

Cyclic nature of the REM sleep-like state in the cuttlefish *Sepia officinalis*

Iglesias, Teresa L. <sup>a,b,\*</sup>, Boal, Jean G. <sup>c</sup>, Frank, Marcos G. <sup>d</sup>, Zeil, Jochen <sup>e</sup>, Hanlon, Roger T. <sup>f</sup>

<sup>a</sup> Animal Behavior Graduate Group, University of California Davis, Davis, CA, U.S.A.

<sup>b</sup> Physics and Biology Unit, Okinawa Institute of Science and Technology, Okinawa, Japan

<sup>c</sup> Department of Biology, Millersville University, Lancaster, Pennsylvania, U.S.A.

<sup>d</sup> College of Medical Sciences, Washington State University, Spokane, WA, USA.

<sup>e</sup> Research School of Biology, The Australian National University, Canberra, ACT, Australia.

<sup>f</sup> Marine Biological Laboratory, Woods Hole, MA, USA

Teresa.L.Iglesias@gmail.com

orcid.org/0000-0002-0237-8539

\*Corresponding author

Key words: invertebrate sleep, REM sleep, rapid-eye movement, cephalopod, cuttlefish

## Summary statement

These data suggest that cephalopods possess a sleep-like state that resembles behaviorally the vertebrate rapid-eye movement sleep (REMS) state.

## Abstract

Sleep is a state of immobility characterized by three key criteria: an increased threshold of arousal, rapid reversal to an alert state, and evidence of homeostatic "rebound sleep" in which there is an increase of time spent in this quiescent state following sleep deprivation. Common European cuttlefish, *Sepia officinalis*, show states of quiescence during which they meet the last two of these three criteria, yet also show spontaneous bursts of arm and eye movements that accompany rapid changes in chromatophore patterns in the skin. Here we report that this rapid-eye movement sleep-like (REMS-like) state is cyclic in nature. Iterations of the REMS-like state last  $2.42 \pm 0.22$  min ( $\pm$ SE) and alternate with  $34.01 \pm 1.49$  min of the quiescent sleep-like state. These states alternate for durations lasting  $176.89 \pm 36.71$  min. We found clear evidence that this REMS-like state (i) occurs in animals younger than previously reported; (ii) follows an ultradian pattern; (iii) includes intermittent dynamic chromatophore patterning, representing fragments of normal patterning seen in the waking state for a wide range of signaling and camouflage; and (iv) shows variability in the intensity of expression of these skin patterns between and within individuals. These data suggest that cephalopods, which are molluscs with an elaborate brain and complex behavior, possess a sleep-like state that resembles behaviorally the vertebrate REM sleep state, although the exact nature and mechanism of this form of sleep may differ from that of vertebrates.

## Introduction

In mammals, birds, and some reptiles (Libourel et al., 2018; Shein-Idelson, Ondracek, Liaw, Reiter, & Laurent, 2016) (but see (Aulsebrook, Jones, Rattenborg, Roth, & Lesku, 2016)) sleep is characterized neurologically by the occurrence of two distinct brain states denoting slow-wave sleep and rapid-eye-movement sleep (REMS). Behaviorally, slow-wave sleep is quiescent, while REMS is accompanied by rapid eye movements and muscular atonia with occasional muscular twitches (Mascetti, 2016; Stickgold & Walker, 2010). Sleep follows a circadian rhythm and within each sleep bout, slow-wave sleep and REMS follow in an ultradian rhythm (Low, Shank, Sejnowski, & Margoliash, 2008; Martinez-Gonzalez, Lesku, & Rattenborg, 2008; Shein-Idelson et al., 2016; I. Tobler, 1995; Walker & Berger, 1972). Slow-wave sleep alternates with REMS and multiple iterations of REMS can occur within one bout of sleep, with slow-wave sleep always preceding REMS (Stickgold & Walker, 2010). The duration of different sleep states and frequency at which these states alternate (*i.e.* the periodicity), varies across species (John A. Lesku, Roth, Amlaner, & Lima, 2006). Periodicity is widely variable in mammals; for example, humans have a 90-minute periodicity whereas rats have a ten-minute periodicity (Stickgold & Walker, 2010; I. Tobler, 1995). Birds have short sleep periodicities that may last less than a minute, with iterations of REMS just 2-10 seconds in duration (J. A. Lesku & Rattenborg, 2014; Stickgold & Walker, 2010; Walker & Berger, 1972), or up to five minutes in ostriches (John A. Lesku et al., 2011), and slow-wave sleep intervals that range from 10-100 seconds (Low et al., 2008; Martinez-Gonzalez et al., 2008). The recent discovery of REMS in a reptile revealed an 80-second sleep periodicity (Shein-Idelson et al., 2016). An ultradian rhythm, therefore, appears

to be an inherent characteristic of sleep in organisms that experience multiple stages of sleep (Beckers & Rattenborg, 2015; Low et al., 2008; Martinez-Gonzalez et al., 2008; I. Tobler, 1995).

The cuttlefish, *Sepia officinalis* (Figure 1), demonstrates several behavioral indicators that its quiescent sleep-like (QS) state may be analogous to sleep in other organisms (Frank, Waldrop, Dumoulin, Aton, & Boal, 2012). Juvenile and senescing adults were recorded to spend, respectively,  $35.7 \pm 10$  and  $30 \pm 10.3$  percent of time over a 24-hour period in the QS state. This species meets two of the three sleep criteria; animals in the QS state show a rapid reversal to an alert state, and QS-deprivation induces a homeostatic response known as "rebound sleep" (Frank et al., 2012). The third criterion, an increase in arousal threshold during the sleep state, has not yet been successfully examined in this species; however, all three criteria have been tested in another cephalopod, *Octopus vulgaris* (Brown, Piscopo, De Stefano, & Giuditta, 2006; Meisel, Byrne, Mather, & Kuba, 2011) and in many other invertebrates (Kaiser & Steiner-Kaiser, 1983; Nath et al., 2017; Raizen et al., 2008; Ramon, Hernandez-Falcon, Nguyen, & Bullock, 2004; Shaw, Cirelli, Greenspan, & Tononi, 2000). In this study, we video recorded adult, non-senescent, cuttlefish day and night to further examine the REMS-like behavior reported in adult, senescing cuttlefish by Frank et al. (2012).

In the present study, multiple iterations of the REMS-like state were observed in adult, non-senescent animals that were in the QS state and the behaviors were similar to those described in Frank et al. (2012): characterized by rapid chromatophore changes (skin brightness and patterning), skin-texture changes, rapid eye-movement, and arm twitching. Our aim here was to determine if there was evidence of an ultradian rhythm — typical periodic cycle between slow-wave sleep and REMS — using the observable state of rapid chromatophore changes as a

proxy for the REMS-like state. Using measures of pixel intensity, we compared chromatophore changes within and between individuals to assess if chromatophore patterns followed a stereotyped sequence of activation within and/or between individuals. We then examined the rate and nature of body patterning changes across the three states: awake, QS, and REMS-like.

## Methods

Six common cuttlefish, one-year-old *Sepia officinalis* (ca. 7.5cm mantle length), hatched from eggs collected from the English Channel and raised in captivity in the Marine Resources Center at the Marine Biological Laboratory, were transferred singly to one of four 19-liter glass tanks (L x W x D: 40.64cm, 20.32cm, 24.5cm). Visual isolation was accomplished by covering the sides of the tanks with black and white patterned cloth, and white grating was used to constrain cuttlefish to one part of the tank for better video viewing. Animals had ample room to swim (L x W x D: 24.5cm, 20.32cm, 24.5cm). Black plastic sheeting was used to isolate the tank area and reduce human disturbance. At least 1cm of gravel substrate was provided as this seemed to calm the animals, along with a 4cm tall, black cone placed in front of the inflow tube to reduce water agitation and cuttlefish movement due to water flow. Animals were provided thawed frozen shrimp every day. Tanks were supplied with flow-through seawater at a temperature of 15°C. At this temperature, animals grew slowly and did not develop secondary sex characteristics; thereby precluding sex-determination of animals used in this study. Room lighting was on a 12-hour light: 12-hour dark time cycle. One overhead camera and one side-view camera, Sanyo CCD camera model VCB-3384 and Everfocus Polestar II model EQ610, were placed such that they could record activity in one tank at a rate of 30 frames per second in black and white. Red lights (to which cuttlefish exhibited rapid pupil expansion indicating weak

sensitivity) were in constant use to illuminate the dark period to enable video recording of body patterns from above, and eye movement and arm twitching from the side. Video was captured and saved in real time to a hard drive.

Video was reviewed at high speed (4X-32X) approximately every 24-48 hours to determine if an animal had experienced a REMS-like state. After REMS-like behavior was recorded at least once, the animal was returned to the regular holding tanks and a new individual was placed in that observation tank. The camera pair was moved to record an animal that had already spent at least 24 hours in an observation tank. The number of days each animal was observed was as follows: Hippo (28), zZ (12), MrChips (9), Medusa (9), Dumbo (7), and Clyde (4).

Durations of REMS-like and QS states were determined from the video using the video time code for each recording. Video clips were made of the REMS-like states including a 30 s lead-in of QS state before the start of the REMS-like state and ending approximately 5 s after the animal expelled water through the siphon and/or repositioned itself on the substrate (Figure 2). These behaviors were performed consistently after the REMS-like state. Following these movements, animals either resumed the QS state or became active with open pupils. The pixel intensity data for this 30 s lead in were excluded when calculating the distance matrix for the hierarchical clustering dendrogram and for the Mantel tests (see below). Overhead and side-view camera footage of REMS-like states were combined in one video using Adobe After Effects (Figure 2, Video S1-S3) so that QS states could be confirmed (animal partially buried in substrate with pupils closed (Frank et al., 2012)).

Pixel value measurements were used to assess chromatophore activity; they were performed using custom-written MATLAB code (MathWorks, Natick, MA, USA). Most animals

were relatively immobile as they were slightly buried in the substrate; however, one animal, zZ, drifted during the REMS-like state because the substrate was too shallow as a result of repeated self-burying in the same spot. Videos for zZ were digitized using Adobe After Effects, using the motion-tracking feature so that the region of interest (head or mantle) was constantly at the center of view. Video clips of REMS-like states were analyzed in MATLAB. A square 5 x 5 pixel region was selected on the head and on the mantle of the cuttlefish (Figure 2). The average pixel values (ranging from 0 to 255 in 8bit grey level images) were collected for each video frame (30fps) over the entire video clip for these two regions (Figure 2).

The absolute pixel values were affected by the amount of ambient light in the video. Because the aim was to compare patterns of pixel value changes throughout the REMS-like state within and between individuals and across different iterations of the REMS-like state, the pixel value at the beginning of sequences was subtracted from all subsequent values so that all REMS-like iterations had a standardized start at zero (see Figure 2). In other words, in the standardized data, positive numbers indicate brightening of pixels and negative numbers indicate darkening of pixels. The REMS-like state began with a rapid darkening of the head and mantle region. We visually determined from the videos that a darkening by approximately 10 mean pixel values coincided with the start of the REMS-like state and only used the subsequent sequence for our analysis (see Figure 2).

The temporal structure of body patterning was compared between and within individuals using the R package *dtwclust* (Sarda-Espinosa, 2016) which calculated all pair-wise distances/dissimilarities between pixel value series for all 55 iterations of the REMS-like state. This package stretches or condenses signals, based on user specified settings, to determine how similar (or different) signals are to one another (Figure S1). The settings disallowing unlimited



stretching and condensing of signals were determined using the R package *dtw* (Giorgino, 2009) and were as follows: `step=symmetric2`, `window.type=slantedband`, and `window.size=30`. In *dtwclust* the function ‘`hclust`’ was used to build a hierarchical dendrogram for head and mantle data with animal and iteration of REMS-like state annotated at the tips to visually assess clustering. The R package *ape* (Paradis, Claude, & Strimmer, 2004) was used to format the hierarchical dendrogram.

The R package *Ecodist* (Goslee & Urban, 2007) was used to perform Mantel tests between the pixel intensity distance matrix and binary matrices for animal identity and REMS-like iteration. Each binary matrix was compared against the pixel distance matrix independently. All Mantel tests were performed using 100,000 permutations, sampling with replacement, and 100,000 iterations for bootstrapped confidence intervals. A Mantel correlogram was examined to understand the structure of the relationship when the Mantel *r* statistic was significant.

The duration of the REMS-like state varied between individuals and REMS-like iteration, resulting in pixel intensity series of differing lengths; however, interpolating data onto the ends of the series to make them of equal length did not change results. Interpolated data were not used, therefore.

The rate and nature of body patterning changes were determined by analyzing video sequences of three cuttlefish during awake, quiescent and REMS-like states. To provide consistency of data gathering, one coauthor (with long experience of this species’ body patterns (RTH)) performed all analyses. The technique was to watch video and count the number of changes in either the whole body pattern, or with the transitory expression of chromatic components of body patterns (as described by Hanlon & Messenger, 1988). Notes were made of components or patterns that have never been seen in naturally behaving animals. Data were

analyzed via model comparison using size-corrected Akaike Information Criterion (AICc), which penalizes the addition of excess parameters. We compared two models: one null model represented by the intercept-only model and one model where state (awake, QS, REMS-like) was the only fixed predictor. Models contained animal identity as a random effect. We used the R package *lme4* (Bates, Maechler, Bolker, & Walker, 2015) to create and compare models using AICc scores. The R package *AICcmodavg* (Mazerolle, 2014) was used to calculate parameter estimates, standard errors, and confidence intervals.

