		
CG33531	Ddr	Discoidin domain receptor	TK		DDR1, DDR2
CG33554	Nipped-A	Nipped-A	Atypical	PIKK	TRRAP
CG33671			Non-Protein Kinase	Carbohydrate Kinase	MVK
CG33981			Kinase-associated		ANAPC13
CG3400	Pfrx	6-phosphofructo-2-kinase	Non-Protein Kinase	Carbohydrate Kinase	PFKFB1, PFKFB2, PFKFB4, PFKFB3
CG34344	rdgA	retinal degeneration A	Non-Protein Kinase	Lipid Kinase	DGKZ, DGKI
CG34356			Unclassified		
CG34357			RGC		NPR2, NPR1
CG34359	IP3K2	Inositol 1,4,5-triphosphate kinase 2	Non-Protein Kinase	Lipid Kinase	ITPK1
CG34361	Dgk	Diacyl glycerol kinase	Non-Protein Kinase	Lipid Kinase	DGKA, DGKG, DGKB

Kinases and Associated factors

Kinases and Associated factors						
			TK			
CG34380	Epac	Exchange protein directly activated by cAMP ortholog	Non-Protein Kinase	Lipid Kinase	DDR1, DDR2	
			Kinase-associated	RAPGEF5, RAPGEF3, RAPGEF4, RAPGEF1		
			Other	TLK	TLK1, TLK2	
CG34412	tlk	Tousled-like kinase	Other	TLK	TLK1, TLK2	
CG3510	CycB	Cyclin B	CMGC	CDK	CCNB2, CCNB1	
CG3525	eas	easily shocked	Non-Protein Kinase	Lipid Kinase	ETNK2, ETNK1	
CG3534			Non-Protein Kinase	Carbohydrate Kinase	XYLB	
CG3544			Non-Protein Kinase	Carbohydrate Kinase	XYLB	
CG3608			Atypical	ABC1	ADCK1	
CG3682	PIP5K59B	PIP5K59B	PKL	PIP5K1A, PIP5K1B, PSMD4, PIP5K1C		
CG3738	Cks30A	Cyclin-dependent kinase subunit 30A	Kinase-associated	CKS2, CKS1B		
CG3809			Non-Protein Kinase	Nucleic Acid Kinase	ADK	
CG3837			Kinase-associated			
CG3915	Drl-2	Derailed 2	TK	Ryk	RYK, TYRO3, AXL, MERTK	
CG3938	CycE	Cyclin E	CMGC	CDK	CCNE2, CCNE1	
CG4006	Akt1	Akt1	AGC	Akt	SGK3, AKT1, SGK1, AKT2, SGK2, AKT3	
CG4007	Nrk	Neurospecific receptor kinase	TK	Musk	MUSK, ROR1, ROR2	
CG4012	gek	genghis khan	AGC	DMPK	CDC42BPA, CDC42BPG, CDC42BPB	
CG40129	Gprk1	G protein-coupled receptor kinase 1	AGC	GRK	ADRBK2, ADRBK1	
CG4026	IP3K1	Inositol 1,4,5-triphosphate kinase 1	PKL	ITPKB, ITPKC, ITPKA		
CG40293	Stlk	Ste20-like kinase	STE	STRADA, STRADB		
CG4032	Abl	Abl tyrosine kinase	TK	Abl	ABL1, ABL2	
CG4041			Other	TBCK	TBCK	
CG40478	Dyrk3	Dyrk3	CMGC	DYRK	DYRK4, DYRK1B, DYRK1A, DYRK3, DYRK2	
CG4132	pkaap	pkaap	Kinase-associated	AKAP10		
CG4141	Pi3K92E	Pi3K92E	PKL	PIK3CB, PIK3CD, PIK3CA, PIK3CG		
CG4201	ird5	immune response deficient 5	Other	IKK	IKBKB, CHUK	
CG42273	mnb	minibrain	CMGC	DYRK	DYRK4, DYRK1B, DYRK1A, DYRK3, DYRK2	
CG42317	Csk	C-terminal Src kinase	TK	Csk	MATK, CSK	
CG42320	Doa	Darkener of apricot	CMGC	CLK	CLK4, CLK2, CLK3, CLK1	
CG42341	Pka-R1	cAMP-dependent protein kinase R1	AGC	PKA	PRKAR1B, PRKAR1A	
CG42347			CAMK	MYLK4, MYLK2, MYLK3		
CG42349	Pkcδ	Protein kinase C δ	AGC	PKC	PRKCG, PRKCB, PRKCE, PRKCQ, PRKCA, PRKCH,	
CG42366			CMGC	CDC2	ICK, MAK	
CG42403	Ca-β	Ca2+-channel-protein-β-subunit	Non-Protein Kinase	Nucleic Acid Kinase	CACNB3, CACNB1, CACNB4, CACNB2	
CG4252	mei-41	meiotic 41	Atypical	PIKK	ATR	
CG42636	Gyc76c	Guanylyl cyclase at 76C	RGC	RGC	NPR2, NPR1	
CG4268	Pitslre	Pitslre	CMGC	CDK	CDC2L2	
CG42783	aPKC	atypical protein kinase C	AGC	PKC	PRK CZ, PRK CI	
CG42856	Sik3	Salt-inducible kinase 3	CAMK	CAMKL	SIK2, SIK3, SIK1	
CG4290	Sik2	Salt-inducible kinase 2	CAMK	CAMKL	SIK2, SIK3, SIK1	
CG43143			Unclassified			
CG4353	hep	hemipterus	STE	STE7	MAP2K7	
CG43729			Kinase-associated	STAC, STAC3, STAC2		
CG43741	Ack-like	Activated Cdc42 kinase-like	TK	Ack		

Kinases and Associated factors

CG4379	Pka-C1	cAMP-dependent protein kinase 1	AGC	PKA	PRKACA, PRKACB, PRKACG, PRKX
CG4488	wee	wee	Other	WEE	WEE2, WEE1
CG4523	Pink1	PTEN-induced putative kinase 1	Other	NKF2	PINK1
CG4527	slik	Sterile20-like kinase	STE	STE20	STK10
CG4546			Unclassified		MYCBP
CG4551	smi35A	smell impaired 35A	CMGC	DYRK	DYRK4, DYRK1B, DYRK1A, DYRK3, DYRK2
CG4583	Ire1	Inositol-requiring enzyme-1	Other	IRE	ERN2, ERN1
CG4629			CAMK	CAMKL	NIM1
CG4720	Pk92B	Protein kinase at 92B	STE	STE11	MAP3K6, MAP3K5, MAP3K15
CG4798	I(2)k01209	lethal (2) k01209	Non-Protein Kinase	Nucleic Acid Kinase	UCKL1
CG4803	Takl2	Tak1-like 2	TKL	MLK	MAP3K7
CG4839			AGC	PKG	PRKG1, PRKG2
CG4926	Ror	Ror	TK	Ror	MUSK, ROR1, ROR2
CG4945			Other	NKF1	
CG5072	Cdk4	Cyclin-dependent kinase 4	CMGC	CDK	CDK4, CDK6
CG5092	Tor	Target of rapamycin	Atypical	PIKK	MTOR
CG5125	ninaC	neither inactivation nor afterpotential C	STE	STE20	
CG5144			Non-Protein Kinase	Nucleic Acid Kinase	CKM, CKB, CKMT2, CKMT1B, CKMT1A
CG5169	GckIII	Germinal centre kinase III	STE	STE20	STK25, STK24, MST4
CG5179	Cdk9	Cyclin-dependent kinase 9	CMGC	CDK	CDK13, CDK9, CDK12
CG5182	Pk34A	PK34A	CMGC	GSK	
CG5288			Non-Protein Kinase	Carbohydrate Kinase	GALK2
CG5310	nmdyn-D6	nmdyn-D6	Atypical		NME6
CG5363	cdc2	cdc2	CMGC	CDK	CDK1, CDK2, CDK3
CG5373	Pi3K59F	Phosphatidylinositol 3 kinase 59F	CMGC	CDK	PIK3C3
CG5387	Cdk5α	Cdk5 activator-like protein	Kinase-associated		CDK5R1
CG5408	trbl	tribbles	CAMK	Trbl	TRIB2, TRIB3, TRIB1
CG5452	dnk	deoxyribonucleoside kinase	Non-Protein Kinase	Nucleic Acid Kinase	TK2
CG5475	Mpk2	Mpk2	CMGC	MAPK	MAPK12, MAPK13, MAPK14, MAPK11
CG5483	Lrrk	Leucine-rich repeat kinase	TKL	LRRK	LRRK2, LRRK1
CG5626			Non-Protein Kinase	Nucleic Acid Kinase	
CG5680	bsk	basket	CMGC	MAPK	MAPK10, MAPK9
CG5725	fbl	fumble	Non-Protein Kinase	Lipid Kinase	PANK2, PANK1, PANK3
CG5757			Non-Protein Kinase	Nucleic Acid Kinase	DTYMK
CG5790			Other	CDC7	CDC7
CG5940	CycA	Cyclin A	CMGC	CDK	CCNA1, CCNA2
CG5974	pll	pelle	TKL	IRAK	IRAK2
CG6027	cdi	center divider	TKL	LISK	TESK1
CG6092	Dak1	Dak1	Non-Protein Kinase	Nucleic Acid Kinase	CMPK
CG6114	sff	sugar-free frosting	CAMK	CAMKL	BRSK1, BRSK2
CG6117	Pka-C3	cAMP-dependent protein kinase 3	AGC	PKA	PRKACG, PRKACA, PRKACB, PRKX
CG6214	MRP	Multidrug-Resistance like Protein 1	Kinase-associated		ABCC1, ABCC2, ABCC4, ABCC3
CG6292	CycT	Cyclin T	CMGC	CDK	CCNT1, CCNT2
CG6297	JIL-1	JIL-1	AGC	RSK	RPS6KA6, RPS6KA5, RPS6KA4, RPS6KA3, RPS6KA2, RPS6KA1
CG6343	ND42	NADH:ubiquinone reductase 42kD subunit precursor	Kinase-associated		NDUFA10

