

# MLL1, a H3K4 methyltransferase, regulates the TNF $\alpha$ -stimulated activation of genes downstream of NF- $\kappa$ B

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## Summary

Genes of the mixed lineage leukemia (MLL) family regulate transcription by methylating histone H3K4. Six members of the MLL family exist in humans, including *SETD1A*, *SETD1B* and *MLL1–MLL4*. Each of them plays non-redundant roles in development and disease genesis. *MLL1* regulates the cell cycle and the oscillation of circadian gene expression. Its fusion proteins are involved in leukemogenesis. Here, we studied the role of MLL1 in innate immunity and found it selectively regulates the activation of genes downstream of NF- $\kappa$ B mediated by tumor necrosis factor (TNF $\alpha$ ) and lipopolysaccharide (LPS). Real-time PCR and genome-wide gene expression profile analysis proved that the deficiency of *MLL1* reduced the expression of a group of genes downstream of nuclear factor  $\kappa$ B (NF- $\kappa$ B). However, the activation of NF- $\kappa$ B itself was not affected. The MLL1 complex is found both in the nucleus and cytoplasm and is associated with NF- $\kappa$ B. CHIP assays proved that the translocation of MLL1 to chromatin was dependent on NF- $\kappa$ B. Our results suggest that MLL1 is recruited to its target genes by activated NF- $\kappa$ B and regulates their transcription.

**Key words:** MLL1, MLL, Histone methylation, Transcription regulation, NF- $\kappa$ B signaling pathway

## Introduction

*MLL1* (mixed lineage leukemia 1) was first identified from leukemia patients ~20 years ago (Krivtsov and Armstrong, 2007; Ziemin-van der Poel et al., 1991). The N-terminal of *MLL1* is able to fuse with other partners upon chromatin translocation and this induces leukemogenesis by regulating the expression of HOX genes (Krivtsov and Armstrong, 2007; Mohan et al., 2010). The biochemical function of MLL1 was not clear until Set1, its homolog in *Saccharomyces cerevisiae*, was defined as an H3K4 methyltransferase (Miller et al., 2001). Six homologs for Set1 exist in human, SETD1A (SET domain containing 1A), SETD1B (SET domain containing 1B) and MLL1–MLL4 (Wu et al., 2008). Each of these six proteins contains a conserved SET domain, the catalytic domain for methylating histone H3K4, and its full function requires the formation of a complicated complex. Each enzyme complex contains four common units, including RBBP5 (retinoblastoma binding protein 5), WDR5 (WD repeat domain 5), ASH2L (ash2-like) and DPY30 (dpy-30 homolog) (Cho et al., 2007; Lee and Skalnik, 2005; Lee and Skalnik, 2008; Lee et al., 2007; Wu et al., 2008; Yokoyama et al., 2004). The six complexes might have different biological functions because each one also contains specific subunits (Eissenberg and Shilatifard, 2010). This idea is also supported by studies within animal models. Deletion of either *Mll1* or *Mll2* in mice is embryonic lethal, suggesting that both *Mll1* and *Mll2* have important and non-redundant roles in embryo development (Glaser et al., 2006; Yu et al., 1995).

Data from a ChIP-Chip assay has indicated that MLL1 might be a common transcription factor and essential for gene transcription (Guenther et al., 2005). However, other groups have come to different conclusions because MEFs (mouse embryonic fibroblasts) derived from *Mll1*<sup>-/-</sup> mice grow normally. Moreover, the global mRNA expression patterns in the *Mll1*<sup>-/-</sup> MEF cell also did not change much compared with that of the wild type. MLL1 has been shown to regulate only the expression of a subset of active genes in vivo (Milne et al., 2005). Genome wide ChIP-Chip assays of H3K4 trimethylation indicate that only ~5% of the actively transcribed genes have reduced methylation in *Mll1*<sup>-/-</sup> cells in comparison with the wild type (Wang et al., 2009). A similar result was also observed for a global gene expression profile (Wang et al., 2009). The function of MLL1 on the majority of gene promoters is still unknown.

How MLL1 fusion proteins regulate leukemogenesis has been heavily investigated in the past 20 years (Krivtsov and Armstrong, 2007; Mohan et al., 2010). In contrast, very few studies have explored other physiological functions of MLL1. Although methylated H3K4 seems to be related to global gene transcription (Barski et al., 2007; Eissenberg and Shilatifard, 2010; Ruthenburg et al., 2007; Shilatifard, 2008; Sims and Reinberg, 2006), deficiency of the methylation does not always lead to gene silencing. Therefore, how H3K4 methylation regulates transcription still remains puzzling. Moreover, multiple H3K4 methyltransferases exist in mammals and their functions are not redundant. Their

functions in development and other physiological events remain to be elucidated.

Recently, an MLL1 complex has been found to associate with condensed chromatin during M phase, on genes that are usually expressed preferentially in interphase (Blobel et al., 2009). It is hypothesized that MLL1 binds to these genes and regulates their methylation and transcription immediately after exit from M phase. Another report has recently suggested that MLL1 is associated with the CLOCK (circadian locomotor output cycles kaput)–BMAL1 (brain and muscle ARNT-like 1) complex and is recruited to circadian promoters (Katada and Sassone-Corsi, 2010).

The MLL1 complex is usually considered to mainly function inside the nucleus; nearly all previous studies have shown that MLL1 functions on the chromatin, and the first complex to be purified was also done from nuclear extract (Yokoyama et al., 2004). However, one study has reported that WDR5, one important subunit of the MLL1 complex, is associated with the VISA-associated complex and regulates the virus-triggered induction of type I interferons (IFNs) and the innate immune response in the cytoplasm (Wang et al., 2010). The discovery raised the possibility that the MLL1 complex might also function in the cytoplasm and methylate substrates other than H3. The NF- $\kappa$ B pathway is one of the key pathways involved in innate immunity and other physiological responses (Baud and Karin, 2009; Ghosh and Hayden, 2008; Smale, 2010; Spehlmann and Eckmann, 2009; Sun and Karin, 2008). This pathway can be activated by many signals, including TNF $\alpha$  (tumor necrosis factor), LPS (lipopolysaccharide), TLRs (TOLL-like receptors) and so on. Usually different signals will trigger different profiles of gene transcription (Ghosh and Hayden, 2008; Smale, 2010). How a single transcription factor achieves selective gene transcription has been an unsolved question for a long time. Epigenetic markers and molecules have emerged as key players in gene transcription (Campos and Reinberg, 2009; Chi et al., 2010; Eissenberg and Shilatifard, 2010; Smith et al., 2011; Sukanuma and Workman, 2011; Trojer and Reinberg, 2006). MLL1 and MLL2 have been shown to be recruited by the p52 subunit of NF- $\kappa$ B to the promoter of *MMP9* (matrix metalloproteinase 9) in a T cell lymphoma cell line (Robert et al., 2009). It is therefore possible MLL1 is involved in the regulation of the NF- $\kappa$ B signaling pathway.

We investigated the relationship between MLL1 and the TNF $\alpha$ - and LPS-stimulated NF- $\kappa$ B signaling pathway in this study. Our data suggest that MLL1 selectively regulates the activation of genes downstream of NF- $\kappa$ B.

## Results

### MLL1 deficiency leads to a reduction of the activation of a subset of genes downstream of NF- $\kappa$ B

We started to investigate the role of Mll1 in NF- $\kappa$ B signaling pathway by checking the altered expression of NF- $\kappa$ B downstream genes in Mll1-deficient cells. Wild-type and *Mll1*<sup>-/-</sup> cells were treated with TNF $\alpha$  (10 ng/ml), and harvested 0.5 hours or 4 hours after treatment. Over thirty known NF- $\kappa$ B target genes were examined. The basal mRNA level of most of the genes did not show a significant change in the *Mll1*<sup>-/-</sup> cells (supplementary material Fig. S1). Only very few genes, such as *Ptges*, showed reduction of the mRNA level upon *Mll1* deficiency (supplementary material Fig. S1). Genes activated significantly by TNF $\alpha$  in MEFs were used for further

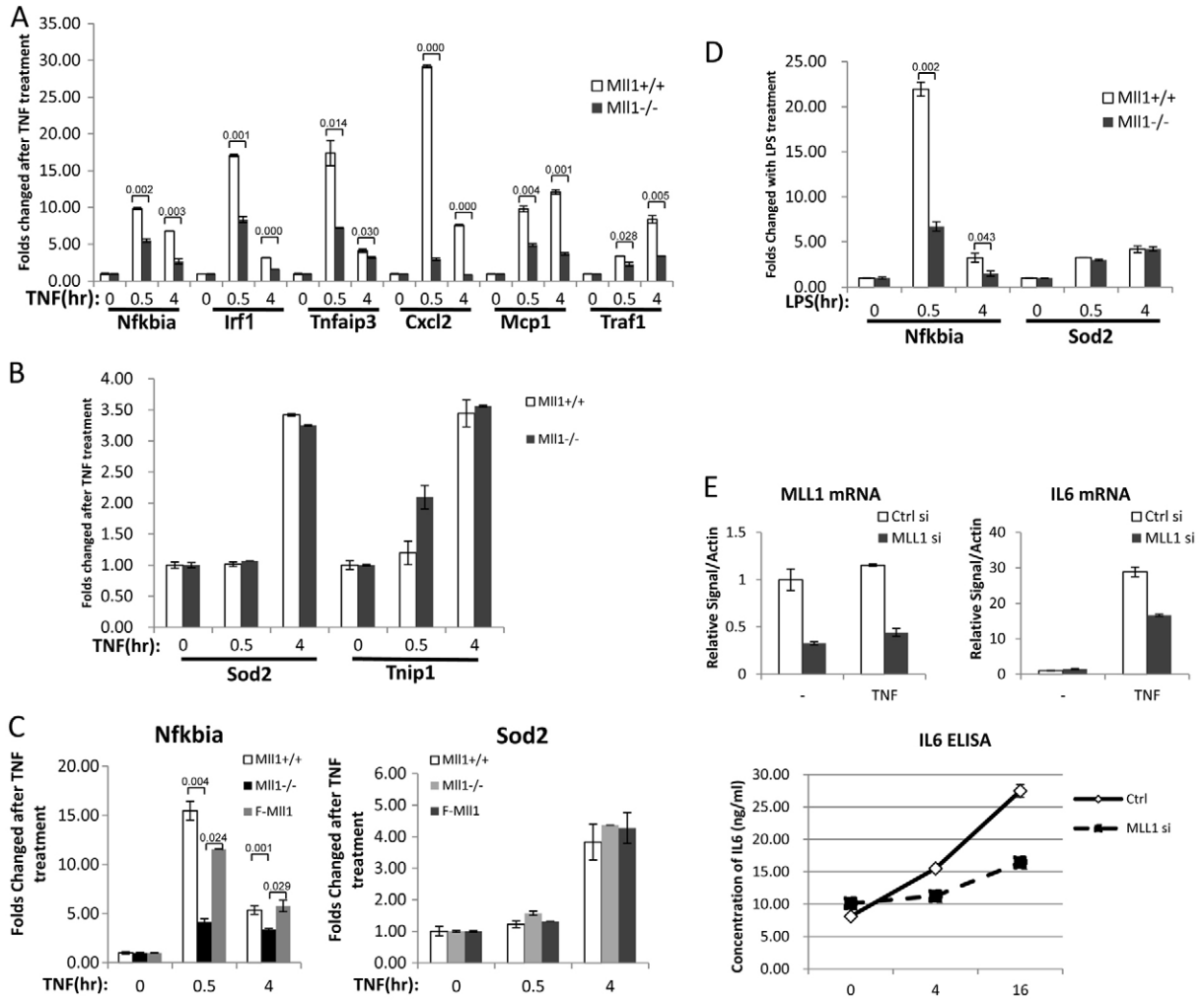
analysis. We found that the activation of many genes was greatly impaired in the *Mll1*<sup>-/-</sup> cells, such as the *Nfkbia* (the gene encoding I $\kappa$ B $\alpha$ ), *Tnfaip3* (also known as A20), *Irf1*, *Cxcl1*, *Ccl2* (also known as Mcp1) and *Trafl* genes (Fig. 1A). However, we also found a group of the NF- $\kappa$ B target genes whose mRNA level was not reduced in *Mll1*<sup>-/-</sup> cells, such as *Sod2* and *Tnip1* (Fig. 1B). In order to further confirm that the reduction of gene expression was the consequence of *Mll1* deficiency, the above experiments were repeated using a cell line in which a Flag-tagged *Mll1* had been integrated into the genome of *Mll1*<sup>-/-</sup> cell (designated F-Mll1). We found that the activation levels of *Nfkbia* (I $\kappa$ B $\alpha$ ) (Fig. 1C), *Tnfaip3* (A20) and *Irf1* were restored to a normal level in the F-Mll1 cell line. The increase of Mll1 amount did not affect the expression of *Sod2*, as judged by comparing the expression in *Mll1*<sup>-/-</sup> and F-Mll1 cells (Fig. 1C). These data suggest that MLL1 is involved in the transcriptional activation of NF- $\kappa$ B target genes and selectively regulates a subset of downstream genes in MEFs. In the following studies, *Nfkbia* (I $\kappa$ B $\alpha$ ) and *Sod2* were used as the representative genes for understanding the molecular mechanisms of this process.

We further tested whether Mll1 also regulates gene expression downstream of NF- $\kappa$ B under other stimuli or in other cell lines. LPS was used to stimulate the above MEF cell lines. The expression of *Nfkbia* (I $\kappa$ B $\alpha$ ) and *Sod2* was similar to that upon TNF $\alpha$  treatment (Fig. 1D). HCT116, a human colon cancer cell line, and HL7702, an immortalized human liver cell line, were used to verify the effect of *Mll1* on activation of genes downstream of NF- $\kappa$ B. siRNA against *Mll1* was transfected into the above cell lines and the mRNA levels of genes were assayed. Data suggest Mll1 also regulates NF- $\kappa$ B target genes in the above cell lines (supplementary material Figs S2, S3). In order to verify whether the protein levels of these genes were also impaired in the MLL1-deficient cells, an ELISA experiment was performed for IL6 in the HL7702 cell line. Similar to the results with mRNA (Fig. 1E, top right), the amount of secreted IL6 in the medium was greatly reduced after *Mll1* knockdown (Fig. 1E, bottom). Thus, we conclude that MLL1 is involved in the activation of downstream genes of NF- $\kappa$ B mediated by TNF $\alpha$  and LPS in multiple cell lines.

### A genome-wide gene expression profile proves Mll1 regulates the NF- $\kappa$ B signaling pathway

In order to identify the genes downstream of NF- $\kappa$ B that are regulated by Mll1 on a genome-wide scale, a gene expression profile was analyzed by next-generation sequencing. *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> MEF cells were treated with TNF $\alpha$  for 4 hours, and then isolated mRNA was reverse transcribed and subjected to next-generation sequencing. The raw data have been uploaded to the GEO database (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE36567>). The identified genes with different expression levels in each cell line are listed in supplementary material Tables S2 and S3 ( $P < 0.01$ ). The numbers of genes upregulated by TNF $\alpha$  were calculated (Fig. 2A). The numbers of genes upregulated more than 1.5-fold were very close in wild-type and knockout cells. However, for genes upregulated more than 2-, 3- or 4-fold, the numbers in the *Mll1*<sup>-/-</sup> cells were much less than those in *Mll1*<sup>+/+</sup> cells. This suggests that some of the genes are stimulated to a lesser extent by TNF $\alpha$  in the absence of *Mll1*.

Genes that were upregulated more than 2-fold in *Mll1*<sup>+/+</sup> cells were picked for clustering analysis in all the four samples ( $P < 0.01$  in all four samples) (Fig. 2B). The genes were clearly



**Fig. 1. MLL1 regulates the expression of genes downstream of NF- $\kappa$ B.** (A) Cells were treated with TNF $\alpha$  and the levels of mRNA for various genes were assayed. The activation of genes downstream of NF- $\kappa$ B [*Nfkb1a* ( $\text{I}\kappa\text{B}\alpha$ ), *Irf1*, *Tnfaip3* (A20), *Cxcl2*, *Mcp1* and *Traf1*] was reduced in *Mll1*<sup>-/-</sup> cells. (B) The activation of *Sod2* and *Tnfp1* was not reduced in *Mll1*<sup>-/-</sup> cells. (C) The activation of *Nfkb1a* was restored in the F-*Mll1* cell line. (D) LPS was used to activate the NF- $\kappa$ B signaling pathway. The activation of the  $\text{I}\kappa\text{B}\alpha$ -encoding gene was also impaired in *Mll1*<sup>-/-</sup> cells, but not that of *Sod2*. (E) *Mll1* was knocked down by siRNA in HL-7702 cells (top left). Levels of IL6 in the medium were assayed by ELISA. MLL1 deficiency caused the reduction of mRNA (top right) and protein of IL6 (bottom). The *P* value of significantly different results is indicated above the corresponding columns.

categorized into two groups, *Mll1*-dependent and *Mll1*-independent genes (Fig. 2B). These data correlated very well with our studies using real-time PCR and further proved that MLL1 selectively regulates TNF $\alpha$ -induced genes downstream of NF- $\kappa$ B on a genome-wide scale. We searched the two groups of genes using the Gene Ontology database. Among the 61 *Mll1*-dependent genes, 27 genes (~45%) are related to the inflammation and immune response. For the *Mll1*-independent genes, 11 out of 37 genes (29%) are related to the inflammation and immune response. We conclude that *Mll1*-dependent genes seem to be enriched in those related with inflammation and immune response.

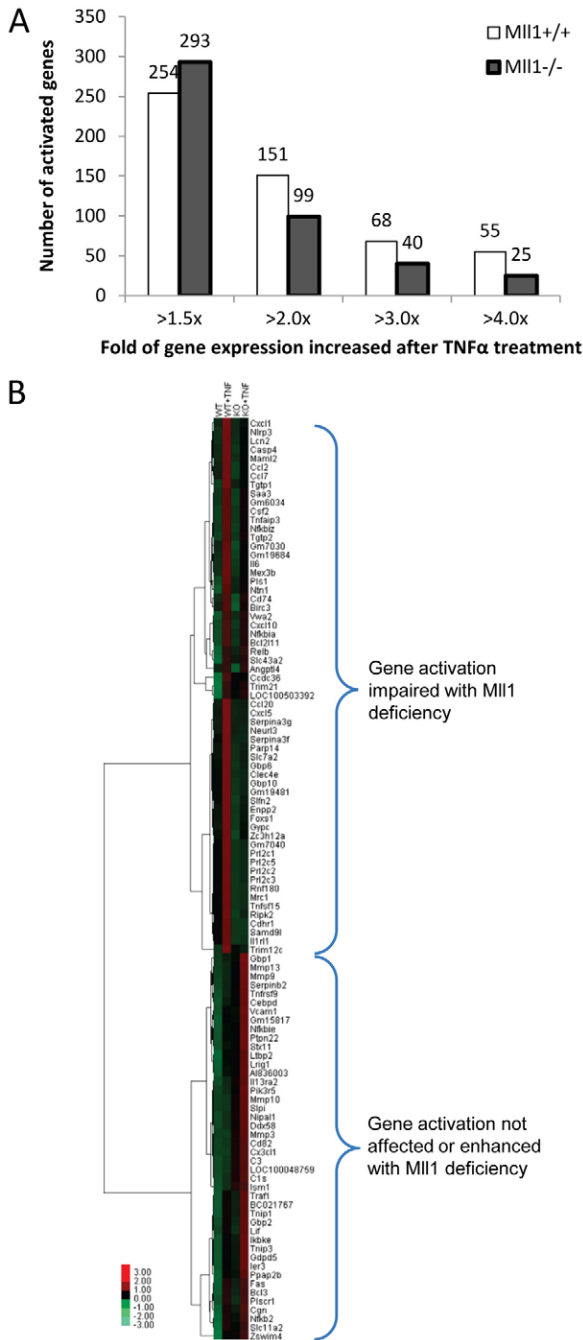
#### MLL1 depletion does not affect the activation of NF- $\kappa$ B by TNF $\alpha$

In order to further understanding of the role of MLL1 in the activation of the NF- $\kappa$ B signaling pathway, we checked whether activation of the pathway was impaired in *Mll1*<sup>-/-</sup> cell lines by

using a luciferase reporter assay. *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> cell lines were treated with TNF $\alpha$  and the luciferase activity was assayed 12 hours later. The result showed that the NF- $\kappa$ B pathway was activated in *Mll1*<sup>-/-</sup> cells to a similar level to that in wild-type cells (Fig. 3A). We also studied the translocation of the p50 subunit of NF- $\kappa$ B into the nucleus by immunofluorescent staining and CHIP assays in the *Mll1*<sup>-/-</sup> cell. Immunofluorescent staining indicated that p50 was still translocated into nucleus after TNF $\alpha$  treatment in the absence of *Mll1* (supplementary material Fig. S4). The results of CHIP assay showed that the ability of p50 to bind to DNA had no obvious difference between *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> cells (Fig. 3B). Taken together, these data suggest that MLL1 deficiency does not affect the activation of NF- $\kappa$ B by TNF $\alpha$ .

