

# Tissue-specific function of Patj in regulating the Crumbs complex and epithelial polarity

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## SUMMARY

Patj is described as a core component of the Crumbs complex. Along with the other components, Crumbs and Stardust, Patj has been proposed as essential for epithelial polarity. However, no proper *in vivo* genetic analysis of Patj function has been performed in any organism. We have generated the first null mutants for *Drosophila Patj*. These mutants are lethal. However, Patj is not required in all epithelia where the Crumbs complex is essential. Patj is dispensable for ectoderm polarity and embryonic development, whereas more severe defects are observed in the adult follicular epithelium, including mislocalisation of the Crumbs complex from the apical domain, as well as morphogenetic defects. These defects are similar to those observed with *crumbs* and *stardust* mutants, although weaker and less frequent. Also, gain-of-function of Crumbs and *Patj* mutation genetically suppress each other in follicular cells. We also show that the first PDZ domain of Patj associated with the Stardust-binding domain are sufficient to fully rescue both *Drosophila* viability and Crumbs localisation. We propose that the only crucial function of Patj hinges on the ability of its first two domains to positively regulate the Crumbs complex, defining a new developmental level of regulation of its dynamics.

**KEY WORDS:** Crumbs, Epithelium, Follicular cells, Polarity

## INTRODUCTION

Epithelial polarity relies on the dynamic interplay of several protein complexes that define the different cortical domains (Laprise and Tepass, 2011; St Johnston and Ahringer, 2010). Among these, the Crumbs (Crb) complex plays key roles in the definition of the apical domain and in the organisation and positioning of the first apical junction, which are adherens junctions in *Drosophila* and tight junctions in mammals (Tepass et al., 1990; Wodarz et al., 1995; Bulgakova and Knust, 2009; Morais-de-Sá et al., 2010; Bazellieres et al., 2009; Hurd et al., 2003). At the molecular level, essential components of the Crb complex are Crb, Stardust (Sdt; PALS1) and Patj. Crb is a transmembrane protein; its cell polarity function depends on a C-terminal PDZ binding motif (ERLI) that binds Sdt (Klebes and Knust, 2000; Bachmann et al., 2001; Hong et al., 2001; Roh et al., 2002). Patj has been isolated as a component of the Crb complex via direct interaction with Sdt, involving heterodimerisation of their L27 domains (Roh et al., 2002; Bulgakova et al., 2008). To mediate its function in polarity, the Crb complex also interacts in multiple ways with other polarity determinants, including the PAR6/aPKC complex (Hurd et al., 2003; Kempkens et al., 2006; Lemmers et al., 2004; Krahn et al., 2010). These interactions together reflect the intricate dynamics of these complexes, which have yet to be fully elucidated.

There are genetic data clearly showing the importance of Crb and Sdt, but less is known about Patj. Reducing PATJ protein levels in mammal cell culture leads to destabilisation of the Crb complex and to defects in tight junction assembly (Lemmers et al., 2002; Michel et al., 2005; Shin et al., 2005). However, this phenotype is mild compared with those of *Crb* or *Sdt* mutants, and one could

assume that the difference reflects hypomorphic conditions. Genetic data in fruit flies have failed to produce any clear picture of *Patj* function, although the gene appears to be essential for viability (Bhat et al., 1999; Nam and Choi, 2006; Pielage et al., 2003; Richard et al., 2006).

Here we present the first null mutants for the *Drosophila Patj* gene. This analysis reveals a tissue-specific requirement for the stability of the Crb complex at the apical membrane.

## MATERIALS AND METHODS

### *Drosophila* genetics

*Patj* mutants were generated by imprecise excision of *P{GSV2}/GS50262*. Excisions that were lethal when crossed with the *Df(3L)My10* deficiency were analysed at the molecular level. Only deletions exclusively affecting *Patj* were retained (*Patj*<sup>53</sup>, *Patj*<sup>58</sup>, *Patj*<sup>63</sup>) and recombined with *FRT2A*. Rescue by a *Patj* transgene was assessed for all alleles.

Clonal analyses were performed using the flp/FRT or MARCM systems with GFP as marker for wild-type or mutant cells, respectively. Flies were dissected 4 days after clone induction by heat shock. Mutant stocks are *FRT2A*, *Patj*, *FRT19A*, *sdt*<sup>K85</sup> (Berger et al., 2007) and *FRT82B*, *crb*<sup>1</sup> (Tepass et al., 1990). Overexpression was performed using Tub:Gal4 flip-out or MARCM systems. UAS transgenes are *UAS:Patj* (Bhat et al., 1999) and *UAS:crb*<sup>WT-30.12e</sup> (Wodarz et al., 1995). For embryo analysis, maternal contribution was removed using the flp/FRT *Ovo*<sup>D1</sup> system and zygotic mutants were selected by the absence of *TM3*, *twi:Gal4*, *UAS:eGFP*.

### Transgenes

The *Patj* genomic rescue transgene was produced by cloning a genomic PCR fragment into the pCASPER4 vector. *Patj* transgenes for structure/function analysis, including a full-length transgene, were produced by PCR on *Patj* cDNA and cloning with and without GFP for C-terminal fusions in pWRpAUBiqP, which contains a *Ubiquitin-63E* promoter. Transgenic flies were generated by Fly Facility.