## Results

Animals primarily maintained some variation of the mottle pattern (Chiao et al., 2009) unless feeding or disturbed by human activity. The number of hours of video recorded for each animal is as follows: Hippo (282), zZ (187), MrChips (159), Medusa (167), Dumbo (70), and Clyde (54). During sleep-like bouts, when the animals were not in the REMS-like state, body patterns consisted mostly of mottle variants. In total, 55 REMS-like iterations were observed (Table S1). All six animals were recorded demonstrating at least one sleep-like bout with the REMS-like state alternating with the QS state (Figure 3, Figure S2). The number of sleep-like bouts and REMS-like iterations for each animal is as follows: Hippo (2, 15), zZ (1, 10), MrChips (2, 6), Medusa (3, 11), Dumbo (1, 9), and Clyde (1, 4). Duration of the REMS-like iteration was on average  $2.42 \pm 0.22$  min ( $\pm$ SE,  $n=55$ ) and the time between REMS-like iterations (*i.e.* QS state intervals,  $n=45$ ) was  $34.01 \pm 1.49$  min (Table S1). We defined the periodicity as the time between onset of one REMS-like iteration to the onset of the next REMS-like iteration (Figure 3B, marked by eye squinting, discussed below); this resulted in a  $36.34 \pm 1.46$  min periodicity. On average, animals spent 13.36 min total (range = 1.93 - 23.07 min) in the REMS-like state

during a  $176.89 \pm 36.71$  min sleep bout (Table S1, n=10). Therefore, cuttlefish spent approximately 7% of the sleep-like state in the REMS-like state.

We observed arm movements by every individual in 55 iterations and eye movements in 54 out of 55 (side view was obscured once by condensation on the tank glass). Eye movements and concurrent chromatophore patterning around the eyes and brain (collectively known as the “head” in cephalopods) included peculiar features that best characterize the REMS-like state of *Sepia officinalis*. The eyes shifted fore and aft (or left and right when viewed from the side) in all animals, and the W-shaped slit pupil sometimes dilated to a round shape. Yet the more common eye behavior was a gross “squinting” of the whole eye socket in which various eye muscles contracted and squeezed the surrounding tissue to occlude the pupil. A close-up example of these strong eye movements can be seen in Video S1. Often this behavior was accompanied by sudden darkening of all the chromatophores around each eye and between the eyes, forming a “dark head bar” that has not been seen in normally behaving cuttlefish during daytime (Hanlon et al., 2009; Hanlon & Messenger, 1988). The darkening of the skin in cuttlefish is a result of direct neural stimulation from lower motor centers in the brain to muscles that contract to stretch the chromatophore pigment sac (Messenger, 2001) while muscle relaxation is concomitant with pale or white skin coloration. Sometimes the whole head was flattened as if the cuttlefish was ducking. It is worth mentioning with regard to the expectation of loss of muscle tone at the onset of REMS that Video S2 shows an animal with arms raised, which relax just as the REMS-like iteration begins. Also a white head bar was always observed to precede the REMS-like state but was not useful in determining whether and when a REMS-like state would occur. These various muscular contractions of the eyes, head, chromatophores, and arms seem akin to the myoclonic twitching and phasic activity in birds and mammals in the REM state.

Arm movements included slight twitching, raising of the 1<sup>st</sup> pair, and movement of all arms (see Video S2 for arm raising). The only arm movement observed in all 55 iterations was slight arm twitching. Collectively, these represent conspicuous dramatic behaviors that are rarely or never seen in day-active cuttlefish; that is, they are REMS-like state specific and their order of appearance was not fixed but variable. The white head bar was present in all animals just prior to the REMS-like state and all animals performed this gross squinting, which coincided with the rapid chromatophore darkening at the start of what we considered the REMS-like state. The eye squint was used to mark the start of the REMS-like state for duration measurements.

The videos revealed episodic expression, during the REMS-like state, of several of the 34 chromatic components of body patterns that *Sepia officinalis* is known to use for signaling and camouflage (Hanlon & Messenger, 1988) (Video S3). For example, cuttlefish showed each of the following: (i) mottle patterns (Chiao et al., 2009), a common camouflage pattern for primary defense; (ii) various expressions of deimatic (threat, startle) patterns, such as the uni- or bilateral paired mantle spots, which are secondary defenses; (iii) and dark flashing of the mantle or whole body, or the Passing Cloud pattern, which are examples of protean defense. These expressions were transitory and were not usually full or normal expressions of the patterns as seen in field or laboratory behavioral studies (Adamo, Kelly, Cheryl, & Ivy, 2006; Hanlon et al., 2009; Hanlon & Messenger, 1988, 2017; Langridge, 2006; Langridge, Broom, & Osorio, 2007; Staudinger et al., 2013). Most of the body patterns exhibited during the REMS-like state were not observed in these animals outside of the REMS-like state.

The rate and nature of change in body patterning differed greatly in the REMS-like state compared to the QS and awake states (summarized in Table 1, raw data Table S2). In ~28 minutes of the REMS-like state, cuttlefish were changing body patterns (either whole or in part)

36 – 65 times per minute (Table S2, see also Notes for Table S2 in Supplementary Materials). By comparison, cuttlefish in the QS state changed appearance only 0.35 – 3.88 times per minute. This is greater than a ~20x difference in rate on average. These changes can be compared to awake cuttlefish, which had a stable camouflage pattern deployed the vast majority of time but did change occasionally at very low rates of 0.39 – 0.99 times per minute. It is particularly noteworthy that in over 90% of the cases, body pattern and chromatic components shown during REMS-like sleep differed from those of naturally behaving, awake (eyes open) animals (Figure 4). In further analysis, AICc model selection strongly supports state (awake, QS, REMS-like) as a predictor over the null model with  $\Delta 69.92$  and an AICc model weight of 1. Standard practice requires a minimum of  $\Delta 2$  to consider a model better at describing the data than others in the set (Burnham & Anderson, 2013). Model averaged parameter estimates (relative to awake state) calculate the rate of pattern change in the REMS-like state as  $50.08 \pm 3.33$  ( $\pm$  unconditional SE) with [43.56 - 56.61] unconditional 95% confidence interval. The parameter estimate for the QS state is  $-0.67 \pm 3.54$ , relative to the awake state, with [-7.61 – 6.28] as the 95% confidence interval.

To quantify the variability of skin chromatophore patterning activity (to corroborate the species-specific skin displays), the distance matrix containing all pair-wise comparisons of REMS-like iterations was visualized in a hierarchical clustering dendrogram with animal ID and iteration tip labels for the two body regions (head - Figure 5; and mantle - Figure S3). No robust clustering by animal or by iteration (1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup>... REMS-like iteration within a cycle) was found for either body region. A Mantel statistical test comparing the pixel series distance matrix to a binary matrix for the head region showed a weak effect of animal identity: Mantel  $r = 0.09$ , two-tailed  $p < 0.05$ , and 95% confidence interval for Mantel  $r$  of [0.06, 0.14]. Results for pixel

intensity distance data from the mantle of the cuttlefish were similar but not significant with Mantel  $r = 0.07$ ,  $p = 0.19$ , and 95% confidence interval of [0.04, 0.12]. Tests showed no effect of iteration for either head or mantle regions: Mantel  $r = 0.002$ , two-tailed  $p = 0.97$ , and 95% confidence interval of [-0.03, 0.04] for the head region and Mantel  $r = -0.006$ ,  $p = 0.92$ , and 95% confidence interval of [-0.11, 0.04] for the mantle region. A Mantel correlogram was computed to examine the underlying structure of the data where pixel series distance correlated significantly with animal identity. We found that REMS-like iterations that were more similar could be attributed to the same animal with Mantel  $r \sim +0.14$  and that iterations that were more dissimilar could be attributed to different animals with Mantel  $r \sim -0.07$  (Figure S4).

## Discussion

Slow-wave sleep and REM sleep (REMS) states have been demonstrated in birds, mammals (Jerome M. Siegel, 2008) and reptiles (Libourel et al., 2018; Shein-Idelson et al., 2016). Here we show that cuttlefish in a quiescent, sleep-like state periodically undergo a REMS-like state, characterized by (i) general immobility with occasional muscle twitching (arms and neuromuscular chromatophore organs), (ii) rapid horizontal movements of the eyes, (iii) alternating quiescent-sleep (QS) and REMS-like states in a predictable ultradian rhythm, and (iv) a significant increase in the number and combinations of neurally controlled skin pattern changes compared to the awake state. In octopuses, similar observations of quiescent behavior have been reported, including (i) circadian rhythms in activity, (ii) quiet periods characterized by closed eyes and a lack of activity, (iii) rebound when quiescence was prevented, and (iv) heightened sensory thresholds during quiet periods (Brown et al., 2006; Meisel et al., 2011). There are also anecdotal reports in octopuses of body patterning inconsistent with camouflage during this

quiescent state (Meisel et al., 2011). If borne out by more quantitative measures (brain as well as behavioral), collectively this would suggest that some cephalopods including the common cuttlefish, *Sepia officinalis*, and possibly the common octopus, *Octopus vulgaris*, experience a state akin to REM sleep.

Frank et al. (2012) described REMS-like behavior in adult, senescing cuttlefish but only observed a single REMS-like iteration per animal. In the present study, we recorded 55 REMS-like iterations (up to 15 in a single cuttlefish) in six non-senescing cuttlefish of the same species. The behaviors exhibited in both studies included spontaneous and rapid changes in patterns and brightness, arm twitching, and eye movements. It is not implausible that the behaviors observed and reported in Frank et al. (2012) of senescing adults could have been a result of the deterioration of physiological systems that can affect sleep behaviors in senescent animals (Anderson, Wood, & Byrne, 2002; Chichery & Chichery, 1992; Pandi-Perumal et al., 2002); especially since, as discussed below, such behavior was not detected in younger cuttlefish. However, our present demonstration that the REMS-like state occurs in adult, non-senescing cuttlefish nullifies this possibility.

Frank et al. (2012) also tested juvenile cuttlefish and observed homeostatic regulation of the QS state; however, they did not observe the REMS-like state in juveniles. One possible explanation presented in Frank et al. 2012 is potential neural immaturity of sleep-specific brain areas. Alternatively, the REMS-like state may be decreased or absent in situations that are stressful and demand increased vigilance. For example, most species show a circadian periodicity of REM; however, marine mammals, such as pinnipeds, show significantly decreased durations of REM sleep while in the water as opposed to when on land (O. Lyamin et al., 2008; O. I. Lyamin et al., 2018) and the frigatebird (*Fregata minor*) decreases REM sleep while in

flight (Rattenborg et al., 2016). Frank et al. (2012) also noted that the juvenile cuttlefish in their study may not have been provided a sufficiently long acclimation period. Technical issues prevented uninterrupted recording of cuttlefish in our study; however, the longest continuous recordings allowed us to observe two animals (Hippo and zZ) for 53 and 40 hours, respectively. Although the animals appear to experience the QS state, they did not show REMS-like states during these continuous recordings. The REMS-like state had been observed in Hippo prior to this time block, suggesting the animal was acclimated. If we allow that stress effects are not responsible for altering the expression of the REMS-like state in these animals, an alternative explanation may be that it is not physiologically necessary for *Sepia officinalis* to experience this state every 24-hours. Elephants (*Loxodonta africana*) and the walrus (*Odobenus rosmarus*) have been reported to forego REM sleep for as long as four days (Gravett et al., 2017; Pryaslova, Lyamin, Siegel, & Mukhametov, 2009). It is important to consider that these rapid chromatophore pattern changes likely interfere with successful camouflage and may be costly in terms of predation risk. This may reflect a trade-off between meeting the presumed physiological need for the REMS-like state while limiting periods of risky conspicuousness. Future behavioral study in juvenile and adult cuttlefish will be necessary to explore these possibilities.

Pixel intensity changes in small patches of chromatophores recorded in REMS-like iterations were not identical within individuals, nor were they identical between animals (Figure 5, Figure S3). Moreover, the expressions of individual chromatic components, as well as whole body patterns, were always highly transitory in the REMS-like state and usually different from the way they are expressed in awake cuttlefish. They were expressed in a seemingly disorganized manner; i.e. a typical camouflage pattern such as Mottle was shown often but very briefly (seconds), whereas in awake animals this chronic pattern is maintained for many minutes or



hours. The Mottle pattern was only shown acutely in cuttlefish in the REMS-like state, however, and was preceded or followed by patterns that function as conspicuous displays such as Deimatic and Passing Cloud (Adamo et al., 2006). There were some similarities among REMS-like iterations (Video S3), however. For example, all animals performed the Passing Cloud display and most performed symmetrical/asymmetrical activation of mantle deimatic spots. These displays are associated with antipredator behavior and are therefore normally expressed in response to a perceived threat (Hanlon & Messenger, 1988; Langridge, 2006). None of the animals were in visual contact with other animals or with humans; it is possible — but highly unlikely — that the body pattern changes seen during the REMS-like state in these experiments were a result of artifactual stimulation outside of the experimental chamber, such as ground-transmitted vibrations. Given that the system had a continuous inflow of local seawater, it is also possible that a chemical cue was responsible for eliciting these body patterns; however, all tanks received the same water and the REMS-like state was not observed in multiple cuttlefish simultaneously. Thus, it is highly likely that the cuttlefish were indeed exhibiting a typical behavior associated with the sleep-like state.

We recognize our methods may underestimate the similarity in gross pattern performance within and between animals because we compared pixel intensity changes for relatively small areas of the animal over temporally short periods of time. The animals actually performed many recognizable components of their patterning repertoire as mentioned above, but did not express these components in the same order or at the same time across REMS-like iterations. For example, all individuals displayed, either bilaterally or unilaterally, the 3<sup>rd</sup> pair of deimatic spots on the mantle; however, the appearance of the spots did not occur at similar time points within the REMS-like iteration or even in all REMS-like iterations by the same individual (Video S3).

The only recognizable common chromatophore pattern that was displayed in a temporally predictable manner was the presence of the white head bar, which turned dark (never before seen) and was accompanied by the eye squint, marking the start of the REMS-like state. All other components were recognizable but the order in which they were performed and the duration of appearance was variable. Many of these components are normally expressed in response to predators or perceived threats or are used in camouflage, yet the testing environment was static with regard to substrate, structures and visual cues.