#### MLL1 depletion affects the oscillation of $\text{I}\kappa\text{B}\alpha$ upon TNF $\alpha$ treatment

To study the effect of MLL1 on the NF- $\kappa$ B signaling pathway, we analyzed the degradation of  $\text{I}\kappa\text{B}\alpha$ , which is one of the



**Fig. 2. The expression profiles of genes downstream of NF-κB regulated by MLL1, as analyzed by RNA sequencing.** *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> cells were treated with TNFα and extracted RNA were subjected to a gene expression profile analysis. (A) The number of genes upregulated by the indicated amount in both cell lines was calculated. The number of genes upregulated by >1.5-fold was not substantially different in the two cell lines. The number of genes upregulated genes by a greater amount (>2-, 3- and 4-fold) was less in the *Mll1*<sup>-/-</sup> cell, which suggests that the level of activation of some genes was reduced in the absence of MLL1. (B) All the genes activated by TNFα in the *Mll1*<sup>+/+</sup> cell (i.e. with a >2-fold increase at *P*<0.01 in all four samples) were subjected to gene clustering. A total of 61 of these 97 genes showed an *Mll1*-dependent regulation.

hallmarks of NF-κB activation. In *Mll1*<sup>-/-</sup> cells, the protein level of IκBα rapidly decreased 30 minutes after TNFα treatment (Fig. 3C). However, at 4 hours, its protein level did not recover to

the same extent as that seen in wild-type cells. Instead, the protein levels of IκBα at 0.5, 4 and 12 hours were almost the same (Fig. 3C). The level of *Nfkbia* mRNA (i.e. that encoding IκBα), and the bound p65 on the *Nfkbia* promoter were also analyzed at the above time points (supplementary material Figs S5, S6). The oscillation of IκBα protein levels is a typical characteristic of prolonged NF-κB activation and is known to be the consequence of degradation and synthesis of IκBα at the same time. The levels of IκBα expression in *Mll1*<sup>-/-</sup> cells is consistent with the above data, and is due to the impaired mRNA synthesis of *Nfkbia* (supplementary material Fig. S5). However, the amount of p65 on the *Nfkbia* promoter was not different between the two cell lines (supplementary material Fig. S6), which is consistent with the above data.

**WDR5 and RBBP5 are present in both the nucleus and cytoplasm**

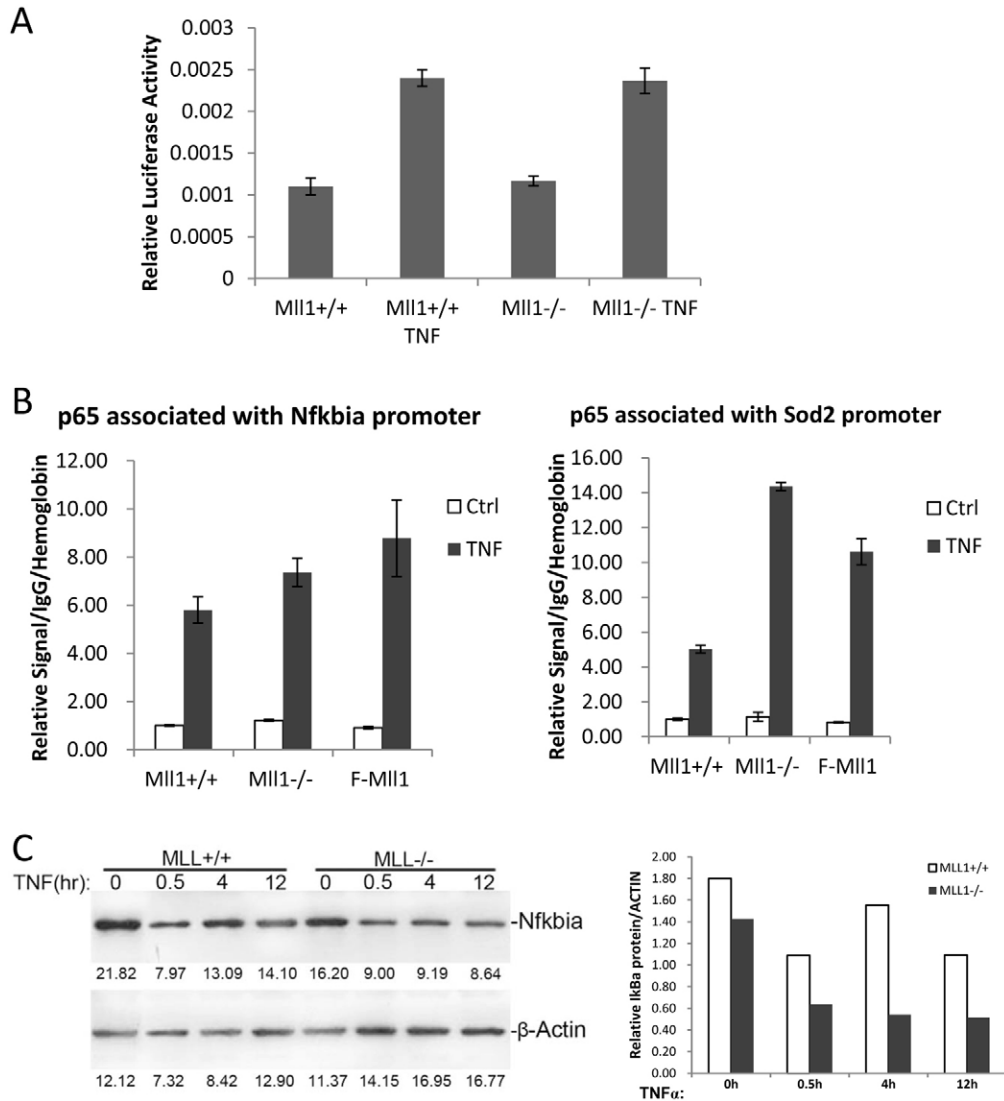
The MLL1 complex exerts its function on the chromatin and until recently no study had reported whether it is present outside the nucleus. Wang et al. reported recently that WDR5 localizes to both the cytosol and nucleus (Wang et al., 2010). We studied the localization of endogenous RBBP5 and WDR5 by western blotting and immunofluorescent staining. Wild-type MEF cells were separated into three fractions, those from the cytosol, soluble and insoluble fraction of nucleus. The results of western blotting showed that RBBP5 and WDR5 are present both in cytosol and nucleus (Fig. 4A). Histone H3 was used as the control for the nuclear fraction. p65, the transactivation subunit of NF-κB, resided in the cytoplasm without upstream signals (Fig. 4A). The results of immunofluorescent staining showed that the majority of WDR5 and RBBP5 are localized inside the nuclear. However, we also observed substantial amount of the above proteins in the cytosol (Fig. 4B). These data suggested that WDR5 and RBBP5, two subunits of the MLL1 complex, are localized both in cytosol and nuclear.

**Generation of a cell line expressing triple-Flag-tagged MLL1**

MLL1 is usually expressed at a very low level in many cell lines. In order to further study the localization of Mll1, we generated a knock-in cell line derived from a colon cancer cell line, HCT116 (Fig. 4C). A triple Flag tag and streptavidin-binding peptide (SBP) were knocked into the C-terminal of the *MLL1* gene in one of the chromosome copies. Because the SET domain, which is responsible for the catalytic activity of MLL1, is located at the very C-terminus of the MLL1 protein, the addition of the tags meant the resulting protein (designated 3F-MLL1) was unable to methylate histone H3 (data not shown). We did not get homozygous clones, which suggests that the complete loss of MLL1 catalytic activity causes cell death. However, the heterozygous clones were still useful.

**MLL1 is associated with RBBP5 in cytoplasm**

Next, we separated nuclear and cytoplasm fractions of the 3F-MLL1-expressing cells and performed immunoprecipitation with anti-Flag antibody for each fraction. We detected 3F-MLL1 in both fractions (Fig. 4D). Interestingly, this result also indicates that cytoplasmic MLL1 is associated with RBBP5 (Fig. 4D), which raises the possibility that the MLL1 complex might be intact and catalytically functional outside of nucleus.



**Fig. 3. MLL1 deficiency affected the oscillation of the level of IκBα protein.** (A) In wild-type and *Mll1*<sup>-/-</sup> cells, TNFα activates NF-κB pathways to the same level in a luciferase reporter assay. (B) A ChIP assay of p65 shows that the recruitment of p65 to the *Nfkbia* (IκBα) (left) and *Sod2* (right) promoters was not reduced in the *Mll1*<sup>-/-</sup> cell, compared with wild-type and Flag-MLL1-expressing MEFs. (C) The oscillation of IκBα was affected in *Mll1*<sup>-/-</sup> cell. With prolonged TNFα treatment in the wild-type cell, levels of IκBα had decreased by 30 minutes, but had recovered at 4 hours and had decreased again at 12 hours. In *Mll1*<sup>-/-</sup> cell, the level of IκBα was very similar at the different time points. The right-hand panel shows a quantification of western blot.

### The cytoplasmic MLL1 complex translocates into nucleus after TNFα treatment

To investigate the role of the MLL1 complex in TNFα-mediated NF-κB signaling, we first studied the localization of MLL1 and RBBP5 upon TNFα treatment. The 3F-MLL1-expressing HCT116 cell line was treated with 10 ng/ml TNFα for 30 minutes before being fractionated to give cytosol and nuclear extracts. The results of immune blotting showed that p65 was translocated into the nuclei after TNFα treatment as expected. The protein levels of 3F-MLL1 in the cytosol also decreased upon TNFα treatment. MLL1 might be translocated from cytosol into nucleus (Fig. 4E). We did not see a significant increase in 3F-MLL1 in the nuclear fraction, which might be owing to the large amount of MLL1 protein in the nucleus before TNFα treatment (Fig. 4E). We also tried immunofluorescent staining experiment with anti-Flag antibody, but we could not see MLL1 signal owing to the low expression level. The localization of RBBP5 was also examined by western blotting and immunofluorescent staining. The pattern was similar to that in MEF cell (Fig. 4A) and only a very tiny change was observed in the amount of this protein that was present in both the cytosol and nuclear fractions. Since RBBP5 is a common subunit of all six H3K4 methyltransferases, a change in the distribution of

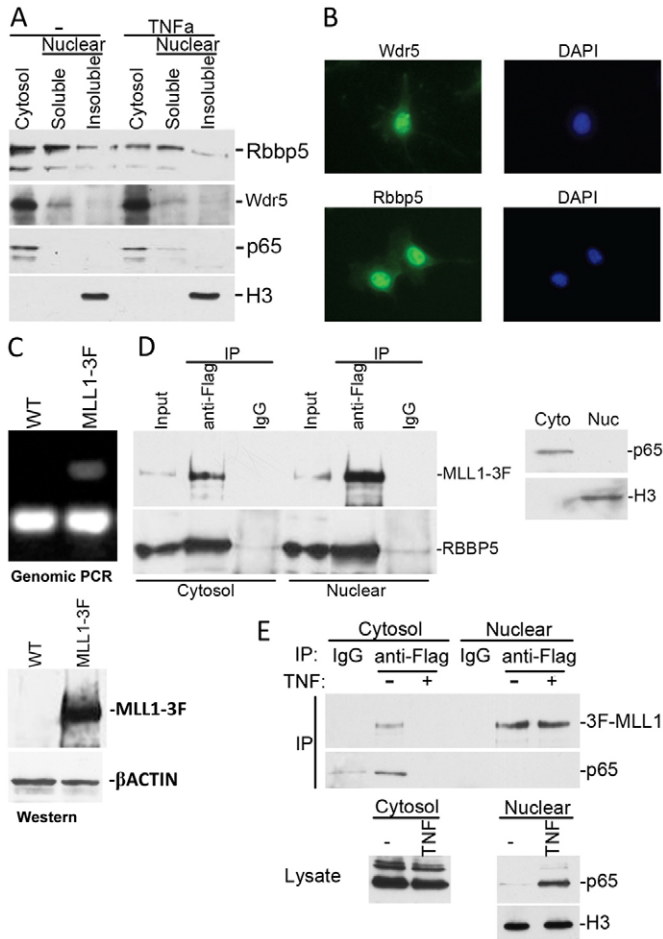
the MLL1 complex might not affect the distribution of RBBP5 greatly in the cell.

### MLL1 is associated with p65

The above studies suggest that there is a correlation between MLL1 localization and TNFα-stimulated signaling transduction. Thus, we further explored the interaction between p65 and MLL1 complex in immunoprecipitation studies. Anti-Flag antibody pulled down 3F-MLL1, as well as p65, in the cytoplasm fraction, which suggests that MLL1 is associated with NF-κB in the cytoplasm (Fig. 4E). After TNFα treatment, MLL1 disappeared from the cytoplasm but a large amount of p65 still remained. However, anti-Flag antibody did not bring down detectable p65 protein, which serves as good control (Fig. 4E). We did not observe obvious interaction in the nuclear fraction. This might be because the high concentration of NaCl (420 mM) used in the fractionation procedure broke the interaction.

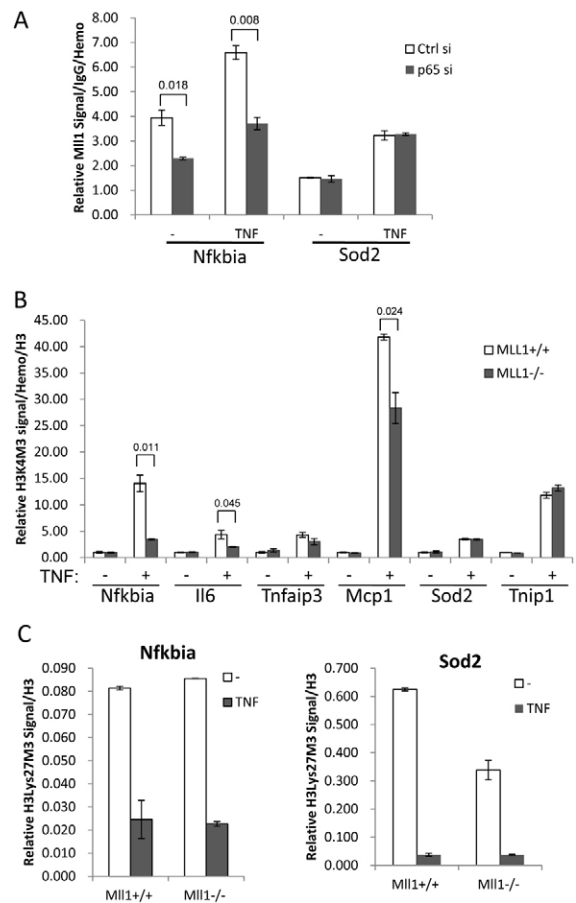
### MLL1 binds to the promoters of NF-κB target genes before activation

Because MLL1 regulates the expression of NF-κB target genes but does not affect the activation of NF-κB, it might function



**Fig. 4. MLL1 translocates into the nucleus upon TNF $\alpha$  treatment.** (A) MEF cell was fractionated into cytoplasmic, and soluble and insoluble nuclear fractions. RBBP5 and WDR5 were found in both the cytoplasm and nucleus. (B) Immunofluorescent staining shows that WDR5 and RBBP5 in MEF cell are localized in both the cytoplasm and nucleus. (C) PCR of genomic DNA (top) and anti-Flag western blotting (bottom) of the 3F-MLL1-expressing HCT116 cell. (D) After fractionation, the cytoplasm and nuclear fractions of the 3F-MLL1-expressing HCT116 cell were used for anti-Flag immunoprecipitation (IP). 3F-MLL1 brought down RBBP5 in both fractions. Western blots of p65 and H3 are shown as the control for fractionation. (E) 3F-MLL1 in the cytoplasm disappeared after TNF $\alpha$  treatment. Immunoprecipitation of 3F-MLL1 brought down p65 in the cytoplasm fraction. Western blots of H3 and p65 are shown as the loading control.

after NF-κB binds to the chromatin. In addition, it is possible that MLL1 functions differently on the promoters of *Nfkbia* (encoding IκB $\alpha$ ) and *Sod2*. We first studied whether MLL1 binds to the promoters of *Nfkbia* and *Sod2* in MEF cell lines by using a CHIP assay. The results showed that MLL1 binds to both of the promoters before treatment (Fig. 5A). Although the signal on the *Sod2* promoter was quite low, we observed it above background in most of the experiments. This result is consistent with the genome-wide chromatin association study of MLL1 reported previously (Guenther et al., 2005). The trimethylation of H3K4 on the promoters of multiple genes were also studied in wild-type and *Mll1*<sup>-/-</sup> cells. All of the promoters were had H3K4 trimethylation, which did not show a significant reduction in *Mll1*<sup>-/-</sup> cells compared that of with the wild type (Fig. 5B). This



**Fig. 5. MLL1 regulates the increased H3K4 trimethylation on the *Nfkbia* (IκB $\alpha$ ) promoter induced by TNF $\alpha$ .** (A) p65 was knocked down in the F-Mll1 MEF cell line by siRNA. A ChIP assay was performed to study the amount of F-Mll1 on chromatin. MLL1 bound to the promoters of *Nfkbia* and *Sod2* in the untreated cell. After knocking down p65, the bound F-Mll1 on the *Nfkbia* promoter was reduced in comparison with that in the control. (B) A ChIP assay indicated that, in wild-type cells, TNF $\alpha$  induced the increase of H3K4 trimethylation on the promoters of *Mll1*-dependent genes [*Nfkbia* (IκB $\alpha$ ), *Il6*, *Tnfaip3* (A20) and *Mcp1*]. In the *Mll1*<sup>-/-</sup> cell, the basal level of H3K4 trimethylation was the same, but the induction of methylation was impaired. The H3K4 trimethylation on *Mll1*-independent genes (*Sod2* and *TNIP1*) promoter was still elevated in the absence of MLL1. (C) H3K27 trimethylation of *Nfkbia* and *Sod2* promoters decreased with TNF $\alpha$  treatment in wild type and *Mll1*<sup>-/-</sup> MEF cells. The *P* value of significantly different results is indicated above the corresponding columns.

is also consistent with our previous report (Wang et al., 2009). Considering that the mRNA of the two genes did not reduce upon *Mll1* deficiency, it seems that MLL1 just pre-binds the promoters and has no function without stimulation.

**p65 is indispensable for MLL1 recruitment to NF-κB target genes**

Because MLL1 pre-binds the promoters of genes downstream of NF-κB, we questioned whether the cytoplasmic MLL1 is still required for the gene activation. We performed an MLL1 CHIP assay to study MLL1 occupancy on promoters after TNF $\alpha$  treatment. MEF cells were treated with TNF $\alpha$  and cells were harvested 30 minutes later. The result showed that the amount of MLL1 bound to both the *Nfkbia* (encoding IκB $\alpha$ ) and *Sod2*

promoters increased substantially (Fig. 5A). When p65 was knocked down by siRNA, the amount of MLL1 bound to the *Nfkbia* promoter substantially reduced (Fig. 5A). However, there was no obvious difference in the amount of MLL1 on the *Sod2* promoter with or without p65 (Fig. 5A). These results suggest that the translocation of MLL1 to the *Nfkbia* promoter is dependent on p65, but that other factors might mediate MLL1 translocation to the *Sod2* promoter.