### Immunostaining

Immunofluorescent staining was performed according to standard protocols using the following primary antibodies: rabbit anti-PKCz (1/500; C-20, Santa Cruz), mouse anti-Dlg (1/50; Developmental Studies Hybridoma Bank, University of Iowa), rabbit anti-Baz (1/1000) (Wodarz et al., 1999),

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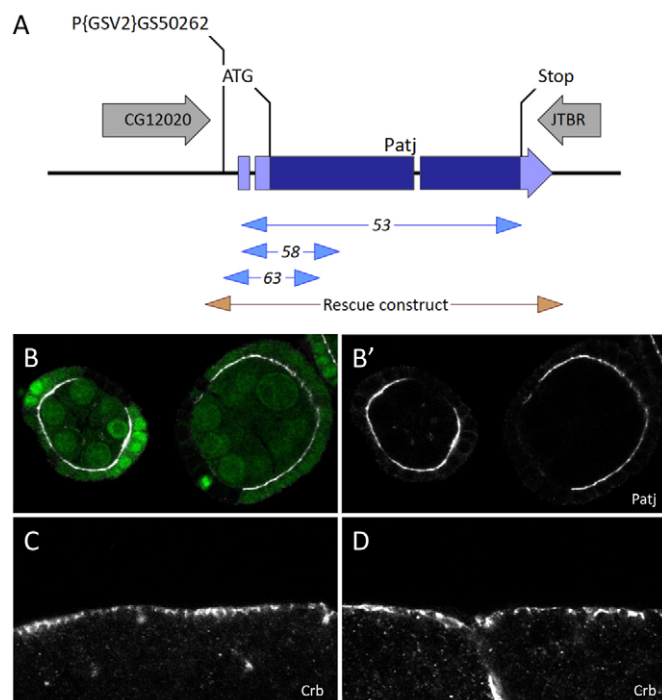
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rat anti-Crb (1/500) (Pellikka et al., 2002), and rabbit anti-Patj (1/500) (Tanentzapf et al., 2000). Images were acquired on a Leica SP5 microscope.

## RESULTS AND DISCUSSION

### *Patj* null mutations are lethal

No proper mutation of the *Patj* gene was available, limiting its genetic manipulation. In order to generate *Patj* mutants, 100 excisions of a *P*-element-based transgene inserted in the 5' region of *Patj* were produced (Fig. 1A). We identified three internal deletions of *Patj* associated with lethality. *Patj*<sup>58</sup> and *Patj*<sup>63</sup> delete start codons, whereas *Patj*<sup>53</sup> shows complete deletion of the coding sequence. A rescue construct containing the *Patj* gene and 300 bp upstream of its transcription start site perfectly rescued the lethality and fertility of *Patj* mutants. We therefore assumed these alleles to be null mutations for *Patj*, and we confirmed the absence of Patj protein in mutant follicle cells as compared with the surrounding wild-type cells (Fig. 1B). Our results confirm previous findings that *Patj* is an essential gene (Nam and Choi, 2006; Richard et al., 2006). However, despite previous reports that *Patj* mutants die during the second instar larval stage, we found that *Patj* homozygotes or transheterozygotes could reach the pupal stage, highlighting the utility of bona fide null mutants. We also produced maternal *Patj* germline clones to test whether maternal contribution could mask embryonic phenotype. However, maternal and zygotic null mutants were able to reach the pupal stage, like the zygotic mutants, indicating that *Patj* is dispensable for embryonic development. In addition, we failed to detect polarity defects in the embryonic ectoderm upon examining the localisation of Crb and other polarity markers (Fig. 1C,D; data not shown).



**Fig. 1. Null mutants for *Patj*.** (A) The *Drosophila Patj* locus. The site of the P(GSV2)GS50262 insertion, deletion mutations and the rescue transgene are indicated. (B, B') Mutant clones for *Patj*<sup>53</sup> in follicular cells stained for Patj (white). Mutant cells are marked by the absence of GFP (green). (C, D) Normal apical localisation of Crb in zygotic (C) or maternal zygotic (D) embryos mutant for *Patj*.

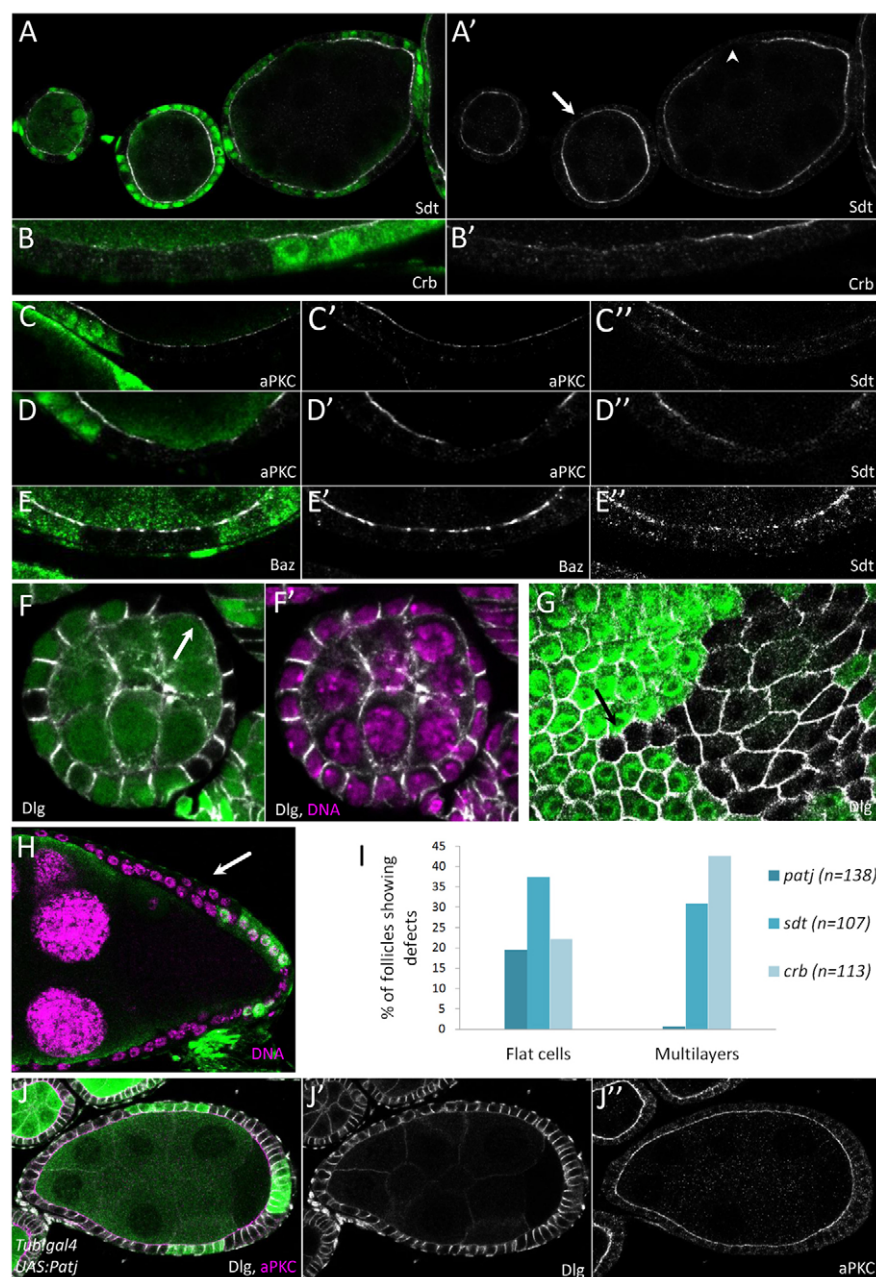
### *Patj* regulates Crb complex stability