Our findings suggest that during the REMS-like state, fragments of normal waking activity in these areas are spontaneously re-activated, but in combinations not observed in wakefulness. This is reminiscent of mammalian REM sleep, which also contains fragments of waking patterns of activity in motor, limbic and cortical circuits (Matarazzo et al., 2011). This is revealed most dramatically in mammals that lack the normal paralysis of REM sleep. Under such conditions, during REM sleep animals will display components of complete waking behaviors, including aggressive displays, orientation and stalking behavior (Dumoulin Bridi et al., 2015; Louie & Wilson, 2001; Morrison, 1979). Moreover, in REM sleep remote and recent memories are accessed and combined in complex ways not typically experienced in the waking state (Hobson, Pace-Schott, & Stickgold, 2000). While speculative, it is possible that the unusual combinations of chromatophore activity during REMS-like states reflect a similar process in *Sepia officinalis*.

Although not the prime target of this study, the dynamics of brightness changes would be worth investigating more quantitatively. Of particular note, (1) all REMS-like iterations begin with a rapid darkening that is synchronized between head and mantle regions of interest (bottom left panel in (Figure 6A), but can also involve delays of several seconds (bottom left panel in

(Figure 6B)); (2) overall, brightness changes in the head and the mantle region are correlated, often with a delay of 30 to 100 ms (insets in top panels in (Figure 6A and B)); (3) such correlations are not present, however, in the fine-scale brightness change on short time scales (Figure 6, bottom panels); (4) rapid brightness changes occur in discrete steps, even before the REMS-like state starts (Figure 6, bottom panels), but (5) can also involve bursts of smooth oscillatory changes (Figure 6, bottom right panels); (6) both head and mantle regions tend to brighten slowly throughout the course of a REMS-like iteration (top panels Figure 6A and B). Given that these brightness changes reflect chromatophore dynamics and the activity of neural networks, it would be interesting to compare the temporal organization of brightness changes in awake and “sleeping” cuttlefish and to eventually relate them to changes in chromatophores and to the neural activity orchestrating them.

If future work reveals the cephalopod QS and REMS-like states to be analogous to vertebrate slow-wave and REM sleep, then it is worth remarking here that the periodicity of the REMS-like state in the cuttlefish appears more similar to the periodicity of REM sleep in small mammals than to birds or reptiles (Shein-Idelson et al., 2016; Stickgold & Walker, 2010; I. Tobler, 1995; Walker & Berger, 1972). The average time an animal spent in the REMS-like state across a single QS-REMS-like cycle (~7%) is similar to small mammals and birds. Regardless of whether sleep in animals resulted from single or multiple evolutionary origins, this phylogenetic distribution of patterns of periodicity, total duration of specific sleep-states, and flexibility to delay REM sleep (or REMS-like states), promises a fruitful avenue towards identifying ecological pressures that shape sleep behavior (Aulsebrook et al., 2016).

The possible occurrence of sleep and REMS-like sleep in a clade of organisms that is distantly related to any clade where REM sleep has been observed previously poses a challenging and important opportunity to broaden our understanding of sleep and REM sleep (Corner, 2013; J. M. Siegel, 1995; Irene Tobler, 1988). Cephalopods have the largest and most complex brains — and the most diverse repertoire of behaviors — of any invertebrate animal (Hanlon & Messenger, 2017; Nixon & Young, 2003). Whether future work demonstrates that cuttlefish adhere to all the criteria currently deemed necessary for a legitimate sleep state, or prove to be an exception to the general pattern described in the literature for vertebrates, examining these questions in an organism that is ecologically distinct and phylogenetically distant from current model organisms should serve to inform the broader questions of how to accurately define sleep, discover its functions, and understand its evolutionary origins.

#### Acknowledgments

We thank the Grass Foundation for support and the summer 2012 MBL Grass Laboratory Directors and Fellows for early feedback and assistance. We thank the staff at the MBL for their indispensable technical and logistical assistance. Thanks to members of the Hanlon lab for culturing the cuttlefish. TLI would like to acknowledge the Physics and Biology Unit at the Okinawa Institute of Science and Technology. JGB thanks the Millersville University Faculty Grants Committee and RTH thanks the Sholley Foundation for partial support.

Funding: The GRASS Foundation, Millersville University Faculty Grants Committee, and The Sholley Foundation.

## Literature cited

- Adamo, S. A., Kelly, E., Cheryl, S., & Ivy, W. (2006). Signaling to the Enemy? Body Pattern Expression and Its Response to External Cues during Hunting in the Cuttlefish *Sepia officinalis* (Cephalopoda). *Biological Bulletin*, 210(3), 192-200. doi: 10.2307/4134557
- Anderson, R. C., Wood, J. B., & Byrne, R. A. (2002). Octopus Senescence: The Beginning of the End. *Journal of Applied Animal Welfare Science*, 5(4), 275-283. doi: 10.1207/S15327604JAWS0504\_02
- Aulsebrook, A. E., Jones, T. M., Rattenborg, N. C., Roth, T. C., 2nd, & Lesku, J. A. (2016). Sleep Ecophysiology: Integrating Neuroscience and Ecology. *Trends Ecol Evol*, 31(8), 590-599. doi: 10.1016/j.tree.2016.05.004
- Bates, D., Maechler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, 67(1), 1-48. doi: 10.18637/jss.v067.i01
- Beckers, G. J. L., & Rattenborg, N. C. (2015). An in depth view of avian sleep. *Neuroscience and Biobehavioral Reviews*, 50, 120-127. doi: 10.1016/j.neubiorev.2014.07.019
- Brown, E. R., Piscopo, S., De Stefano, R., & Giuditta, A. (2006). Brain and behavioural evidence for rest-activity cycles in *Octopus vulgaris*. *Behavioural Brain Research*, 172(2), 355-359. doi: 10.1016/j.bbr.2006.05.009
- Burnham, K. P., & Anderson, D. R. (2013). *Model Selection and Inference: A Practical Information-Theoretic Approach*: Springer New York.
- Chiao, C.-C., Chubb, C., Buresch, K. C., Barbosa, A., Allen, J. J., Mäthger, L. M., & Hanlon, R. T. (2009). Mottle camouflage patterns in cuttlefish: quantitative characterization and visual background stimuli that evoke them. *The Journal of Experimental Biology*, 213(2), 187.
- Chichery, M. P., & Chichery, R. (1992). Behavioural and neurohistological changes in aging *Sepia*. *Brain Res*, 574(1-2), 77-84.
- Corner, M. A. (2013). Call it sleep -- what animals without backbones can tell us about the phylogeny of intrinsically generated neuromotor rhythms during early development. *Neurosci Bull*, 29(3), 373-380. doi: 10.1007/s12264-013-1313-3
- Dumoulin Bridi, M. C., Aton, S. J., Seibt, J., Renouard, L., Coleman, T., & Frank, M. G. (2015). Rapid eye movement sleep promotes cortical plasticity in the developing brain. *Science Advances*, 1(6).
- Frank, M. G., Waldrop, R. H., Dumoulin, M., Aton, S., & Boal, J. G. (2012). A preliminary analysis of sleep-like states in the cuttlefish *Sepia officinalis*. *PLoS ONE*, 7(6), e38125. doi: 10.1371/journal.pone.0038125
- Giorgino, T. (2009). Computing and Visualizing Dynamic Time Warping Alignments in R: The dtw Package. 2009, 31(7), 24. doi: 10.18637/jss.v031.i07
- Goslee, S. C., & Urban, D. L. (2007). The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software*, 22(7), 1-19.
- Gravett, N., Bhagwandin, A., Sutcliffe, R., Landen, K., Chase, M. J., Lyamin, O. I., . . . Manger, P. R. (2017). Inactivity/sleep in two wild free-roaming African elephant matriarchs – Does large body size make elephants the shortest mammalian sleepers? *PLOS ONE*, 12(3), e0171903. doi: 10.1371/journal.pone.0171903
- Hanlon, R. T., Chiao, C. C., Mäthger, L. M., Barbosa, A., Buresch, K. C., & Chubb, C. (2009). Cephalopod dynamic camouflage: bridging the continuum between background matching and disruptive coloration. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1516), 429.
- Hanlon, R. T., & Messenger, J. B. (1988). Adaptive coloration in young cuttlefish (*Sepia-officialis* L) - the morphology and development of body patterns and their relation to behavior. *Philosophical*

- Transactions of the Royal Society of London Series B-Biological Sciences*, 320(1200), 437-&. doi: 10.1098/rstb.1988.0087
- Hanlon, R. T., & Messenger, J. B. (2017). *Cephalopod behaviour* (2nd ed.). Cambridge: Cambridge University Press.
- Hobson, J. A., Pace-Schott, E. F., & Stickgold, R. (2000). Dreaming and the brain: toward a cognitive neuroscience of conscious states. *Behav Brain Sci*, 23(6), 793-842; discussion 904-1121.
- Kaiser, W., & Steiner-Kaiser, J. (1983). Neuronal correlates of sleep, wakefulness and arousal in a diurnal insect. *Nature*, 301, 707. doi: 10.1038/301707a0
- Langridge, K. V. (2006). Symmetrical crypsis and asymmetrical signalling in the cuttlefish *Sepia officinalis*. *Proceedings of the Royal Society B: Biological Sciences*, 273(1589), 959-967. doi: 10.1098/rspb.2005.3395
- Langridge, K. V., Broom, M., & Osorio, D. (2007). Selective signalling by cuttlefish to predators. *Current Biology*, 17(24), R1044-R1045.
- Lesku, J. A., Meyer, L. C. R., Fuller, A., Maloney, S. K., Dell'Omo, G., Vyssotski, A. L., & Rattenborg, N. C. (2011). Ostriches Sleep like Platypuses. *PLOS ONE*, 6(8), e23203. doi: 10.1371/journal.pone.0023203
- Lesku, J. A., & Rattenborg, N. C. (2014). Avian sleep. *Current Biology*, 24(1), R12-R14.
- Lesku, J. A., Roth, T. C., II, Amlaner, C. J., & Lima, S. L. (2006). A phylogenetic analysis of sleep architecture in mammals: The integration of anatomy, physiology, and ecology. *American Naturalist*, 168(4), 441-453. doi: 10.1086/506973
- Libourel, P.-A., Barrillot, B., Arthaud, S., Massot, B., Morel, A.-L., Beuf, O., . . . Luppi, P.-H. (2018). Partial homologies between sleep states in lizards, mammals, and birds suggest a complex evolution of sleep states in amniotes. *PLoS biology*, 16(10), e2005982-e2005982. doi: 10.1371/journal.pbio.2005982
- Louie, K., & Wilson, M. A. (2001). Temporally structured replay of awake hippocampal ensemble activity during rapid eye movement sleep. *Neuron*, 29(1), 145-156. doi: 10.1016/s0896-6273(01)00186-6
- Low, P. S., Shank, S. S., Sejnowski, T. J., & Margoliash, D. (2008). Mammalian-like features of sleep structure in zebra finches. *Proceedings of the National Academy of Sciences*, 105(26), 9081-9086. doi: 10.1073/pnas.0703452105
- Lyamin, O., Kosenko, P., Lapierre, J., Pryaslova, J., Vyssotski, A., Lipp, H., . . . Mukhametov, L. (2008). Study of sleep in a walrus. *Sleep*, 31, A24-A25.
- Lyamin, O. I., Kosenko, P. O., Korneva, S. M., Vyssotski, A. L., Mukhametov, L. M., & Siegel, J. M. (2018). Fur Seals Suppress REM Sleep for Very Long Periods without Subsequent Rebound. *Curr Biol*, 28(12), 2000-2005.e2002. doi: 10.1016/j.cub.2018.05.022
- Martinez-Gonzalez, D., Lesku, J. A., & Rattenborg, N. C. (2008). Increased EEG spectral power density during sleep following short-term sleep deprivation in pigeons (*Columba livia*): evidence for avian sleep homeostasis. *Journal of Sleep Research*, 17(2), 140-153. doi: 10.1111/j.1365-2869.2008.00636.x
- Mascetti, G. G. (2016). Unihemispheric sleep and asymmetrical sleep: behavioral, neurophysiological, and functional perspectives. *Nature and Science of Sleep*, 8, 221-237. doi: 10.2147/nss.s71970
- Matarazzo, L., Foret, A., Mascetti, L., Muto, V., Shaffii, A., & Maquet, P. (2011). A systems-level approach to human REM sleep. In A. R. Morrison, B. N. Mallick, R. W. McCarley & S. R. Pandi-Perumal (Eds.), *Rapid Eye Movement Sleep: Regulation and Function* (pp. 71-79). Cambridge: Cambridge University Press.
- Mazerolle, M. J. (2014). AICcmodavg: Model selection and multimodel inference based on (Q)AIC(c). R package version 2.00. <http://CRAN.R-project.org/package=AICcmodavg>.
- Meisel, D. V., Byrne, R. A., Mather, J. A., & Kuba, M. (2011). Behavioral sleep in *Octopus vulgaris*. *Vie Et Milieu-Life and Environment*, 61(4), 185-190.