### MLL1 mediates the induced H3K4 methylation on the *Nfkbia* promoter

MLL1 is a histone H3K4 methyltransferase and, hence, we analyzed the levels of histone H3K4 trimethylation on promoters. In wild-type cells, the H3K4 trimethylation on the *Nfkbia* promoter increased dramatically upon TNF $\alpha$  treatment. In *Mll1*<sup>-/-</sup> cells, the basal level of H3K4 trimethylation at this promoter was the same as that of wild-type cells, but after TNF $\alpha$  treatment, the increase in H3K4 trimethylation was much less than that of wild-type cells (Fig. 5B). Other MLL1-dependent genes also showed similar results (Fig. 5B). This suggests that MLL1 is not required for the basal level of H3K4 trimethylation on the *Nfkbia* promoter, but is necessary for the trimethylation induced by TNF $\alpha$ . In the case of the *Sod2* and *Tnip1* promoters, the H3K4 trimethylation still increased in *Mll1*<sup>-/-</sup> cells upon TNF $\alpha$  treatment (Fig. 5B), which suggests other enzymes might regulate their H3K4 trimethylation levels.

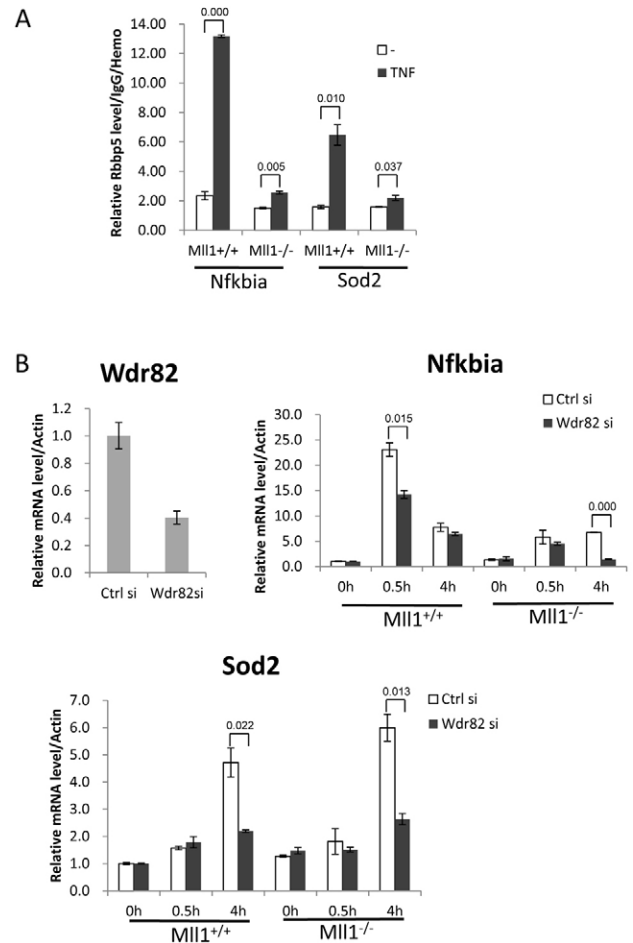
H3K27 trimethylation was also analyzed on the *Nfkbia* and *Sod2* promoters. When treated with TNF $\alpha$ , the amount of the H3K27 trimethylation reduced dramatically on both promoters, which suggests that MLL1 functions independently of the regulation of H3K27 trimethylation on these sites (Fig. 5C). We also analyzed H3K9 dimethylation on these promoters and no obvious signals were detected with or without TNF $\alpha$ . This suggests that H3K9 dimethylation might not be involved in the regulation of *Nfkbia* and *Sod2* expression.

### The H3K4 methyltransferases synergistically regulate the activation of the NF- $\kappa$ B signaling pathway

Given that the H3K4 trimethylation on the *Sod2* promoter did not change in *Mll1*<sup>-/-</sup> cells, we were aware that there might be other H3K4 methyltransferases involved in the process. The closest homologs of *Mll1* include the five genes *Setd1a*, *Setd1b*, *Mll2* (also called Wbp7), *Mll3* and *Mll4* (also called Alr). It is possible that one or several of these five genes is responsible for the H3K4 trimethylation on *Sod2* promoter. We analyzed the bound RBBP5 and WDR5 proteins on the promoters. RBBP5 and WDR5 are two subunits that are common to all of the six methyltransferase complexes. RBBP5 was recruited to both *Nfkbia* and *Sod2* promoters upon TNF $\alpha$  treatment in wild-type cells. In *Mll1*<sup>-/-</sup> cells, RBBP5 was still recruited to the promoters but the amount was greatly reduced (Fig. 6A). WDR5 behaved very similarly to RBBP5 in the above cell lines (supplementary material Fig. S7). This suggests that some other methyltransferases might also be involved.

### SETD1A and SETD1B regulate the expression of *Nfkbia* and *Sod2*

We investigated the functions of *Mll2*, *Setd1a* and *Setd1b* in NF- $\kappa$ B signaling by using RNA interference. Small interfering RNAs (siRNAs) were transfected into MEFs and, 72 hours after transfection, cells were treated with TNF $\alpha$  before harvesting.



**Fig. 6. SETD1A regulates the activation of genes downstream of NF- $\kappa$ B.** (A) ChIP results showing that the increased amount of RBBP5 on *Nfkbia* ( $\text{I}\kappa\text{B}\alpha$ ) and *Sod2* promoters was less in *Mll1*<sup>-/-</sup> cell than in wild-type cells. However, this level still increased slightly upon TNF $\alpha$  treatment. (B) *Wdr82* was knocked down by siRNA in wild-type MEFs (top left), which mimics deficiency of *Setd1a*. The basal mRNA levels of *Nfkbia* (top right) and *Sod2* (bottom) were not changed upon *Wdr82* deficiency. However, the elevation of their mRNA after TNF $\alpha$  stimulation was significantly impaired. The *P* value of significantly different results is indicated above the corresponding columns.

*Mll2* deficiency did not affect the basal level or activation of genes downstream of NF- $\kappa$ B (supplementary material Fig. S8). Because *Wdr82* deficiency is known to lead to a reduction of SETD1A protein, knockdown *Wdr82* will mimic the effect of SETD1A and/or SETD1B reduction (Wu et al., 2008). siRNA against *Wdr82* was transfected into MEFs, and similar experiments to those described above were performed. The knockdown of *Wdr82* did not affect the basal level of NF- $\kappa$ B target gene expression; however, both the *Nfkbia* and *Sod2* genes showed impaired expression upon treatment with TNF $\alpha$  (Fig. 6B). By using specific siRNAs against the methyltransferases, we also confirmed that SETD1A and SETD1B were involved in the transcriptional activation of *Nfkbia* and *Sod2* (supplementary material Fig. S9). These data suggest that SETD1A and SETD1B, but not MLL2, also regulate the activation of genes downstream of NF- $\kappa$ B.

### Discussion

The NF- $\kappa$ B signaling pathway can be activated by numerous upstream signals. Usually, the activated gene expression patterns

vary depending on the signals and cell lines, but the mechanisms involved are not clear. Epigenetics has emerged as a key player that is involved in regulation of gene transcription and cell identity. Different combination of chromatin modifications and their enzymes might be crucial in 'tuning' gene expression.

MLL1 and SETD1A are two well-studied histone H3K4 methyltransferases in mammals. The role of MLL1 in leukemogenesis has been extensively studied; however, its functions in other physiological events require more work. Previously, MLL1 has been reported to be associated with CLOCK and to regulate the oscillation of circadian gene expression. We found that MLL1 is associated with p65, the transactivation subunit of NF- $\kappa$ B. When the NF- $\kappa$ B pathway is activated, MLL1 is translocated onto the promoters of NF- $\kappa$ B target genes in a p65-dependent manner. Moreover, MLL1 deficiency also causes the change in the pattern of I $\kappa$ B $\alpha$  oscillation. Interestingly, MLL1 only regulates a subgroup of genes downstream of NF- $\kappa$ B. SETD1A and SETD1B also seem to be important in the regulation of gene transcription. These methyltransferases might act synergistically in gene regulation and it is possible that all the six members behave in a similar manner. The different combination of H3K4 methyltransferases might be another regulatory step in selective transcriptional regulation.

Surprisingly, none of the H3K4 methyltransferases tested in this study regulated the basal expression of *Nfkb* and *Sod2*. Therefore, further experiments are required to determine whether any other H3K4 tri-methyltransferases, such as MLL5, are required for the basal level expression of *Nfkb* and *Sod2*. Another possibility is that the regulatory step of H3K4 trimethylation is not necessary for some genes. Set1 is the only H3K4 methyltransferase in *S. cerevisiae*; however, the  $\Delta$ set1 strain is still viable under certain conditions. This means many genes are still transcribed without H3K4 methylation. It is possible that although H3K4 trimethylation is always accompanied with active gene transcription, it is dispensable for some genes under certain circumstances.

Most of the previous studies on MLL1 focused on its role in the nucleus, such as in histone modification and transcriptional regulation. We found that MLL1 exists both in the cytoplasm and nucleus. Moreover, MLL1 is still associated with RBBP5 in the cytoplasm, which suggests that the active catalytic complex might exist in both fractions. Many methyltransferases have been reported to be able to modify multiple subunits. We performed an in vitro histone methyltransferase (HMT) assay with <sup>3</sup>H-labeled S-adenosylmethionine (SAM) and purified proteins. Although we did not detect a catalytic activity of MLL1 against p65, it is still possible that MLL1 might have other substrates, both inside and outside of the nucleus.

Previous studies reported that MLL1 binds to a large number of gene promoters, but that the methylation status and gene expression profile was affected too much by its absence. If this is the case, then why does MLL1 bind to these regions? We found that MLL1 binds to the promoter of *Nfkb* and does not affect its expression and methylation status before activation. However, upon activation, the amount of MLL1 on the *Nfkb* promoter increased immediately, as did the level of H3K4 trimethylation. The behavior of MLL1 is very much like the poised RNA polymerase II (RNA pol II). It has been reported that RNA pol II is associated with some silenced genes. However, when the transcription program is initiated, the amount of RNA pol II on the promoter increases dramatically and leads to rapid

transcription of the gene. We hypothesized that the poised MLL1 pre-binds chromatin and will methylate H3 immediately upon gene activation, which facilitates a quick response to signal transduction. A previous report categorized TNF $\alpha$ -induced genes into three groups (Zhou et al., 2003). The first group elevates rapidly and decreases fast, such as *Il6* and *Cxcl1*; the second elevates rapidly and has sustained upregulation, such as *Nfkb* and *Tnfrsf3* (A20); the third elevates slowly, such as *Sod2*. The rapid elevated genes were all *MLL1*-dependent genes in our experiments, which supports our hypothesis.

## Materials and Methods

### Cell lines and antibodies

MEF wild type, MLL1 knockout and F-MLL1 cell lines were a gift from Jay L. Hess (University of Pennsylvania). HL-7702 cell line was purchased from the Cell Bank of Chinese Science Academy. The antibodies against Flag (Sigma), trimethylated H3K4 (Millipore), trimethylated H3K27 (Millipore), H3 (Abcam), NF- $\kappa$ B p65 (Abcam), WDR5 (Bethyl) and RBBP5 (Bethyl) were purchased from indicated companies. The siRNAs against p65 (5'-GCGACAAGGTGCAGAAAGA-3'), Wdr82 (5'-AGAGAACCUGUACAGUAA-3'), MLL1 (5'-GGACAAGAGTAGAGAGA-3') and MLL2 (5'-AGGAGAAGGAAGGCAAA-3') were synthesized by Genepharma. The PCR primers are available in the supplementary material Table S1.

### Cell fractionation

Cells were harvested and spun down in cold PBS. Approximately 5 volumes of buffer A (10 mM Hepes pH 7.9, 1.5 mM MgCl<sub>2</sub>, 10 mM KCl, 0.5 mM DTT, plus proteinase inhibitors; compared with the volume of the cell pellet) was added to the cells, which were then incubated on ice for 15 minutes. Cells were spun down and the supernatant was discarded. Another 2 volumes of buffer A was added to the pellet and the resuspended cells were homogenized with a homogenizer (Wheaton). The mixture was then centrifuged at 25,000 g and the supernatant was taken as the cytoplasm fraction. The pellet was resuspended in buffer C (20 mM Hepes pH 7.9, 25% glycerol, 1.5 mM MgCl<sub>2</sub>, 0.2 mM EDTA, 0.5 mM DTT, plus proteinase inhibitors) and a solution containing 5 M NaCl was dropped into the suspension followed by immediate homogenization. The final NaCl concentration was 0.42 M. The mixture was incubated on ice for 30 minutes and ultracentrifuged at 40,000 g for 1 hour. The resulting supernatant was taken as the soluble fraction of the nucleus and the pellet was the insoluble fraction of nucleus.

### Immunofluorescent staining

Cells were cultured on coverslips and fixed with frozen methanol after washing twice in PBS. The coverslips were then washed three times with PBS and blocked in PBS with 1% BSA for 10 minutes. The coverslips were hybridized with primary and second antibodies for 1 hour, respectively. Then the coverslips were mounted with prolong anti-fade kit (Invitrogen) and observed using fluorescent microscopy.

### Immunoprecipitation

The cells were harvested and lysed in NP40 lysis buffer (50 mM Tris-HCl pH 7.4, 150 mM NaCl, 0.5% NP40) or high-salt lysis buffer (20 mM Hepes pH 7.4, 10% glycerol, 0.35 M NaCl, 1 mM MgCl<sub>2</sub>, 0.5% Triton X-100, 1 mM DTT) with proteinase inhibitors. The supernatant was then incubated with Protein G beads (GE Healthcare) and the desired antibody at 4°C for 4 hours. The beads were spun down and washed three times with lysis buffer. The final drop of wash buffer was vacuumed out and SDS loading buffer was added to the beads, which were then used for western blotting.

### Luciferase reporter assay

HEK293T cells (approximately  $1 \times 10^5$ ) were seeded on 24-well plates and transfected by calcium phosphate precipitation. The reporter plasmid (pRL-TK or pRL-SV40 *Renilla* luciferase) was added to each transfection to normalize for transfection efficiency. A dual-specific luciferase assay kit (Promega) was used for the luciferase assays. Assays were repeated at least three times. Data shown are means  $\pm$  s.d. for one representative experiment.

### ChIP assay

The ChIP assay was performed as previously described (Wu et al., 2008). Briefly,  $\sim 1 \times 10^7$  cells were fixed with 1% formaldehyde and quenched by glycine. The cells were washed three times with PBS and then harvested in ChIP lysis buffer (50 mM Tris-HCl pH 8.0, 1% SDS, 5 mM EDTA). DNA was sonicated to 400–600 bp before extensive centrifugation. Four volumes of ChIP dilution buffer (20 mM Tris-HCl, pH 8.0, 150 mM NaCl, 2 mM EDTA, 1% Triton X-100) was added to the supernatant. The resulted lysate was then incubated with Protein G beads and antibodies at 4°C overnight. The beads were washed five times, and



DNA was eluted using the Chip elution buffer (0.1 M NaHCO<sub>3</sub>, 1% SDS, 30 µg/ml proteinase K). The elution was incubated at 65°C overnight, and DNA was extracted with a DNA purification kit (Sangon). The purified DNA was assayed by quantitative PCR with Biorad MyIQ. Assays were repeated at least three times. Data shown are means±s.d. for representative experiments.

#### Reverse transcription and quantitative PCR

Cells were scraped down and collected by centrifugation. Total RNA was extracted using an RNA extraction kit (Yuanpinghao) according to the manufacturer's manual. Approximately 1 µg of total RNA was used for reverse transcription with a first-strand cDNA synthesis kit (Toyobo). The amount of mRNA was assayed by quantitative PCR. β-actin was used to normalize the amount of each sample. Assays were repeated at least three times. Data shown are means±s.d. for one representative experiment.

#### ELISA

Experiments were carried out with an IL6 ELISA kit (Boster), according to the manufacturer's procedure. Assays were repeated at least three times. Data shown are means±s.d. for representative experiments.

#### Next generation sequencing and data analysis

The cells were treated with TNFα for 4 hours before collection. Total RNA was extracted and reverse transcribed. Then, the cDNA were analyzed by Sinogenomax CO. The raw reads containing low-quality data were cleaned by removing those contain either a base of N or over half qualities below 20. Then, the resulting clean reads were mapped to the mouse mRNA sequences with TopHat software. The RPKM value, which is the normalized number of reads of each mRNA, was calculated and used as the expression level. Genes expressed differently between every two samples were analyzed by the DESeq R package using a cutoff of  $P < 0.01$ . To study the relationship of the differentially expressed genes, the values of selected genes were submitted for cluster analysis by using Cluster3.0 and the heatmap was presented using Java Treeview.

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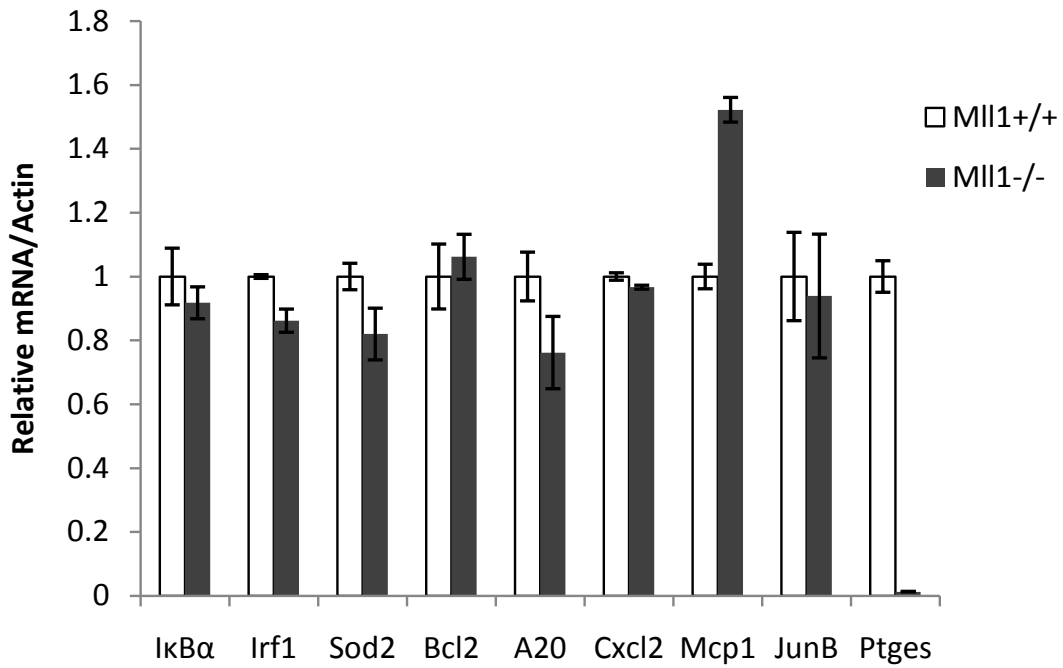
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<http://jcs.biologists.org/lookup/suppl/doi:10.1242/jcs.103531/-DC1>