We therefore analysed *Patj* function in another tissue; we looked at the follicular epithelium that surrounds the germline cysts of the ovary. We first investigated the localisation of the other Crb complex members Crb and Sdt. Both showed partial mislocalisation from the apical domain in *Patj* mutant as compared with wild-type cells. This phenotype is usually weak during early stages (1-5), but tends to gain strength later, leading in some cases to a complete apical loss of both Sdt and Crb, thus indicating that *Patj* is required for Crb complex stability at the apical domain (Fig. 2A,B). Absence of Crb and Sdt from the apical domain is not associated with a strong cytoplasmic accumulation of these proteins, suggesting that they might be degraded. Because Crb and Sdt are important for defining the apical domain of epithelial cells, we also looked at the localisation of other polarity markers. aPKC is also less apically enriched in *Patj* mutant cells than in wild-type cells, although the effect on this key polarity determinant is weaker than that observed for Sdt and Crb (Fig. 2C,D). Baz protein is also globally absent from the apical domain, but remains in subapical spots that might correspond to adherens junctions (Fig. 2E) (Moraes-de-Sá et al., 2010). Also, we did not observe any lateral markers, such as Discs large (Dlg), extending apically in *Patj* mutant cells, which is usually the case when the apical domain is no longer specified (Fig. 2F).

In agreement with these results, we found extremely rare cases (<1%) in which a loss of polarity of *Patj* mutant cells was indicated by the formation of multilayers or round cells (Fig. 2H). However, we observed a cell flattening in *Patj* mutant clones compared with wild-type cells in ~20% of the follicle-containing clones (Fig. 2F,G). This flattening always affected cells on the lateral side of the follicles. We compared the *Patj* phenotype with those of *crb* and *sdt* mutants (Fig. 2I; supplementary material Figs S1, S2). As previously described, *crb* mutation frequently induced multilayers (42%) and cell flattening (22%) (Tanentzapf et al., 2000). *sdt* mutation also induced the same defects, with multilayers (31%) and some flat cells in the epithelium (37%). Moreover, *crb* and *sdt* mutant cells showed a greater loss of apical markers than *Patj* cells (supplementary material Figs S1, S2). Taken together, our data suggest that the flattening is a mild phenotype, whereas multilayer formation corresponds to a stronger polarity phenotype. Our results therefore suggest that the loss of any one of the three core components of the Crb complex leads to similar phenotypic traits, but with the severity of defects decreasing in the order *crb*, *sdt*, *Patj*. The importance of Crb as a polarity determinant is also apparent through the extension of the apical domain and disruption of epithelial architecture when Crb is overexpressed (Tanentzapf et al., 2000; Wodarz et al., 1995). Mirroring the loss-of-function experiments, Sdt overexpression in follicle cells leads to less pronounced defects than Crb overexpression (Horne-Badovinac and Bilder, 2008). Similar clonal overexpression of *Patj* did not induce any polarity defect in follicle cells (Fig. 2J; data not shown), confirming that *Patj* plays a more subtle role in epithelial polarity than Crb and Sdt.

### The L27 and first PDZ domain are sufficient for *Patj* function

It has been shown that the L27 domain and the first of the four PDZ domains of *Patj* are able to rescue lethality associated with *Patj* deletion (Nam and Choi, 2006). We produced several *Patj* transgenes to provide a more accurate structure-function analysis. A full-length *Patj* transgene compared with a transgene encoding these two domains exhibited no differences; both were able to fully