- Messenger, J. B. (2001). Cephalopod chromatophores: neurobiology and natural history. *Biological Reviews*, 76(4), 473-528. doi: 10.1017/S1464793101005772
- Morrison, A. R. (1979). Relationships between phenomena of paradoxical sleep and their counterparts in wakefulness. *Acta Neurobiol Exp (Wars)*, 39(6), 567-583.
- Nath, R. D., Bedbrook, C. N., Abrams, M. J., Basinger, T., Bois, J. S., Prober, D. A., . . . Goentoro, L. (2017). The jellyfish *Cassiopea* exhibits a sleep-like state. *Current Biology*. doi: 10.1016/j.cub.2017.08.014
- Nixon, M., & Young, J. Z. (2003). *The brains and lives of cephalopods*. Oxford: Oxford University Press.
- Pandi-Perumal, S. R., Seils, L. K., Kayumov, L., Ralph, M. R., Lowe, A., Moller, H., & Swaab, D. F. (2002). Senescence, sleep, and circadian rhythms. *Ageing Research Reviews*, 1(3), 559-604. doi: 10.1016/s1568-1637(02)00014-4
- Paradis, E., Claude, J., & Strimmer, K. (2004). APE: analyses of phylogenetics and evolution in R language. *Bioinformatics*, 20, 289-290.
- Pryaslova, J. P., Lyamin, O. I., Siegel, J. M., & Mukhametov, L. M. (2009). Behavioral sleep in the walrus. *Behavioural Brain Research*, 201(1), 80-87. doi: <https://doi.org/10.1016/j.bbr.2009.01.033>
- Raizen, D. M., Zimmerman, J. E., Maycock, M. H., Ta, U. D., You, Y. J., Sundaram, M. V., & Pack, A. I. (2008). Lethargus is a *Caenorhabditis elegans* sleep-like state. *Nature*, 451(7178), 569-572. doi: 10.1038/nature06535
- Ramon, F., Hernandez-Falcon, J., Nguyen, B., & Bullock, T. H. (2004). Slow wave sleep in crayfish. *Proceedings of the National Academy of Sciences of the United States of America*, 101(32), 11857-11861. doi: 10.1073/pnas.0402015101
- Rattenborg, N. C., Voirin, B., Cruz, S. M., Tisdale, R., Dell'Omo, G., Lipp, H. P., . . . Vyssotski, A. L. (2016). Evidence that birds sleep in mid-flight. *Nature Communications*, 7. doi: 10.1038/ncomms12468
- Sarda-Espinosa, A. (2016). dtwclust: Time series clustering along with optimizations for the dynamic time warping distance. Retrieved from <http://CRAN.R-project.org/package=dtwclust>
- Shaw, P. J., Cirelli, C., Greenspan, R. J., & Tononi, G. (2000). Correlates of sleep and waking in *Drosophila melanogaster*. *Science*, 287(5459), 1834-1837.
- Shein-Idelson, M., Ondracek, J. M., Liaw, H.-P., Reiter, S., & Laurent, G. (2016). Slow waves, sharp waves, ripples, and REM in sleeping dragons. *Science*, 352(6285), 590-595. doi: 10.1126/science.aaf3621
- Siegel, J. M. (1995). Phylogeny and the function of REM sleep. *Behavioural Brain Research*, 69(1-2), 29-34. doi: 10.1016/0166-4328(95)00023-m
- Siegel, J. M. (2008). Do all animals sleep? *Trends in Neurosciences*, 31(4), 208-213. doi: 10.1016/j.tins.2008.02.001
- Staudinger, M. D., Buresch, K. C., Mäthger, L. M., Fry, C., McNulty, S., Ulmer, K. M., & Hanlon, R. T. (2013). Defensive responses of cuttlefish to different teleost predators. *Biological Bulletin*, 225(3), 161-174.
- Stickgold, R., & Walker, M. P. (2010). *The Neuroscience of Sleep*: Elsevier Science.
- Tobler, I. (1988). Evolution and Comparative Physiology of Sleep in Animals. In R. Lydic & J. F. Biebuyck (Eds.), *Clinical Physiology of Sleep* (pp. 21-30). New York, NY: Springer New York.
- Tobler, I. (1995). Is sleep fundamentally different between mammalian-species? *Behavioural Brain Research*, 69(1-2), 35-41. doi: 10.1016/0166-4328(95)00025-o
- Walker, J. M., & Berger, R. J. (1972). Sleep in the domestic pigeon (*Columba livia*). *Behavioral Biology*, 7(2), 195-203. doi: [https://doi.org/10.1016/S0091-6773\(72\)80199-8](https://doi.org/10.1016/S0091-6773(72)80199-8)

animal ID	state	total minutes	total changes	# changes per minute
Clyde	awake	41.00	25	0.61
	REMS-like	6.58	257	39.04
	QS	36.76	96	2.61
Dumbo	awake	40.63	16	0.39
	REMS-like	10.82	613	56.67
	QS	45.96	39	0.85
MrChips	awake	41.20	36	0.87
	REMS-like	10.90	618	56.70
	QS	41.75	23	0.55

Table 1. Rate of body pattern changes per minute in three different states: awake state, quiescent sleep-like state (QS), and rapid eye movement sleep-like state (REMS-like). At least 3 different iterations of the REMS-like state were analyzed for each of three animals and approximately 40 minutes were analyzed per animal for the QS and awake states.



## Figures

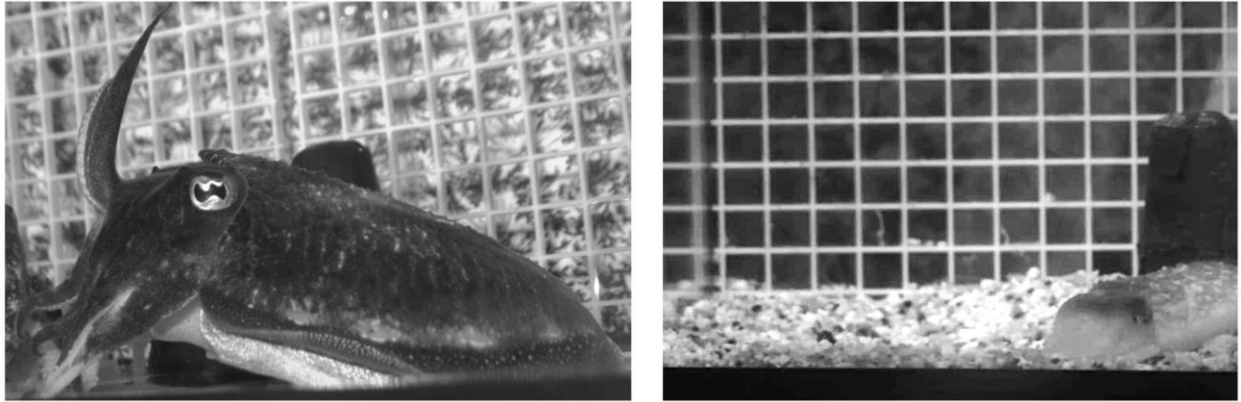


Figure 1. Cuttlefish in awake state with pupils partly dilated, arms raised in an alarm pattern (left). Cuttlefish in quiescent sleep-like (QS) state, partly buried in substrate with pupils closed (right).

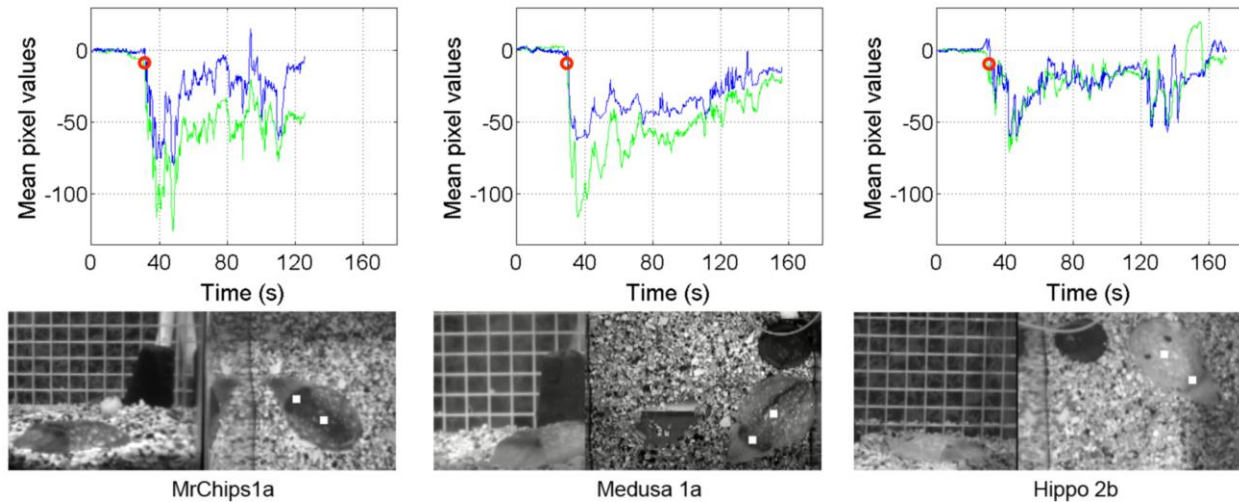


Figure 2. Stills of video analyzed next to pixel intensity measures for three cuttlefish (MrChips, Medusa, and Hippo). The white squares indicate regions of interest measured. The graphs show the pixel intensity measures for head and mantle regions for these video clips (head is the green line, mantle is blue line). The first 30 seconds of pixel intensity at the beginning is the QS state where pixel intensity was standardized at 0 to allow comparisons. Lower pixel values (more negative) indicate darker pigmentation. Our criterion for the start of a REMS-like iteration is marked by a red circle (rapid darkening of pigmentation in both regions by 10 mean pixel values).

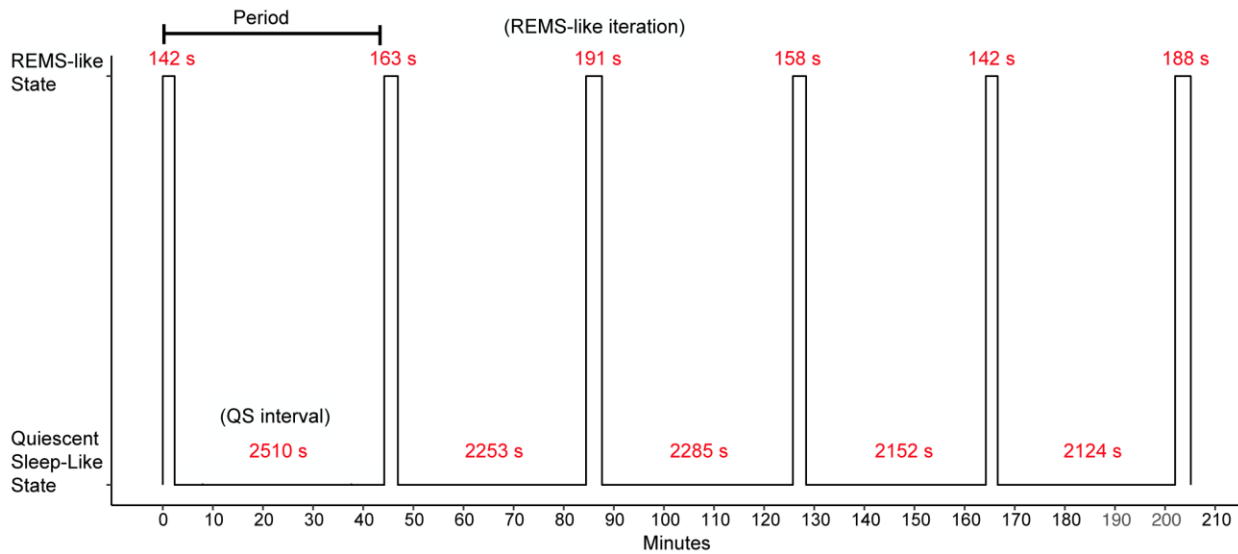
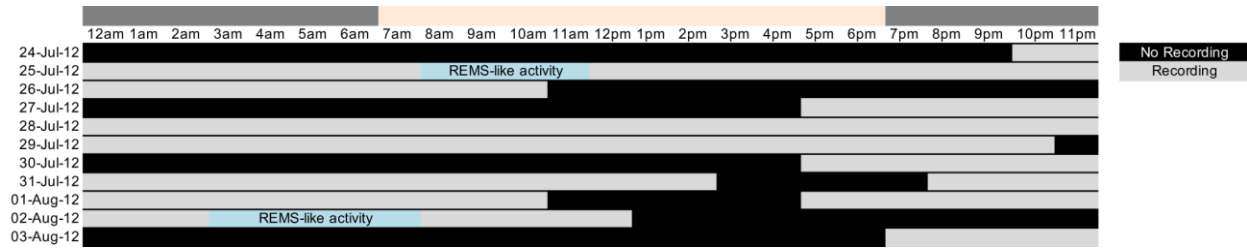
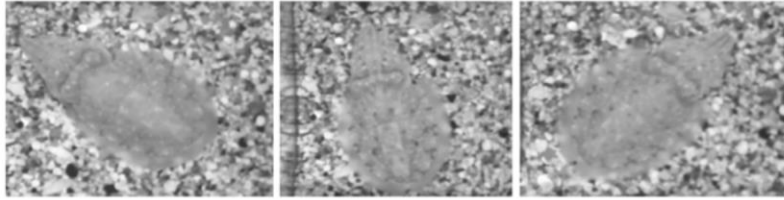
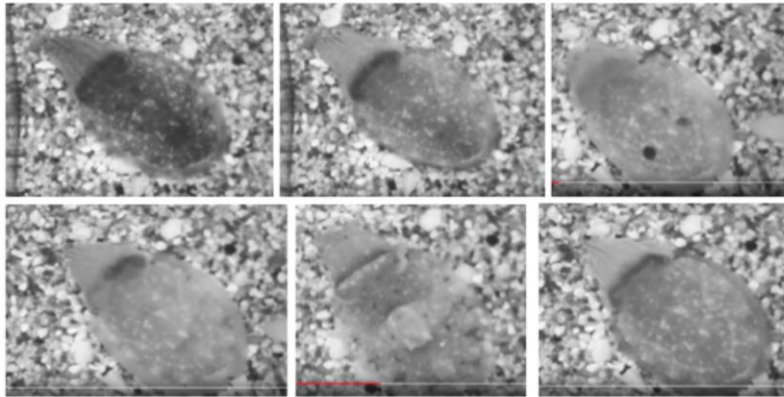


Figure 3. Video recording timeline and hypnogram for one sleep bout. A) Abbreviated timeline for one animal (Hippo) indicating when it was video recorded (gray) and when a sleep-like bout with REMS-like cycling was observed (blue) (see Supplementary Materials S3A for full timeline). Black areas indicate intentional breaks in the recording or where video was lost due to technical issues with the hardware. B) Hypnogram for one animal (Hippo) for one sleep-like bout demonstrating the ultradian rhythm with durations of REMS-like iterations and quiescent sleep-like intervals in seconds.