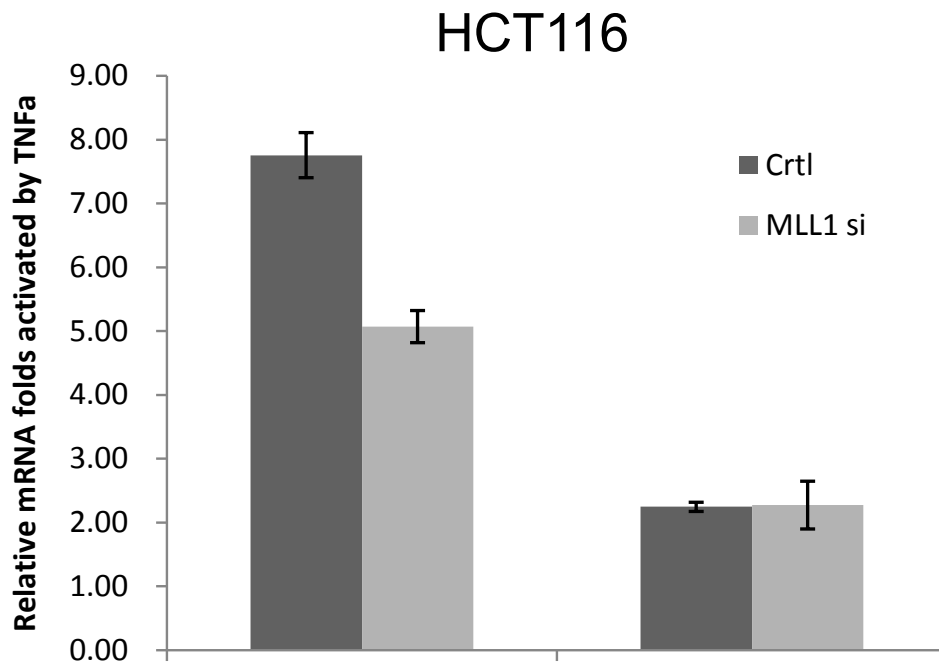
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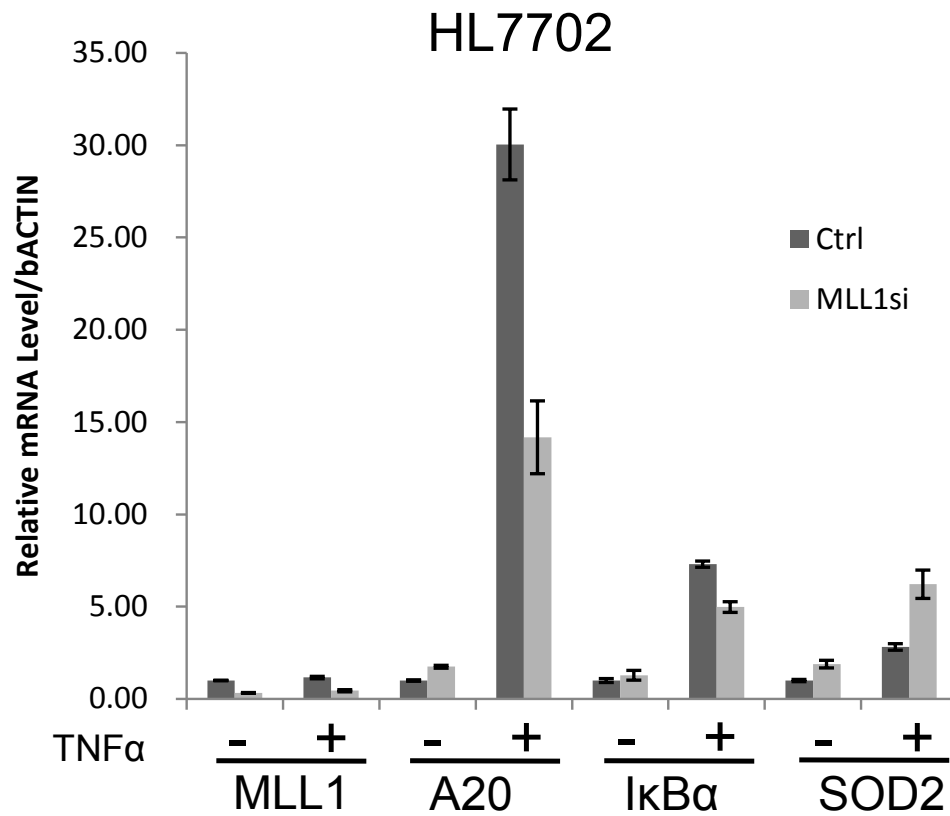


**Sup. Fig. 1** The basal mRNA level of NF-κB targeting genes in wide type and *Mll1*<sup>-/-</sup> cells.

The wide type and *Mll1*<sup>-/-</sup> cell were harvested and RT-PCR was performed to analyze the expression of indicated genes. Totally 40 reported NF-κB targeting genes were assayed (only part of data were presented). Among them, the expression of Ptges was significantly reduced.

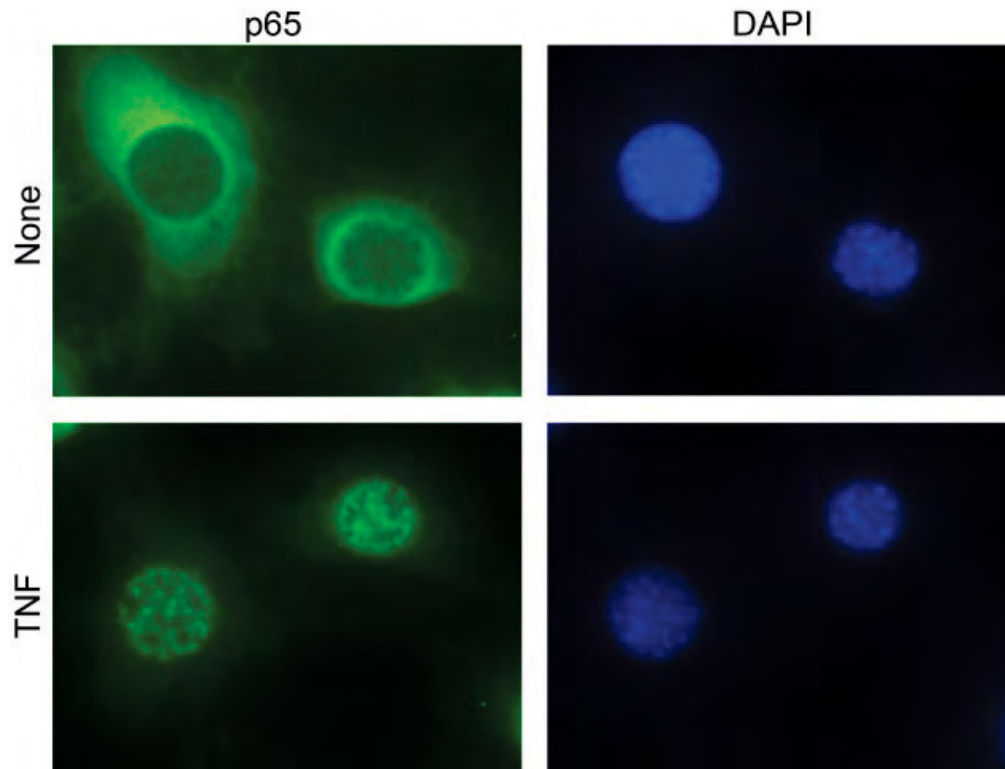


**Sup. Fig. 2 MLL1 regulated the activation of I $\kappa$ B $\alpha$  in HCT116 cell.** HCT116 cell was transiently transfected with MLL1 siRNA and treated by TNF $\alpha$  72hr after transfection. The elevated levels of I $\kappa$ B $\alpha$  and SOD2 mRNA were monitored by real time PCR.

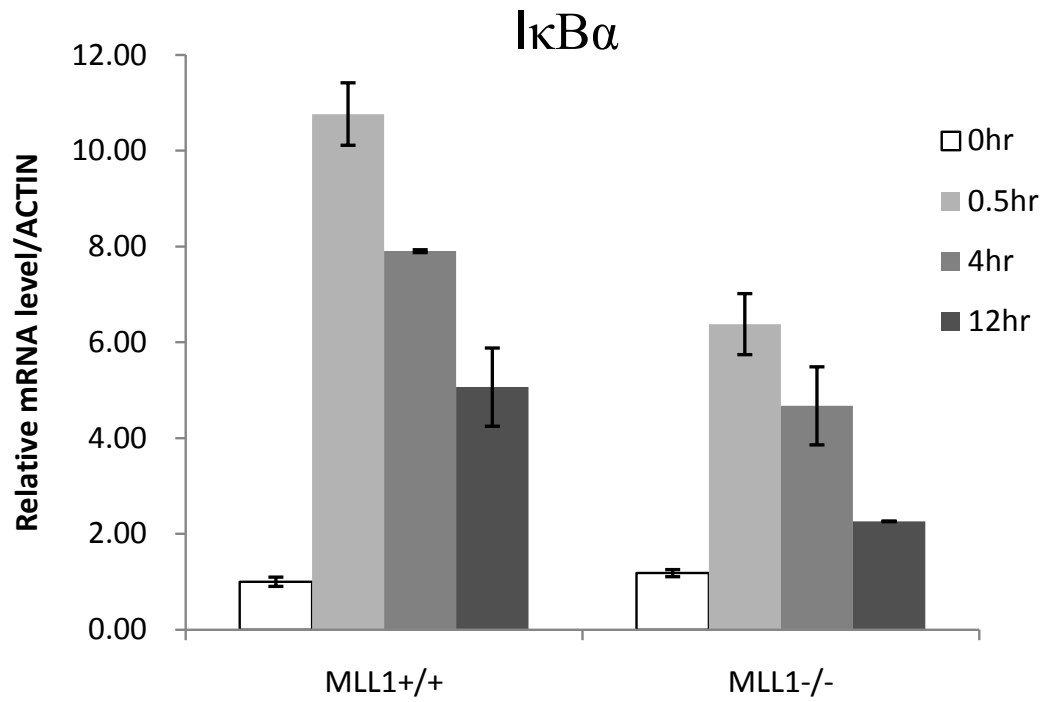


**Sup. Fig. 3 MLL1 regulated the activation of NFκB downstream genes in HL7702 cell.**

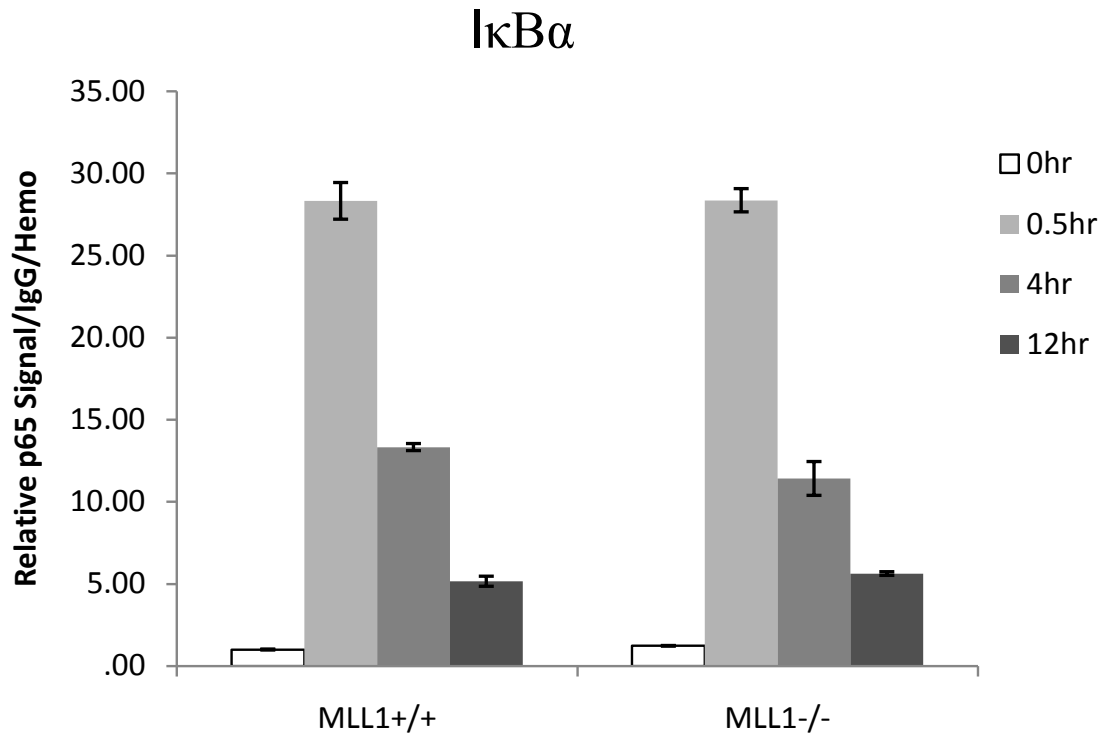
Immortalized liver cell HL7702 was transiently transfected with MLL1 siRNA and treated by TNFα 72hr after transfection. The elevated mRNA levels of indicated genes were assayed by real time PCR.



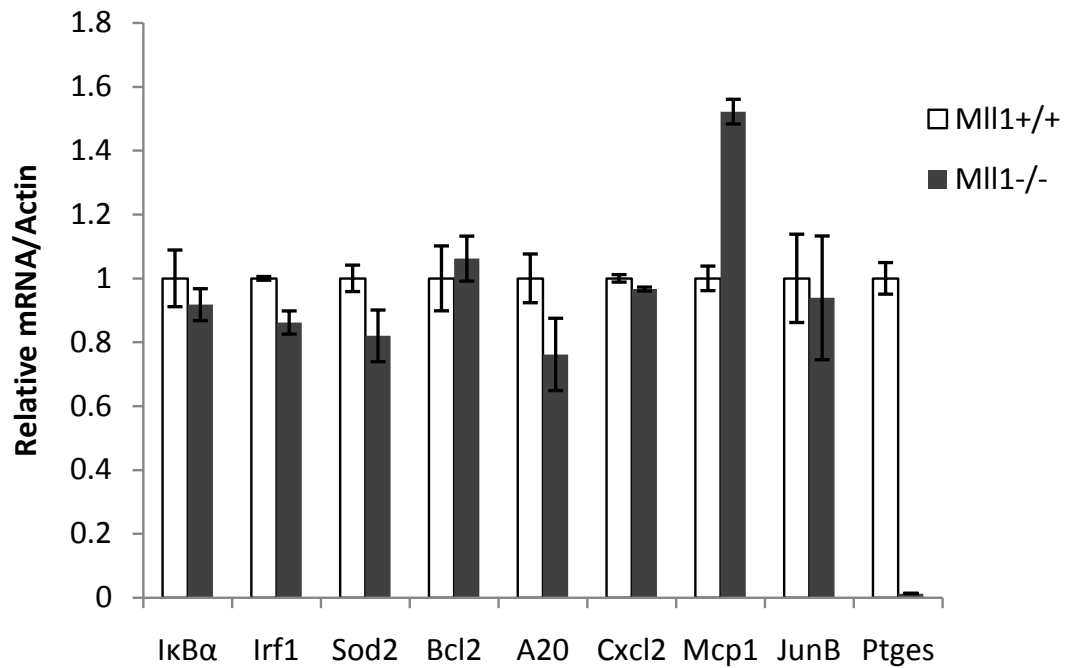
**Sup. Fig. 4 NF- $\kappa$ B was translocated into nuclear in *Mll1*<sup>-/-</sup> cell.** Immunofluorescent staining experiment was performed to study the localization of p65 in *Mll1*<sup>-/-</sup> cell. p65 is localized in the cytoplasm (upper) and translocated into nuclear (lower) after TNF $\alpha$  treatment, which is no difference from wide type cell.



**Sup. Fig. 5** The *IκBα* mRNA level at different time points after TNF $\alpha$  treatment. *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> cells were treated with TNF and harvested at 0hr, 0.5hr, 4hr and 12hr. The *IκBα* mRNA levels were analyzed.



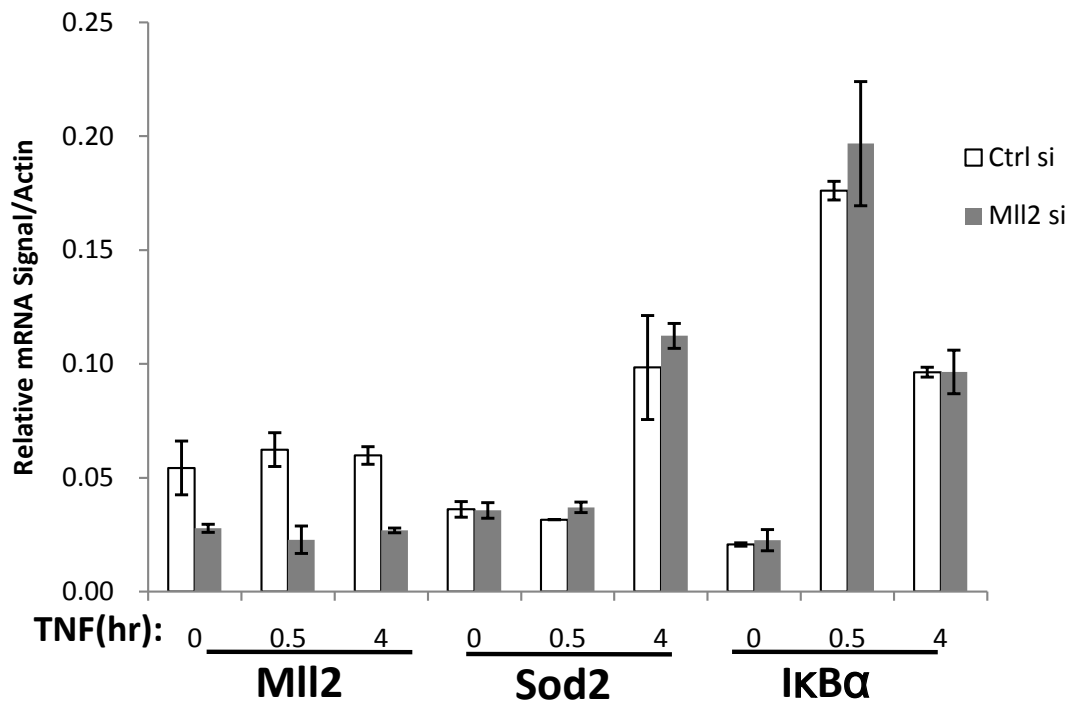
**Sup. Fig. 6 The bound p65 amount on  $\text{I}\kappa\text{B}\alpha$  promoter at different time points after  $\text{TNF}\alpha$  treatment.**  $\text{Mll1}^{+/+}$  and  $\text{Mll1}^{-/-}$  cells were treated with TNF and harvested at 0hr, 0.5hr, 4hr and 12hr. The amount of p65 protein on  $\text{I}\kappa\text{B}\alpha$  promoter was analyzed by CHIP assay.



**Sup. Fig. 7** The amount of Wdr5 on Ikbα increased upon TNFα treatment in *Mll1*<sup>-/-</sup> cell.

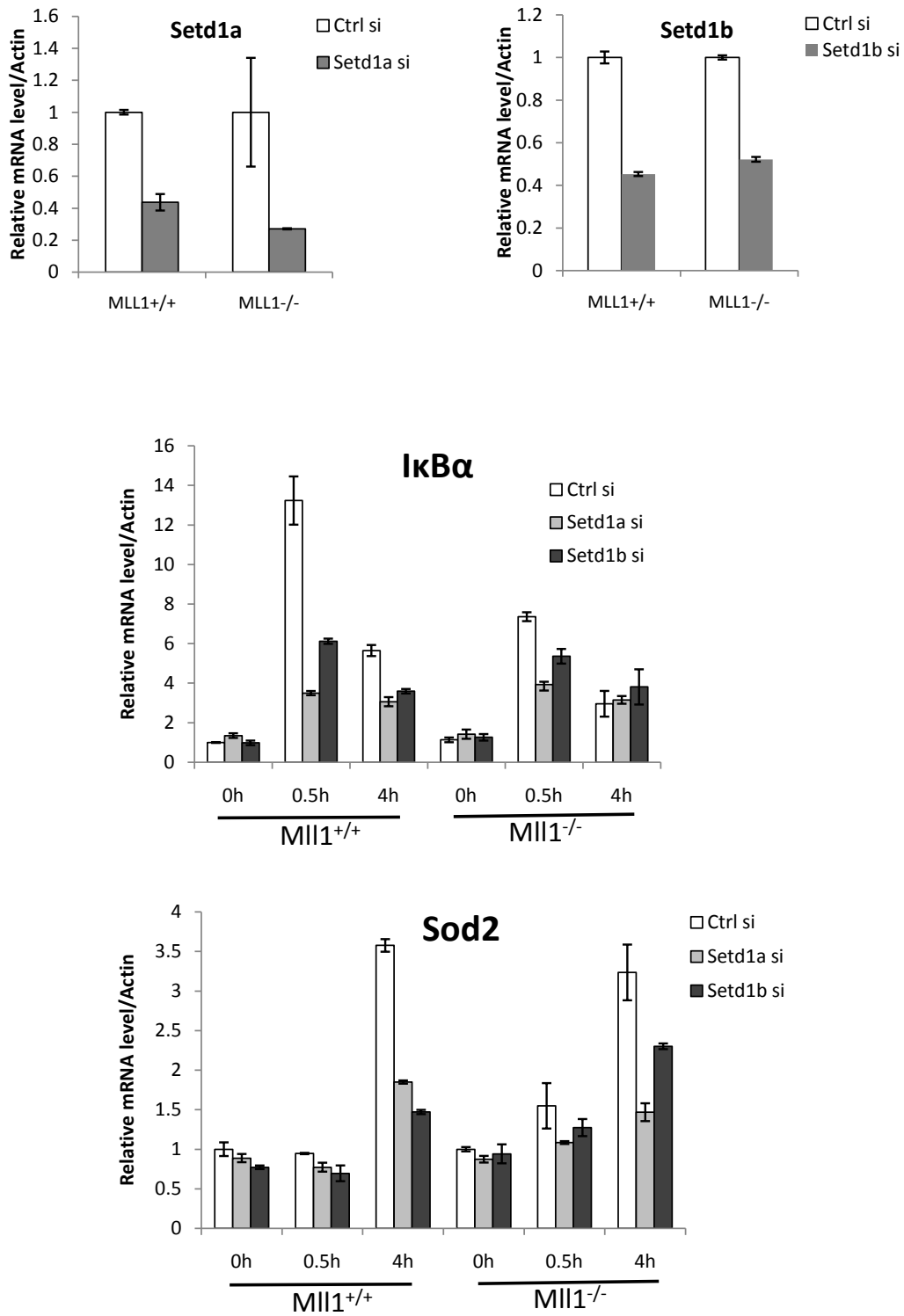
ChIP assay was performed to analyze the amount of Wdr5 on Ikbα promoter. Wdr5 bound Ikbα promoter in wide type MEF and increased upon TNFα treatment. The bound Wdr5 decreased in *Mll1*<sup>-/-</sup> cell, but still increased after treatment.