Kinases and Associated factors

CG6355	fab1	fab2	PKL		PIKFYVE
CG6364			Other	UK	UCK2, UCK1
CG6386	ball	ballchen	CK1	VRK	VRK3, VRK1, VRK2
CG6498			AGC	MASR	MAST1, MAST3, MAST4, MAST2
CG6509			Kinase-associated		DLG5
CG6518	inaC	inactivation no afterpotential C	AGC	PKC	PRKCG, PRKCB, PRKCE, PRKCQ, PRKCA, PRKCH,
CG6521	Stam	Signal transducing adaptor molecule	Kinase-associated		STAM2, STAM
CG6535	tefu	telomere fusion	Atypical	PIKK	ATM
CG6551	fu	fused	Other	ULK	STK36
CG6612	Adk3	Adenylate kinase-3	Non-Protein Kinase	Nucleic Acid Kinase	AK4, AK2, AK3
CG6620	ial	IplI-aurora-like kinase	Other	AUR	AURKC, AURKB, AURKA
CG6622	Pkc53E	Protein C kinase 53E	AGC	PKC	PRKCG, PRKCB, PRKCE, PRKCQ, PRKCA, PRKCH,
CG6703	CASK	CASK ortholog	CAMK	CAMKL	CASK, MPP4, MPP5, MPP7, MPP2, MPP1, MPP3
CG6715	KP78a	KP78a	CAMK	CAMKL	MARK2, MARK4, MARK3, MARK1
CG6767			Non-Protein Kinase	Carbohydrate Kinase	PRPS1L1, PRPS1
CG6772	Slob	Slowpoke binding protein	Other	Slob	
CG6775	rg	rugose	Kinase-associated		NBEA, LRBA
CG6800			CMGC	CDK	CDK7, CDK20
CG6875	asp	abnormal spindle	Kinase-associated		ASPM
CG6963	gish	gilgamesh	CK1	CK1	CSNK1G1, CSNK1G3, CSNK1G2
CG7001	Pk17E	Protein kinase-like 17E	AGC	RSKR	
CG7004	fwd	four wheel drive	PKL		PIK4B
CG7028	PRP4	PRP4	CMGC	DYRK	PRPF4B
CG7069			Non-Protein Kinase	Carbohydrate Kinase	PKM2, PKLR
CG7070	PyK	Pyruvate kinase	Non-Protein Kinase	Carbohydrate Kinase	PKM2, PKLR
CG7094			CK1	CK1	CSNK1E, CSNK1A1, CSNK1A1L, CSNK1D
CG7097	hppy	happyhour	STE	STE20	MAP4K2, MAP4K3, MAP4K1, MAP4K5
CG7103	Pvf1	PDGF- and VEGF-related factor 1	Kinase-associated		VEGFA, VEGFB, VEGFD
CG7125	PKD	Protein Kinase D	CAMK	PKD	PRKD1, PRKD2, PRKD3
CG7156			AGC	RSKL	
CG7177	Wnk	WNK homolog	Other	WNK	WNK1, WNK4, WNK2, WNK3
CG7186	SAK	Sak kinase	Other	PLK	PLK4
CG7207	cert	ceramide transfer protein	Kinase-associated		COL4A3BP
CG7223	htl	heartless	TK	FGFR	FGFR4, FGFR1, FGFR3, RET, FGFR2
CG7236			CMGC	CDKL	CDKL3, CDKL5, CDKL1, CDKL2
CG7281	CycC	Cyclin C	CMGC	CDK	CCNC
CG7328			Non-Protein Kinase	Carbohydrate Kinase	KHK
CG7335			Non-Protein Kinase	Carbohydrate Kinase	KHK
CG7362			Non-Protein Kinase	Carbohydrate Kinase	PKLR, PKM2
CG7393	p38b	p38b	CMGC	MAPK	MAPK12, MAPK13, MAPK14, MAPK11
CG7405	CycH	Cyclin H	CMGC	CDK	CCNH
CG7470			Unclassified		ALDH18A1
CG7524	Src64B	Src oncogene at 64B	TK	Src	HCK, LCK, FRK, SRC, BLK, YES1, FGR, FYN, LYN
CG7525	Tie	Tie-like receptor tyrosine kinase	TK		TIE1
CG7551			Non-Protein Kinase	Carbohydrate Kinase	KHK

Kinases and Associated factors

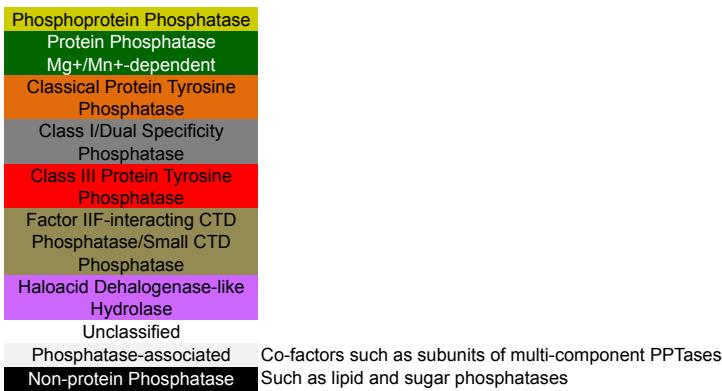
CG7597	Cdk12	Cdk12	CMGC	CDK	CDK13, CDK9, CDK12
CG7616			Atypical	ABC1	ADCK5
CG7643	ald	altered disjunction	Other	TTK	TTK
CG7693	fray	frayed	STE	STE20	OXSR1
CG7717	Mekk1	Mekk1	STE	STE11	MAP3K4
CG7719	gwl	greatwall	AGC	MAST	MASTL
CG7766			Kinase-associated		PHKA1, PHKA2
CG7838	BubR1	Bub1-related kinase	Other	BUB	BUB1B
CG7873	Src42A	Src oncogene at 42A	TK	Src	HCK, LCK, FRK, SRC, BLK, YES1, FGR, FYN, LYN
CG7892	nmo	nemo	CMGC	MAPK	NLK
CG7904	put	punt	TKL	STKR	BMPR2, ACVR2A, TGFBR2, ACVR2B
CG7995			Non-Protein Kinase	Lipid Kinase	GK, GK2
CG8049	Btk29A	Btk family kinase at 29A	TK	Tec	TEC, ITK, TXK, BMX, BTK
CG8057	alc	alicorn	Non-Protein Kinase	Nucleic Acid Kinase	PRKAB2, PRKAB1
CG8094	Hex-C	Hexokinase C	Non-Protein Kinase	Carbohydrate Kinase	HK3, HK2, HKDC1, HK1
CG8173			Other	TOPK	PBK
CG8174	SRPK	SRPK	CMGC	SRPK	SRPK1, SRPK3, SRPK2
CG8201	par-1	par-1	CAMK	CAMKL	MARK2, MARK4, MARK3, MARK1
CG8203	Cdk5	Cyclin-dependent kinase 5	CMGC	CDK	CDK5
CG8222	Pvr	PDGF- and VEGF-receptor related	TK	VEGFR	VEGFR3, VEGFR2, VEGFR1
CG8224	babo	baboon	TKL	STKR	ACVR1, BMPR1A, ACVR1C, ACVR1B, BMPR1B, ACVRL1
CG8239			Non-Protein Kinase	Carbohydrate Kinase	MVD
CG8250	Alk	Alk	TK	ALK	ALK, LTK, ROS1
CG8286	P58IPK	P58IPK	Kinase-associated		DNAJC3, DNAJC7
CG8298			Non-Protein Kinase	Lipid Kinase	GK, GK2
CG8351	Tcp-1 η	Tcp-1 η	Kinase-associated		CCT7
CG8362	nmdyn-D7	nmdyn-D7	Atypical		NME7
CG8363	Papss	PAPS synthetase	Non-Protein Kinase	Nucleic Acid Kinase	PAPSS1, PAPSS2
CG8475			CAMK		PHKB
CG8485			CAMK	CAMKL	SNRK
CG8565			CMGC	SRPK	SRPK1, SRPK3, SRPK2
CG8637	trc	tricornered	AGC	NDR	STK38
CG8657	Dgk ϵ	Diacyl glycerol kinase ϵ	Non-Protein Kinase	Lipid Kinase	DGKE
CG8726			Other	Slob	PXK
CG8767	mos	mos	Other	MOS	MOS
CG8789	wnd	wallenda	TKL	MLK	MAP3K12, MAP3K13
CG8808	Pdk	Pyruvate dehydrogenase kinase	Atypical	PDHK	PDK4, PDK3, PDK1, PDK2
CG8866			Other	ULK	ULK3
CG8874	Fps85D	Fps oncogene analog	TK	Fer	FER
CG8878			CK1	VRK	
CG8914	CK11 β 2	Casein kinase II β 2 subunit	CMGC		CSNK2B
CG8948	Graf	GTPase regulator associated with focal adhesion kinase ortholog	Kinase-associated		OPHN1, ARHGAP42, ARHGAP10, ARHGAP26
CG8967	otk	off-track	TK	CCK4	PTK7
CG9096	CycD	Cyclin D	CMGC	CDK	CCND1, CCND2, CCND3
CG9222			CAMK	TSSK	TSSK6, TSSK2, TSSK4, TSSK3,

Kinases and Associated factors

			CAKIN	IISIN	
CG9326	vari	varicose	Non-Protein Kinase	Nucleic Acid Kinase	TSSK1 MPP4, MPP5, MPP7, MPP2, MPP1, MPP3
CG9358	Phk-3	Pherokine 3	Non-Protein Kinase	Lipid Kinase	
CG9374	Ikb1	Ikb1	CAMK	CAMKL	STK11
CG9541			Non-Protein Kinase	Nucleic Acid Kinase	
CG9738	Mkk4	MAP kinase kinase 4	STE	STE7	MAP2K4, MAP2K6, MAP2K3
CG9746	ird1	immune response deficient 1	Other	VPS15	PIK3R4
CG9774	rok	Rho-kinase	AGC	DMPK	ROCK1, ROCK2
CG9961			Non-Protein Kinase	Lipid Kinase	PGK1, PGK2
CG9962			CK1	CK1	
CG9985	sktl	skittles	PKL		PIP5K1A, PIP5K1B, PSMD4, PIP5K1C

Fig S2

Classifications of phosphatases and associated factors



Drosophila Phosphatase			Gene Group	Gene Family	Human ortholog(s)
Annotation Symbol	Gene Symbol	Gene Name			
CG10089			PTP	Class I/DSP	DUSP22, DUSP15
CG10138	PpD5	Protein phosphatase D5	STP	PPP	PPP1CC, PPP1CA
CG10371	Plip	PTEN-like phosphatase	PTP	Classical	PTPMT1
CG10376			STP	PPM	PPM1F, PPM1E
CG10417			STP	PPM	PPM1G
CG10426			Non-Protein Phosphatase	Lipid Phosphatase	INPP5E
CG10443	Lar	Leukocyte-antigen-related-like	PTP	Classical	PTPRS, PTPRD, PTPRF
CG10493	Phipp	PH domain leucine-rich repeat protein phosphatase	STP	PPM	PHLPP1, PHLPP2
CG10574	I-2	Inhibitor-2		Phosphatase-associated	PPP1R2
CG10592				Unclassified	ALPL, ALPPL2, ALPP
CG10827				Unclassified	ALPL, ALPPL2, ALPP
CG10930	PpY-55A	Protein phosphatase Y at 55A	STP	PPP	
CG10975	Ptp69D	Protein tyrosine phosphatase 69D	PTP	Classical	PTPRG, PTPRZ1
CG11217	CanB2	Calcineurin B2	STP	PPP	PPP3R1, WDR92, PPP3R2
CG11425			Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG11426			Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG11437			Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG11438			Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG11440	laza	lazaro	Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG11516	Ptp99A	Protein tyrosine phosphatase 99A	PTP	Classical	PTPRG, PTPRZ1
CG11597			STP	PPP	
CG11883				Phosphatase-associated	
CG12034				Phosphatase-associated	SMPD2
CG12078			Asp-based catalysis	HAD	
CG12091			STP	PPM	PPTC7
CG12151	Pdp	Pyruvate dehydrogenase phosphatase	STP	PPM	PDP1, PDP2
CG12169	Ppm1	Ppm1	STP	PPM	
CG12173			Asp-based catalysis	HAD	ENOPH1
CG12217	PpV	Protein phosphatase V	STP	PPP	PPP4C, PPP6C, PPP2CA, PPP2CB
CG12252	Fcp1		Asp-based catalysis	FCP/SCP	CTDP1
CG1228	Ptpmeg	Ptpmeg	PTP	Classical	PTPN3, PTPN4