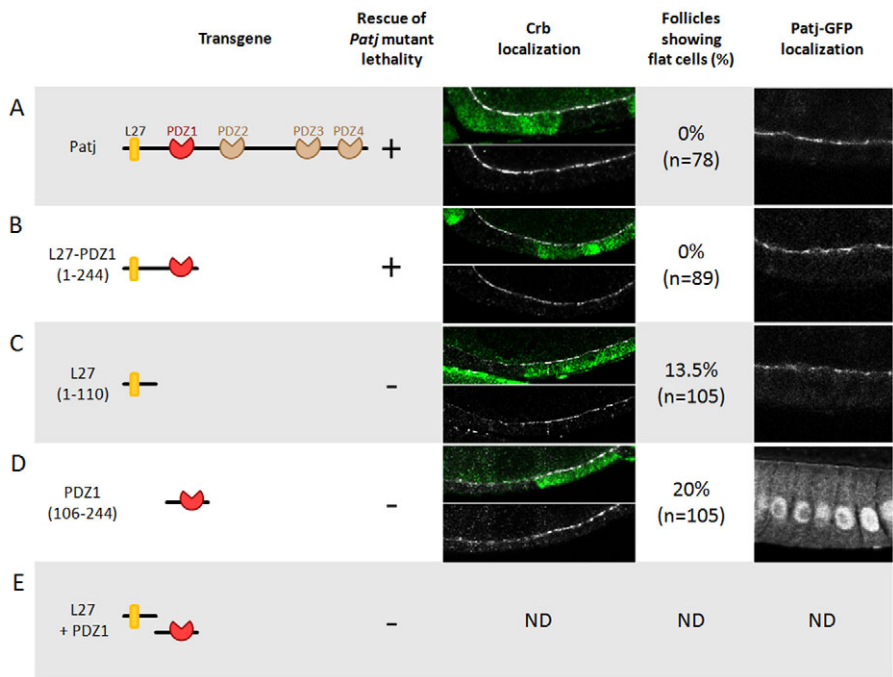
**Fig. 2. *Patj* mutant phenotype in the follicular epithelium.** (A-H) *Patj* null mutation clones viewed in transverse (A-F,H) or planar (G) section stained (white) for Sdt (A,A',C'',D'',E''), Crb (B,B'), aPKC (C,C',D,D'), Baz (E,E'), Dlg (F-G) and for DNA (purple, F',H). Mutant cells are marked by the absence of GFP (green). (A,A') Sdt loss from the apical domain can be partial (arrows) or complete (arrowhead). (B-E'') Reduction in the apical domain of Crb, aPKC and Baz is also observed. (F-G) *Patj* mutant clones showing a cell flattening defect. This defect is not observed in all the cells, which is in part owing to the genotype of the surrounding cells (arrow in G). (H) *Patj* mutant clone forming a multilayer (arrow). (I) Quantification of *Patj*, *sdt* and *crb* phenotypes in follicular cells. Percentages of follicles containing mutant clones and showing multilayers or cell flattening. (J-J'') Clonal overexpression of *Patj* (green cells) does not induce any polarity defect in follicular cells as visualised with Dlg (J', white in J) and aPKC (J'', purple in J).

rescue *Patj* mutant-associated lethality (Fig. 3A,B). In addition, we investigated Crb localisation in follicle cells mutant for *Patj* in flies carrying these transgenes. The presence of full-length *Patj* or L27-PDZ1 proteins fully rescued the Crb complex localisation and cell flattening phenotype (Fig. 3A,B). By contrast, transgenes encoding only L27 or only PDZ1 failed to rescue lethality, Crb localisation or cell flattening in follicle cells mutant for endogenous *Patj* (Fig. 3C,D). A similar inability to rescue the lethality was observed in flies expressing L27 and PDZ1 as encoded by two distinct transgenes (Fig. 3E). Taken together, these results show that all the essential functions of *Patj* are concentrated in these two domains and that they have to be linked together. Also, the ability of *Patj* transgenes to rescue lethality is correlated with their ability to restore apical localisation of the Crb complex, suggesting that this constitutes the main function of *Patj*.

Analysis of the subcellular localisation of the different *Patj* domains fused to GFP indicates that the PDZ1 domain localises

on its own to the apical domain, although less efficiently than proteins containing the L27 domain, suggesting that PDZ1 can bind an apical protein (Fig. 3A-D). Whereas L27 domain binding to Sdt is well established, what binds to the PDZ1 domain is far less clear. No partner has been identified in flies, and only a few proteins are proposed to link to the mammalian PATJ PDZ2 domain, which corresponds to fly PDZ1. One of these is TSC2 (Massey-Harroche et al., 2007), but we failed to find similar interaction between the *Drosophila* proteins. Mammalian PATJ PDZ2 also interacts with angiomin (AMOT), and this interaction seems robust because it is conserved with the other members of the AMOT protein family and the mammalian paralogue of PATJ called MUPP1 (MPDZ) (Sugihara-Mizuno et al., 2007; Wells et al., 2006). However, the *Drosophila* genome does not contain any member of the AMOT family. These data suggest the existence of an as yet unidentified apical ligand of PDZ1 that is essential for *Patj* function.



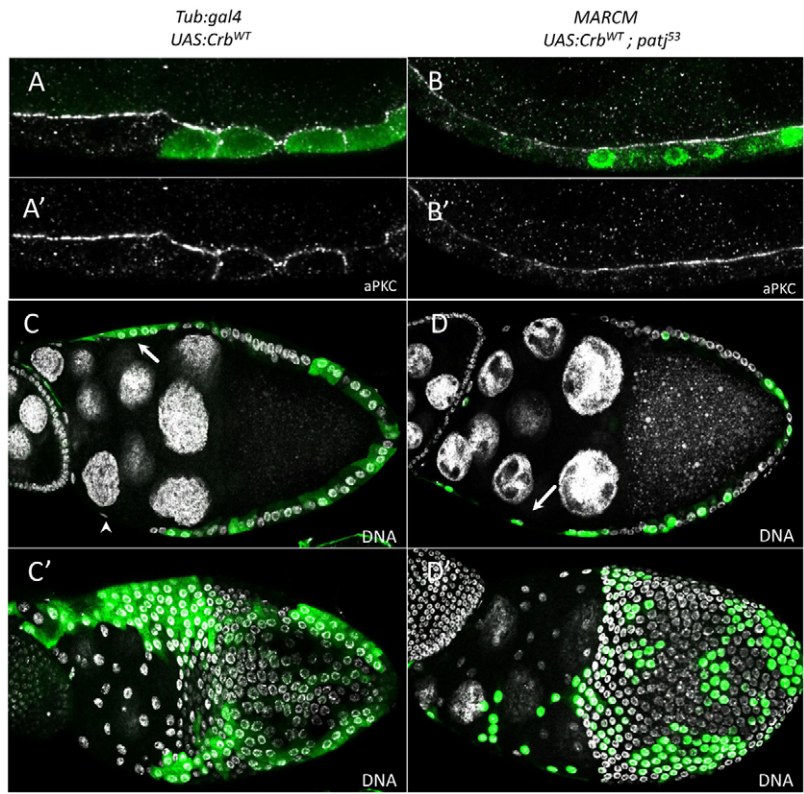


**Fig. 3. Structure-function analysis of *Patj*.** (A) Full-length *Patj*, (B) L27 + PDZ1 domains, (C) L27 domain, (D) PDZ1 domain, (E) L27 and PDZ1 domains encoded by two independent transgenes. The ability of these transgenes to rescue *Patj* mutant lethality is indicated. *Patj* mutant clones in the follicular epithelium are marked by the absence of GFP and stained for Crb (white) in flies containing the different transgenes. Quantification of the cell flattening phenotype is shown. The L27 and PDZ1 domains are necessary and sufficient for rescue of Crb localisation and lethality. The final column illustrates the subcellular localisation in the follicular cells of the different *Patj* proteins as visualised by GFP. ND, not determined.