**Sup. Fig. 8 Mll2/HRX2 did not regulate the expression of *Ikbα* and *Sod2* in MEF cell.**

Mll2 was knocked down by siRNA in MEF cell and the mRNA levels of indicated genes were assayed by quantitative PCR following reverse transcription.



**Sup. Fig. 9** Setd1a and Setd1b regulated the expression of the expression of *IκBα* and *Sod2* in MEF cell. *Setd1a* and *Setd1b* were knocked down in *Mll1*<sup>+/+</sup> and *Mll1*<sup>-/-</sup> cells by siRNA respectively. The up panels showed the knockdown effect assayed by real time PCR. The middle and bottom showed that the expression of both *IκBα* (middle) and *Sod2* (bottom) were impaired with Setd1a or Setd1b deficiency.

## Supplementary Table 1

### The sequences of primers used in the study

Primers for RT PCR		
Gene	Forward primer	Reverse Primer
mb-Actin	aggatcatcactattggcaac	agaggtctttacggatgtca
mWdr82	CTCTAGCAGCGATGATGACT	GGTCCACACCGTACTTCTTA
mM111	ATGGGGAATGATGACAAGT	GGTGACAGGCTTAATTGGT
mM112	AGCACAGTGTGGAGCTG	AGCTGGTACAGAAGAGCAAG
mM113	CAAGCCTTATTTGATTCCAC	GTTCTTCCATTTGGCATATT
mM114	CATGGTGCCTGAAGATGT	TCTCTGATGCTGATGACGTA
mSet1A	GTCATGGGCAACATCATT	TGAGGAGTGTAAAGCCATT
mp65	ATGGCTACTATGAGGCTGAC	GTCTCGCTTCTTCACACACT
mB94	ATACCTACTTGCTGCTGCTC	GACACCTTGAAGCTCCTGT
mPGES	TACAGGAGTGACCCAGATGT	CGAGGAAGAGGAAAGGATAG
mikBa	TCCTCAACTCCAGAACAAC	GGGTATTTCTCGAAAGTCT
mSOD2	ACACCATTTTCTGGACAAAC	CAAAGTCACGCTTGATAGC
mMCP1	CTGGAGCATCCACGTGTT	CATCTTGCTGGTGAATGAG
mIAP1	ATAGAACACGCCAAATGGT	GCTTGAATCTCATCAACAAAC
mCox2	GTACCCGGACTGGATTCTAT	GGCTTCAGCAGTAATTTGAT
mJunB	CAGCTACTTTTCGGGTCAG	GATCAAGCGCTCCAGTTC
mC-myc	tagtgctgcatgaggagac	caccacatcaatttcttctct
mIL6	cacatgttctctctggaaatc	catcgttgttcatacaatcag
mTap1	ACTCCTGCTTATCTTGATG	CTTGGGGCTCTCATACAG
mTAPBP	TCTACCTGGCTACGGTACAC	GTGCTGGTGTAGAGACTC
mTraf1	ATGAGAGGAGAATACGATGC	GGTTGTTCTGGTCAAGTAGC
mTraf2	ACCGCTACTGCTCCTTCT	TACAGGCCTTCATAGACACA
mTraf3	AAGCATCATCAAAGACAAGG	ATTCCGACAGTAGACCTGAA
mGch1	TAGTGATTGAAGCGACACAC	TGGTGTAGTGACAGTCTTG
mGfpt2	CCGAGGTTATGATGTTGACT	GGTGACGACAGTCTTGTGAT
mKlrc2	GCTGAACTGAAGAAGCAGAT	TTGGACAATGAGGACAAG
mIcam1	CACGCTACCTCTGCTCCT	GGATGGATGGATACCTGAG
mI127ra	TTCTGCTGTCGCTGATGT	GACCGACGCTGTAGCACT
mNFkB1	CTGAGTCCTGCTCCTTCTAA	CTGTGTAGCCCATCTGTTG
mNFkB2	TGGACACATACAGGAAGACC	CAATAGACCCACCAAGTCC
mcRel	AGAACTGTGGAAGTGTGAGG	GTCATTCAACACAAAACGAA
mTrim16	GAACTCAAGTGTCTGCCATT	TGTCACAGGCATATTGAAGA
mTnip1	CTGTCACCACCGACATCT	GTAGCAGGATGTACCTGGAC
mCyb5	GTCTAATTCCAGTTGGTGGA	AAGTCAATCTTCTGCCATGT

mPsmb9	ACGGAAGAAGTCCACACC	GAATCAGAGCCCACCAC
mCxc12	ACGGAAGAACCAAAGAGAA	AAATAAGTGAAGTCTCAGACAGC
mTNFa	TACTGAACTTCGGGGTGAT	GCCATAGAAGTATGAGAGG
mBID	CCTTCAACCAAGGAAGAATA	CGAGATGTCTGGCAATGT
mCtgf	GTGTGTGACGAGCCCAAG	GCCAAATGTGTCTTCCAGT
hb-Actin	GATCCACATCTGCTGGAAG	CAGCACAATGAAGATCAAGA
hWdr82	CTCCATCGTGCTCTATGACT	GATGAGGTCCACACCATATT
hMLL1	<b>cgggaaaagtattacgacag</b>	<b>cacacgagtgattgatgaag</b>
hMl12	TGTGCTCTATGCCAACATTA	TTCTCCAGAGCTTCATGATT
hMLL3	<b>gtttggagtatcgacagcat</b>	<b>tatccagaaagaccattgct</b>
hMl14	GACTGGAAAGCAGAAGTCTC	AGCCACACGTTCTCTAAATC
hSetd1A	<b>acagtgacctgctgaaactc</b>	<b>acgtattcgatgacctctc</b>
hSetd1b	<b>acagtgacctgctcaagttc</b>	<b>acgtactcgatgacctctc</b>
hp65	CTGGAGCAGGCTATCAGTC	ACAGCATTGAGTTCGATGTC
hB94	ACCTACATGCTGCTGCTC	CCATACCCTGCAGCTCAC
hIkBa	GTCTTGGGTGCTGATGT	GAGAATAGCCCTGGTAGGTAA
hSOD2	GCACGTTACTACCTTCAGT	CTCCAGTTGATTACATTCC
hI16	AACCTGAACCTTCAAAGAT	ACTCCAAAAGACCAGTGATG
hcIAP1	GCCATCTAGTGTCCAGTTC	AGCATTTGACATCATCATTG
hcIAP2	TCAAGTTCAAGCCAGTTACC	GACTCTGCATTTTCATCTCC
hCox2	CCAGACAAGCAGGCTAATAC	TGATAGCCACTCAAGTGTTG
hJunB	CGACGACTCATACACAGCTA	TCGGTTTCAGGAGTTGTAG
hTap1	GTGGTCCTCTCCTCTTTG	TGAGCCATCTGTAGAAATCC
hTAPBP	GTCCCTGTCTCTGCTCT	CTCCACGAACCAACACTC
hTraf1	CCACCTCTATCCACCAGAG	CAGGGTCTGCTGAAGCTC
hTraf2	CAAGATTGAAGCCCTGAGTA	ATCTCCAAGACCTTCTGCTC
hTraf3	GGAGAGCGTGGACAAGAG	CACACTCAGCATCTGGTCAT
hGch1	CTACCAGGAGACCATCTCAG	TATGTCTTCACAATCACCA
hGfpt2	ACAAGCTCTCCACAGAACAG	CCTCTTCATCCGTGTCTTAC
hKlrc2	TATGACTGCCAAGGTTACTG	ATCAGGACAATGCAAATGAT
hIcam1	GTGACCATCTACAGCTTCC	TCACACTTACTGTACCTC
hI127ra	CTCTCACCAACCTCTCTTTG	ACCAGTAGCTCCGTGCTC
hNFkB1	TGGTATCAGACGCCATCTA	GCTGTCTGTCCATTCTTAC
hNFkB2	AATTGAACTCCTCCATTGTG	CCTCTCTGCTTAGGCTGTT
hcRel	TTCTGACCAGGAAGTTAGTGA	TTTGCTTTATTGCCGTAAGT
hTrim16	ACACCATAGTCTCCCTGGAT	GATGGCATTTCATTCAACT
hTnip1	AATGCAAGGGATAAAGATGT	TTGTCTTCACTAGCTCCTC
hCyb5	ACAAGGTGTACGATTTGACC	GTTCTCAGTAGCGTCACCTC
hPsmb9	ATCGAGAGGACTTGCTGTC	GGTTCCATATACCTGACCT
hCxc12	CAAGAACATCCAAAGTGTGA	CCATTCTTGAGTGTGGCTAT

hCtgf	GAGTGGGTGTGTGACGAG	CGTGTCTTCCAGTCGGTA
Primers for ChIP		
Name	Forward	Reverse
mHemob	CTTTGGGCATCTAGCTTTTA	AATCCTTGCAAGAAACAAAA
mIkBapro	GAGGACTTTCAGCCACTCA	GCTCGTCCTCCACTGAGAAG
mSOD2proF	AGTCTCAGGGGCAACAAAGA	GCCCCTCTGACCCAGTTAAT
mI16pro	gggatgtctgtagctcattc	gcagagaggaacttcatagc
mTnip1pro	CCTGCAGAAGCTCAGAAA	ACGAGGTGATCTGAAGATGT
mA20pro	catggatgtgacgtggaa	cggagaaactcctaggtc
mMcp1pro	ccacagtttctctcttccac	atcacctggataagtgatg

Supplementray Table 2 Differential expressed genes of MEF wide type cell with or without TNF  $\alpha$ 

ID	ACCESSION NUMBER	NAME	TNF	Ctrl	FOLDCHANGE
17260	NM 001170537.1	myocyte enhancer factor 2C	0.0000	0.0449	0.0000
73887	XR 106601.1	RIKEN cDNA 4930417022 gene	0.0000	0.2437	0.0000
73887	XR 106411.1	RIKEN cDNA 4930417022 gene	0.0000	0.2437	0.0000
100504427	XR 104672.1	predicted gene 13404	0.0182	0.2388	0.0762
100504427	XR 107364.1	predicted gene 13404	0.0183	0.2211	0.0826
78445	XR 106969.1	RIKEN cDNA C330013E15 gene	0.1018	0.7192	0.1416
78445	XR 105710.1	RIKEN cDNA C330013E15 gene	0.1018	0.7192	0.1416
442801	NM 177566.3	Rho guanine nucleotide exchange factor (GEF) 15	0.0255	0.1455	0.1749
69903	NM 028544.1	Ras interacting protein 1	0.0564	0.2390	0.2360
100342	NM 175307.6	family with sequence similarity 46, member B	0.1406	0.5044	0.2788
56379	NM 019659.3	potassium inwardly-rectifying channel, subfamily J, member 1	0.0931	0.3286	0.2832
56379	NM 001168354.1	potassium inwardly-rectifying channel, subfamily J, member 1	0.0930	0.3284	0.2832
11606	NM 007428.3	angiotensinogen (serpin peptidase inhibitor, clade A, member 8)	0.1926	0.6801	0.2832
14747	NM 008153.3	chemokine-like receptor 1	0.8669	2.5226	0.3437
22403	NM 016873.2	WNT1 inducible signaling pathway protein 2	0.4007	1.0714	0.3740
11754	NM 009675.2	amine oxidase, copper containing 3	0.1870	0.4676	0.3999
100503611	XM 003084596.1	-	0.6688	1.4808	0.4516
100039781	NM 027511.1	histidine rich carboxyl terminus 1	2.1003	4.5465	0.4620
64929	NM 022886.2	sciellin	0.6556	1.2876	0.5092
22417	NM 009523.1	wingless-related MMTV integration site 4	2.9543	5.6667	0.5213
56745	NM 019959.2	Clq and tumor necrosis factor related protein 1	1.6079	3.0580	0.5258
100042198	XR 104937.1	predicted gene 3716	0.9085	1.7203	0.5281
100042198	XR 107688.1	predicted gene 3716	1.0012	1.8815	0.5321
15902	NM 010496.3	inhibitor of DNA binding 2	12.2842	22.8301	0.5381
105844	NM 130859.2	caspase recruitment domain family, member 10	1.7540	3.2205	0.5446
70788	NM 027551.2	kelch-like 30 (Drosophila)	2.9826	5.4573	0.5465
23882	NM 011817.2	growth arrest and DNA-damage-inducible 45 gamma	12.7631	23.2863	0.5481
68169	NM 172399.3	RIKEN cDNA A930038C07 gene	3.8765	7.0064	0.5533

14181	NM 008009.3	fibroblast growth factor binding protein 1	3.7718	6.8084	0.5540
329540	NM 001001986.2	RIKEN cDNA 8430427H17 gene	1.1553	2.0751	0.5567
13653	NM 007913.5	early growth response 1	1.1868	2.1249	0.5585
12159	NM 007554.2	bone morphogenetic protein 4	4.9474	8.7537	0.5652
71093	NM 153778.3	atonal homolog 8 (Drosophila)	4.1587	7.3353	0.5669
17130	NM 008542.3	MAD homolog 6 (Drosophila)	2.3821	4.1757	0.5705
329540	NM 001134300.2	RIKEN cDNA 8430427H17 gene	1.0585	1.8549	0.5706
68404	NM 153529.1	neuritin 1	9.8921	17.1447	0.5770
76969	NM 023850.2	carbohydrate (keratan sulfate Gal-6) sulfotransferase 1	10.1686	17.5478	0.5795
231861	NM 001122730.1	trinucleotide repeat containing 18	3.2867	5.5961	0.5873
223650	NM 144848.2	epiplakin 1	0.6346	1.0706	0.5928
231861	NM 178242.2	trinucleotide repeat containing 18	2.4112	4.0547	0.5947
271221	NM 198642.2	RIKEN cDNA 5031414D18 gene	3.7752	6.2986	0.5994
74760	NM 144538.2	RAB3A interacting protein (rabin3)-like 1	5.1150	8.3631	0.6116
380912	NM 199029.2	zinc finger protein 395	2.6201	4.2724	0.6133
14200	NM 010212.3	four and a half LIM domains 2	13.6753	22.2197	0.6155
15426	NM 010466.2	homeobox C8	6.8197	11.0521	0.6171
77889	NM 029999.4	limb-bud and heart	3.1456	5.0193	0.6267
19252	NM 013642.3	dual specificity phosphatase 1	15.8758	25.1264	0.6318
70445	NM 054042.2	CD248 antigen, endosialin	11.5292	18.1946	0.6337
21825	NM 011580.3	thrombospondin 1	51.2370	80.6369	0.6354
16582	NM 010631.2	kinesin family member C3	4.3592	6.7345	0.6473
16582	NM 001145831.1	kinesin family member C3	4.1775	6.4013	0.6526
16582	NM 001145832.1	kinesin family member C3	4.7334	7.1883	0.6585
170676	NM 001040611.1	paternally expressed 10	3.8115	5.7253	0.6657
170676	NM 130877.2	paternally expressed 10	3.8115	5.7253	0.6657
67306	NM 173181.3	family with sequence similarity 164, member A	24.2472	36.0316	0.6729
104445	NM 027219.3	CDC42 effector protein (Rho GTPase binding) 1	18.9825	27.8494	0.6816
53881	NM 017391.3	solute carrier family 5 (inositol transporters), member 3	13.5437	19.4589	0.6960
18596	NM 001146268.1	platelet derived growth factor receptor, beta polypeptide	13.0414	18.5416	0.7034

18596	NM 008809.2	platelet derived growth factor receptor, beta polypeptide	13.0684	18.5585	0.7042
57265	NM 020510.2	frizzled homolog 2 (Drosophila)	11.0146	15.6363	0.7044
94242	NM 001168333.1	tubulointerstitial nephritis antigen-like 1	43.7188	61.8658	0.7067
14313	NM 008046.2	follistatin	26.5054	37.4681	0.7074
94242	NM 023476.3	tubulointerstitial nephritis antigen-like 1	40.5717	57.2028	0.7093
20410	NM 011366.2	sorbin and SH3 domain containing 3	18.2764	25.7422	0.7100
20346	NM 009152.3	sema domain, immunoglobulin domain (Ig), short basic domain, secr	11.6199	16.1153	0.7210
11826	NM 007472.2	aquaporin 1	78.5188	108.0738	0.7265
100503984	XR 107221.1	-	704.9197	524.9652	1.3428
75788	NM 001038627.1	SMAD specific E3 ubiquitin protein ligase 1	30.0263	22.2618	1.3488
75788	NM 029438.3	SMAD specific E3 ubiquitin protein ligase 1	29.9830	22.2181	1.3495
100503946	XR 107210.1	-	736.1257	543.3139	1.3549
15364	NM 010441.2	high mobility group AT-hook 2	151.6343	110.8795	1.3676
83946	NM 001081216.1	pleckstrin homology domain interacting protein	11.8800	8.6521	1.3731
328949	NM 001085373.1	mutated in colorectal cancers	10.4393	7.5425	1.3841
328949	NM 001085374.1	mutated in colorectal cancers	10.6870	7.7204	1.3843
19791	NR 003278.1	18S ribosomal RNA	499.2789	360.4836	1.3850
17118	NM 008538.2	myristoylated alanine rich protein kinase C substrate	27.3839	19.6516	1.3935
67073	NM 025951.2	phosphatidylinositol 4-kinase type 2 beta	21.1863	15.2003	1.3938
218454	NM 172589.2	lipoma HMGIC fusion partner-like 2	26.8897	19.2346	1.3980
67073	NM 028744.2	phosphatidylinositol 4-kinase type 2 beta	20.6771	14.7820	1.3988
215449	NM 024457.2	RAS related protein 1b	314.0320	224.2397	1.4004
16880	NM 013584.2	leukemia inhibitory factor receptor	10.2795	7.3299	1.4024
18828	NM 008880.2	phospholipid scramblase 2	56.8543	40.5030	1.4037
26921	NM 008696.2	mitogen-activated protein kinase kinase kinase kinase 4	39.3240	27.9976	1.4046
20348	NM 013657.5	sema domain, immunoglobulin domain (Ig), short basic domain, secr	60.1780	42.8094	1.4057
12977	NM 001113530.1	colony stimulating factor 1 (macrophage)	124.2404	87.9714	1.4123
226525	NM 177644.5	RAS protein activator like 2	4.7036	3.3242	1.4150
16476	NM 010591.2	Jun oncogene	19.2680	13.5904	1.4178
12977	NM 007778.4	colony stimulating factor 1 (macrophage)	159.7792	112.5273	1.4199