CG12538			Non-Protein Phosphatase	Nucleic Acid Phosphatase	
CG12746			Non-Protein Phosphatase	Lipid Phosphatase	
CG13125	TbCMF46	TbCMF47		Phosphatase-associated	
CG13197			PTP	Class I/DSP	
CG13311				Phosphatase-associated	
CG13570	spag	spaghetti		Phosphatase-associated	
CG1395	stg	string	PTP	Class I/DSP	CDC25A, CDC25B
CG14022			Non-Protein Phosphatase	Lipid Phosphatase	ACYP1, ACYP2
CG14080	Mkp3	Mitogen-activated protein kinase phosphatase 3	PTP	Class I/DSP	DUSP7, DUSP6, DUSP9
CG14211	MKP-4	MAPK Phosphatase 4	PTP	Class I/DSP	DUSP12
CG14297			PTP	Class III	ACP1
CG14411			PTP	Class I/DSP	MTMR11, MTMR10, MTMR12
CG1455	CanA1	Calcineurin A1	STP	PPP	PPP3CB, PPP3CC
CG14616	I(1)G0196	lethal (1) G0196	Non-Protein Phosphatase	Lipid Phosphatase	PPIP5K2, PPIP5K1
CG1462	Aph-4	Alkaline phosphatase 4		Unclassified	ALPL, ALPPL2, ALPP
CG14719	I-t	inhibitor-t		Phosphatase-associated	
CG14903			PTP	Class III	PTRHD1
CG15035			STP	PPM	PPTC7
CG15385			Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPL2
CG15400			Non-Protein Phosphatase	Carbohydrate Phosphatase	G6PC
CG15528			PTP	Class I/DSP	DUSP14, DUSP21
CG15533				Phosphatase-associated	SMPD1
CG15534				Phosphatase-associated	SMPD1
CG15743			Non-Protein Phosphatase	Lipid Phosphatase	IMPAD1
CG1637			Non-Protein Phosphatase	Nucleic Acid Phosphatase	PAPL
CG16717				Phosphatase-associated	MPPED2, MPPED1
CG16757	Spn	Spinophilin		Phosphatase-associated	
CG16771				Unclassified	ALPL, ALPPL2, ALPP
CG16870	Acyp	Acylphosphatase	Non-Protein Phosphatase	Lipid Phosphatase	ACYP1, ACYP2
CG16928	mre11	meiotic recombination 11		Phosphatase-associated	MRE11A
CG1696	Dd	Dullard	Asp-based catalysis	FCP/SCP	CTDNEP1
CG17026			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA2
CG17027			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA2
CG17028			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA2
CG17029			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA2
CG17124				Phosphatase-associated	PPP1R14C, PPP1R14B, PPP1R14D, PPP1R14A
CG17291	Pp2A-29B	Protein phosphatase 2A at 29B	STP	PPP	PPP2R1B, PPP2R1A
CG17598			STP	PPM	PPM1M, PPM1J, PPM1H
CG17746			STP	PPM	
CG1809				Unclassified	ALPL, ALPPL2, ALPP
CG1810	mRNA-cap	mRNA-capping enzyme	PTP	Class I/DSP	RNGTT
CG1817	Ptp10D	Protein tyrosine phosphatase 10D	PTP	Classical	PTPRB, PTPRJ, PTPRO
CG18243	Ptp52F	Ptp52F	PTP	Classical	
CG1906	alph	alphabet	STP	PPM	PPM1A, PPM1B

CG2096	flw	flapwing	STP	PPP	PPP1CB, PPP1CA, PPP1CC
CG2104			PTP	Unclassified	PPP2R4
CG2890	PPP4R2r	Protein phosphatase 4 regulatory subunit 2-related	STP	PPP	PPP4R2
CG2984	Pp2C1	Protein phosphatase 2C	STP	PPM	PP2CM
CG30103				Phosphatase-associated	NT5E
CG30104				Phosphatase-associated	NT5E
CG3028	Ipp	Inositol polyphosphate 1-phosphatase	Non-Protein Phosphatase	Lipid Phosphatase	INPP1
CG3059	NTPase	NTPase	Non-Protein Phosphatase	Nucleic Acid Phosphatase	ENTPD6, ENTPD5
CG31137	twin	twin		Phosphatase-associated	CNOT6L, CNOT6
CG31299	cu	curled		Phosphatase-associated	CCRN4L
CG31469			PTP	Class III	ACP1
CG31692	fbp	fructose-1,6-bisphosphatase	Non-Protein Phosphatase	Carbohydrate Phosphatase	FBP2, FBP1
CG31717			Non-Protein Phosphatase	Lipid Phosphatase	PPAPDC3, PPAPDC2
CG31759				Phosphatase-associated	PDE12
CG3178	Rrp1	Recombination repair protein 1		Phosphatase-associated	APEX1
CG31795	IA-2	IA-2 ortholog	PTP	Classical	PTPRN2, PTPRN
CG32156	Mbs	Myosin binding subunit	STP	PPP	PPP1R12A, PPP1R12B
CG3245	PpN-58A	Protein phosphatase N at 58A	STP	PPP	
CG32487			Asp-based catalysis	HAD	PDXP, PGP
CG32488			Asp-based catalysis	HAD	PDXP, PGP
CG32505	Pp4-19C	Protein phosphatase 19C	STP	PPP	PPP4C, PPP6C, PPP2CA, PPP2CB
CG32568			STP	PPP	
CG3264				Unclassified	ALPL, ALPPL2, ALPP
CG32697	I(1)G0232	lethal (1) G0232	PTP	Classical	PTPN9
CG32812			STP	PPP	
CG3289	Ptpa	Phosphotyrosyl phosphatase activator	PTP	Unclassified	PPP2R4
CG3290				Unclassified	ALPL, ALPPL2, ALPP
CG3292				Unclassified	ALPL, ALPPL2, ALPP
CG33747	primo-2	primo-2	PTP	Class III	ACP2
CG33748	primo-1	primo-1	PTP	Class III	ACP1
CG3376				Phosphatase-associated	SMPD1
CG34099	Mkp	MAP kinase-specific phosphatase	PTP	Class I/DSP	DUSP19
CG34140				Phosphatase-associated	PUSL1
CG3530			PTP	Class I/DSP	MTMR6, MTMR7, MTMR8
CG3573	Ocrl	Oculocerebrorenal syndrome of Lowe ortholog	Non-Protein Phosphatase	Lipid Phosphatase	INPP5B, OCRL
CG3632			PTP	Class I/DSP	MTMR3, MTMR4
CG3705	aay	astray	Asp-based catalysis	HAD	PSPH
CG3954	csw	corkscrew	PTP	Classical	PTPN11, PTPN6
CG3980	Cep97			Phosphatase-associated	CEP97
CG40448	Pp1-Y2	Pp1-Y2	STP	PPP	PPP1CB, PPP1CA, PPP1CC
CG4123	Mipp1	Multiple inositol polyphosphate phosphatase 1	Non-Protein Phosphatase	Lipid Phosphatase	
CG41534	Pp1-Y1	Pp1-Y1	STP	PPP	PPP1CB, PPP1CA, PPP1CC

CG4209	CanB	Calcineurin B	STP	PPP Phosphatase-associated	PPP3R1, WDR92, PPP3R2 NT5E
CG42249					
CG42271			Non-Protein Phosphatase	Lipid Phosphatase	INPP4A, INPP4B
CG42283	5Ptasel	5Ptasel	Non-Protein Phosphatase	Lipid Phosphatase	INPP5A
CG42327			PTP	Classical	
CG4317	Mipp2	Multiple inositol polyphosphate phosphatase 2	Non-Protein Phosphatase	Lipid Phosphatase	
CG4733	PR72	PR72	STP	PPP	PPP2R3A, PPP2R3B
CG4827	veil	veil		Phosphatase-associated	NT5E
CG4965	twe	twine	PTP	Class I/DSP	
CG4993	PRL-1	PRL-2	PTP	Class I/DSP	PTP4A1, PTP4A2, PTP4A3
CG5026			PTP	Class I/DSP	MTMR9
CG5150				Unclassified	ALPL, ALPPL2, ALPP
CG5171			Non-Protein Phosphatase	Carbohydrate Phosphatase	
CG5177			Non-Protein Phosphatase	Carbohydrate Phosphatase	
CG5276			Non-Protein Phosphatase	Nucleic Acid Phosphatase	CANT1
CG5361				Unclassified	
CG5567			Asp-based catalysis	HAD	PDXP, PGP
CG5577			Asp-based catalysis	HAD	PDXP, PGP
CG5643	wdb	widerborst	STP	PPP	PPP2R5D, PPP2R5E, PPP2R5C, PPP2R5A
CG5650	Pp1-87B	Protein phosphatase 1 at 87B	STP	PPP	PPP1CB, PPP1CA, PPP1CC
CG5656				Unclassified	ALPL, ALPPL2, ALPP
CG5671	Pten	Pten	PTP	Class I/DSP	PTEN, TPTE, TPTE2
CG5784	Mapmodulin	Mapmodulin		Phosphatase-associated	ANP32E, ANP32D, ANP32A, ANP32B
CG5820	Gp150			Phosphatase-associated	
CG5830			Asp-based catalysis	FCP/SCP	CTDSP1, CTDSP2, CTDSP1
CG5851	sds22	sds22	STP	PPP	PPP1R7
CG6036			STP	PPM	PPM1A, PPM1B
CG6235	tws	twins	STP	PPP	PPP2R2C, PPP2R2B, PPP2R2D, PPP2R2A
CG6238	ssh	slingshot	PTP	Class I/DSP	SSH2, SSH3, SSH1
CG6380			STP	PPP	PPP1R2
CG6542	EDTP	Egg-derived tyrosine phosphatase	PTP	Class I/DSP	MTMR14
CG6562	synj	synaptjanin	Non-Protein Phosphatase	Lipid Phosphatase	SYNJ2, SYNJ1
CG6571	rdgC	retinal degeneration C	STP	PPP	PPEF1, PPEF2
CG6593	Pp1α-96A	Protein phosphatase 1α at 96A	STP	PPP	PPP1CB, PPP1CA, PPP1CC
CG6656			Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPP, ACP2, ACPT
CG6746			PTP	Unclassified	PTPLA, PTPLB
CG6805			Non-Protein Phosphatase	Lipid Phosphatase	INPP5J, INPP5K
CG6896	MYPT-75D	MYPT-75D	STP	PPP	PPP1R16B, PPP1R16A
CG6899	Ptp4E	Protein tyrosine phosphatase 4E	PTP	Classical	PTPRB, PTPRJ, PTPRO
CG6939	Sbf	SET domain binding factor	PTP	Class I/DSP	SBF2, SBF1