Genetic interaction between *Patj* and Crb

All the defects caused by *Patj* mutation are therefore also present in other mutants of the Crb complex, and at least one of the two key domains of *Patj* interacts directly with Sdt. Taken together, these results suggest that the main function of *Patj* is to regulate the Crb complex. We took advantage of the moderate effect of *Patj* mutation on follicle cell polarity to look for genetic interaction with a Crb gain-of-function. We analysed the phenotype of *Patj*

MARCM mutant clones overexpressing Crb, and compared their effects with both *Patj* mutant and Crb overexpression on their own. As previously described, Crb overexpression disrupts follicular cell polarity, as seen by extension of aPKC localisation to the lateral domain and from the spheroidal shape of the cells, and can lead to multilayer formation (Fig. 4A; data not shown) (Tanentzapf et al., 2000). Crb overexpression also blocks the cuboid-to-squamous transition of a subpopulation of follicular cells, termed stretched



**Fig. 4. Genetic interaction between *Patj* and *crb*.** Clonal overexpression (green, positive cells) of Crb in wild-type (A,C) or *Patj* mutant (B,D) cells. (A,A') Crb overexpression induces a loss of cell polarity that can be visualised by an extension of aPKC cortical staining and by the spheroidal shape of the cells. (B,B') This phenotype is suppressed when the cells are mutant for *Patj*. (C,C') Transverse section (C) and top view (C'). Crb overexpression blocks cell flattening of anterior cells at stage 9 of oogenesis (arrow compared with arrowhead). (D,D') Transverse section (D) and top view (D'). This defect is suppressed when the cells are mutant for *Patj*. Also, Crb overexpression rescues the various *Patj* mutant defects observed in follicular cells (see Fig. 2).

cells, at stage 9 of oogenesis (Fig. 4C) (Grammont, 2007). *Patj* MARCM mutant clones overexpressing Crb exhibit normal morphology, and aPKC no longer extends to the lateral domain of the cells, even though it is slightly more enriched on the apical domain (Fig. 4B). Thus, we never observed a reduction in aPKC at the apical domain, in contrast to *Patj* mutant cells. In addition, Crb overexpression also fully rescues the cell flattening that results from the loss of Patj, whereas the absence of Patj restores the normal stretching of anterior cells at stage 9 (Fig. 4D). Thus, Crb overexpression and null mutation for *Patj* suppress each other for all the defects that we observed in follicle cells. These genetic data lead to the conclusion that the essential function of Patj is to positively regulate the Crb complex.

The *Patj* mutant phenotype suggests that it might be important for delivering or stabilising Crb at the apical membrane or for promoting its recycling from endosomes. PATJ knockdown in human cells leads to CRB mislocalisation from the apical cortex and its accumulation in early endosomes (Michel et al., 2005). Interestingly, Rab11 and the retromer are both important for Crb recycling in *Drosophila* (Pocha et al., 2011; Roeth et al., 2009; Zhou et al., 2011). However, they exercise this function in the embryo, where Patj is largely dispensable, indicating that Patj is not essential for Crb recycling.

Patj requirement appears stronger in some epithelia than in others, with no impact on embryonic ectoderm polarity but stronger defects in the follicular epithelium. Structure-function analyses of Sdt and Crb lead to similar conclusions, with the requirements for Sdt or Crb domains seeming to differ from one epithelial tissue to another (Wodarz et al., 1995; Bit-Avragim et al., 2008; Bulgakova and Knust, 2009; Fletcher et al., 2012). The reason for such differences remains to be elucidated. The Crb complex is also involved in the control of cell proliferation and cell morphogenetic processes other than the establishment or maintenance of epithelial polarity (Bulgakova and Knust, 2009; Grusche et al., 2010; Kerman et al., 2008; Laprise et al., 2010; Letizia et al., 2011; Xu et al., 2008). Modulation of its activity by Patj might participate in these developmental functions of the Crb complex.

### Note added in proof

During the editorial process, another article was published describing *Drosophila Patj* mutants and their phenotypic analysis (Zhou and Hong, 2012).

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### Competing interests statement

The authors declare no competing financial interests.

### Supplementary material

Supplementary material available online at <http://dev.biologists.org/lookup/suppl/doi:10.1242/dev.085449/-/DC1>