14182	NM 001079909.1	fibroblast growth factor receptor 1	20.2126	14.2323	1.4202
14182	NM 010206.2	fibroblast growth factor receptor 1	19.9201	13.9416	1.4288
24136	NM 015753.3	zinc finger E-box binding homeobox 2	6.3854	4.4639	1.4304
14182	NM 001079908.1	fibroblast growth factor receptor 1	19.8939	13.9006	1.4312
13486	NM 026106.4	down-regulator of transcription 1	29.4534	20.5493	1.4333
208846	NM 172464.2	dishevelled associated activator of morphogenesis 1	7.5518	5.2661	1.4341
12977	NM 001113529.1	colony stimulating factor 1 (macrophage)	163.4552	113.8839	1.4353
208846	NM 026102.2	dishevelled associated activator of morphogenesis 1	7.4237	5.1657	1.4371
17936	NM 008667.3	Ngfi-A binding protein 1	18.9093	13.1578	1.4371
78388	NM 080638.3	major vault protein	18.0370	12.4491	1.4489
20912	NM 011504.1	syntaxin binding protein 3A	13.6674	9.4132	1.4520
140780	NM 080708.1	BMP2 inducible kinase	6.9804	4.8030	1.4533
54216	NM 001122758.1	protocadherin 7	11.4650	7.8501	1.4605
21664	NM 009344.3	pleckstrin homology-like domain, family A, member 1	32.8883	22.4914	1.4623
12457	NM 009834.2	CCR4 carbon catabolite repression 4-like ( <i>S. cerevisiae</i> )	9.8646	6.7216	1.4676
22762	NM 011766.5	zinc finger protein, multitype 2	17.5195	11.8835	1.4743
20971	NM 011521.2	syndecan 4	59.1218	39.9927	1.4783
30953	NM 001113419.1	schwannomin interacting protein 1	11.1351	7.5303	1.4787
268396	NM 177364.3	SH3 and PX domains 2B	12.3986	8.3803	1.4795
15979	NM 010511.2	interferon gamma receptor 1	44.6664	30.1706	1.4805
16195	NM 010560.3	interleukin 6 signal transducer	62.4263	42.1626	1.4806
30953	NM 001113421.1	schwannomin interacting protein 1	11.9842	8.0936	1.4807
14042	NM 010162.2	exostoses (multiple) 1	51.4466	34.5395	1.4895
16423	NM 010581.3	CD47 antigen (Rh-related antigen, integrin-associated signal tran	49.6476	33.2580	1.4928
380711	NM 001015046.2	RAP1 GTPase activating protein 2	6.7583	4.5178	1.4959
18412	NM 011018.2	sequestosome 1	389.4593	260.1461	1.4971
23872	NM 011809.3	E26 avian leukemia oncogene 2, 3' domain	33.3136	22.1853	1.5016
21928	NM 009396.2	tumor necrosis factor, alpha-induced protein 2	28.1665	18.7552	1.5018
20496	NM 009194.3	solute carrier family 12, member 2	53.6863	35.5881	1.5085
30953	NM 001113420.1	schwannomin interacting protein 1	15.3918	10.1965	1.5095

30953	NM 013928.5	schwannomin interacting protein 1	14.5393	9.6318	1.5095
54216	NM 018764.2	protocadherin 7	21.5416	14.2519	1.5115
16362	NM 001159396.1	interferon regulatory factor 1	16.5818	10.9424	1.5154
18933	NM 001025570.1	paired related homeobox 1	10.5276	6.9171	1.5220
268515	NM 198423.3	BAH domain and coiled-coil containing 1	4.7554	3.1148	1.5267
71409	NM 172409.2	formin-like 2	8.1921	5.3624	1.5277
110521	NM 007772.2	human immunodeficiency virus type I enhancer binding protein 1	3.0667	1.9942	1.5379
52552	NM 001081009.1	poly (ADP-ribose) polymerase family, member 8	13.5426	8.7941	1.5400
18481	NM 008778.2	p21 protein (Cdc42/Rac)-activated kinase 3	7.0763	4.5827	1.5441
170737	NM 133206.3	zinc and ring finger 1	6.7605	4.3767	1.5447
16362	NM 008390.2	interferon regulatory factor 1	17.1987	11.0824	1.5519
102626	NM 178907.3	mitogen-activated protein kinase-activated protein kinase 3	9.9259	6.3906	1.5532
99382	NM 178890.3	ankyrin repeat and BTB (POZ) domain containing 2	5.5385	3.5475	1.5612
433022	NM 001134480.1	phosphatidylinositol-specific phospholipase C, X domain containing	2.1635	1.3833	1.5640
98376	NM 178883.5	golgin, RAB6-interacting	10.4566	6.6407	1.5746
100463512	NM 001190732.1	-	113.7580	72.2355	1.5748
18933	NM 175686.3	paired related homeobox 1	22.8892	14.5037	1.5782
52118	NM 027514.2	poliovirus receptor	18.4506	11.6816	1.5795
21356	NM 001025313.1	TAP binding protein	24.8124	15.6451	1.5859
12608	NM 009883.3	CCAAT/enhancer binding protein (C/EBP), beta	30.1458	19.0011	1.5865
21356	NM 009318.2	TAP binding protein	24.8915	15.6753	1.5879
18933	NM 011127.2	paired related homeobox 1	22.7663	14.3271	1.5890
14538	NM 133219.1	glucosaminyl (N-acetyl) transferase 2, I-branching enzyme	3.2357	2.0359	1.5893
67338	NM 001164569.1	ring finger and FYVE like domain containing protein	10.5859	6.6496	1.5920
170737	NM 001168622.1	zinc and ring finger 1	6.1730	3.8729	1.5939
16177	NM 008362.2	interleukin 1 receptor, type I	2.8510	1.7883	1.5942
12837	NM 007739.2	collagen, type VIII, alpha 1	32.0221	20.0786	1.5948
67338	NM 001164570.1	ring finger and FYVE like domain containing protein	10.7568	6.7268	1.5991
67338	NM 001007465.3	ring finger and FYVE like domain containing protein	11.0411	6.9021	1.5997
16177	NM 001123382.1	interleukin 1 receptor, type I	2.8175	1.7552	1.6053

67338	NM 026097.3	ring finger and FYVE like domain containing protein	10.6633	6.6350	1.6071
12192	NM 007564.5	zinc finger protein 36, C3H type-like 1	13.5355	8.4117	1.6091
20620	NM 152804.2	polo-like kinase 2 (Drosophila)	226.8847	140.7800	1.6116
67338	NM 001164571.1	ring finger and FYVE like domain containing protein	10.9996	6.8126	1.6146
15980	NM 008338.3	interferon gamma receptor 2	48.4175	29.8441	1.6223
18033	NM 008689.2	nuclear factor of kappa light polypeptide gene enhancer in B-cell	34.4185	21.1595	1.6266
58244	NM 021433.3	syntaxin 6	21.9780	13.5074	1.6271
231549	NM 178701.3	leucine rich repeat containing 8D	12.1910	7.4796	1.6299
231549	NM 001122768.1	leucine rich repeat containing 8D	12.1168	7.4340	1.6299
75234	NM 029219.1	ring finger protein 19B	20.7091	12.7049	1.6300
74194	NM 028810.2	Rho family GTPase 3	29.4216	17.9490	1.6392
192657	NM 138953.2	elongation factor RNA polymerase II 2	22.6277	13.6865	1.6533
22031	NM 001048206.1	TNF receptor-associated factor 3	10.4963	6.3396	1.6557
20352	NM 013659.4	sema domain, immunoglobulin domain (Ig), transmembrane domain (TM)	6.6758	4.0122	1.6639
14573	NM 010275.2	glial cell line derived neurotrophic factor	5.8571	3.5149	1.6663
22031	NM 011632.2	TNF receptor-associated factor 3	10.5627	6.3387	1.6664
16362	NM 001159393.1	interferon regulatory factor 1	10.8842	6.5247	1.6682
236285	NM 173414.3	LanC lantibiotic synthetase component C-like 3 (bacterial)	6.3554	3.8088	1.6686
75296	NM 201230.4	Fgfr1 oncogene partner	13.8136	8.2592	1.6725
114774	NM 054056.2	PRKC, apoptosis, WT1, regulator	17.2460	10.2983	1.6746
19288	NM 008987.3	pentraxin related gene	48.8220	29.0910	1.6783
320924	NM 178793.4	collagen and calcium binding EGF domains 1	11.1861	6.6199	1.6898
16155	NM 008349.5	interleukin 10 receptor, beta	17.5100	10.3314	1.6948
19225	NM 011198.3	prostaglandin-endoperoxide synthase 2	27.0702	15.9376	1.6985
19734	NM 011267.3	regulator of G-protein signaling 16	7.4345	4.3718	1.7005
75599	NM 029357.3	protocadherin 1	3.0870	1.8115	1.7041
387609	NM 199449.2	zinc fingers and homeoboxes 2	3.1821	1.8640	1.7072
672511	XM 001476651.2	ring finger protein 213	5.8450	3.4128	1.7127
672511	XM 001477846.2	ring finger protein 213	5.8450	3.4128	1.7127
100504142	XR 106553.1	predicted gene, 20084	240.3367	140.3107	1.7129

100504142	XR 105374.1	predicted gene, 20084	240.3367	140.3107	1.7129
380855	NM 001013769.1	regulator of sex limited protein 1	4.7169	2.7497	1.7154
260315	NM 001081035.1	neuron navigator 3	0.9002	0.5237	1.7191
100504290	XR 105381.1	predicted gene, 20152	21.0056	12.0925	1.7371
100504290	XR 106558.1	predicted gene, 20152	21.1812	12.1505	1.7432
100125931	NR 030676.1	RIKEN cDNA A130049A11 gene	3.6301	2.0793	1.7458
72747	NM 028341.4	tetratricopeptide repeat domain 39C	7.9645	4.5361	1.7558
15273	NM 010437.2	human immunodeficiency virus type I enhancer binding protein 2	5.4886	3.1134	1.7629
408065	NM 001001186.3	zinc finger protein 456	1.9103	1.0698	1.7856
229675	NM 172684.2	rosbin, round spermatid basic protein 1	5.7704	3.1845	1.8120
20308	NM 011338.2	chemokine (C-C motif) ligand 9	17.2538	9.4835	1.8193
26399	NM 011943.2	mitogen-activated protein kinase kinase 6	7.2214	3.9488	1.8287
53970	NM 017395.2	regulatory factor X, 5 (influences HLA class II expression)	2.0734	1.1278	1.8385
16477	NM 008416.2	Jun-B oncogene	10.6419	5.7279	1.8579
319622	NM 001033380.3	inositol 1,4,5-triphosphate receptor interacting protein-like 2	44.3796	23.7390	1.8695
73683	NM 001111111.1	autophagy related 16 like 2 (S. cerevisiae)	2.2595	1.2060	1.8736
231830	NM 174850.3	MICAL-like 2	28.4579	15.1649	1.8766
20656	NM 013671.3	superoxide dismutase 2, mitochondrial	28.3965	15.0895	1.8819
13874	NM 007950.2	epiregulin	17.2754	9.1729	1.8833
108767	NM 001033225.2	proline-rich nuclear receptor coactivator 1	9.7834	5.1865	1.8863
223881	NM 172612.3	Rho family GTPase 1	63.4554	33.5264	1.8927
75985	NM 029494.2	RAB30, member RAS oncogene family	7.7293	4.0790	1.8949
671535	NM 001163576.1	poly (ADP-ribose) polymerase family, member 10	5.1870	2.7172	1.9090
18174	NM 008732.2	solute carrier family 11 (proton-coupled divalent metal ion trans	28.1269	14.7291	1.9096
671535	NM 001163575.1	poly (ADP-ribose) polymerase family, member 10	5.3393	2.7918	1.9125
64292	NM 022415.3	prostaglandin E synthase	6.7331	3.4511	1.9510
67844	NM 026405.3	RAB32, member RAS oncogene family	67.8149	34.7469	1.9517
12125	NM 009754.3	BCL2-like 11 (apoptosis facilitator)	4.2829	2.1795	1.9651
100503984	XR 104558.1	-	24.2867	12.2887	1.9763
100503984	XR 104553.1	-	25.6072	12.9568	1.9763

100503984	XR 104554.1	-	24.1888	12.2392	1.9763
100503984	XR 104561.1	-	25.5255	12.9155	1.9763
100503984	XR 104562.1	-	27.3260	13.8265	1.9763
100503984	XR 104560.1	-	24.7879	12.5423	1.9763
100503984	XR 104557.1	-	25.0726	12.6863	1.9763
100503984	XR 104555.1	-	24.7369	12.5165	1.9763
100503984	XR 104563.1	-	28.2008	14.2479	1.9793
100503984	XR 104559.1	-	24.3228	12.2887	1.9793
100503984	XR 104556.1	-	24.0312	12.1414	1.9793
230073	NM 172689.3	DEAD (Asp-Glu-Ala-Asp) box polypeptide 58	1.7282	0.8609	2.0074
12125	NM 207681.2	BCL2-like 11 (apoptosis facilitator)	4.3432	2.1615	2.0094
74732	NM 001163590.1	syntaxin 11	3.1775	1.5791	2.0123
270118	NM 001013813.3	mastermind like 2 (Drosophila)	2.6256	1.2998	2.0200
74732	NM 001163591.1	syntaxin 11	3.3312	1.6469	2.0227
319909	NM 001126490.1	isthmin 1 homolog (zebrafish)	1.1413	0.5635	2.0253
12125	NM 207680.2	BCL2-like 11 (apoptosis facilitator)	4.2339	2.0894	2.0264
212168	NM 172503.3	zinc finger, SWIM domain containing 4	4.6611	2.2975	2.0287
108797	NM 175366.3	mex3 homolog B (C. elegans)	6.2886	3.0933	2.0330
70701	NM 001081205.1	NIPA-like domain containing 1	0.9605	0.4717	2.0363
17082	NM 001025602.2	interleukin 1 receptor-like 1	31.0010	15.1780	2.0425
67916	NM 080555.2	phosphatidic acid phosphatase type 2B	17.1256	8.3487	2.0513
74732	NM 029075.1	syntaxin 11	3.1328	1.5202	2.0607
100503392	XR 107188.1	-	3.8623	1.8678	2.0679
100503392	XR 105978.1	-	3.8623	1.8678	2.0679
22038	NM 011636.2	phospholipid scramblase 1	44.7919	21.5776	2.0758
100502974	XR 107727.1	predicted gene, 19481	4.4408	2.1310	2.0839
18174	NM 001146161.1	solute carrier family 11 (proton-coupled divalent metal ion trans	32.4984	15.5207	2.0939
16878	NM 001039537.1	leukemia inhibitory factor	6.5125	3.0480	2.1366
17082	NM 010743.2	interleukin 1 receptor-like 1	53.3054	24.9395	2.1374
16878	NM 008501.2	leukemia inhibitory factor	6.2228	2.9103	2.1382

100503392	XR 107187.1	-	4.1181	1.9161	2.1492
100503392	XR 105977.1	-	4.1181	1.9161	2.1492
57875	NM 020581.2	angiopoietin-like 4	1.9028	0.8847	2.1509
100503984	XR 104572.1	-	3.1527	1.4563	2.1648
100503984	XR 104565.1	-	3.1695	1.4641	2.1648
192656	NM 138952.3	receptor (TNFRSF)-interacting serine-threonine kinase 2	45.2043	20.8356	2.1696
209086	NM 010156.3	sterile alpha motif domain containing 9-like	14.2532	6.5460	2.1774
100503984	XR 104570.1	-	12.4206	5.6963	2.1805
100503984	XR 104569.1	-	12.3266	5.6532	2.1805
100503984	XR 104567.1	-	11.8513	5.3866	2.2001
11796	NM 007464.3	baculoviral IAP repeat-containing 3	3.6884	1.6753	2.2016
100503984	XR 104568.1	-	12.2280	5.5361	2.2088
100503984	XR 104566.1	-	12.1348	5.4939	2.2088
326623	NM 177371.3	tumor necrosis factor (ligand) superfamily, member 15	0.7176	0.3218	2.2300
270118	NM 173776.3	mastermind like 2 (Drosophila)	3.0188	1.3528	2.2315
50908	NM 001097617.1	complement component 1, s subcomponent	1.7196	0.7684	2.2380
239650	NM 177716.3	expressed sequence AI836003	2.8046	1.2457	2.2514
50908	NM 144938.2	complement component 1, s subcomponent	1.6859	0.7465	2.2584
15937	NM 133662.2	immediate early response 3	34.3661	15.1534	2.2679
100503551	XR 107722.1	predicted gene 15817	2.1234	0.9284	2.2872
100503551	XR 106291.1	predicted gene 15817	2.1234	0.9284	2.2872
70737	NM 001037711.2	cingulin	1.3052	0.5654	2.3084
16206	NM 008377.2	leucine-rich repeats and immunoglobulin-like domains 1	25.1269	10.8787	2.3097
170677	NM 130878.2	cadherin-related family member 1	0.4505	0.1901	2.3701
20312	NM 009142.3	chemokine (C-X3-C motif) ligand 1	0.7390	0.3097	2.3860
319236	NM 001146007.1	tripartite motif-containing 12C	0.5071	0.2124	2.3877
12521	NM 001136055.1	CD82 antigen	2.6123	1.0928	2.3903
12521	NM 007656.4	CD82 antigen	2.4983	1.0321	2.4206
20821	NM 001082552.1	tripartite motif-containing 21	1.5322	0.6264	2.4462
215113	NM 173388.1	solute carrier family 43, member 2	2.7445	1.1175	2.4559

18034	NM 019408.3	nuclear factor of kappa light polypeptide gene enhancer in B-cell	58.9402	23.9000	2.4661
18034	NM 001177370.1	nuclear factor of kappa light polypeptide gene enhancer in B-cell	62.8664	25.4739	2.4679
12609	NM 007679.4	CCAAT/enhancer binding protein (C/EBP), delta	4.0962	1.6596	2.4683
20821	NM 009277.3	tripartite motif-containing 21	1.5457	0.6238	2.4778
18034	NM 001177369.1	nuclear factor of kappa light polypeptide gene enhancer in B-cell	62.2524	25.1053	2.4797
230738	NM 153159.2	zinc finger CCCH type containing 12A	15.0109	6.0299	2.4894
71683	NM 001048207.1	glycophorin C	15.8227	6.3333	2.4983
21929	NM 001166402.1	tumor necrosis factor, alpha-induced protein 3	13.3463	5.3201	2.5087
21929	NM 009397.3	tumor necrosis factor, alpha-induced protein 3	13.1228	5.2182	2.5148
100048759	XM 003086792.1	-	5.9885	2.3270	2.5735
434438	NM 001135198.1	coiled-coil domain containing 36	0.5216	0.2005	2.6017
12266	NM 009778.2	complement component 3	5.8334	2.2281	2.6180
18812	NM 011118.2	prolactin family 2, subfamily c, member 3	33.4373	12.7137	2.6300
17533	NM 008625.2	mannose receptor, C type 1	1.5246	0.5760	2.6469
107849	NM 181852.1	prolactin family 2, subfamily c, member 5	14.0089	5.2485	2.6691
20568	NM 011414.3	secretory leukocyte peptidase inhibitor	4.1181	1.5329	2.6864
18811	NM 031191.1	prolactin family 2, subfamily c, member 2	36.9197	13.7408	2.6869
19260	NM 008979.1	protein tyrosine phosphatase, non-receptor type 22 (lymphoid)	2.8108	1.0421	2.6973
71816	NM 027934.2	ring finger protein 180	0.4627	0.1715	2.6980
80859	NM 001159395.1	nuclear factor of kappa light polypeptide gene enhancer in B-cell	6.1356	2.2726	2.6998
80859	NM 030612.3	nuclear factor of kappa light polypeptide gene enhancer in B-cell	5.6188	2.0768	2.7055
18208	NM 008744.2	netrin 1	4.8428	1.7853	2.7125
80859	NM 001159394.1	nuclear factor of kappa light polypeptide gene enhancer in B-cell	5.8570	2.1589	2.7130
666317	NM 001045532.1	Prolactin family 2, subfamily c, member 1	5.0628	1.8389	2.7531
57783	NM 021327.2	TNFAIP3 interacting protein 1	30.0159	10.7610	2.7893
545551	NR 033629.1	cDNA sequence BC021767	1.0428	0.3723	2.8010
630663	XM 904245.4	predicted gene 7040	9.9369	3.4792	2.8561
11988	NM 007514.3	solute carrier family 7 (cationic amino acid transporter, y+ syst	0.2404	0.0840	2.8632
11988	NM 001044740.1	solute carrier family 7 (cationic amino acid transporter, y+ syst	0.2411	0.0842	2.8632
20556	NM 011408.1	schlafen 2	2.7381	0.9423	2.9058