CG7067	NitFhit	Nitrilase and fragile histidine triad fusion protein	Non-Protein Phosphatase	Nucleic Acid Phosphatase	NIT1, NIT2, FHIT
CG7109	mts	microtubule star	STP	PPP	PPP4C, PPP6C, PPP2CA, PPP2CB
CG7115			STP	PPM	PPM1L
CG7134	cdc14	cdc14	PTP	Class I/DSP	CDC14B, CDC14A
CG7180			PTP	Classical	PTPRK
CG7378			PTP	Class I/DSP	DUSP3, DUSP26, DUSP13, DUSP1, DUSP27
CG7615	fig	fos intronic gene	STP	PPM	PPTC7
CG7789			PTP	Class I/DSP	BPNT1
CG7850	puc	puckered	PTP	Class I/DSP	DUSP10
CG7899	Acph-1	Acid phosphatase 1	Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPP, ACP2, ACPT
CG7913	Pp2A-B'	Pp2A-B'	STP	PPP	PPP2R5D, PPP2R5E, PPP2R5C, PPP2R5A
CG7942	ldbr	lariat debranching enzyme		Phosphatase-associated	DBR1
CG8105				Unclassified	ALPL, ALPPL2, ALPP
CG8147				Unclassified	ALPL, ALPPL2, ALPP
CG8402	PpD3	Protein phosphatase D3	STP	PPP	PPP5C
CG8455			Non-Protein Phosphatase	Lipid Phosphatase	MPPE1
CG8509			PTP	Unclassified	PPP2R4
CG8584			Asp-based catalysis	FCP/SCP	
CG8804	wun	wunen	Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG8805	wun2	wunen-2	Non-Protein Phosphatase	Lipid Phosphatase	PPAP2C, PPAP2B, PPAP2A
CG8822	PpD6	Protein phosphatase D6	STP	PPP	PPP1CC, PPP1CA
CG8889	Mppe	Metallophosphoesterase	Non-Protein Phosphatase	Lipid Phosphatase	MPPE1
CG8980	NiPp1	Nuclear inhibitor of Protein phosphatase 1		Phosphatase-associated	PPP1R8
CG9115	mtm	myotubularin	PTP	Class I/DSP	MTMR1, MTM1, MTMR2
CG9128	Sac1	Sac1	Non-Protein Phosphatase	Lipid Phosphatase	SACM1L
CG9151	acj6	abnormal chemosensory jump 6		Phosphatase-associated	POU4F2, POU4F3
CG9156	Pp1-13C	Protein phosphatase 1 at 13C	STP	PPP	PPP1CB, PPP1CA, PPP1CC
CG9181	Ptp61F	Protein tyrosine phosphatase 61F	PTP	Classical	PTN2, PTPN1
CG9236	Cib2	CIB2 ortholog		Phosphatase-associated	CIB1, CIB4, CIB2
CG9238			STP	PPP	PPP1R3C
CG9267			PTP	Classical	PTPLAD1, PTPLAD2
CG9311	mop	myopic	PTP	Classical	PTPN23
CG9351	flf	falafel	STP	PPP	SMEK2, SMEK1
CG9389			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA1
CG9391			Non-Protein Phosphatase	Lipid Phosphatase	IMPA1, IMPA2
CG9449			Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPP, ACP2, ACPT
CG9451			Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPP, ACP2, ACPT
CG9452			Non-Protein Phosphatase	Nucleic Acid Phosphatase	ACPP, ACP2, ACPT
CG9493	Pez	Pez	PTP	Classical	PTPN14

CG9554	eya	eyes absent	Asp-based catalysis	HAD	EYA1, EYA4, EYA3, EYA2
CG9601			Non-Protein Phosphatase	Nucleic Acid Phosphatase	PNKP
CG9619	Gbs-76A	Glycogen binding subunit 76A	STP	PPP	PPP1R3A, PPP1R3C, PPP1R3B
CG9764	yrt	yurt		Phosphatase-associated	EPB41L4B, EPB41L5
CG9784			Non-Protein Phosphatase	Lipid Phosphatase	
CG9801			STP	PPM	
CG9819	CanA-14F	Calcineurin A at 14F	STP	PPP	
CG9842	Pp2B-14D	Protein phosphatase 2B at 14D	STP	PPP	PPP3CB, PPP3CC
CG9856	PTP-ER	Protein tyrosine phosphatase-ERK/Enhancer of Ras1	PTP	Class I/DSP	

Fig. S3

Drosophila Gene			PILOT SCREEN								
Annotation Symbol	Gene Symbol	Gene Name	RNAi Stock	UAS-dcr2::dpp-Gal4			RNAi Stock	UAS-dcr2;hh-Gal4			
				Wing disc				Wing disc			
				Wg	Sens	DII		Wg	Sens	DII	
CG4889	wg	wingless	13352				104579				
CG6205	por	porcupine	47864				100780				
CG6210	wls	wntless	103812				5215				
CG5912	arr	arrow	36286				6708				
CG11579	arm	armadillo	107344				7767				
CG7926	Axn	Axin	7748								
CG6193	Apc2	Adenomatous polyposis coli tumor suppressor homolog 2	22290				100104				
CG34403	pan	pangolin	108679				25940				

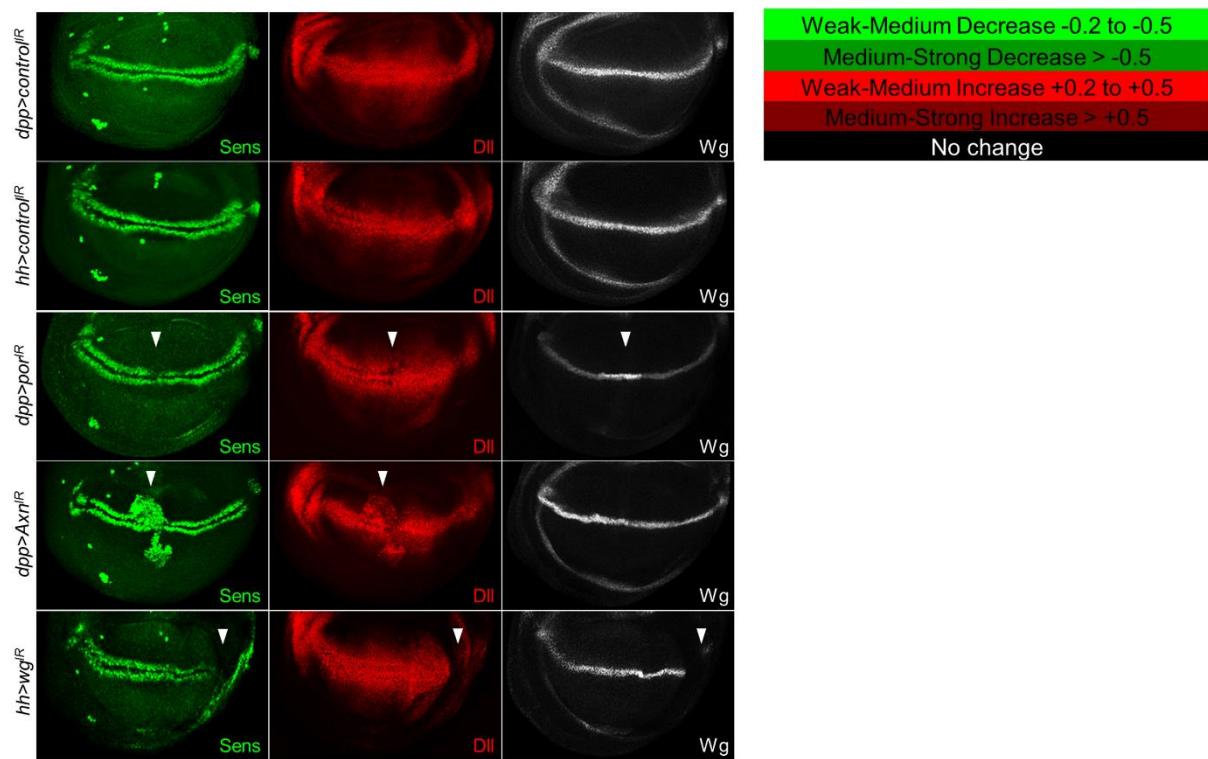


Fig. S4

Kinase Data

Weak-Medium Decrease -0.2 to -0.5
 Medium-Strong Decrease > -0.5
 Weak-Medium Increase +0.2 to +0.5
 Medium-Strong Increase > +0.5
 No change

Wing phenotype any visible wing phenotype including notches, blisters, vein spacing, altered size
 Lethal No adults recovered

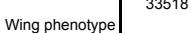
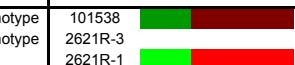
Kinase Gene		PRIMARY SCREEN									SECONDARY SCREEN						HEDGEHOG SCREEN			
Annotation Symbol	Gene Symbol	RNAi Stock	UAS-dcr2::dpp-Gal4			RNAi Stock	UAS-dcr2;hh-Gal4			RNAi Stock	UAS-dcr2;C5-Gal4 or UAS-dcr2; hh-Gal4			RNAi Stock	MS1096-Gal4;UAS-dcr2					
			Wing disc	Adult	Wg Sens Dll		Wing disc	Adult	Wg Sens Dll		Wing disc	Adult	Wing disc		Cut	Wing disc	Adult	Ci	Ptc	
CG10023	Fak56D	17957 108608																		
CG10033	for	38320 108293	Wing phenotype				10033R-1	Lethal		38320	Wing phenotype				38320 108293	Wing phenotype	Wing phenotype			
CG10079	Egfr	10033R-2 43268 43267 107130 10079R-2	Wing phenotype	Lethal						43268	Wing phenotype				43268 107130	Wing phenotype	Wing phenotype			
CG10082		38327 103749																		
CG10155	Spred	18024 18025																		
CG10177		38349 107848																		
CG10244	Cad96Ca	1091 8402 15993	Wing phenotype	Wing phenotype	Wing phenotype															
CG10260		105614 110640 10268R-2	Wing phenotype	Wing phenotype	Wing phenotype															
CG10268		12553 108937	Wing phenotype	Wing phenotype																
CG10295	Pak	31217 110222																		
CG10308	CycJ	104959 10498R-2 34856	Lethal	Wing phenotype																
CG10498	cdc2c	16062 47280																		
CG10504	Ilk	100265 18126	Wing phenotype	Wing phenotype																
CG10522	sticky	104369 45370	Wing phenotype	Wing phenotype																
CG10539	S6k	107187	Wing phenotype																	
CG10572	Cdk8																			
CG10579	Eip63E	47860 106824 50599 10579R-3	Wing phenotype	Wing phenotype	Wing phenotype	Wing phenotype	47860	Lethal	47860	Wing phenotype					47860 50599	Wing phenotype	Wing phenotype			
CG10637	Nak	35482 109507 27301																		
CG10673		27052 3691 100842	Wing phenotype	Wing phenotype	3691	Lethal	3691								27052 3691	Wing phenotype	Wing phenotype			
CG10738		28580 10738R-1																		

CG10776	wit	42244 865 103808 10776R-2				
CG10895	lok	44980 110342 46726	Wing phenotype			
CG10933		46727 104282	Wing phenotype			
CG10951	niki	16120 100823	Wing phenotype			
CG10967	Atg1	16133 10967R-1				
CG1098	Madm	27347 101758	Wing phenotype			
CG1107	aux	16182 103426	Wing phenotype Wing phenotype		16182	16182 103426
CG11221		100163 42947 29603	Wing phenotype			
CG11228	hpo	7823 104169 27661	Wing phenotype			
CG11249		18196 108319	Wing phenotype			
CG11255		17533 17534	Wing phenotype			
CG11420	png	31500 11420R-3				
CG11486		106497 51711 11486R-3 11486R-4	Wing phenotype Wing phenotype 11486R-3 11486R-4	Wing phenotype	106497	106497 11486R-3
CG11489	srpk79D	47544 102632				
CG11525	CycG	13655 106846				
CG11533	Asator	45120 45121				
CG11594		28311 105487				
CG11621	Pi3K68D	16240 109582				
CG11660		18526 105395	Wing phenotype			
CG11811		30915 110740	Wing phenotype			
CG11859		47401 109296	Wing phenotype Wing phenotype	11859R-2 11859R-3 31580	Lethal Lethal Lethal	109296
CG12019	Cdc37	47776 110727	Lethal Lethal		110727	47776 110727
CG12066	Pka-C2	30658 108424				
CG12069		23719 100886				
CG12072	warts	9928 106174 12072R-1	Wing phenotype Lethal Wing phenotype			
CG1210	Pdk1	18736 109812	Wing phenotype Wing phenotype			