**Awake state: chronic stable camouflage patterns**



**REMS-like state: acute non-camouflage patterns**



**QS state: acute camouflage pattern variations**

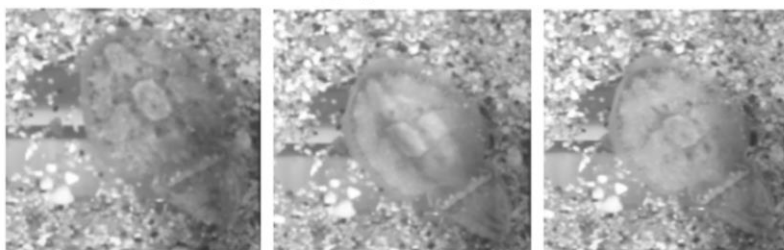


Figure 4. Representative body patterns produced in each of the three states. In awake they showed a normal light mottle camouflage pattern. In REMS-like the vast majority of patterns were fragmentary and different from those ever seen in awake normal animals. For example, in top left the odd dark pattern is not complete. In middle top, the dark head bar has not been seen except in this state. In right top, the deimatic mantle spots are normal but not in combination with the slightly dark head. In bottom left the head bar is only expressed unilaterally (never seen before). In bottom middle the disruptive White mantle square and Head bar are inappropriate type of camouflage for this gravel background. In bottom right there is an odd mix of dark

mantle and light arms/head. The QS state patterns somewhat normal – they are more camouflaged in pattern and brightness than REMS-like. In the right image, the head bar, large mantle spots and dark scalloping along the posterior mantle are a bit odd.

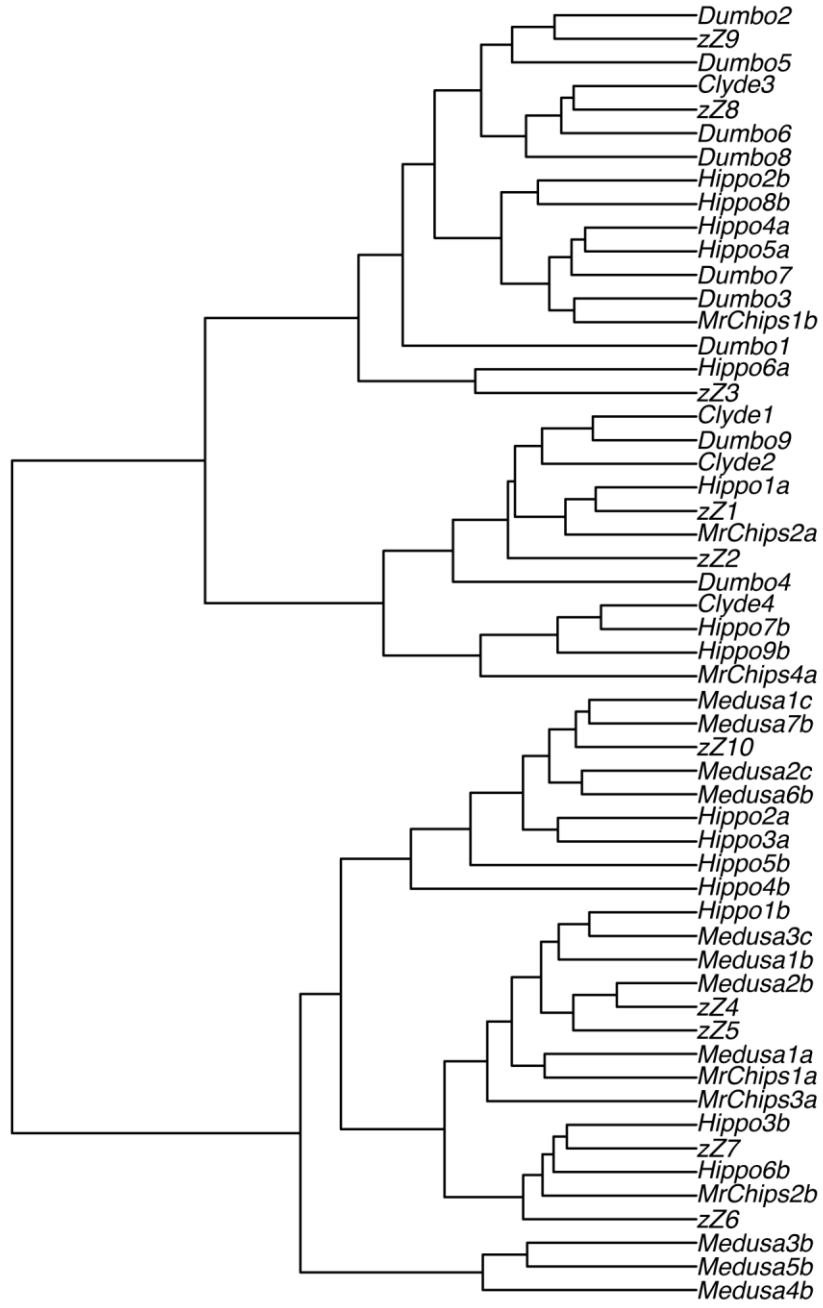


Figure 5. Cluster dendrogram of similarity between all REMS-like iterations using head data. Tip labels indicate the animal (Hippo, zZ, MrChips, Medusa, Dumbo, and Clyde) and REMS-like iteration (1=1<sup>st</sup>, 2=2<sup>nd</sup> ... 10= 10<sup>th</sup>). Letters following the number indicate the sleep-like bout (a= 1<sup>st</sup> bout, b=2<sup>nd</sup> bout, c=3<sup>rd</sup> bout).

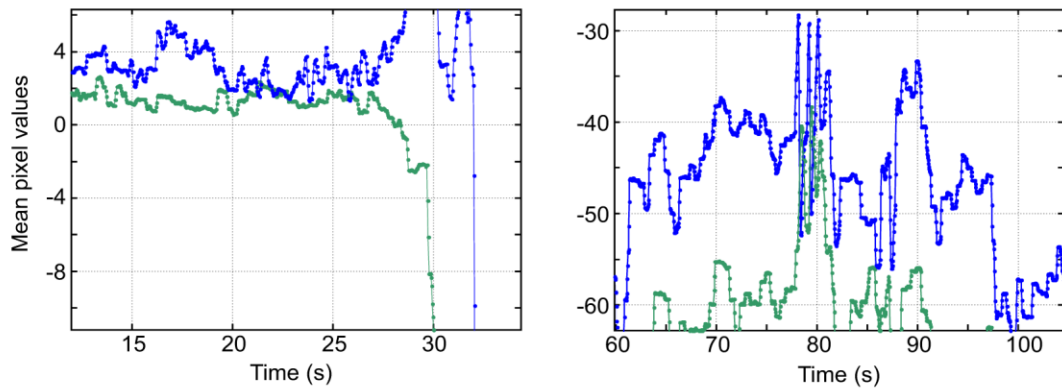
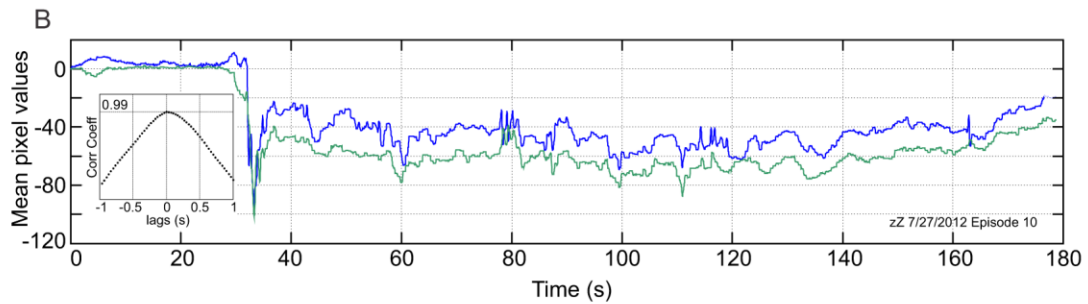
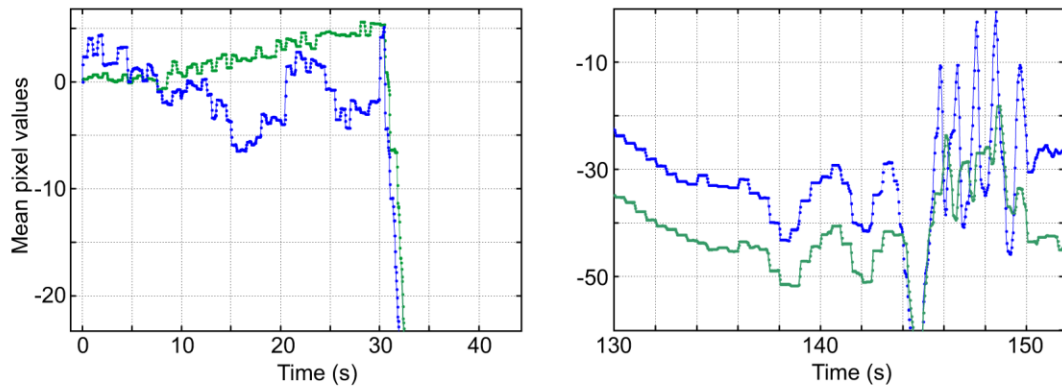
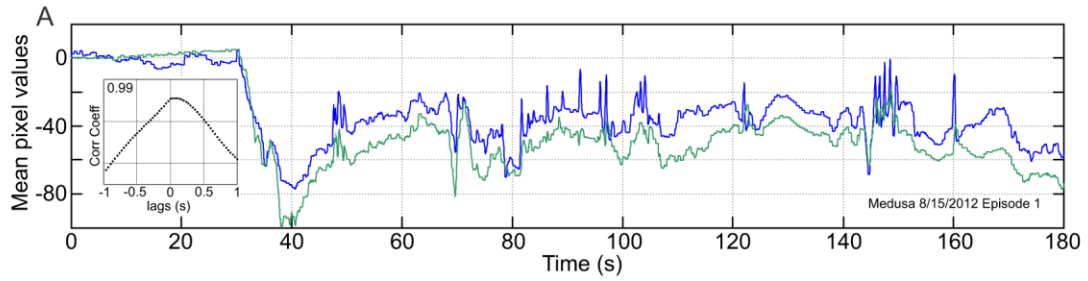


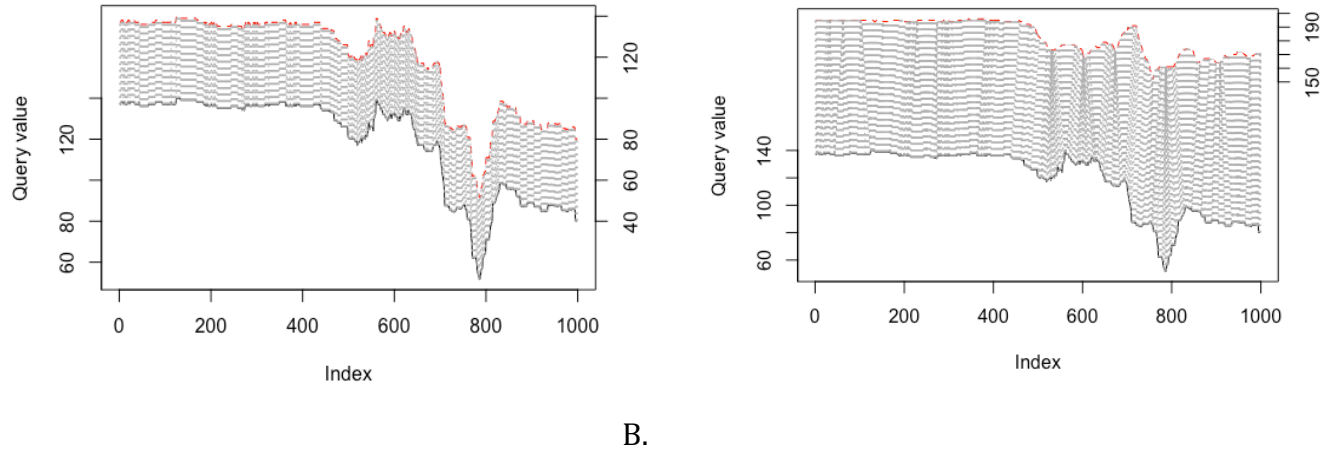


Figure 6. Two REMS-like iterations demonstrating common patterns of rapid brightness changes of skin patterning. Top panels in (A) and (B) show the full-length sequence for the head (green) and the mantle (blue) and bottom panels show expanded time series of the rapid onset of the REMS-like state (left panels) and later in the REMS-like state (right panels) as indicated by the x-axis time scale. Insets in top panels show the cross-correlation function between head and mantle time series. See Results for detailed explanation.