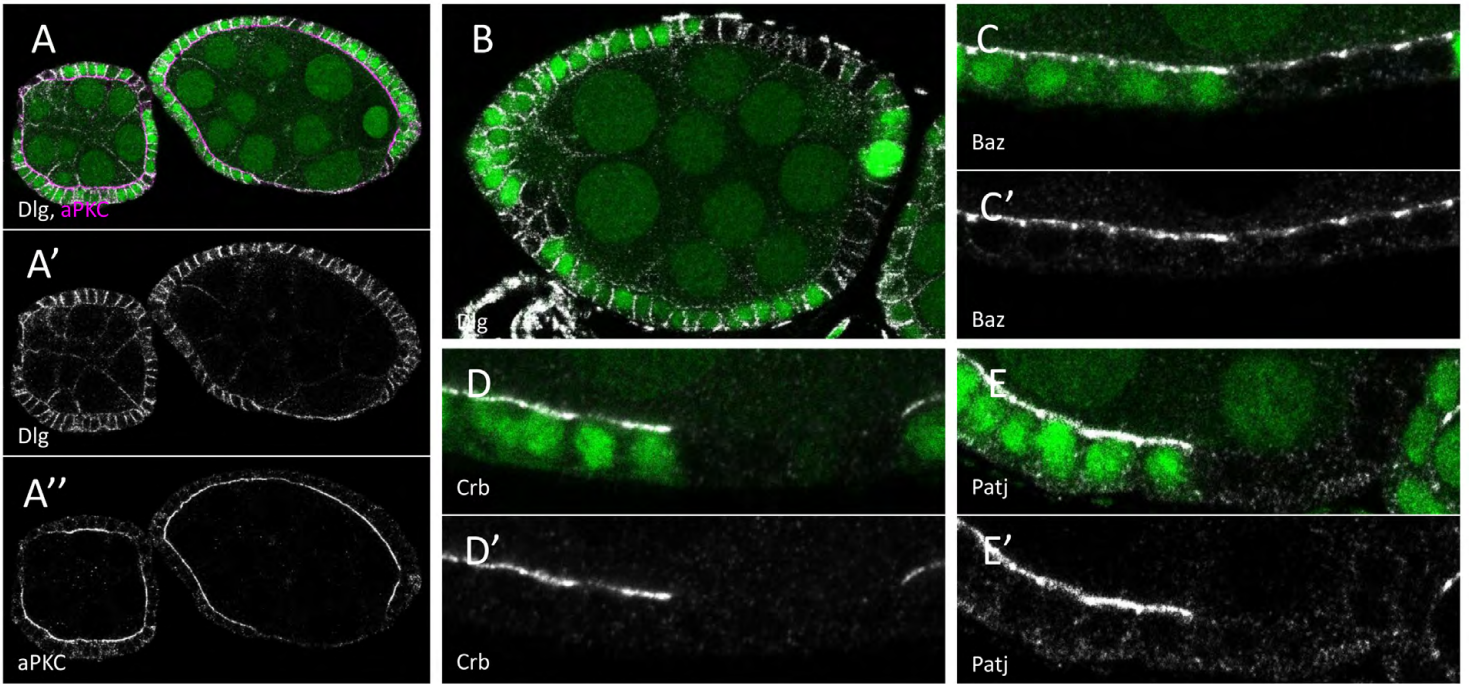
### References

- Bachmann, A., Schneider, M., Theilenberg, E., Grawe, F. and Knust, E. (2001). *Drosophila* Stardust is a partner of Crumbs in the control of epithelial cell polarity. *Nature* **414**, 638–643.
- Bazellieres, E., Assemat, E., Arsanto, J. P., Le Bivic, A. and Massey-Harroche, D. (2009). Crumbs proteins in epithelial morphogenesis. *Front. Biosci.* **14**, 2149–2169.
- Berger, S., Bulgakova, N. A., Grawe, F., Johnson, K. and Knust, E. (2007). Unraveling the genetic complexity of *Drosophila* stardust during photoreceptor morphogenesis and prevention of light-induced degeneration. *Genetics* **176**, 2189–2200.
- Bhat, M. A., Izaddoost, S., Lu, Y., Cho, K. O., Choi, K. W. and Bellen, H. J. (1999). Discs Lost, a novel multi-PDZ domain protein, establishes and maintains epithelial polarity. *Cell* **96**, 833–845.
- Bit-Avragim, N., Hellwig, N., Rudolph, F., Munson, C., Stainier, D. Y. and Abdellah-Seyfried, S. (2008). Divergent polarization mechanisms during vertebrate epithelial development mediated by the Crumbs complex protein Nagie oko. *J. Cell Sci.* **121**, 2503–2510.
- Bulgakova, N. A. and Knust, E. (2009). The Crumbs complex: from epithelial-cell polarity to retinal degeneration. *J. Cell Sci.* **122**, 2587–2596.
- Bulgakova, N. A., Kempkens, O. and Knust, E. (2008). Multiple domains of Stardust differentially mediate localisation of the Crumbs-Stardust complex during photoreceptor development in *Drosophila*. *J. Cell Sci.* **121**, 2018–2026.
- Fletcher, G. C., Lucas, E. P., Brain, R., Tournier, A. and Thompson, B. J. (2012). Positive feedback and mutual antagonism combine to polarize crumbs in the *Drosophila* follicle cell epithelium. *Curr. Biol.* **22**, 1116–1122.
- Grammont, M. (2007). Adherens junction remodeling by the Notch pathway in *Drosophila* melanogaster oogenesis. *J. Cell Biol.* **177**, 139–150.
- Grusche, F. A., Richardson, H. E. and Harvey, K. F. (2010). Upstream regulation of the hippo size control pathway. *Curr. Biol.* **20**, R574–R582.
- Hong, Y., Stronach, B., Perrimon, N., Jan, L. Y. and Jan, Y. N. (2001). *Drosophila* Stardust interacts with Crumbs to control polarity of epithelia but not neuroblasts. *Nature* **414**, 634–638.
- Horne-Badovinac, S. and Bilder, D. (2008). Dynein regulates epithelial polarity and the apical localization of stardust mRNA. *PLoS Genet.* **4**, e8.
- Hurd, T. W., Gao, L., Roh, M. H., Macara, I. G. and Margolis, B. (2003). Direct interaction of two polarity complexes implicated in epithelial tight junction assembly. *Nat. Cell Biol.* **5**, 137–142.
- Kempkens, O., Médina, E., Fernandez-Ballester, G., Özüyan, S., Le Bivic, A., Serrano, L. and Knust, E. (2006). Computer modelling in combination with in vitro studies reveals similar binding affinities of *Drosophila* Crumbs for the PDZ domains of Stardust and DmPar-6. *Eur. J. Cell Biol.* **85**, 753–767.
- Kerman, B. E., Cheshire, A. M., Myat, M. M. and Andrew, D. J. (2008). Ribbon modulates apical membrane during tube elongation through Crumbs and Moesin. *Dev. Biol.* **320**, 278–288.
- Klebes, A. and Knust, E. (2000). A conserved motif in Crumbs is required for E-cadherin localisation and zonula adherens formation in *Drosophila*. *Curr. Biol.* **10**, 76–85.
- Krahn, M. P., Bückers, J., Kastrup, L. and Wodarz, A. (2010). Formation of a Bazooka-Stardust complex is essential for plasma membrane polarity in epithelia. *J. Cell Biol.* **190**, 751–760.
- Laprise, P. and Tepass, U. (2011). Novel insights into epithelial polarity proteins in *Drosophila*. *Trends Cell Biol.* **21**, 401–408.
- Laprise, P., Paul, S. M., Boulanger, J., Robbins, R. M., Beitel, G. J. and Tepass, U. (2010). Epithelial polarity proteins regulate *Drosophila* tracheal tube size in parallel to the luminal matrix pathway. *Curr. Biol.* **20**, 55–61.
- Lemmers, C., Médina, E., Delgrossi, M. H., Michel, D., Arsanto, J. P. and Le Bivic, A. (2002). hNADL/PATJ, a homolog of discs lost, interacts with crumbs and localizes to tight junctions in human epithelial cells. *J. Biol. Chem.* **277**, 5408–5415.
- Lemmers, C., Michel, D., Lane-Guermonprez, L., Delgrossi, M. H., Médina, E., Arsanto, J. P. and Le Bivic, A. (2004). CRB3 binds directly to Par6 and regulates the morphogenesis of the tight junctions in mammalian epithelial cells. *Mol. Biol. Cell* **15**, 1324–1333.