CG12147		31659 101875 17043 101345				
CG1216	mri	31704 101097 106822	Wing phenotype			
CG12229		20166 38648 105610				
CG12244	lic	31725 105685	Wing phenotype			
CG1227						
CG12289						
CG12306	polo	20177 12306R-2 12306R-3	Wing phenotype Wing phenotype	12306R-2	20177 Wing phenotype	20177 Wing phenotype
CG12559	rl	43123 109108 13044 110178	Wing phenotype			12306R-3 Wing phenotype
CG1271						
CG13369		17161 17158 100747 13369R-1	Wing phenotype Wing phenotype Wing phenotype	17161 Lethal 13369R-1	100747 Wing phenotype	17161 Wing phenotype
CG13388	Akap200	5647 102374 108838	Wing phenotype Wing phenotype			
CG1344		17282				
CG13591	Ssl	108692				
CG1362	cdc2rk	32249 100299				
CG13688	lpk2	43824 43825 36280				
CG1389	tor	101154				
CG14026	tkv	3059 105834	Lethal Lethal			3059 Wing phenotype
CG14030	Bub1	101096 14030R-1 14030R-2	Wing phenotype Wing phenotype	101096 24833 14030R-1 14030R-2	105834 Wing phenotype Lethal Lethal	105834 Wing phenotype
CG14217	Tao	17432 107645 17477	Wing phenotype Wing phenotype			
CG14305		17478 14305R-3				
CG14396	Ret	30832 107648				
CG14895	Pak3	44607 107260				
CG14939	CycY	107010 32335 34009	Wing phenotype Wing phenotype	107010 32335 32334	107010 Wing phenotype	107010 Wing phenotype
CG1495	CaMK1	101380 1495R-2 39857				
CG14992	Ack	14992R-1 33899	Wing phenotype Wing phenotype/Lethal			
CG1511	Eph	4771 110448				
CG15218	CycK	36216 110774	Wing phenotype Wing phenotype		36216 Wing phenotype	36216 Wing phenotype

CG15224	Ckll β	32378 106845	Wing phenotype/Lethal Wing phenotype		106845	Wing phenotype	32378 106845	Wing phenotype Wing phenotype	
CG15547		32621 104520							
CG15793	Dsor1	40026 40025 107276	Wing phenotype Wing phenotype Wing phenotype	32920	Lethal	40026	Wing phenotype	40026 32920	Wing phenotype Wing phenotype
CG15862	Pka-R2	39437 101763 102830							
CG1594	hop	40037 32966	Wing phenotype Wing phenotype						
CG1609	Gcn2	32664 103976 1609R-2	Wing phenotype	32664 1609R-2		103976		32664 103976 1609R-4 1609R-2	Wing phenotype
CG16708	Cerk	43412 101550 37570 37572	Wing phenotype						
CG16903		7723	Wing phenotype						
CG16910	key	100257 101517	Lethal						
CG16973	msn	16973R-2 28791	Lethal						
CG17010		17010R-2 17010R-3	Wing phenotype						
CG17090	hipk	108254 32855 17090R-1	Wing phenotype Lethal	108254 32855	Wing phenotype Wing phenotype	32855	Wing phenotype	108254 32855	Wing phenotype Wing phenotype
CG17146	Adk1	25214 104475 12680							
CG17161	grp	110076							
CG17216	KP78b	51996 105265 27220							
CG17245	plexB	12167 46687 17245R-2							
CG1725	dlg1	41134 109274 1725R-2 1725R-1	Lethal	109274 41136	Lethal	41136	Wing phenotype	41134 109274	Wing phenotype
CG17256	Nek2	40052 103408 17299R-1							
CG17299	SNF4Ay	17299R-3 34726							
CG17342	Lk6	32885 109663	Wing phenotype						
CG17348	drl	27053 100039							
CG1747	Sk1	32932 32930							
CG17471		1747R-3							
CG17520	Cklla	17520R-2 31645	Wing phenotype Wing phenotype	17520R-2 31645	Lethal Lethal	31645		17520R-2 31645	Wing phenotype Wing phenotype
CG17528	dot	26292 CG17550 27057							

CG17359	uril	106056 5702 101451 41099 106119 Lethal Wing phenotype				
CG17596	S6kII	101451 41099 106119 35634 105884 36720				
CG17603	Taf1					
CG17698						
CG1772	dap					
CG17998	Gprk2	1835 101463 34843	1835 101463	Wing phenotype	101463	1835 101463 34843
CG18069	CaMKII	47280 100265 49925 107048 105706 25304 18255R-1 18255R-3 26736 31891				
CG18085	sev					
CG18247	shark					
CG18255	Strn-Mlck					
CG1830	PhKy	110638 33054 52478 110806 18374R-3 991 992	Wing phenotype/Lethal			
CG18374	Gyk					
CG18402	InR	25343 25344 101357 1388R-2 33133 1851R-2 109880 29379 31732	Wing phenotype	Wing phenotype		
CG1848	LIMK1					
CG18492	Tak1					
CG1851	Ady43A					
CG18582	mbt	46044 29379 31732				
CG18854						
CG1891	sax	42457 46358 47298 47301 44765 100276 33430 33431 108151 19275 110214	Wing phenotype	Wing phenotype		
CG1915	sls					
CG1939	Dpck					
CG1951						
CG1954	Pkc98E					
CG1973	yata					
CG2028	Ckla	13664 110768 25786	Lethal Wing phenotype/Lethal Lethal		110768	13664 110768
CG2048	dco	9241 2048R-1 2048R-3 27719	Wing phenotype	9241 2048R-1 2048R-3	Lethal	9241 2048R-1 2048R-3
CG2049	Pkn	108870 2055R-2 28335	Lethal			
CG2087	PEK	16427 110278	Wing phenotype			

CG2201		33502 108958					
CG2210	awd	110782 2210R-2	 Wing phenotype	110782 2210R-1 33712	 Wing phenotype Lethal	2210R-2	 Wing phenotype
CG2246		48878 110389					
CG2252	fs(1)h	51227 51305 108662 2252R-2	 Wing phenotype/Lethal Lethal Lethal Lethal			51305 * 108662	 Wing phenotype Wing phenotype
CG2272	slpr	33518 106449 2272R-1	 Wing phenotype	106449 2272R-1	 Wing phenotype	33518	 Wing phenotype 2272R-1
CG2577		41694 105471					
CG2615	ik2	49366 103748					
CG2621	sgg	101538 2621R-3 31308	 Wing phenotype Wing phenotype	101538 2621R-3 2621R-1	 Lethal Wing phenotype Wing phenotype	101538	 Wing phenotype 101538 2621R-3
CG2699	Pi3K21B	33556 13369 109682					
CG2794		20909 107766 100266					
CG2845	pfl	45041 110621					
CG2846							
CG2899	ksr						
CG2929	Pi4KIIα	40995 110687		40995 110687	 Wing phenotype	40995	 Wing phenotype 40995 110687
CG2964		42293 109509					
CG3001	Hex-A	21054 104680					
CG30021	metro	29965 110814					
CG3008		52634 103828					
CG30184		3539 104976					
CG30295	lpk1	47790 109497					
CG3051	SNF1A	1827 106200					
CG3068	aur	108446 35763					
CG3086	MAPk-Ak2	3170 110317					
CG31003	gskt	25640 107429					
CG3105	Pask	25661 107025					
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CG31183		859 4773					

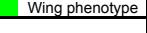
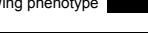
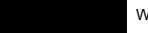
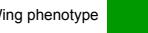
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CG31349	pyd	104159				
CG3140	Adk2	25046 107326 25760	Wing phenotype Wing phenotype			
CG31421	Takl1	110765				
CG31643		31643R-1				
CG3172	twf	31643R-2 25817 21415				
CG31751		110319				
CG31873		105621 36375				
CG32019	bt	46252 46253				
CG32031	Argk	34036				
CG32134	btl	34037 27106 110277				
CG3216		29915				
CG32417	Myt1	107985 34547				
CG32484	Sk2	105157 101018 41905				
CG32649		26534 26536				
CG32666	Drak	107263 32961 44374 29449	Lethal	44374 29449	107263 Wing phenotype	107263 Lethal
CG32703		44374 29449	Wing phenotype	44374 29449	44374 Wing phenotype	44374 Wing phenotype
CG32717	sdt	13444 109661 29844 100685				
CG32742	I(1)G0148	40716 104902				
CG32743	nonC	41990 108450				
CG3277		7271 108052				
CG32849	Hex-t2	47331 100218				
CG32944		29286 103642				
CG33102	Hex-t1	46573 46574				
CG33114	Gyc32E	108142				
CG3319	Cdk7	10442 103413	Wing phenotype Wing phenotype			
CG3324	Pkg21D	34594 103513				
CG33338	p38c	105173				
CG33519	Unc-89	29412 106267	Wing phenotype			
CG33531	Ddr	52126 101831				
CG33554	Nipped-A	44781 52487 52486 2905R-3	Wing phenotype Lethal Lethal Wing phenotype	44781 Lethal	52487 Wing phenotype	44781 Wing phenotype
CG33671		49772				

CG33981		49773					
CG3400	Pfrx	25958 25959					
CG34344	rdgA	102909 3024 5201		Wing phenotype			
CG34356		28745 109790		Wing phenotype			
CG34357		105185 31983					
CG34359	IP3K2	19159 102772	Wing phenotype	1630R-2	Wing phenotype	19159 1630R-2	Wing phenotype
CG34361	Dgk	38239 105753 29459	Wing phenotype/Lethal		102772		102772
CG34380		39447 39446					
CG34384		102481 43739					
CG34392	Epac	110077 50373	Wing phenotype				
CG34412	tlk	105732 20905 2829R-1 33983	Wing phenotype/Lethal Lethal Lethal Lethal		105732	Wing phenotype	105732 20905
CG3510	CycB	43772 109611 103784					
CG3525	eas	34286	Wing phenotype				
CG3534		109666 41276					
CG3544		21685 105459					
CG3608		106695 3608R-2					
CG3682	PIP5K59B	47027 108104					
CG3738	Cks30A	108401					
CG3809		108975 43780					
CG3837		44576 105549	Wing phenotype				
CG3915	Drl-2	40484 102192					
CG3938	CycE	52662 110204					
CG4006	Akt1	2902 103703	Wing phenotype	Wing phenotype			
CG4007	Nrk	103804 42442					
CG4012	gek	107207 28367					
CG40129	Gprk1	28354					
CG4026	IP3K1	31733					
CG40293	Stlk	27305 34620					
CG4032	Abl	2897 110186					
CG4041		108887 34780					