16997	NM 013589.3	latent transforming growth factor beta binding protein 2	0.2106	0.0708	2.9733
630663	XM 001473551.2	predicted gene 7040	13.9888	4.6690	2.9961
16149	NM 001042605.1	CD74 antigen (invariant polypeptide of major histocompatibility c	1.3238	0.4284	3.0899
16149	NM 010545.3	CD74 antigen (invariant polypeptide of major histocompatibility c	1.4713	0.4657	3.1592
19698	NM 009046.2	avian reticuloendotheliosis viral (v-rel) oncogene related B	14.9871	4.7315	3.1675
233552	NM 201352.2	glycerophosphodiester phosphodiesterase domain containing 5	1.9709	0.6096	3.2333
12051	NM 033601.3	B-cell leukemia/lymphoma 3	3.6036	1.0416	3.4595
21822	NM 011579.3	T-cell specific GTPase 1	0.3563	0.1027	3.4689
14239	NM 010226.2	forkhead box S1	0.9542	0.2751	3.4689
20306	NM 013654.3	chemokine (C-C motif) ligand 7	342.8538	98.8195	3.4695
56489	NM 019777.3	inhibitor of kappaB kinase epsilon	6.0930	1.7114	3.5602
20296	NM 011333.3	chemokine (C-C motif) ligand 2	#####	433.7946	3.7885
100039796	NM 001145164.1	T-cell specific GTPase 2	0.3427	0.0896	3.8229
21942	NM 001077508.1	tumor necrosis factor receptor superfamily, member 9	0.6437	0.1624	3.9645
14102	NM 007987.2	Fas (TNF receptor superfamily member 6)	9.7176	2.4512	3.9645
626578	NM 001039646.2	guanylate-binding protein 10	0.3242	0.0818	3.9645
18035	NM 010907.2	nuclear factor of kappa light polypeptide gene enhancer in B-cell	66.4125	16.4009	4.0493
16193	NM 031168.1	interleukin 6	4.9981	1.2276	4.0716
18606	NM 001136077.1	ectonucleotide pyrophosphatase/phosphodiesterase 2	0.9115	0.2233	4.0811
56619	NM 019948.2	C-type lectin domain family 4, member e	0.7095	0.1718	4.1296
320207	NM 177320.2	phosphoinositide-3-kinase, regulatory subunit 5, p101	1.8842	0.4464	4.2214
21942	NM 011612.2	tumor necrosis factor receptor superfamily, member 9	0.6532	0.1521	4.2948
238393	NM 001033335.3	serine (or cysteine) peptidase inhibitor, clade A, member 3F	0.4199	0.0978	4.2948
18606	NM 015744.2	ectonucleotide pyrophosphatase/phosphodiesterase 2	1.0805	0.2447	4.4150
102502	NM 001033210.3	plastin 1 (I-isoform)	0.6961	0.1561	4.4600
238393	NM 001168294.1	serine (or cysteine) peptidase inhibitor, clade A, member 3F	0.4282	0.0960	4.4600
238393	NM 001168295.1	serine (or cysteine) peptidase inhibitor, clade A, member 3F	0.4726	0.1060	4.4600
100702	NM 194336.2	guanylate binding protein 6	0.4128	0.0916	4.5051
14102	NM 001146708.1	Fas (TNF receptor superfamily member 6)	3.5421	0.7798	4.5426
18788	NM 011111.4	serine (or cysteine) peptidase inhibitor, clade B, member 2	1.0686	0.2336	4.5744



21942	NM 001077509.1	tumor necrosis factor receptor superfamily, member 9	0.7267	0.1535	4.7353
12363	NM 007609.2	caspase 4, apoptosis-related cysteine peptidase	12.0475	2.5167	4.7871
18788	NM 001174170.1	serine (or cysteine) peptidase inhibitor, clade B, member 2	1.0608	0.2177	4.8730
214854	NM 153408.2	neutralized homolog 3 homolog (Drosophila)	7.3203	1.4660	4.9933
14468	NM 010259.2	guanylate binding protein 1	0.2915	0.0512	5.6989
547253	NM 001039530.3	poly (ADP-ribose) polymerase family, member 14	0.1528	0.0249	6.1449
18037	NM 008690.3	nuclear factor of kappa light polypeptide gene enhancer in B-cell	6.8181	1.1019	6.1878
17384	NM 019471.2	matrix metallopeptidase 10	1.4331	0.2305	6.2170
14469	NM 010260.1	guanylate binding protein 2	1.6490	0.2627	6.2771
22029	NM 009421.3	TNF receptor-associated factor 1	10.6692	1.6442	6.4890
630294	NM 001177467.1	predicted gene 7030	0.7952	0.1203	6.6074
240675	NM 172840.2	von Willebrand factor A domain containing 2	0.1288	0.0186	6.9378
100503422	XM 003085129.1	predicted gene, 19684	0.4659	0.0672	6.9378
100503422	XM 003085162.1	predicted gene, 19684	0.4665	0.0672	6.9378
547347	NM 001034909.3	predicted gene 6034	0.3064	0.0442	6.9378
17392	NM 010809.1	matrix metallopeptidase 3	0.3270	0.0388	8.4245
17386	NM 008607.2	matrix metallopeptidase 13	1.1358	0.1348	8.4245
20715	NM 009251.1	serine (or cysteine) peptidase inhibitor, clade A, member 3G	2.0601	0.2310	8.9200
16819	NM 008491.1	lipocalin 2	18.0602	1.9871	9.0887
216799	NM 145827.3	NLR family, pyrin domain containing 3	0.1689	0.0179	9.4156
17395	NM 013599.2	matrix metallopeptidase 9	0.1347	0.0113	11.8934
414084	NM 001001495.2	TNFAIP3 interacting protein 3	3.2149	0.2488	12.9226
15945	NM 021274.1	chemokine (C-X-C motif) ligand 10	5.3800	0.4071	13.2149
20311	NM 009141.2	chemokine (C-X-C motif) ligand 5	337.7799	24.9941	13.5144
22329	NM 011693.3	vascular cell adhesion molecule 1	21.2693	1.3054	16.2930
14825	NM 008176.3	chemokine (C-X-C motif) ligand 1	319.9473	18.5556	17.2426
12981	NM 009969.4	colony stimulating factor 2 (granulocyte-macrophage)	0.6228	0.0349	17.8401
408066	XR 001629.2	cDNA sequence BC067074	0.1007	0.0044	22.7956
16165	NM 008356.3	interleukin 13 receptor, alpha 2	0.5735	0.0231	24.7779
20297	NM 016960.2	chemokine (C-C motif) ligand 20	56.3419	1.6931	33.2767

20210	NM 011315.3	serum amyloid A 3	4.6446	0.1358	34.1934
20297	NM 001159738.1	chemokine (C-C motif) ligand 20	55.8253	1.6142	34.5847
71132	NM 001042418.1	calcium-binding tyrosine-(Y)-phosphorylation regulated (fibroushe	0.1741	0.0000	Inf
14728	NM 013532.2	leukocyte immunoglobulin-like receptor, subfamily B, member 4	0.2769	0.0000	Inf
20310	NM 009140.2	chemokine (C-X-C motif) ligand 2	0.4951	0.0000	Inf
13982	NM 007956.4	estrogen receptor 1 (alpha)	0.0662	0.0000	Inf
18126	NM 010927.3	nitric oxide synthase 2, inducible	1.2452	0.0000	Inf
16365	NM 008392.1	immunoresponsive gene 1	0.1381	0.0000	Inf
15894	NM 010493.2	intercellular adhesion molecule 1	0.1407	0.0000	Inf
408066	XR 035217.2	cDNA sequence BC067074	0.0759	0.0000	Inf
100503281	XR 108224.1	-	0.4655	0.0000	Inf

Supplementary Table 3 Differential expressed genes of MLL1-/- MEF cell with or without TNF $\alpha$ 

ID	ACCESSION NUMBER	NAME	TNF	Ctrl	FOLDCHANGE
12695	NM_001005787.1	InaD-like (Drosophila)	0.0000	0.1546	0.0000
100045736	XM_001474842.2	-	0.0722	1.2501	0.0578
100039589	XM_003084688.1	predicted gene 2329	0.0722	1.1112	0.0650
380698	NM_199152.2	obscurin, cytoskeletal calmodulin and titin-interacting RhoGEF	0.0023	0.0260	0.0867
245607	NM_001163016.1	G protein-coupled receptor associated sorting protein 2	0.0274	0.2769	0.0991
245607	NM_001163017.1	G protein-coupled receptor associated sorting protein 2	0.0275	0.2780	0.0991
245607	NM_001163015.1	G protein-coupled receptor associated sorting protein 2	0.0272	0.2742	0.0991
100503347	XM_003086491.1	predicted gene, 19648	0.0400	0.2308	0.1734
19784	NR_004439.1	ribonuclease P RNA-like 2	0.2223	1.2111	0.1836
24131	NM_011918.4	LIM domain binding 3	0.2466	0.6916	0.3566
24131	NM_001039072.2	LIM domain binding 3	0.2533	0.7001	0.3618
24131	NM_001039071.2	LIM domain binding 3	0.2561	0.7079	0.3618
24131	NM_001039074.2	LIM domain binding 3	0.2509	0.6936	0.3618
24131	NM_001039073.2	LIM domain binding 3	0.2624	0.7148	0.3671
19017	NR_027710.1	peroxisome proliferative activated receptor, gamma, coactivator 1 alpha	0.1675	0.4523	0.3702
19017	NM_008904.2	peroxisome proliferative activated receptor, gamma, coactivator 1 alpha	0.1683	0.4545	0.3702
11754	NM_009675.2	amine oxidase, copper containing 3	0.2003	0.5324	0.3762
100047224	XM_003086950.1	-	1.6493	4.3521	0.3790
195727	NM_001081052.1	Nance-Horan syndrome (human)	0.3465	0.7773	0.4458
223650	NM_144848.2	epiplakin 1	0.2347	0.5250	0.4471
668066	XM_001004338.2	predicted gene 14418	1.5995	3.4546	0.4630
243385	NM_183183.2	GPRIN family member 3	1.6040	3.3894	0.4732
244867	NM_175535.3	Rho GTPase activating protein 20	1.5055	3.0903	0.4872
625703	XM_904090.4	-	1.7629	3.5234	0.5003
13838	NM_007936.3	Eph receptor A4	1.0641	2.1247	0.5008

231861	NM_178242.2	trinucleotide repeat containing 18	3.0532	6.0066	0.5083
242481	NM_172868.2	paralemmin 2	1.5201	2.9836	0.5095
240427	NM_053099.2	SET binding protein 1	0.7680	1.4917	0.5149
66395	NM_009643.1	AHNAK nucleoprotein (desmoyokin)	96.8238	183.5819	0.5274
67393	NM_133687.2	CXXC finger 5	2.4654	4.5681	0.5397
330938	NM_178118.2	DIX domain containing 1	6.9659	12.5231	0.5562
100503159	XR_108047.1	-	6.5701	11.6379	0.5645
26408	NM_008580.4	mitogen-activated protein kinase kinase kinase 5	1.8536	3.2225	0.5752
231861	NM_001122730.1	trinucleotide repeat containing 18	3.9304	6.6259	0.5932
17309	NM_010795.3	mannoside acetylglucosaminyltransferase 3	5.4516	9.0626	0.6016
100042862	XM_003085879.1	predicted gene 4076	234.2826	388.2562	0.6034
100042862	XM_001478943.2	predicted gene 4076	246.8104	408.8011	0.6037
11630	NM_172393.2	absent in melanoma 1	11.4334	18.8011	0.6081
214137	NM_172525.2	Rho GTPase activating protein 29	6.1049	9.9728	0.6122
271564	NM_173028.3	vacuolar protein sorting 13A (yeast)	3.3181	5.4093	0.6134
68861	NM_001033145.2	RIKEN cDNA 1190002N15 gene	18.2794	28.9826	0.6307
77057	NM_029858.2	stonin 1	13.2659	20.9501	0.6332
54127	NM_016844.2	ribosomal protein S28	718.3538	452.7200	1.5868
667682	XM_991853.3	predicted gene 8759	1085.2008	683.1858	1.5884
14455	NR_002840.2	growth arrest specific 5	65.1405	40.7494	1.5986
671641	XM_001479052.2	predicted gene 10063	1981.0625	1234.7892	1.6044
100042561	XM_003086748.1	predicted gene 10177	175.4771	109.3654	1.6045
100042561	XM_001478520.2	predicted gene 10177	175.4771	109.3654	1.6045
100046297	XM_001477362.2	-	703.1160	436.4319	1.6111
100043718	XM_001480721.2	predicted gene 4604	236.1249	146.3981	1.6129
14775	NM_008160.5	glutathione peroxidase 1	92.2788	57.2039	1.6132
12859	NM_009942.2	cytochrome c oxidase, subunit Vb	141.0200	87.3991	1.6135
54217	NM_018730.3	ribosomal protein L36	284.0859	175.7512	1.6164
30055	NM_013895.4	translocase of inner mitochondrial membrane 13 homolog (yeast)	59.4914	36.7882	1.6171

635470	XM_003086282.1	predicted gene 14407	157.9373	97.5871	1.6184
100046079	XM_001475417.2	-	166.5536	102.7729	1.6206
27370	NM_013765.2	ribosomal protein S26	1431.4740	881.8388	1.6233
67673	NM_026305.2	transcription elongation factor B (SIII), polypeptide 2	147.6390	90.6721	1.6283
665533	XM_003086398.1	predicted gene 13004	529.7444	324.9924	1.6300
665533	XM_977600.2	predicted gene 13004	529.7444	324.9924	1.6300
546695	XM_003086080.1	predicted gene, 16519	106.6450	65.3828	1.6311
545487	XM_619852.4	predicted gene 14439	156.3150	95.8047	1.6316
16206	NM_008377.2	leucine-rich repeats and immunoglobulin-like domains 1	49.2302	30.1324	1.6338
20335	NM_011343.3	SEC61, gamma subunit	104.4851	63.9293	1.6344
19981	NM_009084.4	ribosomal protein L37a	536.2309	328.0250	1.6347
18034	NM_001177370.1	nuclear factor of kappa light polypeptide gene enhancer in B-cells 2, p49/p100	74.7410	45.7093	1.6351
15980	NM_008338.3	interferon gamma receptor 2	35.3064	21.5783	1.6362
18034	NM_001177369.1	nuclear factor of kappa light polypeptide gene enhancer in B-cells 2, p49/p100	73.8610	45.0676	1.6389
18034	NM_019408.3	nuclear factor of kappa light polypeptide gene enhancer in B-cells 2, p49/p100	70.1387	42.7890	1.6392
72338	NM_028203.1	WD repeat domain 89	77.4768	47.2237	1.6406
57294	NM_027015.4	ribosomal protein S27	683.5674	415.9998	1.6432
100043813	NM_001190258.1	predicted gene 9846	676.2565	411.5506	1.6432
100048613	XM_001480380.2	-	107.3612	65.2260	1.6460
67126	NM_025983.3	ATP synthase, H <sup>+</sup> transporting, mitochondrial F1 complex, epsilon subunit	97.9056	59.4226	1.6476
12857	NM_009941.2	cytochrome c oxidase subunit IV isoform 1	312.4543	189.3574	1.6501
27425	NM_013795.5	ATP synthase, H <sup>+</sup> transporting, mitochondrial F0 complex, subunit g	227.3541	137.6219	1.6520
100042348	XM_001478455.2	predicted gene 10221	234.8841	142.1616	1.6522
230075	NM_001033305.2	NADH dehydrogenase (ubiquinone) 1 beta subcomplex, 6	89.4306	54.0857	1.6535
100503897	XR_107153.1	predicted gene, 19947	161.3130	97.2955	1.6580
100503897	XR_105996.1	predicted gene, 19947	161.3130	97.2955	1.6580

69878	NM_027246.1	small nuclear ribonucleoprotein polypeptide F	105.8570	63.8354	1.6583
17133	NM_010755.3	v-maf musculoaponeurotic fibrosarcoma oncogene family, protein F (avian)	22.5600	13.5280	1.6677
12010	NM_009735.3	beta-2 microglobulin	363.5380	217.7566	1.6695
99382	NM_178890.3	ankyrin repeat and BTB (POZ) domain containing 2	7.1917	4.3023	1.6716
12866	NM_009945.3	cytochrome c oxidase, subunit VIIa 2	156.6245	93.5164	1.6748
666642	XM_985134.3	predicted pseudogene 8210	93.1370	55.5507	1.6766
629750	NR_033523.1	predicted gene 11517	130.3621	77.7199	1.6773
13014	NM_007793.3	cystatin B	234.4187	139.6702	1.6784
751556	NR_030451.1	microRNA 682	435.4102	259.3550	1.6788
100040259	NM_001111330.1	predicted pseudogene 16379	248.8759	147.5966	1.6862
57294	NR_033727.1	ribosomal protein S27	344.5862	204.2557	1.6870
192657	NM_138953.2	elongation factor RNA polymerase II 2	15.9917	9.4558	1.6912
66491	NM_025593.1	polymerase (RNA) II (DNA directed) polypeptide L	17.0794	10.0725	1.6956
67945	NM_018860.4	ribosomal protein L41	2637.6470	1554.9638	1.6963
17392	NM_010809.1	matrix metalloproteinase 3	42.4850	24.9785	1.7009
233016	NM_144923.3	biliverdin reductase B (flavin reductase (NADPH))	32.8141	19.2852	1.7015
19944	NM_009082.2	ribosomal protein L29	110.8143	64.9454	1.7063
21929	NM_001166402.1	tumor necrosis factor, alpha-induced protein 3	9.5689	5.5609	1.7208
67184	NM_023312.2	NADH dehydrogenase (ubiquinone) 1 alpha subcomplex, 13	19.4337	11.2887	1.7215
20656	NM_013671.3	superoxide dismutase 2, mitochondrial	29.2301	16.9286	1.7267
223881	NM_172612.3	Rho family GTPase 1	87.7922	50.8359	1.7270
21929	NM_009397.3	tumor necrosis factor, alpha-induced protein 3	9.4206	5.4543	1.7272
665964	XM_980604.3	predicted gene 7866	522.4457	302.1944	1.7288
22186	NM_019883.3	ubiquitin A-52 residue ribosomal protein fusion product 1	1030.7802	595.3682	1.7313
100039782	XM_003086738.1	predicted gene 10709	131.6384	76.0086	1.7319
665964	XM_003086743.1	predicted gene 7866	530.0628	305.8558	1.7330
11927	NM_009720.2	ATX1 (antioxidant protein 1) homolog 1 (yeast)	63.5819	36.6762	1.7336
67916	NM_080555.2	phosphatidic acid phosphatase type 2B	23.7194	13.6538	1.7372
17319	NM_010798.2	macrophage migration inhibitory factor	600.4224	343.1800	1.7496