## Supplementary Materials:

Date	Jul-25	Aug-02	Jul-29	Aug-10		Aug-14	Aug-16		Aug-26	Aug-28
Time	8am	3am	10am	8am	2pm	6pm	5am	11am	6am	10am
Cuttlefish ID	Hippo	Hippo	zZ	MrChips	MrChips	Medusa	Medusa	Medusa	Dumbo	Clyde
REMS-like iteration (minutes)	2.37	1.55	1.30	1.43	3.18	1.93	2.37	2.68	1.67	1.45
QS interval (minutes)	39.47	29.43	82.90	52.50	29.05	NA	48.00	30.90	32.27	56.08
REMS-like	2.72	2.18	1.30	2.15	3.08		2.40	2.60	2.25	2.32
QS	34.83	30.30	30.12	29.27			35.95	33.78	30.85	36.53
REMS-like	3.18	2.65	2.25	2.37			2.30	2.70	2.58	2.50
QS	34.90	30.40	29.67	28.33			30.65		32.02	50.03
REMS-like	2.63	2.90	1.97	2.97			3.17		2.60	1.37
QS	35.87	29.38	26.43				32.62		31.18	
REMS-like	2.37	2.92	1.98				2.85		2.82	
QS	33.03	29.10	34.27				31.62		29.37	
REMS-like	3.13	2.67	2.78				2.62		2.63	
QS		27.73	27.23				30.07		31.53	
REMS-like		2.83	2.45				3.08		2.77	
QS		30.18	27.53						29.85	
REMS-like		2.93	1.60						2.62	
QS		26.00	28.43						36.37	
REMS-like		2.38	1.62						3.13	
QS			24.47							
REMS-like			2.30							

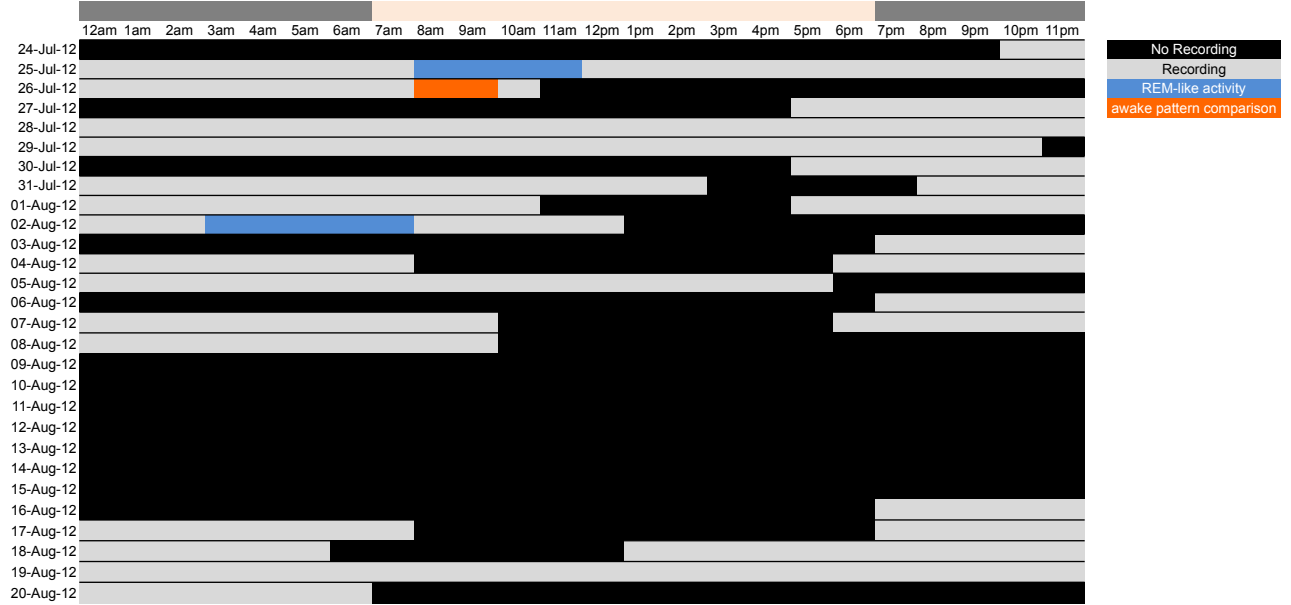
**Table S1.** Duration for all 55 REMS-like iterations and QS intervals in minutes including the date and time of day (to the nearest hour) the sleep-like bout occurred for each cuttlefish. Sleep-like bouts are in separate columns.



**Figure S1.** Graphic representation of two signals aligned to calculate signal distance using Dynamic Time Warp (*dtw*). The y-axis indicates pixel intensity; however, to visualize the signal matching, spacing is introduced along the y-axis. The x-axis denotes the video frame from a video recording at 30 frames per second. A) Compares a signal to itself. The difference measure returned is 0. B) Aligns two different signals by stretching and compressing to find a best match between the sequences. The difference measure returned is 122887.1.

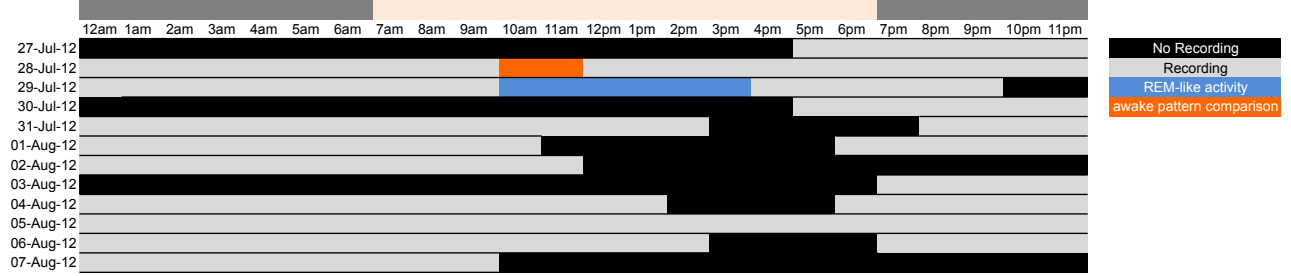
**A.**

Animal ID: Hippo



**B.**

Animal ID: zZ

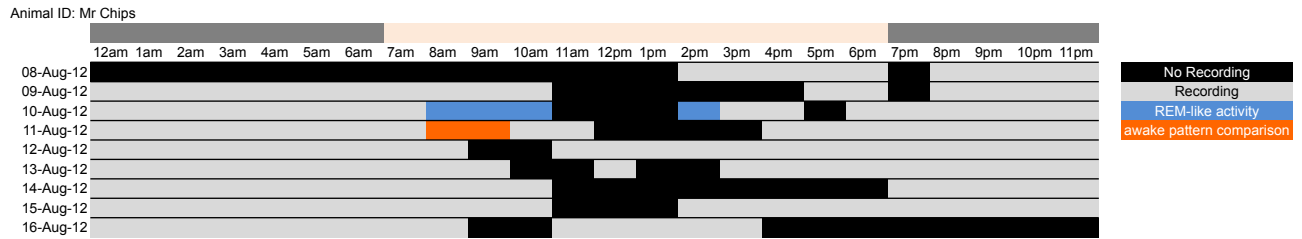


**C.**

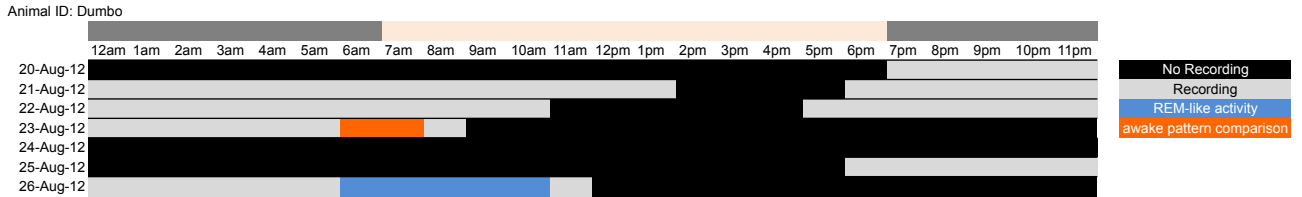
Animal ID: Medusa



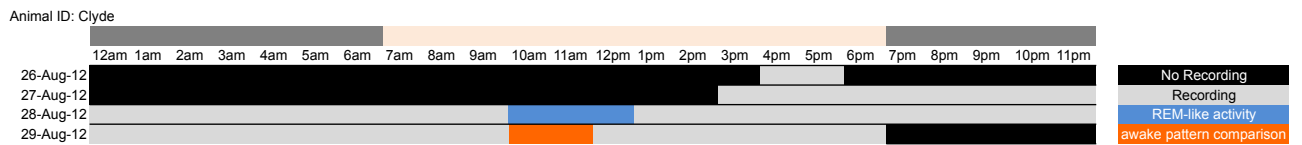
**D.**



**E.**



**F.**



**Figure S2.** Timetable of all animals indicating when they were video recorded (gray) and when a sleep-like bout with REMS-like activity was observed (blue). Black areas indicate intentional breaks in the recording or where video was lost due to technical issues with the hardware. Orange areas indicate where footage of animals in the awake state were used for pattern change counts, however, footage of animals following a sleep-like bout were also used when possible. A) Hippo, B) zZ, C) Medusa (August 14 at 6pm REM-like activity was interrupted by adding food to the tank), D) Mr Chips, E) Dumbo, and F) Clyde.

Rates of body pattern changes in three different states

animal ID	state	# changes	# minutes	changes/minute
clyde	awake	10	10.07	0.99
clyde	awake	10	18.08	0.55
clyde	awake	5	12.85	0.39
clyde	qs	15	15.90	0.94
clyde	qs	81	20.87	3.88
clyde	rem	71	1.67	42.60
clyde	rem	88	2.23	39.40
clyde	rem	98	2.68	36.52
mrchips	awake	18	21.20	0.85
mrchips	awake	18	20.00	0.90
mrchips	qs	10	14.10	0.71
mrchips	qs	7	10.40	0.67
mrchips	qs	6	17.25	0.35
mrchips	rem	86	1.80	47.78
mrchips	rem	131	2.82	46.51
mrchips	rem	194	2.95	65.76
mrchips	rem	207	3.33	62.10
dumbo	awake	16	40.63	0.39
dumbo	qs	8	9.58	0.83
dumbo	qs	20	24.82	0.81
dumbo	qs	11	11.57	0.95
dumbo	rem	97	1.57	61.91
dumbo	rem	145	3.02	48.07
dumbo	rem	187	3.05	61.31
dumbo	rem	184	3.18	57.80

Table S2. Raw count data used in AICc model selection analysis of body pattern changes in each video clip for each state: awake, QS, REMS-like for three animals. Ranges of rates given in the main text originate from this dataset.

**MrCHIPS**

**REMS-like**

86 changes in first clip

131 in next clip

194 in next clip

207 in last clip

OVERALL: 618

**QS**

10 changes in clip QS1 -only one pattern was “full”  
7 changes in clip QS2 -all changes very subtle - no “full” pattern  
6 changes in clip QS3-all changes very subtle - no “full” patterns  
OVERALL: 23

**AWAKE**

18 changes in first clip -Note: fin fluttering constantly in between patterns  
18 changes in next clip all camouflage, very subtle, all same pattern (not as jumpy as first clip)  
OVERALL: 36

=====

**DUMBO**

**REMS-like**  
97 changes in first clip  
145 in next clip  
187 in next clip  
184 in last clip  
OVERALL: 613

**QS**

8 changes in clip QS1 -very subtle tiny changes - amazingly stable overall  
20 changes in clip QS2 -again, very subtle changes -just a faint overall blush hardly noticeable; nearly all pale w just a few small dark spots  
11 changes in clip QS3  
OVERALL: 39

**AWAKE**

16 changes in clip  
OVERALL: 16

=====

**CLYDE**

**REMS-like**  
71 changes in first clip  
88 in next clip  
98 in next clip  
OVERALL: 257

**QS**

15 changes in clip QS1 -only 1 overall pattern change ... all the others are very tiny blushes of components  
81 changes in clip QS2 -many more changes than other QS clips  
OVERALL: 96

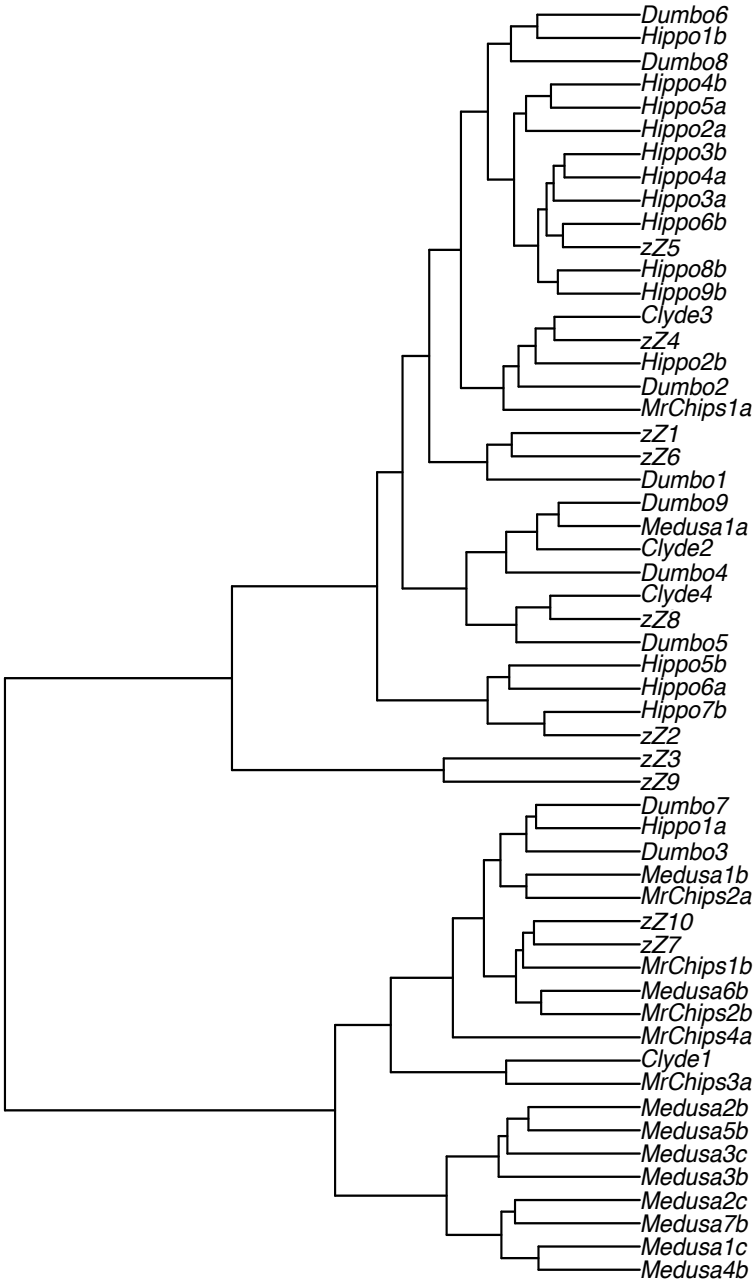
**AWAKE**

10 minor changes in first clip -changes were very subtle  
10 minor changes in next clip  
5 minor changes in next clip  
OVERALL: 25