CG40478	Dyrk3						
CG4132	pkaap	21792 106808	Wing phenotype				
CG4141	Pi3K92E	38985 107390	Wing phenotype				
CG4201	ird5	26427 28628	Wing phenotype				
CG42273	mnb	107066					
CG42317	Csk	109813 32877	Lethal				
CG42320	Doa	20120 102520	Wing phenotype				
CG42341	Pka-R1	103720 26329					
CG42347		101640 44737					
CG42349	Pkcδ	33838 101421	Wing phenotype				
CG42366		108102 39904	Lethal				
CG42403	Ca-β	36112 27581					
CG4252	mei-41	105748 103624					
CG42636	Gyc76c	11251 106525					
CG4268	Pitslre	27094 107303 45127	Wing phenotype	107303 Lethal	107303 Lethal	107303 45127	Wing phenotype Lethal
CG42783	aPKC	34332 2907 105624	Wing phenotype Wing phenotype	2907 Lethal	105624 Wing phenotype	2907 105624	Lethal Wing phenotype
CG42856	Sik3	39866 107458					
CG4290	Sik2	103739 26497	Wing phenotype				
CG43143		16334 106088					
CG4353	hep	109277 47509					
CG43729		35848 35850					
CG43741	Ack-like	26065 100169					
CG4379	Pka-C1	3969R-2 31599	Wing phenotype	101524 Wing phenotype			
CG4488	wee	106329 26543	Wing phenotype				
CG4523	Pink1	109614 21860	Wing phenotype				
CG4527	slik	43783 43784	Wing phenotype				
CG4546		34869 34868	Wing phenotype				
CG4551	smi35A	40535 101376 4551R-1 4551R-3	 	40535 101376 4551R-1	 	101376 4551R-1	 
CG4583	Ire1	39561 39562					
CG4629		26574 26573					
CG4720	PkaR	110228					

CG4780	PK92D	34892 106934 26628 104701 34898 100999 26642 932 935 24683 40577 40576 5092R-1 5092R-2 34639 110702 27360 30946 104240 22024 107158	Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype/Lethal Wing phenotype			
CG5179	Cdk9	30449 103561	Lethal Wing phenotype		30449	Wing phenotype
CG5182	Pk34A	101146 27368 24440 103656 39402 110565				
CG5288						
CG5310	nmdyn-D6		Wing phenotype			
CG5363	cdc2	106130 41839	Lethal Wing phenotype		106130	Wing phenotype
CG5373	PI3K59F	100296 5373R-2 33384	Wing phenotype Wing phenotype			
CG5387	Cdk5α	34990 108656	Wing phenotype Wing phenotype			
CG5408	tbl	22113 106774				
CG5452	dnk	39137 103385	Wing phenotype			
CG5475	Mpk2	102484 52277				
CG5483	Lrrk	105630 22140				
CG5626		27469 5626R-1	Wing phenotype			
CG5680	bsk	104569 34138	Wing phenotype			
CG5725	fbl	44157 101437	Wing phenotype			
CG5757		110460 27504				
CG5790		110683 45045				
CG5940	CycA	32421 103595	Wing phenotype Wing phenotype/Lethal		32421	Wing phenotype
CG5974	pll	103774 2889				
CG6027	cdi	109409 43634				
CG6092	Dak1	44165 104861				

CG6114	sff	22224 100717 6117R-1				
CG6117	Pka-C3	6117R-2 27569				
CG6214	MRP	105419 6214R-3				
CG6292	CycT	37562 103387 6292R-3	Lethal Wing phenotype		103387 [redacted] Wing phenotype [green]	37562 [green] [black] 103387 [green] Wing phenotype
CG6297	JIL-1	107001 6297R-4				
CG6343	ND42	14444 110787	Wing phenotype Wing phenotype			
CG6355	fab1	27591 27592	Wing phenotype			
CG6364		11693 108949	Wing phenotype Wing phenotype		108494 [black] Wing phenotype [green]	11693 [red] [black] 108949 Wing phenotype
CG6386	ball	48980 108630 109282	Wing phenotype			
CG6498		35101				
CG6509		101596 46234	Wing phenotype Wing phenotype			
CG6518	inaC	101719 2895				
CG6521	Stam	22497 35016	Wing phenotype Wing phenotype			
CG6535	tefu	22502 108074				
CG6551	fu	27662 27663	Wing phenotype Wing phenotype			
CG6612	Adk3	110382 42064				
CG6620	ial	104051 35107 6620R-3	Wing phenotype Lethal Wing phenotype/Lethal		35107 [black] [red] Wing phenotype [black]	104051 [red] 35107 Wing phenotype
CG6622	Pkc53E	27696 27699				
CG6703	CASK	104793 34184				
CG6715	KP78a	26722 47657	Wing phenotype			
CG6767		35111 35112 109894 6767R-1	Wing phenotype Wing phenotype	35112	109894 [black] Wing phenotype [black]	35112 109894 [black] Lethal
CG6772	Slob	100987 30674				
CG6775	rg	107056 36404				
CG6800		40394 104255				
CG6875	asp	2911 110177	Wing phenotype			
CG6963	gish	106826 26003 28066	Wing phenotype Wing phenotype Wing phenotype	106826 26003 [green] Wing phenotype	106826 [black] [green] [green]	106826 26003 Wing phenotype
CG7001	Pk17E	101951 102179 27786	Wing phenotype			
CG7004	fwd	110159				
		27808	Wing phenotype		27808 [black] Wing phenotype [green]	27808 [red] Lethal

CG7028	PRP4	107042		Wing phenotype				107042		Wing phenotype/Lethal
CG7069		101116 27834								
CG7070	PyK	49533 35165		Wing phenotype						
CG7094		27843 108273								
CG7097	hppy	103580 35166								
CG7103	Pvf1	6175 102699		Wing phenotype						
CG7125	PKD	22344 106255		Wing phenotype						
CG7156		26035 26036								
CG7177	Wnk	35193 35194 106928	 	Wing phenotype	35193 35194 106928		Wing phenotype Lethal	35194	 	Wing phenotype Wing phenotype
CG7186	SAK	27904 105102								
CG7207	cert	103563 27914								
CG7223	htl	27180 6692								
CG7236		27505								
CG7281	CycC	27937 48834 110610		Wing phenotype						
CG7328		27952 7328R-1		Wing phenotype						
CG7335		29035 100062								
CG7362		104218 7557								
CG7393	p38b	108099 7393R-3								
CG7405	CycH	10398 104312		Wing phenotype						
CG7470		101476 38955								
CG7524	Src64B	35252 7524R-3								
CG7525	Tie	27087 26879								
CG7551		7551R-1 7551R-3								
CG7597	Cdk12	25508		Lethal	34838		Lethal	25508	 	Wing phenotype/Lethal
		25510		Wing phenotype				34838		Wing phenotype
								25510		Lethal
CG7616		41408 104663								
CG7643	ald	110572 7643R-3		Wing phenotype						
CG7693	fray	106919 41719								
CG7717	Mekk1	110339 25529								
CG7719	gwl	21046 7719R-1		Wing phenotype						
CG7766		52572 110184		Wing phenotype						
CG7838	BubR1	26109 7838R-1		Wing phenotype						

CG7873	Src42A	26019 100708 3002	Wing phenotype Wing phenotype			
CG7892	nmo	101545 25793				
CG7904	put	37279 107071 7904R-2 7904R-3	Lethal Lethal Lethal		37279 Wing phenotype	37279 Wing phenotypye
CG7995		101869 22652				
CG8049	Btk29A	106962 22675				
CG8057	alc	104489 8057R-4				
CG8094	Hex-C	35338 35337				
CG8173		35846 105661	Wing phenotype			
CG8174	SRPK	26933 103416	Wing phenotype			
CG8201	par-1	52553 52556 32410	Wing phenotype	52553 52556 32410 Lethal	52556 Wing phenotype	52556 32410 Wing phenotype
CG8203	Cdk5	104491 35856				
CG8222	Pvr	43459 105353				
CG8224	babo	3825 106092	Wing phenotype			
CG8239		24254 107034				
CG8250	Alk	107083 11446				
CG8286	P58IPK	109649 14154				
CG8298		43541 8298R-3				
CG8351	Tcp-1 η	43539 108585 28895	Wing phenotypye			
CG8362	nmdyn-D7	35903 105161				
CG8363	Papss	35904 110544				
CG8475		2801 110591	Wing phenotype			
CG8485		35939 35940	Wing phenotype	35940 Wing phenotype	35939 Wing phenotype	35939 35940 Wing phenotype
CG8565		100449 8565R-2				
CG8637	trc	35988 107923	Wing phenotype			
CG8657	Dgk ϵ	4659 8657R-2	Wing phenotype Wing phenotype			
CG8726		109451 40719				
CG8767	mos	110435 43526				
CG8789	wnd	103410 26910				
CG8808	Pdk	106641 37968				
CG8866		103725				

CG8874 Fps85D	44859 36053 107266					
CG8878	28971 100985	Wing phenotype Wing phenotype		100985	Wing phenotype	28971 100985 Wing phenotype Wing phenotype
CG8914 CK11β2	102633 26915					
CG8948 Graf	42165 42166					
CG8967 otk	42566 30834 104688 8967R-2	42566 30834 104688 8967R-2 Wing phenotype	104688	Wing phenotype	30834 104688	Wing phenotype
CG9096 CycD	29024 105361					
CG9222	104259 27010					
CG9326 vari	104548 24157	Wing phenotype				
CG9358 Phk-3	106938 49008 34362					
CG9374 lkb1	108356 49212	Wing phenotype				
CG9541	102912					
CG9738 Mkk4	108561 26929					
CG9746 ird1	9746R-1 110706	Wing phenotype Wing phenotype				
CG9774 rok	3793 104675	Wing phenotype Wing phenotype				
CG9961	101702 36176					
CG9962	108721 36473					
CG9985 sktl	6229 101624	Wing phenotype Wing phenotype				

Weak-Medium Decrease -0.2 to -0.5
 Medium-Strong Decrease > -0.5
 Weak-Medium Increase +0.2 to +0.5
 Medium-Strong Increase > +0.5
 No change

Transgenic RNAi Project strains are in blue font
 National Institute of Genetics strains have the letter R in the stock number
 All other strains are from Vienna Drosophila RNAi Center

Fig. S5

Phosphatase Data

			PRIMARY SCREEN			SECONDARY SCREEN			HEDGEHOG SCREEN		
Annotation Symbol	Gene Symbol	Gene Name	RNAi Stock	UAS-dcr2; dpp-Gal4	RNAi Stock	UAS-dcr2; hh-Gal4	RNAi Stock	UAS-dcr2; C5-Gal4	UAS-dcr2; dpp-Gal4 or UAS-dcr2; hh-Gal4	RNAi Stock	MS1096-Gal4; UAS-dcr2
				Wing disc	Adult		Wing disc	Adult	Wing disc	Wing disc	Wing disc
				Wg	Sens	Dll	Wg	Sens	Dll	Cut	Ci Ptc
CG10089			17991 108744								
CG10138	PpD5	Protein phosphatase D5	18016 104452	Wing phenotype							
CG10371	Plip	PTEN-like phosphatase	47624 104774	Wing phenotype Wing phenotype							
CG10376			101335 35475	Wing phenotype Wing phenotype							
CG10417			27259 106180	Wing phenotype							
CG10426			16048 34037								
CG10443	Lar	Leukocyte-antigen-related-like	107996 36270 36269								
CG10493	Phlpp	PH domain leucine-rich repeat protein phosphatase	110360 45365								
CG10574	I-2	Inhibitor-2	101547 39054	Wing phenotype Wing phenotype			101547			101547 39054	Wing phenotype Wing phenotype
CG10592			104767 38171 30822								
CG10827			100073								
CG10930	PpY-55A	Protein phosphatase Y-at 55A	102021 16096								
CG10975	Ptp69D	Protein tyrosine phosphatase 69D	104761 40631								
CG11217	CanB2	Calcineurin B2	104370 28762								
CG11425			103037 8458								
CG11426			42599 42600 9452	Wing phenotype							
CG11437											