230073	NM_172689.3	DEAD (Asp-Glu-Ala-Asp) box polypeptide 58	9.9966	5.7019	1.7532
20422	NM_009169.2	split hand/foot malformation (ectrodactyly) type 1	189.0396	107.7058	1.7551
629957	XM_003086273.1	predicted gene 14303	507.7795	289.1612	1.7560
100504654	XR_105586.1	predicted gene 10736	1840.2606	1046.0660	1.7592
100504654	XR_105587.1	predicted gene 10736	1911.2104	1086.3963	1.7592
100504654	XR_106826.1	predicted gene 10736	1778.6007	1010.8844	1.7595
100504654	XR_106827.1	predicted gene 10736	1844.7812	1048.4987	1.7595
100038969	XM_001472037.2	predicted gene 14958	1600.4838	908.9360	1.7608
626048	XM_001473380.2	predicted pseudogene 6646	1601.3925	908.9360	1.7618
100503567	XM_003085762.1	predicted gene 11694	209.1386	118.6965	1.7620
100503567	XM_003085509.1	predicted gene 11694	209.1386	118.6965	1.7620
434460	XM_001477611.2	predicted gene 5623	96.9986	54.7627	1.7713
434460	XM_001477346.2	predicted gene 5623	96.9986	54.7627	1.7713
20200	NM_011313.2	S100 calcium binding protein A6 (calyclin)	729.7546	411.8745	1.7718
67067	NM_001164216.1	reactive oxygen species modulator 1	41.9669	23.6771	1.7725
12864	NM_053071.2	cytochrome c oxidase, subunit VIc	211.6069	119.3407	1.7731
665032	XM_974069.2	predicted gene 13841	123.4288	69.4725	1.7767
665032	XM_991641.2	predicted gene 13841	123.1820	69.3078	1.7773
66379	NM_183256.3	RIKEN cDNA 2310016M24 gene	34.5307	19.4212	1.7780
434843	XM_486761.5	predicted pseudogene 5642	1693.9502	952.3672	1.7787
665463	XM_977110.3	predicted pseudogene 7643	1693.9502	952.3672	1.7787
665611	XM_978135.3	predicted pseudogene 7711	1693.9502	952.3672	1.7787
665522	XM_977540.3	predicted pseudogene 7671	1693.9502	952.3672	1.7787
665579	XM_977918.3	predicted pseudogene 7698	1693.9502	952.3672	1.7787
665032	XM_003085349.1	predicted gene 13841	123.6729	69.4848	1.7799
665032	XM_003086489.1	predicted gene 13841	123.4220	69.3181	1.7805
382265	XM_356374.5	predicted gene 5167	99.8413	56.0350	1.7818
100043714	XM_003086243.1	predicted pseudogene 10774	55.6842	31.2394	1.7825
100043714	XM_001480774.2	predicted pseudogene 10774	55.6842	31.2394	1.7825
629957	XM_894924.4	predicted gene 14303	818.9727	458.8247	1.7849

54405	NM_019443.2	NADH dehydrogenase (ubiquinone) 1 alpha subcomplex, 1	40.6538	22.7750	1.7850
20198	NM_011311.2	S100 calcium binding protein A4	2385.4436	1331.7804	1.7912
17991	NM_010885.4	NADH dehydrogenase (ubiquinone) 1 alpha subcomplex, 2	116.5165	64.9515	1.7939
66142	NM_025379.2	cytochrome c oxidase subunit VIIb	88.5289	49.2034	1.7992
100039782	XM_001473311.2	predicted gene 10709	98.9061	54.9447	1.8001
13198	NM_007837.3	DNA-damage inducible transcript 3	21.3312	11.8400	1.8016
11839	NM_009704.3	amphiregulin	13.9534	7.7248	1.8063
319238	XR_107318.1	RIKEN cDNA 9330123L03 gene	5.1955	2.8542	1.8203
20090	NM_009093.2	ribosomal protein S29	601.5951	330.0521	1.8227
433941	XM_003086189.1	predicted gene 5561	108.9998	59.7879	1.8231
433941	XM_001478364.2	predicted gene 5561	108.9998	59.7879	1.8231
56187	NM_019519.2	Rab geranylgeranyl transferase, a subunit	6.1415	3.3628	1.8263
100503055	XM_003085327.1	-	50.2746	27.5242	1.8266
100043718	XM_003086523.1	predicted gene 4604	237.7590	130.0606	1.8281
320415	NM_177157.4	GTP cyclohydrolase I feedback regulator	84.2406	46.0753	1.8283
11983	NM_007512.3	ATPase inhibitory factor 1	56.6753	30.9422	1.8317
622707	XM_001475711.2	predicted gene 6344	71.2296	38.8333	1.8342
102644	NM_178644.3	OAF homolog (Drosophila)	35.9188	19.5807	1.8344
17873	NM_008655.1	growth arrest and DNA-damage-inducible 45 beta	19.2499	10.4934	1.8345
66169	NM_025394.3	translocase of outer mitochondrial membrane 7 homolog (yeast)	68.1720	36.9712	1.8439
68949	NM_001081005.1	RIKEN cDNA 1500012F01 gene	60.4516	32.7572	1.8454
72655	XM_986352.2	small nucleolar RNA host gene 5	46.3329	25.0715	1.8480
217344	NM_172572.3	rhomboid 5 homolog 2 (Drosophila)	4.4054	2.3712	1.8578
13197	NM_007836.1	growth arrest and DNA-damage-inducible 45 alpha	9.3944	5.0447	1.8622
66915	NR_028108.1	myeloma overexpressed 2	27.9426	15.0044	1.8623
217344	NM_001167680.1	rhomboid 5 homolog 2 (Drosophila)	4.6732	2.5079	1.8634
72655	XM_925560.3	small nucleolar RNA host gene 5	46.0753	24.7247	1.8635
12609	NM_007679.4	CCAAT/enhancer binding protein (C/EBP), delta	11.3447	6.0813	1.8655
69920	NM_027259.1	polymerase (RNA) II (DNA directed) polypeptide I	22.6379	12.1145	1.8687



18036	NM_010908.4	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, beta	11.1049	5.9397	1.8696
20308	NM_011338.2	chemokine (C-C motif) ligand 9	17.4949	9.3368	1.8738
100503653	XR_104731.1	predicted gene 14036	102.2717	54.5805	1.8738
13167	NM_007830.4	diazepam binding inhibitor	152.6526	81.3535	1.8764
14790	NM_013535.1	gene rich cluster, C10 gene	20.6090	10.9587	1.8806
66915	NM_001163425.1	myeloma overexpressed 2	47.8378	25.2243	1.8965
68040	NM_024215.2	zinc finger protein 593	5.6453	2.9763	1.8968
17384	NM_019471.2	matrix metalloproteinase 10	36.3866	19.1835	1.8968
66481	NM_025587.2	ribosomal protein S21	735.2156	386.8005	1.9008
69895	NR_028574.1	small nucleolar RNA host gene 8	27.6465	14.5265	1.9032
83673	NR_002896.3	small nucleolar RNA host gene (non-protein coding) 1	128.6195	67.4759	1.9062
18049	NM_013609.2	nerve growth factor	16.0144	8.3903	1.9087
22272	NM_025352.2	ubiquinol-cytochrome c reductase, complex III subunit VII	90.2122	47.2625	1.9087
30059	NM_013899.2	translocase of inner mitochondrial membrane 10 homolog (yeast)	56.7326	29.7171	1.9091
100041286	XR_106636.1	predicted gene 11974	25.8980	13.5425	1.9123
100041286	XR_105435.1	predicted gene 11974	25.8980	13.5425	1.9123
74732	NM_001163590.1	syntaxin 11	7.2620	3.7881	1.9170
100504319	XM_003086228.1	-	207.0361	107.9865	1.9172
100504319	XM_003084494.1	-	207.0361	107.9865	1.9172
100503669	XM_003086525.1	-	83.2977	43.3896	1.9198
100503669	XM_003084644.1	-	83.2977	43.3896	1.9198
13167	NM_001037999.2	diazepam binding inhibitor	135.1238	70.3696	1.9202
74732	NM_001163591.1	syntaxin 11	7.5787	3.9447	1.9212
20568	NM_011414.3	secretory leukocyte peptidase inhibitor	83.3139	43.3349	1.9226
80859	NM_030612.3	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta	3.9502	2.0538	1.9233
19698	NM_009046.2	avian reticuloendotheliosis viral (v-rel) oncogene related B	14.7822	7.6782	1.9252
100041859	XM_003086090.1	predicted gene 3550	55.8534	28.9648	1.9283
100041859	XM_001477476.2	predicted gene 3550	55.8534	28.9648	1.9283

22038	NM_011636.2	phospholipid scramblase 1	49.7638	25.7935	1.9293
15040	NM_010398.3	histocompatibility 2, T region locus 23	10.7368	5.5581	1.9317
68763	NR_015536.1	RIKEN cDNA 1110038B12 gene	160.2223	82.5510	1.9409
18049	NM_001112698.1	nerve growth factor	18.6172	9.5834	1.9427
74732	NM_029075.1	syntaxin 11	7.0690	3.6370	1.9436
80859	NM_001159395.1	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta	4.3422	2.2293	1.9478
100503826	XR_107649.1	predicted gene 13075	22.0599	11.3206	1.9487
100503826	XR_104900.1	predicted gene 13075	22.0599	11.3206	1.9487
66141	NM_025378.2	interferon induced transmembrane protein 3	157.5462	80.6882	1.9525
80859	NM_001159394.1	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta	4.1437	2.1155	1.9587
68763	NR_027943.1	RIKEN cDNA 1110038B12 gene	142.2872	72.5673	1.9608
12578	NM_001040654.1	cyclin-dependent kinase inhibitor 2A	33.7588	17.1650	1.9667
100045766	XM_001474415.2	predicted gene 10126	123.5835	62.6092	1.9739
100045766	XM_003084933.1	predicted gene 10126	123.5835	62.6092	1.9739
434394	XM_915072.3	predicted gene 5614	78.8342	39.9309	1.9743
436081	XM_001004069.3	predicted gene 5745	26.9668	13.5951	1.9836
436081	XM_488179.5	predicted gene 5745	26.9668	13.5951	1.9836
100503585	XM_003086438.1	-	334.0994	168.0256	1.9884
100503585	XM_003084617.1	-	334.0994	168.0256	1.9884
12578	NM_009877.2	cyclin-dependent kinase inhibitor 2A	34.1776	17.0990	1.9988
381155	XR_035375.1	RIKEN cDNA 9630014M24 gene	3.0137	1.5066	2.0003
66293	NR_027819.1	RIKEN cDNA 1810032008 gene	8.9850	4.4602	2.0145
11435	NM_007389.4	cholinergic receptor, nicotinic, alpha polypeptide 1 (muscle)	1.8119	0.8826	2.0529
100504928	XR_106975.1	-	1.9112	0.9253	2.0655
230738	NM_153159.2	zinc finger CCCH type containing 12A	6.5553	3.1600	2.0745
108153	NM_001003911.2	a disintegrin-like and metallopeptidase (reprolysin type) with thrombospondin type 1 motif, 7	4.0967	1.9739	2.0755
320207	NM_177320.2	phosphoinositide-3-kinase, regulatory subunit 5, p101	15.2432	7.3159	2.0836
19260	NM_008979.1	protein tyrosine phosphatase, non-receptor type 22 (lymphoid)	6.3087	2.9870	2.1120

668092	XM_994269.1	predicted gene 8973	58.7021	27.6645	2.1219
15937	NM_133662.2	immediate early response 3	58.4487	27.5021	2.1252
434394	XM_486208.5	predicted gene 5614	73.9478	34.7634	2.1272
76964	NM_029816.2	RIKEN cDNA 2610028H24 gene	3.8487	1.8077	2.1291
12916	NM_001110853.1	cAMP responsive element modulator	2.4338	1.1359	2.1427
12916	NM_001110857.1	cAMP responsive element modulator	2.2933	1.0690	2.1453
72275	NM_028179.1	RIKEN cDNA 2200002D01 gene	25.1042	11.6382	2.1571
56695	NM_025580.2	paroxysmal nonkinesigenic dyskinesia	8.5520	3.9558	2.1619
668092	XM_003085977.1	predicted gene 8973	58.2089	26.8741	2.1660
100504742	XM_003086332.1	-	117.6074	52.9206	2.2223
18037	NM_008690.3	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, epsilon	18.0281	8.0801	2.2312
12856	NM_001017429.2	cytochrome c oxidase, subunit XVII assembly protein homolog (yeast)	25.4228	11.3069	2.2484
66594	NM_025650.2	ubiquinol-cytochrome c reductase, complex III subunit XI	101.8100	44.6566	2.2798
57783	NM_021327.2	TNFAIP3 interacting protein 1	43.2784	18.8918	2.2909
18788	NM_011111.4	serine (or cysteine) peptidase inhibitor, clade B, member 2	5.7035	2.4563	2.3220
20361	NM_011352.2	sema domain, immunoglobulin domain (Ig), and GPI membrane anchor, (semaphorin) 7A	3.0227	1.3017	2.3222
18788	NM_001174170.1	serine (or cysteine) peptidase inhibitor, clade B, member 2	5.7580	2.4547	2.3457
546663	XM_915258.4	predicted pseudogene 5963	278.1877	118.0584	2.3564
546663	XM_001002128.3	predicted pseudogene 5963	278.1877	118.0584	2.3564
545551	NR_033629.1	cDNA sequence BC021767	1.5344	0.6482	2.3673
100504629	XM_003086098.1	predicted gene, 17669	12.2776	5.1640	2.3775
319689	XM_003086426.1	predicted gene, 17739	1.6810	0.7050	2.3844
319689	XM_003084614.1	predicted gene, 17739	1.6810	0.7050	2.3844
12051	NM_033601.3	B-cell leukemia/lymphoma 3	4.2058	1.7639	2.3844
100503758	XR_106535.1	predicted gene, 19876	75.6369	31.6424	2.3904
100503758	XR_105417.1	predicted gene, 19876	75.6369	31.6424	2.3904
26434	NM_023043.2	prion protein dublet	1.4915	0.6234	2.3924
17748	NM_013602.3	metallothionein 1	125.7142	52.4440	2.3971

69221	NR_030738.1	RIKEN cDNA 2410006H16 gene	63.5934	26.4643	2.4030
257632	NM_145857.2	nucleotide-binding oligomerization domain containing 2	1.7822	0.7220	2.4684
18035	NM_010907.2	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha	54.3012	21.8327	2.4871
16193	NM_031168.1	interleukin 6	2.7637	1.0995	2.5137
16878	NM_001039537.1	leukemia inhibitory factor	9.2464	3.6711	2.5187
233552	NM_201352.2	glycerophosphodiester phosphodiesterase domain containing 5	3.5449	1.4074	2.5188
17395	NM_013599.2	matrix metalloproteinase 9	1.1058	0.4378	2.5261
16878	NM_008501.2	leukemia inhibitory factor	8.9014	3.5099	2.5361
110332	NM_173449.3	RIKEN cDNA 4921523A10 gene	1.8618	0.7240	2.5715
20306	NM_013654.3	chemokine (C-C motif) ligand 7	144.0293	55.3655	2.6014
69749	XR_107112.1	RIKEN cDNA 2410004N09 gene	15.6704	5.9801	2.6204
69749	XR_105940.1	RIKEN cDNA 2410004N09 gene	15.6704	5.9801	2.6204
26434	NM_001126338.1	prion protein dublet	1.7468	0.6616	2.6404
11796	NM_007464.3	baculoviral IAP repeat-containing 3	2.7367	1.0242	2.6721
17750	NM_008630.2	metallothionein 2	39.0333	14.5985	2.6738
12363	NM_007609.2	caspase 4, apoptosis-related cysteine peptidase	4.9157	1.8070	2.7204
56489	NM_019777.3	inhibitor of kappaB kinase epsilon	10.8508	3.9800	2.7264
17386	NM_008607.2	matrix metalloproteinase 13	8.4810	3.0902	2.7445
74013	NM_028713.1	raftlin family member 2	0.7582	0.2700	2.8084
14727	NM_008147.1	glycoprotein 49 A	3.6912	1.2974	2.8451
20296	NM_011333.3	chemokine (C-C motif) ligand 2	752.1989	255.4061	2.9451
57875	NM_020581.2	angiopoietin-like 4	1.7842	0.5978	2.9848
227659	NM_001177627.1	solute carrier family 2 (facilitated glucose transporter), member 6	2.6249	0.8495	3.0899
76773	NM_029734.1	WDYHV motif containing 1	8.4159	2.7215	3.0924
22329	NM_011693.3	vascular cell adhesion molecule 1	58.3048	18.8018	3.1010
22029	NM_009421.3	TNF receptor-associated factor 1	17.6218	5.5043	3.2014
24088	NM_011905.3	toll-like receptor 2	4.2713	1.3168	3.2437
751530	NR_030494.1	-	48.9708	14.9388	3.2781

227659	NM_172659.2	solute carrier family 2 (facilitated glucose transporter), member 6	2.9099	0.8859	3.2847
75614	XR_105882.1	RIKEN cDNA 2610019E17 gene	6.6720	2.0256	3.2938
414084	NM_001001495.2	TNFAIP3 interacting protein 3	6.7889	2.0214	3.3584
100217453	NR_028548.1	small nucleolar RNA, C/D box 16A	18.7832	5.4721	3.4325
14468	NM_010259.2	guanylate binding protein 1	1.7816	0.5121	3.4792
16149	NM_010545.3	CD74 antigen (invariant polypeptide of major histocompatibility complex, class II antigen-associated)	1.1287	0.3215	3.5105
14102	NM_007987.2	Fas (TNF receptor superfamily member 6)	13.0013	3.6861	3.5271
14728	NM_013532.2	leukocyte immunoglobulin-like receptor, subfamily B, member 4	6.6386	1.8586	3.5719
16819	NM_008491.1	lipocalin 2	7.4081	2.0432	3.6257
14469	NM_010260.1	guanylate binding protein 2	2.7460	0.6852	4.0077
14528	NM_008102.3	GTP cyclohydrolase 1	1.8177	0.4504	4.0358
103142	NM_153133.2	retinol dehydrogenase 9	0.4608	0.1107	4.1606
100047734	XM_003086912.1	-	2.0998	0.4282	4.9036
408066	XR_035217.2	cDNA sequence BC067074	0.1158	0.0223	5.2008
408066	XR_001629.2	cDNA sequence BC067074	0.1333	0.0244	5.4608
15945	NM_021274.1	chemokine (C-X-C motif) ligand 10	3.9956	0.7027	5.6862
24108	NM_023137.3	ubiquitin D	1.2580	0.2103	5.9809
20311	NM_009141.2	chemokine (C-X-C motif) ligand 5	14.5530	1.9858	7.3284
14825	NM_008176.3	chemokine (C-X-C motif) ligand 1	67.9688	9.1431	7.4339
20297	NM_001159738.1	chemokine (C-C motif) ligand 20	2.3793	0.2933	8.1133
20297	NM_016960.2	chemokine (C-C motif) ligand 20	2.4925	0.2922	8.5293
80287	NM_030255.3	apolipoprotein B mRNA editing enzyme, catalytic polypeptide 3	0.7682	0.0844	9.1014
80287	NM_001160415.1	apolipoprotein B mRNA editing enzyme, catalytic polypeptide 3	0.7372	0.0810	9.1014
55932	NM_018734.3	guanylate binding protein 3	0.9028	0.0755	11.9618
73474	NR_027900.2	small nucleolar RNA host gene (non-protein coding) 9	3.6795	0.2721	13.5221
434775	XR_001556.2	predicted gene 5636	0.2217	0.0142	15.6024
20210	NM_011315.3	serum amyloid A 3	2.6337	0.0000	Inf
18606	NM_001136077.1	ectonucleotide pyrophosphatase/phosphodiesterase 2	0.2264	0.0000	Inf

16069	NM_152839.3	immunoglobulin joining chain	0.2944	0.0000	Inf
630294	NM_001177467.1	predicted gene 7030	0.4033	0.0000	Inf
399570	XR_104919.1	RIKEN cDNA A730085A09 gene	0.1518	0.0000	Inf
229320	NM_153385.2	clarin 1	0.3093	0.0000	Inf
229320	NM_153384.2	clarin 1	0.3005	0.0000	Inf
229320	NM_153386.2	clarin 1	0.3428	0.0000	Inf
12642	NM_009890.1	cholesterol 25-hydroxylase	0.4174	0.0000	Inf