NOTES for Table S2. Observational notes regarding body pattern changes in three animals in three different states (awake, quiescent sleep-like (QS), and rapid eye movement sleep-like (REMS-like) states). (notes by RTH, edited by TLI)

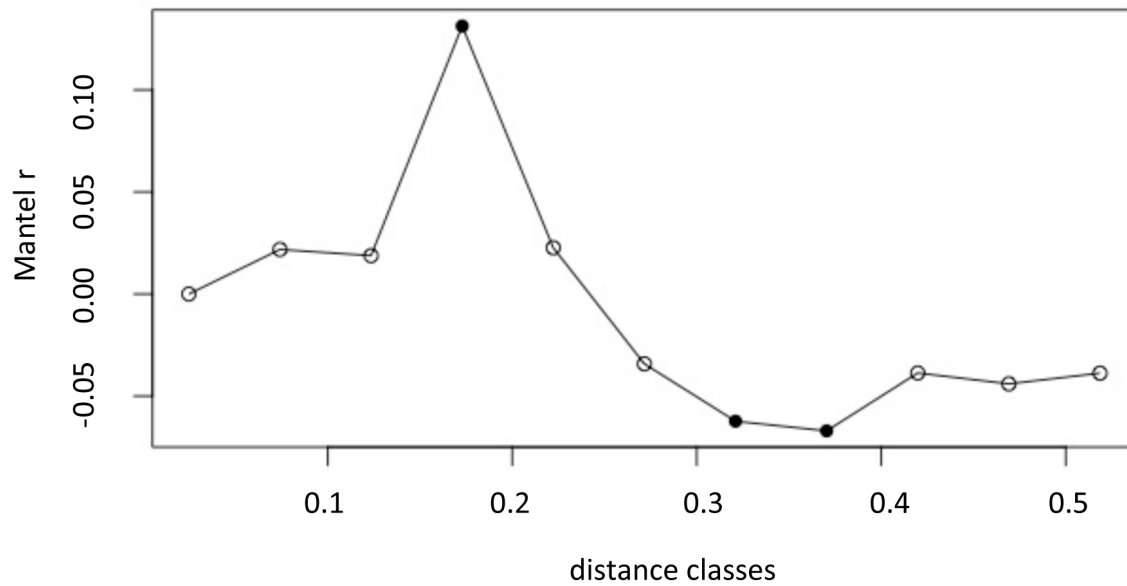
---

---



**Figure S3.** Cluster dendrogram, indicating similarity between all REMS-like iterations using data from mantle region. Tip labels indicate the animal (Hippo, zZ, MrChips, Medusa, Dumbo, and Clyde) and REMS-like iteration within a sleep-like bout (1=1<sup>st</sup>, 2=2<sup>nd</sup> ...10=10<sup>th</sup>). Letters following the number indicate the sleep-like bout (a= 1<sup>st</sup> bout, b=2<sup>nd</sup> bout, c=3<sup>rd</sup> bout).





**Figure S4.** Mantel correlogram demonstrating the correlations between an individual animal and the pixel sequence distance matrix for all REMS-like iterations. Dark circles indicate  $p < 0.05$  significance. The points of significance indicate that REMS-like state sequences with higher similarity (larger Mantel  $r$ ) originate from the same animal and sequences more dissimilar to each other originate from different animals. This relationship (Mantel  $r$ ) is not very strong, however.

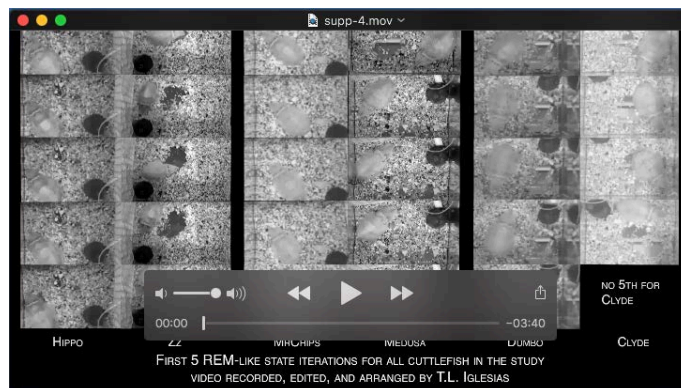
## Movies of animals in the REMS-like state



**Movie S1** Hippo 1a (close up of eye and clear view of skin texture changes)



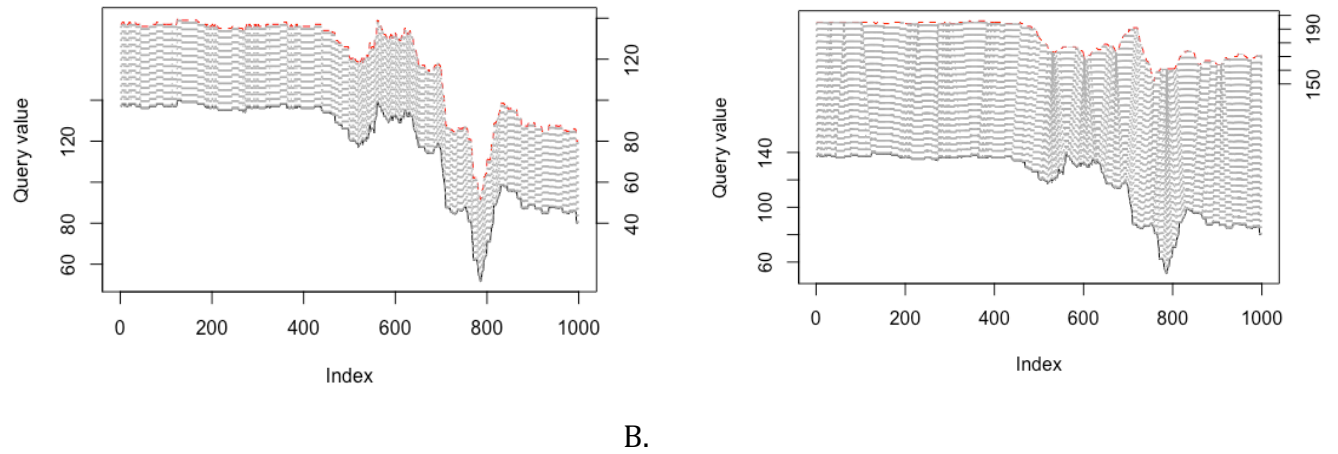
**Movie S2** Clyde 4 (1<sup>st</sup> pair arms raised during QS state; relax at start of REMS-like state)



**Movie S3** (top view of 5 different REMS-like iterations for all individuals in study)

**Table S1.** Duration for all 55 REMS-like iterations and QS intervals in minutes including the date and time of day (to the nearest hour) the sleep-like bout occurred for each cuttlefish. Sleep-like bouts are in separate columns.

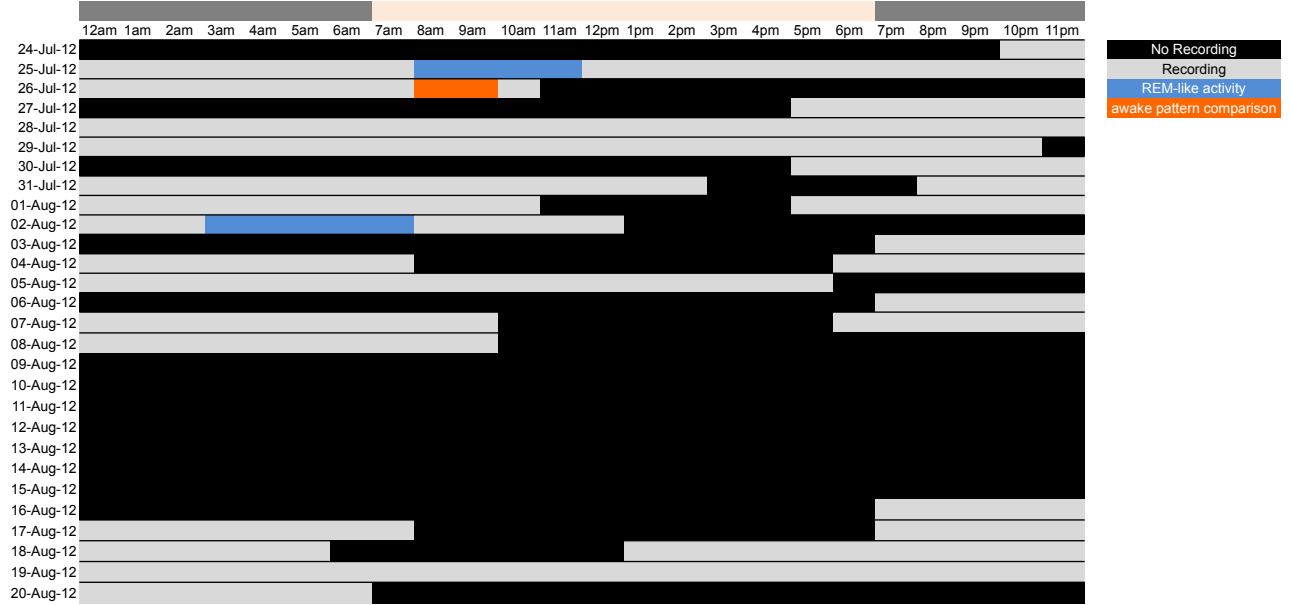
Date	Jul-25	Aug-02	Jul-29	Aug-10		Aug-14	Aug-16		Aug-26	Aug-28
Time	8am	3am	10am	8am	2pm	6pm	5am	11am	6am	10am
Cuttlefish ID	Hippo	Hippo	zZ	MrChips	MrChips	Medusa	Medusa	Medusa	Dumbo	Clyde
REMS-like iteration (minutes)	2.37	1.55	1.30	1.43	3.18	1.93	2.37	2.68	1.67	1.45
QS interval (minutes)	39.47	29.43	82.90	52.50	29.05	NA	48.00	30.90	32.27	56.08
REMS-like	2.72	2.18	1.30	2.15	3.08		2.40	2.60	2.25	2.32
QS	34.83	30.30	30.12	29.27			35.95	33.78	30.85	36.53
REMS-like	3.18	2.65	2.25	2.37			2.30	2.70	2.58	2.50
QS	34.90	30.40	29.67	28.33			30.65		32.02	50.03
REMS-like	2.63	2.90	1.97	2.97			3.17		2.60	1.37
QS	35.87	29.38	26.43				32.62		31.18	
REMS-like	2.37	2.92	1.98				2.85		2.82	
QS	33.03	29.10	34.27				31.62		29.37	
REMS-like	3.13	2.67	2.78				2.62		2.63	
QS		27.73	27.23				30.07		31.53	
REMS-like		2.83	2.45				3.08		2.77	
QS		30.18	27.53						29.85	
REMS-like		2.93	1.60						2.62	
QS		26.00	28.43						36.37	
REMS-like		2.38	1.62						3.13	
QS			24.47							
REMS-like			2.30							



**Figure S1.** Graphic representation of two signals aligned to calculate signal distance using Dynamic Time Warp (*dtw*). The y-axis indicates pixel intensity; however, to visualize the signal matching, spacing is introduced along the y-axis. The x-axis denotes the video frame from a video recording at 30 frames per second. A) Compares a signal to itself. The difference measure returned is 0. B) Aligns two different signals by stretching and compressing to find a best match between the sequences. The difference measure returned is 122887.1.