CG11437						
CG11438			31721 11438R-3 11438R-2 42594 109898			
CG11440	laza	lazaro				
CG11516	Ptp99A	Protein tyrosine phosphatase 99A	103457 27208 38540 104729			
CG11597						
CG11883			106744 38590	Wing phenotype	106744 38590	Wing phenotype
CG12034			107062 12034R-1 101274 15288 100800 13987			
CG12078						
CG12091						
CG12151	Pdp	Pyruvate dehydrogenase phosphatase	31661 107271 38631 101257 31676 31674			
CG12169	Ppm1	Ppm1				
CG12173						
CG12217	PpV	Protein phosphatase V	101997 31690	Wing phenotype Wing phenotype	101997	Wing phenotype Wing phenotype
CG12252	Fcp1		38640 106253	Wing phenotype Wing phenotype	106253	Lethal Wing phenotype
CG1228	Ptpmeg	Ptpmeg	38652 103740 102895 31957			
CG12538						
CG12746			104989 33314 17123			
CG13125	TbCMF46	TbCMF47	36423 105122 51574 101655	Wing phenotype		
CG13197						
CG13311						
CG13570	spag	spaghetti	103353 23895			
CG1395	stg	string	17760 1395R-1 1395R-2 34831	Wing phenotype Lethal	17760 Wing phenotype	17760 Wing phenotype 34831 Wing phenotype
CG14022	Mitogen-activated		110777 31795 45415			

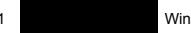
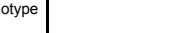
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CG14211	MKP-4	MAPK Phosphatase 4	104884 3146 3147 14211R-4	Wing phenotype Lethal Wing phenotype		104884	Wing phenotype
						3147	Wing phenotype
CG14297			102071 14297R-4				
CG14411			17579				
CG1455	CanA1	Calcineurin A1	109622 32283 32284	Wing phenotype			
CG14616	I(1)G0196	lethal (1) G0196 Alkaline	106973				
CG1462	Aph-4	phosphatase 4	6862 110043 105565 32314				
CG14719	I-t	inhibitor-t	49108 105752 100241 32353				
CG14903			51859 110458 7261				
CG15035			15400R-2 14865 105484 42520				
CG15385			102842 108181 30187				
CG15400			42686 42685	Wing phenotype			
CG15528			101368 33733	Wing phenotype			
CG15533			29199 105831				
CG15534			19658 105888 6386				
CG15743			16771R-2 16870R-2 16870R-1				
CG1637			30474 30476				
CG16717			104785 12941 32813				
CG16757	Spn	Spinophilin	49199 103270				
CG16771							
CG16870	Acyp	Acylphosphatase meiotic					
CG16928	mre11	recombinant 11					
CG1696	Dd	Dullard					
CG17026							
CG17027							

CG17027							
CG17028		50075 100100 32819 32823 49565 19078 17124R-3					
CG17029							
CG17124							
CG17291	Pp2A-29B	Protein phosphatase 2A at 29B	49672 [green] Lethal 23886 [green] Wing phenotype		49672 [black] [green] Wing phenotype [green]	49672 [red] Lethal 23886 [red] Wing phenotype	
CG17598			32956 32955 100178 40102 43231 1809R-1	Wing phenotype			
CG17746							
CG1809							
CG1810	mRNA-cap	mRNA-capping enzyme	108809 [green] Wing phenotype 32847 [green] Wing phenotype	32847 [green] Lethal	108809 [black] [green] Wing phenotype [green]	108809 [green] Wing phenotype/ 32847 [green] Lethal Wing phenotype	
CG1817	Ptp10D	Protein tyrosine phosphatase 10D	1102 8010				
CG18243	Ptp52F	Ptp52F	39175 3116				
CG1906	alph	alphabet	105483 32476				
CG2096	flw	flapwing	29622 104677 15305R-1	[green] Wing phenotype	29622 104677 15305R-1 [green] Wing phenotype	29622 104677 [green] Wing phenotype	
CG2104			30018 100216				
CG2890	PPP4R2r	Protein phosphatase 4 regulatory subunit 2-related protein	105399 [green] Wing phenotype 25445 [green] Wing phenotype			105399 [red] Wing phenotype 25445 [green] Wing phenotype/ Lethal	
CG2984	Pp2C1	Protein phosphatase 2C	105249 2984R-4	105249 [green] Wing phenotype 2984R-3 [green] Wing phenotype	105249 [black] [green] Wing phenotype	105249 [black] Wing phenotype 2984R-4 [black] Wing phenotype	
CG30103			105458 4950				
CG30104			10051 106640				
CG3028	Ipp	Inositol polyphosphate 1-phosphatase	110775 25615				
CG3059	NTPase	NTPase	110510 7265 13365				
CG31137	twin	twin					

CG31137	LWIII	LWIII	104442 45442 109759 102474 23397 108554 21396				
CG31299	cu	curled					
CG31469							
CG31692	fbp	fructose-1,6-bisphosphatase					
CG31717			7663 7662 105379 31717R-1	Lethal Wing phenotype/Lethal	105379 31717R-1	Wing phenotype	7663 7662 105379 31717R-1 31717R-3
CG31759			106772 47321				
CG3178	Rrp1	Recombination repair protein 1	108661 19819				
CG31795	IA-2	IA-2 ortholog	7560 110595				
CG32156	Mbs	Myosin binding subunit Protein	105762 32516	Wing phenotype Wing phenotype	32516	Wing phenotype	105762 32516
CG3245	PpN-58A	Protein phosphatase N at 58A	41901 102060 26154 2077R-1				
CG32487			46570 100129				
CG32488							
CG32505	Pp4-19C	Protein phosphatase 19C	25317 27726	Wing phenotype Lethal	38372	Lethal	25317 27726
CG32568			103317 46651 43250 108783				
CG3264							
CG32697	I(1)G0232	lethal (1) G0232	104427 21276		21276 3101R-1 33744	Lethal Lethal	104427 21276
CG32812			45397 104081				
CG3289	Ptpa	Phosphotyrosyl phosphatase activator	41913 41912 104586 52378 19989 3292R-2				
CG3290							
CG3292							
CG33747	primo-2	primo-2	23081 23079 39004	Wing phenotype Wing phenotype	23079 39004 50642	Lethal Lethal	23081 23079 39004
							Wing phenotype/ Lethal Lethal

CG33748	primo-1	primo-1					
CG3376			12226 12227				
CG34099	Mkp	MAP kinase-specific phosphatase	104374				
CG34140			23452 23454	Wing phenotype			
CG3530			110786 26217 26216	Wing phenotype	110786 26216	Wing phenotype	110786 26216
CG3573	Ocrl	Oculocerebro-renal syndrome of Lowe ortholog	34649				
CG3632			110796				
CG3705	aay	astray	110167 26254 23179 110661				
CG3954	csw	corkscrew	108352 21757	Wing phenotype Wing phenotype	21757	Wing phenotype	108352 21757
CG3980	Cep97		34774				
CG40448	Pp1-Y2	Pp1-Y2 Multiple inositol polyphosphate phosphatase 1	103357 109147 101634 8493 110123 21607				
CG4123	Mipp1		21611 52390 101200 47397 41672 100176	Wing phenotype			
CG41534	Pp1-Y1	Pp1-Y1					
CG4209	CanB	Calcineurin B					
CG42249							
CG42271							
CG42283	5Ptasel	5Ptasel	33768 33769 25030 100802	Wing phenotype Wing phenotype	33768 33769 100802	Lethal Lethal Wing phenotype/ Lethal	33768 100802
CG42327			106630 47455 14163 108018				
CG4317	Mipp2	Multiple inositol polyphosphate phosphatase 2					
CG4733	PR72	PR72	107621 34894		107621 34894	Wing phenotype	107621 34894
CG4827	veil	veil	49359 100050 46064 104147				
CG4965	twe	twine					

CG4993	PRL-1	PRL-2	107836 45518 105674 34916 102646 15298 108686 5171R-3 27367 103024 5177R-1 5177R-2 47995 106691 51984 51985 44319 106981 22163 5577R-1				
CG5026							
CG5150							
CG5171							
CG5177				Wing phenotype			
CG5276							
CG5361				Wing phenotype			
CG5567							
CG5577							
CG5643	wdb	widerborst	27470 101406 5643R-3	27470 101406 5643R-3	Wing phenotype/ Wing phenotype/ Lethal	101406	27470 101406 5643R-3
CG5650	Pp1-87B	Protein phosphatase 1 at 87B	35024 35025 32414		Wing phenotype/ Lethal Wing phenotype Wing phenotype		35024 35025 32414
CG5656			110733 18119				
CG5671	Pten	Pten	101475 35731	Wing phenotype			
CG5784	Mapmodulin	Mapmodulin	100283 49385				
CG5820	Gp150		36301 100134	Wing phenotype Wing phenotype		36301	36301 100134
CG5830			101539 5930R-2				
CG5851	sds22	sds22	42051 5851R-1	Lethal Wing phenotype/Lethal		42051	42051 5851R-1
CG6036			21023 105568				
CG6235	tws	twins	104167 34340	Wing phenotype Wing phenotype		104167	104167 6235R-2 34340
CG6238	ssh	slingshot	107998 30136 29950 100121	Wing phenotype			
CG6380							
CG6542	EDTP	Egg-derived tyrosine phosphatase	6542R-1 6542R-2	Wing phenotype			
			46070 34378	46070 34378			34378
							Wing phenotype

CG6562	synj	synaptojanin	6562R-1	 Wing phenotype/ Lethal	27489	6562R-1	 Wing phenotype	 6562R-1	 Wing phenotype
			6562R-2	 Wing phenotype/ Lethal					
CG6571	rdgC	retinal degeneration C	35105 6571R-2						
CG6593	Pp1α-96A	Protein phosphatase 1α at 96A	27673 105525	 Lethal  Lethal		27673	  Wing phenotype	27673 105525	 Lethal  Lethal
CG6656			104175 1633						
CG6746			46513 103625						
CG6805			107728 34615						
CG6896	MYPT-75D	MYPT-75D	109909 6896R-1 34688	  Wing phenotype   Wing phenotype		109909	 	109909 6896R-1	  Wing phenotype Wing phenotype
CG6899	Ptp4E	Protein tyrosine phosphatase 4E	1012 27232						
CG6939	Sbf	SET domain binding factor	22317 32419						
CG7067	NitFhit	Nitrilase and fragile histidine triad fusion protein	27831 108545						
CG7109	mts	microtubule star	41924 35171 35172	 Wing phenotype/ Lethal  Wing phenotype	41924 35171	 Wing phenotype	41924	  Lethal	41924 35171
CG7115			9404 103354 39065	 Wing phenotype	9404 103354 39065	 Wing phenotype	9404	 	9404 103354 39065
CG7134	cdc14	cdc14	103627 7134R-6						
CG7180			102397 34369						
CG7378			106098						
CG7615	fig	fos intronic gene	35226 47314 47312						
CG7789			3019 3018						
CG7850	puc	puckered	110470 3579						
CG7899	AcpH-1	Acid phosphatase 1	22614 107057 110582						
CG7913	Pp2A-B'	Pp2A-B'							
CG7942	Idhr	lariat debranching							

		9311R-4		Lethal				
CG9351	ffl	falafel	24143 103793	Wing phenotype Wing phenotype	103793	Wing phenotype	24143	Wing phenotype
CG9389			44663					
CG9391			23723					
CG9449			23725					
CG9451			12895					
CG9452			102991					
			109721					
			14344					
			100887					
			51202					
CG9493	Pez	Pez	108301 9493R-3	Wing phenotype	108301		108301	
			40743	Wing phenotype	9493R-2 40743		9493R-3 40743	
CG9554	eya	eyes absent	43911 108071 28733	Wing phenotype	40743	Lethal	43911	
CG9601			24070					
CG9619	Gbs-76A	Glycogen binding subunit 76A	108251					
CG9764	yrt	yurt	103044					
CG9784			36121					
CG9801			28674					
CG9819	CanA-14F	Calcineurin A at 14F	107016					
CG9842	Pp2B-14D	Protein phosphatase 2B at 14D	108075					
CG9856	PTP-ER	Protein tyrosine phosphatase-ERK/Enhancer of Ras1	30098 104525 109858 30105 46873 103144 9856R-1 9856R-2 32359	Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype Wing phenotype				