- Letizia, A., Sotillos, S., Campuzano, S. and Llimargas, M. (2011). Regulated Crb accumulation controls apical constriction and invagination in *Drosophila* tracheal cells. *J. Cell Sci.* **124**, 240–251.
- Massey-Harroche, D., Delgrossi, M. H., Lane-Guermonprez, L., Arsanto, J. P., Borg, J. P., Billaud, M. and Le Bivic, A. (2007). Evidence for a molecular link between the tuberous sclerosis complex and the Crumbs complex. *Hum. Mol. Genet.* **16**, 529–536.
- Michel, D., Arsanto, J. P., Massey-Harroche, D., Béclin, C., Wijnholds, J. and Le Bivic, A. (2005). PATJ connects and stabilizes apical and lateral components of tight junctions in human intestinal cells. *J. Cell Sci.* **118**, 4049–4057.
- Morais-de-Sá, E., Mirouse, V. and St Johnston, D. (2010). aPKC phosphorylation of Bazooka defines the apical/lateral border in *Drosophila* epithelial cells. *Cell* **141**, 509–523.
- Nam, S. C. and Choi, K. W. (2006). Domain-specific early and late function of Dpatj in *Drosophila* photoreceptor cells. *Dev. Dyn.* **235**, 1501–1507.
- Pellikka, M., Tanentzapf, G., Pinto, M., Smith, C., McGlade, C. J., Ready, D. F. and Tepass, U. (2002). Crumbs, the *Drosophila* homologue of human CRB1/RP12, is essential for photoreceptor morphogenesis. *Nature* **416**, 143–149.
- Pielage, J., Stork, T., Bunse, I. and Klämbt, C. (2003). The *Drosophila* cell survival gene discs lost encodes a cytoplasmic Codanin-1-like protein, not a homolog of tight junction PDZ protein Patj. *Dev. Cell* **5**, 841–851.

- Pocha, S. M., Wassmer, T., Niehage, C., Hoflack, B. and Knust, E. (2011). Retromer controls epithelial cell polarity by trafficking the apical determinant Crumbs. *Curr. Biol.* **21**, 1111-1117.
- Richard, M., Grawe, F. and Knust, E. (2006). DPATJ plays a role in retinal morphogenesis and protects against light-dependent degeneration of photoreceptor cells in the *Drosophila* eye. *Dev. Dyn.* **235**, 895-907.
- Roeth, J. F., Sawyer, J. K., Wilner, D. A. and Peifer, M. (2009). Rab11 helps maintain apical crumbs and adherens junctions in the *Drosophila* embryonic ectoderm. *PLoS ONE* **4**, e7634.
- Roh, M. H., Makarova, O., Liu, C. J., Shin, K., Lee, S., Laurinec, S., Goyal, M., Wiggins, R. and Margolis, B. (2002). The Maguk protein, Pals1, functions as an adapter, linking mammalian homologues of Crumbs and Discs Lost. *J. Cell Biol.* **157**, 161-172.
- Shin, K., Straight, S. and Margolis, B. (2005). PATJ regulates tight junction formation and polarity in mammalian epithelial cells. *J. Cell Biol.* **168**, 705-711.
- St Johnston, D. and Ahringer, J. (2010). Cell polarity in eggs and epithelia: parallels and diversity. *Cell* **141**, 757-774.
- Sugihara-Mizuno, Y., Adachi, M., Kobayashi, Y., Hamazaki, Y., Nishimura, M., Imai, T., Furuse, M. and Tsukita, S. (2007). Molecular characterization of angiomotin/JEAP family proteins: interaction with MUPP1/Patj and their endogenous properties. *Genes Cells* **12**, 473-486.
- Tanentzapf, G., Smith, C., McGlade, J. and Tepass, U. (2000). Apical, lateral, and basal polarization cues contribute to the development of the follicular epithelium during *Drosophila* oogenesis. *J. Cell Biol.* **151**, 891-904.
- Tepass, U., Theres, C. and Knust, E. (1990). crumbs encodes an EGF-like protein expressed on apical membranes of *Drosophila* epithelial cells and required for organization of epithelia. *Cell* **61**, 787-799.
- Wells, C. D., Fawcett, J. P., Traweger, A., Yamanaka, Y., Goudreaux, M., Elder, K., Kulkarni, S., Gish, G., Virag, C., Lim, C. et al. (2006). A Rich1/Amot complex regulates the Cdc42 GTPase and apical-polarity proteins in epithelial cells. *Cell* **125**, 535-548.
- Wodarz, A., Hinz, U., Engelbert, M. and Knust, E. (1995). Expression of crumbs confers apical character on plasma membrane domains of ectodermal epithelia of *Drosophila*. *Cell* **82**, 67-76.
- Wodarz, A., Ramrath, A., Kuchinke, U. and Knust, E. (1999). Bazooka provides an apical cue for Inscuteable localization in *Drosophila* neuroblasts. *Nature* **402**, 544-547.
- Xu, N., Keung, B. and Myat, M. M. (2008). Rho GTPase controls invagination and cohesive migration of the *Drosophila* salivary gland through Crumbs and Rho-kinase. *Dev. Biol.* **321**, 88-100.
- Zhou, B., Wu, Y. and Lin, X. (2011). Retromer regulates apical-basal polarity through recycling Crumbs. *Dev. Biol.* **360**, 87-95.
- Zhou, W. and Hong, Y. (2012). *Drosophila* Patj plays a supporting role in apical-basal polarity but is essential for viability. *Development* **139**, 2891-2896.