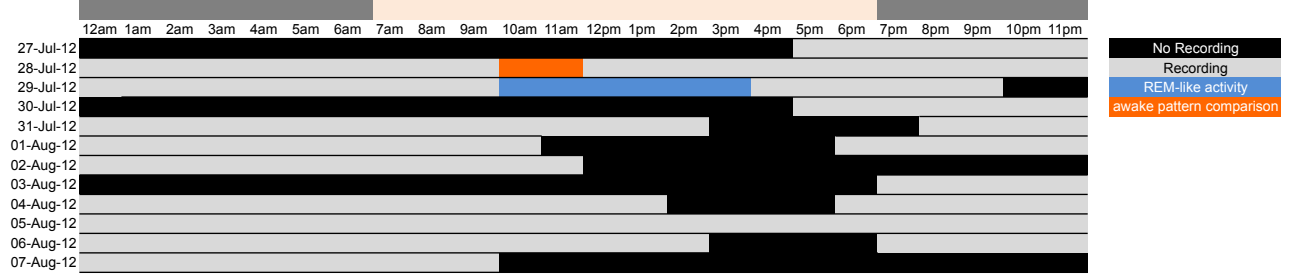
**A.**

Animal ID: Hippo



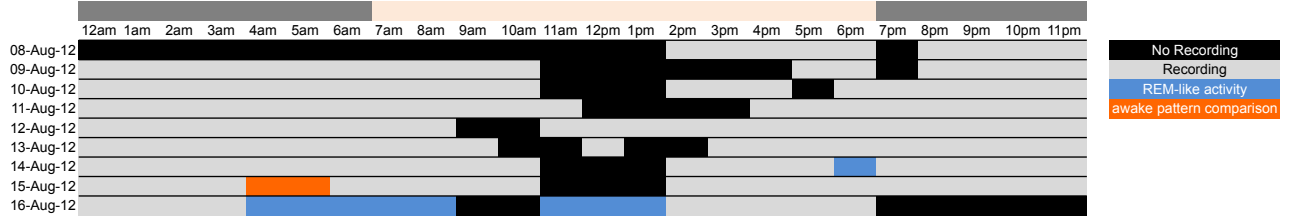
**B.**

Animal ID: zZ

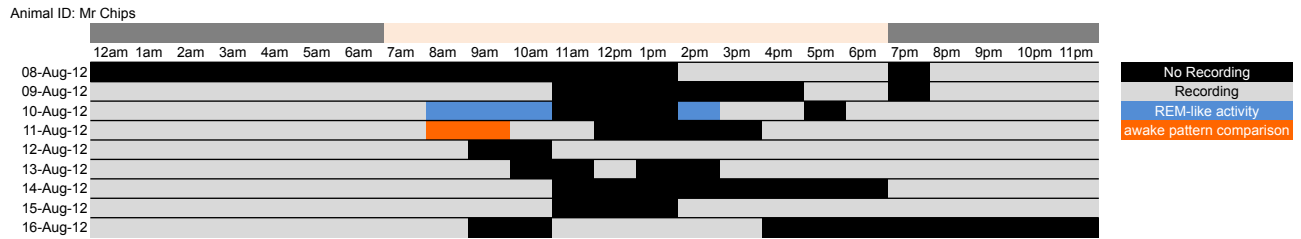


**C.**

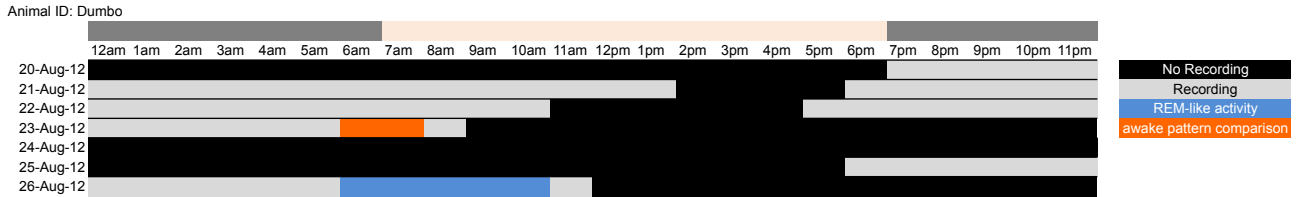
Animal ID: Medusa



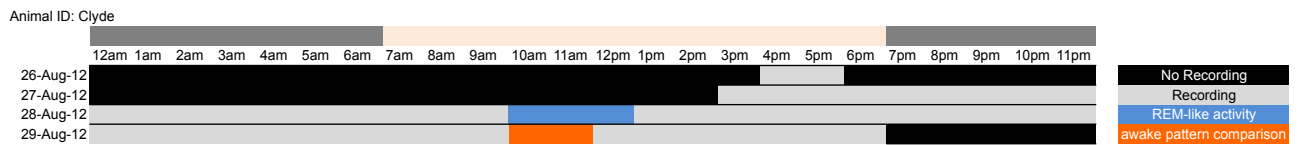
**D.**



**E.**



**F.**



**Figure S2.** Timetable of all animals indicating when they were video recorded (gray) and when a sleep-like bout with REMS-like activity was observed (blue). Black areas indicate intentional breaks in the recording or where video was lost due to technical issues with the hardware. Orange areas indicate where footage of animals in the awake state were used for pattern change counts, however, footage of animals following a sleep-like bout were also used when possible. A) Hippo, B) zZ, C) Medusa (August 14 at 6pm REM-like activity was interrupted by adding food to the tank), D) Mr Chips, E) Dumbo, and F) Clyde.

Rates of body pattern changes in three different states

animal ID	state	# changes	# minutes	changes/minute
clyde	awake	10	10.07	0.99
clyde	awake	10	18.08	0.55
clyde	awake	5	12.85	0.39
clyde	qs	15	15.90	0.94
clyde	qs	81	20.87	3.88
clyde	rem	71	1.67	42.60
clyde	rem	88	2.23	39.40
clyde	rem	98	2.68	36.52
mrchips	awake	18	21.20	0.85
mrchips	awake	18	20.00	0.90
mrchips	qs	10	14.10	0.71
mrchips	qs	7	10.40	0.67
mrchips	qs	6	17.25	0.35
mrchips	rem	86	1.80	47.78
mrchips	rem	131	2.82	46.51
mrchips	rem	194	2.95	65.76
mrchips	rem	207	3.33	62.10
dumbo	awake	16	40.63	0.39
dumbo	qs	8	9.58	0.83
dumbo	qs	20	24.82	0.81
dumbo	qs	11	11.57	0.95
dumbo	rem	97	1.57	61.91
dumbo	rem	145	3.02	48.07
dumbo	rem	187	3.05	61.31
dumbo	rem	184	3.18	57.80

**Table S2.** Raw count data used in AICc model selection analysis of body pattern changes in each video clip for each state: awake, QS, REMS-like for three animals. Ranges of rates given in the main text originate from this dataset.

**MrCHIPS**

**REMS-like**

86 changes in first clip

131 in next clip

194 in next clip

207 in last clip

OVERALL: 618

**QS**

10 changes in clip QS1 -only one pattern was “full”  
7 changes in clip QS2 -all changes very subtle - no “full” pattern  
6 changes in clip QS3-all changes very subtle - no “full” patterns  
OVERALL: 23

**AWAKE**

18 changes in first clip -Note: fin fluttering constantly in between patterns  
18 changes in next clip all camouflage, very subtle, all same pattern (not as jumpy as first clip)  
OVERALL: 36

=====

**DUMBO**

**REMS-like**  
97 changes in first clip  
145 in next clip  
187 in next clip  
184 in last clip  
OVERALL: 613

**QS**

8 changes in clip QS1 -very subtle tiny changes - amazingly stable overall  
20 changes in clip QS2 -again, very subtle changes -just a faint overall blush hardly noticeable; nearly all pale w just a few small dark spots  
11 changes in clip QS3  
OVERALL: 39

**AWAKE**

16 changes in clip  
OVERALL: 16

=====

**CLYDE**

**REMS-like**  
71 changes in first clip  
88 in next clip  
98 in next clip  
OVERALL: 257

**QS**

15 changes in clip QS1 -only 1 overall pattern change ... all the others are very tiny blushes of components  
81 changes in clip QS2 -many more changes than other QS clips  
OVERALL: 96

**AWAKE**

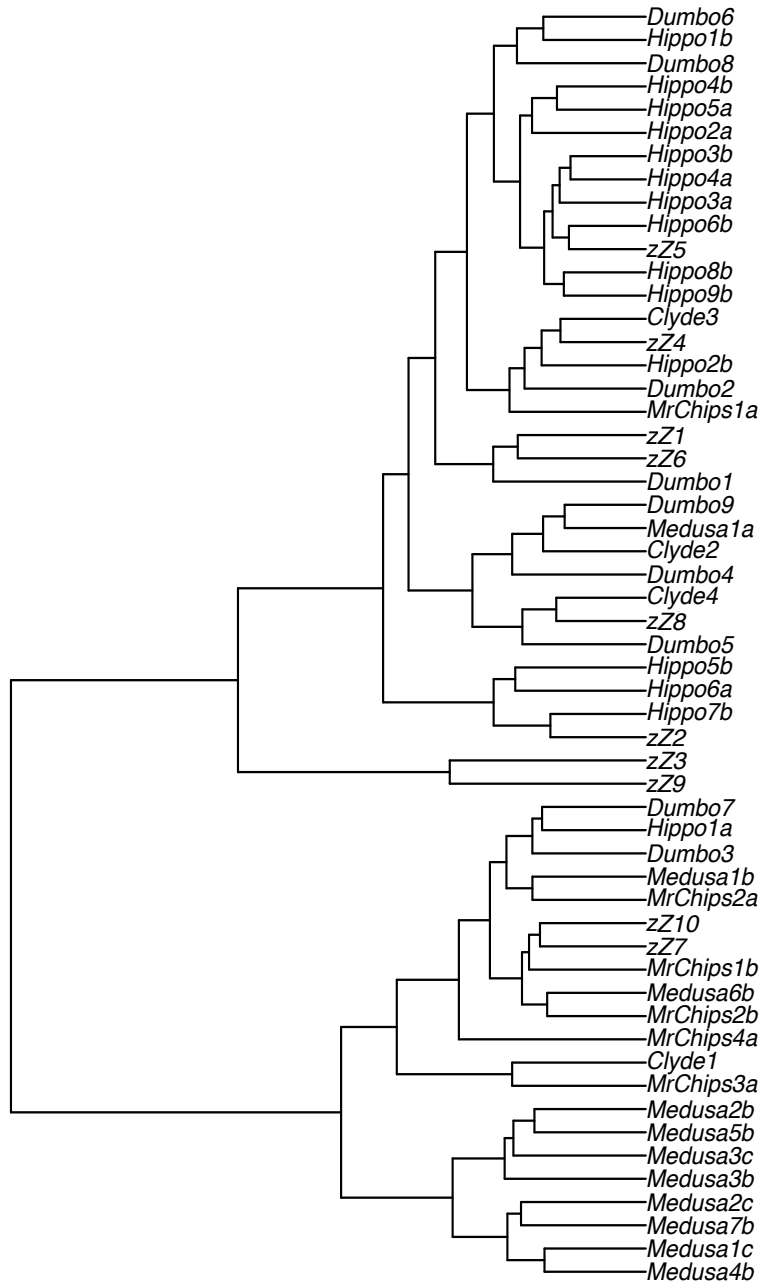
10 minor changes in first clip -changes were very subtle  
10 minor changes in next clip  
5 minor changes in next clip  
OVERALL: 25

NOTES for Table S2. Observational notes regarding body pattern changes in three animals in three different states (awake, quiescent sleep-like (QS), and rapid eye movement sleep-like (REMS-like) states). (notes by RTH, edited by TLI)

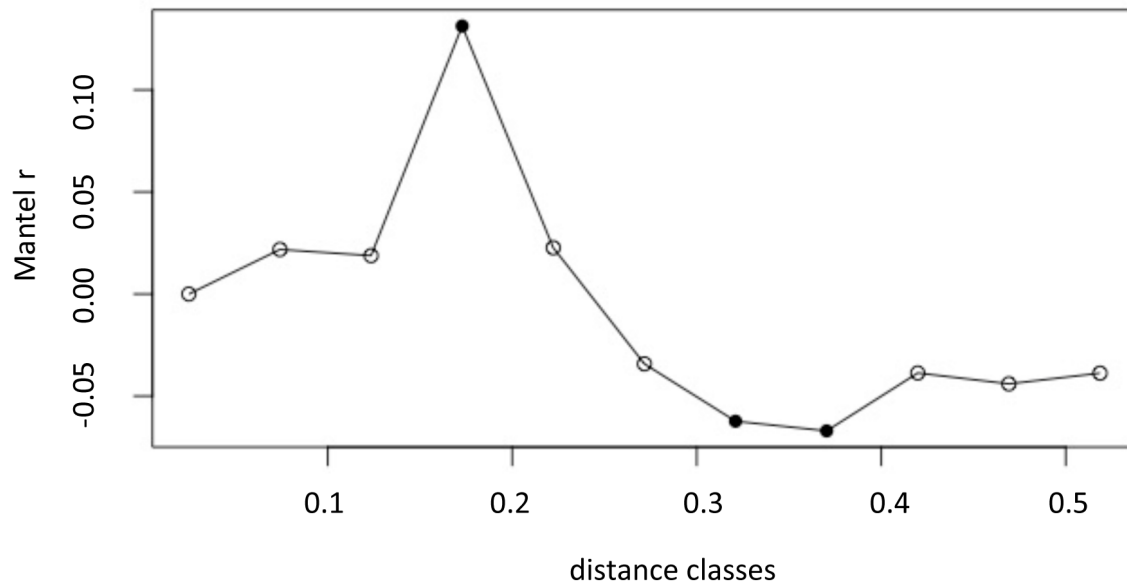
---

---





**Figure S3.** Cluster dendrogram, indicating similarity between all REMS-like iterations using data from mantle region. Tip labels indicate the animal (Hippo, zZ, MrChips, Medusa, Dumbo, and Clyde) and REMS-like iteration within a sleep-like bout (1=1<sup>st</sup>, 2=2<sup>nd</sup> ...10=10<sup>th</sup>). Letters following the number indicate the sleep-like bout (a= 1<sup>st</sup> bout, b=2<sup>nd</sup> bout, c=3<sup>rd</sup> bout).



**Figure S4.** Mantel correlogram demonstrating the correlations between an individual animal and the pixel sequence distance matrix for all REMS-like iterations. Dark circles indicate  $p < 0.05$  significance. The points of significance indicate that REMS-like state sequences with higher similarity (larger Mantel  $r$ ) originate from the same animal and sequences more dissimilar to each other originate from different animals. This relationship (Mantel  $r$ ) is not very strong, however.

## Movies of animals in the REMS-like state



**Movie 1.** Hippo 1a (close up of eye and clear view of skin texture changes)



**Movie 2.** Clyde 4 (1<sup>st</sup> pair arms raised during QS state; relax at start of REMS-like state)



**Movie 3.** Hippo 2b (pupils expand during the REM-like state)



**Movie 4.** Medusa 1a (eye movement and skin texture changes)



**Movie 5.** MrChips 1b (full arm movement, raising 1st pair)



**Movie 6.** (top view of 5 different REMS-like iterations for all individuals in study)