Weak-Medium Decrease -0.2 to -0.5
 Medium-Strong Decrease > -0.5
 Weak-Medium Increase +0.2 to +0.5
 Medium-Strong Increase > +0.5
 No change

Transgenic RNAi Project strains are in blue font
 National Institute of Genetics strains have the letter R in the stock number
 All other strains are from Vienna Drosophila RNAi Center

Fig. S6

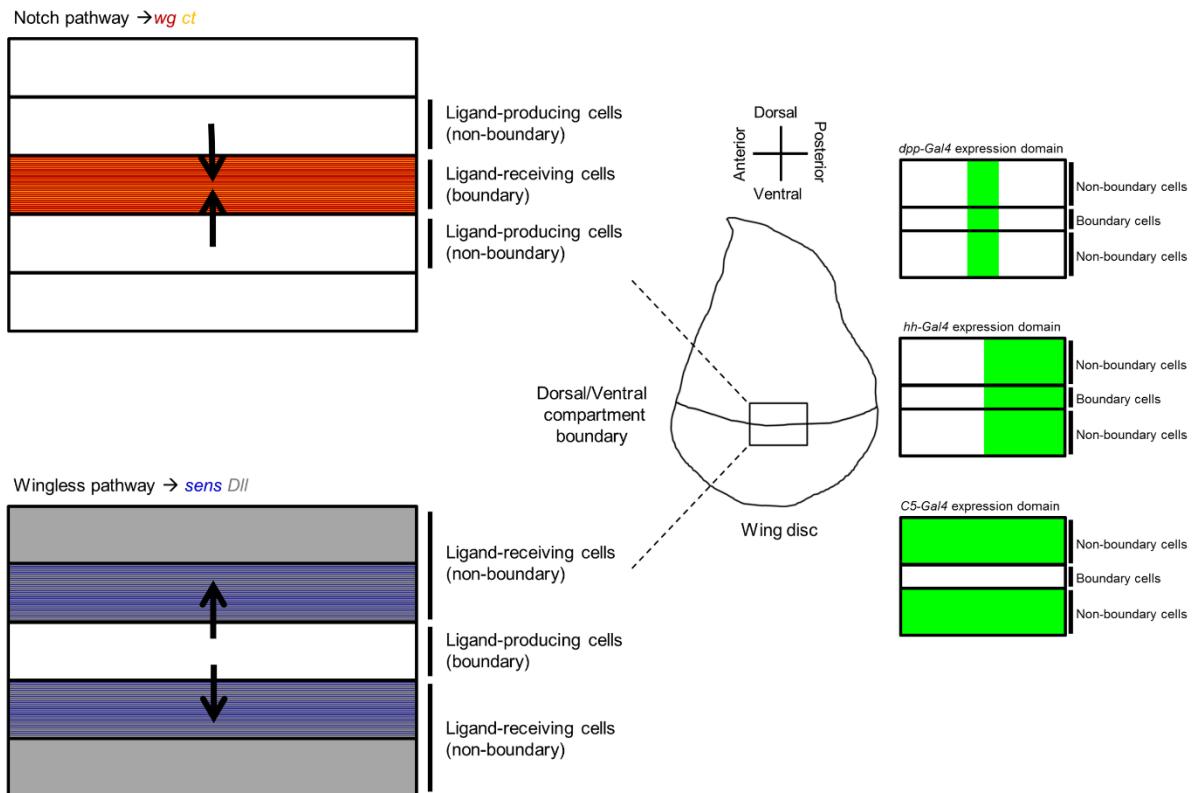


Fig. S7

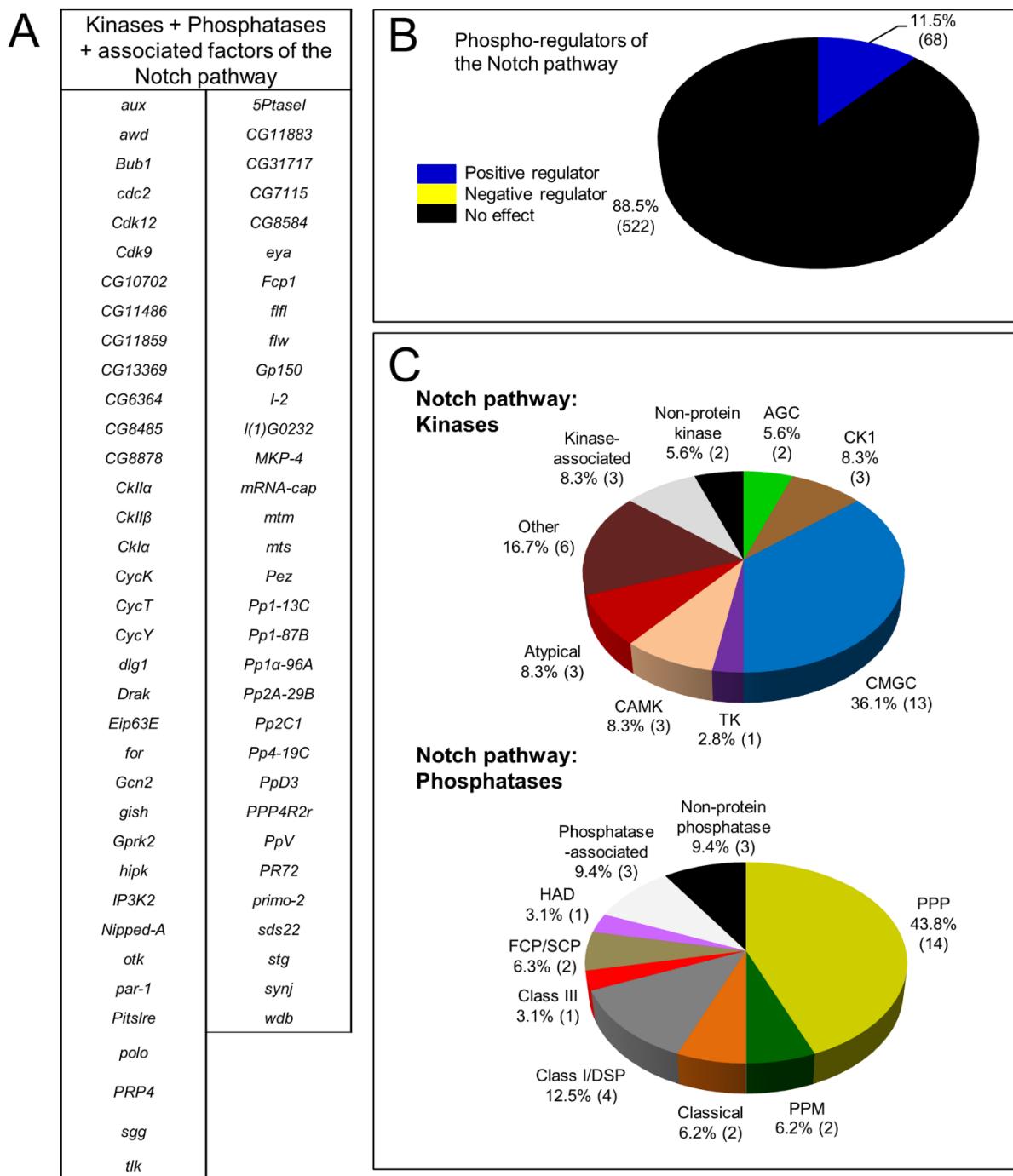


Fig. S8

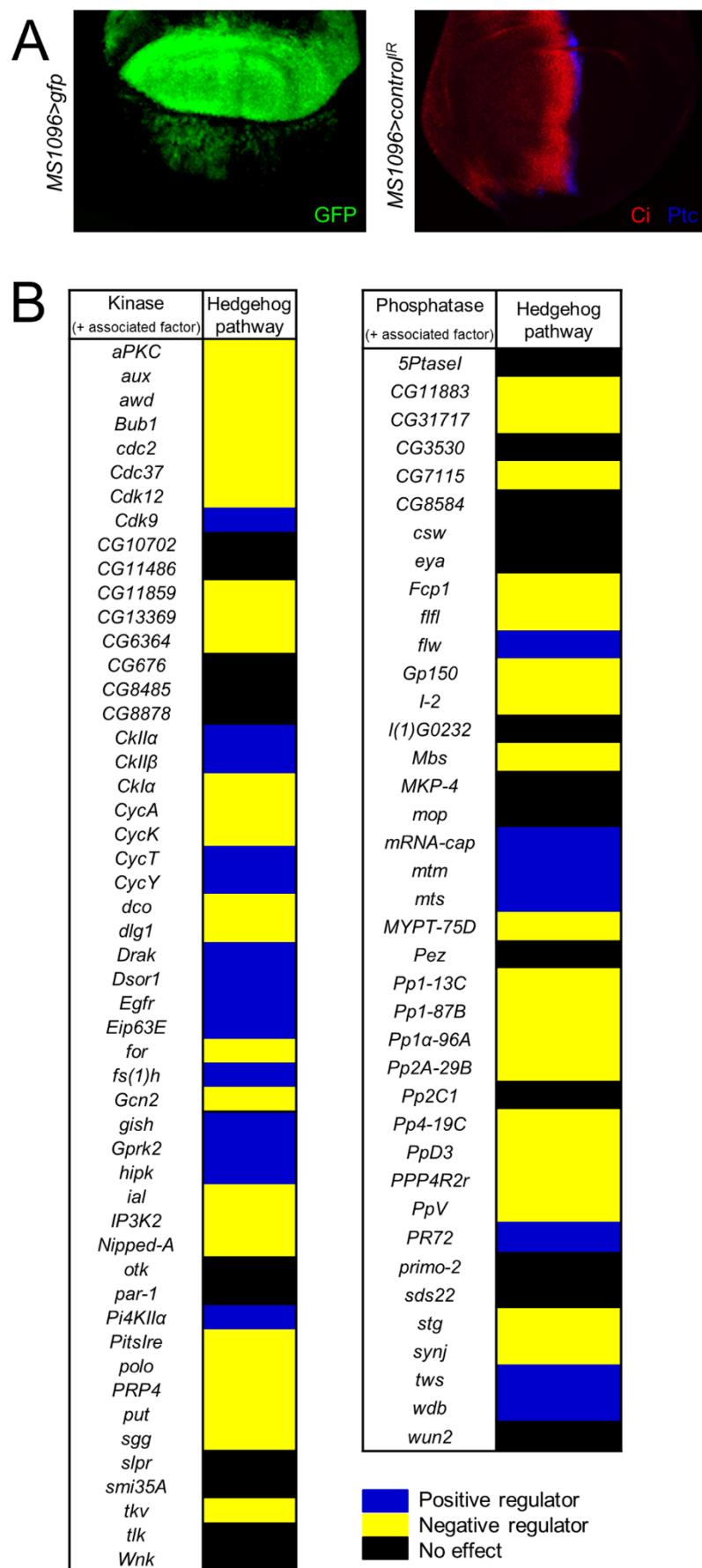


Fig. S9