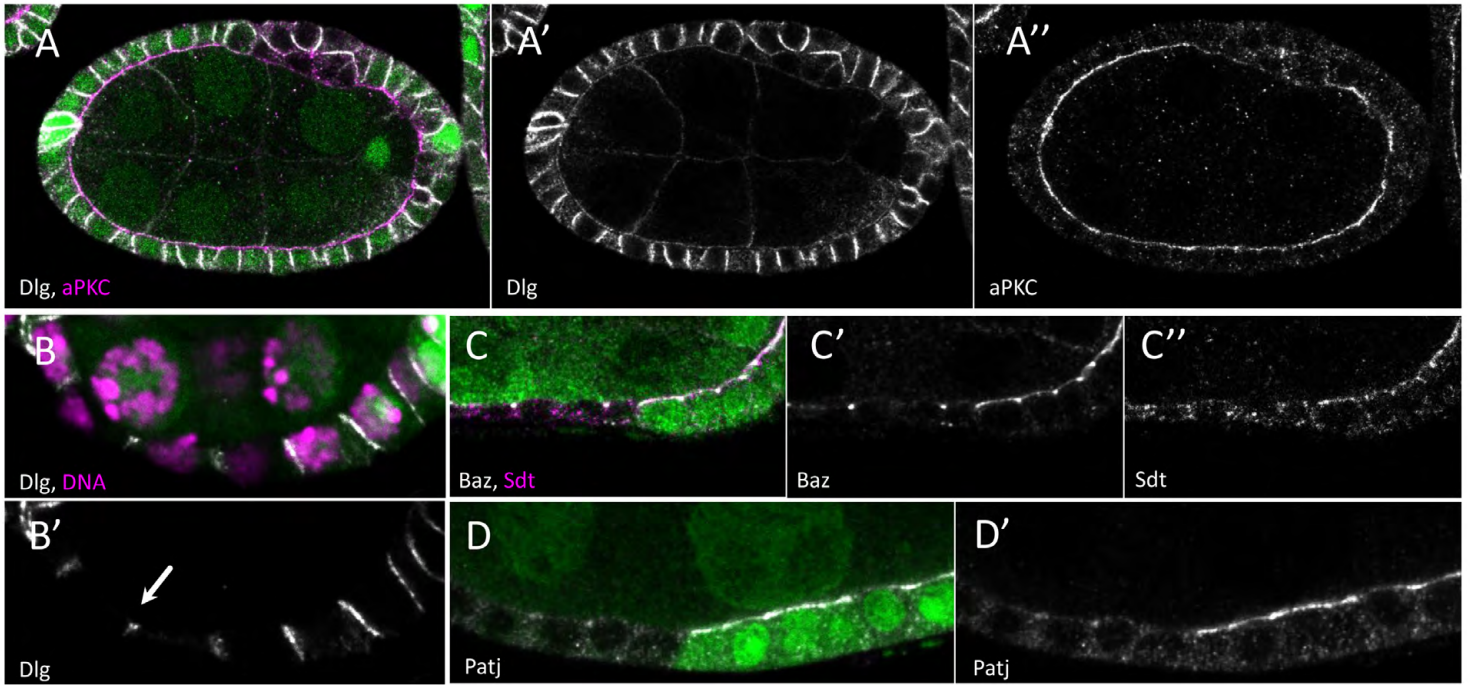


*sdt*<sup>K85</sup>



**Fig. S1. Defects observed in *sdt* mutants in follicular cells.** Follicular cell mutant clones for *sdt*<sup>K85</sup> marked by the absence of GFP. (A-A'') Clones showing a cell stretching defect and decrease of aPKC from the apical domain. (B) One of the mutant clones shows a multilayer and Dlg spreading all around the cells. (C,C') Decrease in Baz from the apical domain in *sdt* mutant cells. (D,D') Complete loss of Crb from the apical domain in *sdt* mutant cells. (E,E') Complete loss of Patj from the apical domain in *sdt* mutant cells.

*crb*<sup>8F105</sup>



**Fig. S2. Defects observed in *crb* mutants in follicular cells.** Follicular cell mutant clones for *crb*<sup>8F105</sup> marked by the absence of GFP. (A-A'') Mutant clone showing multilayers of cells, mislocalisation of aPKC and spreading of Dlg. (B,B') Mutant clones with flat cells that can be seen by the reduced size of the lateral domain stained for Dlg (arrow on B'). (C-C'') Loss of apical Baz (white on C, and C') and Sdt (pink on C, and white on C'') in *crb* mutant cells. (D,D') Complete loss of Patj from the apical domain in *crb* mutant cells.