

## Comparative sequence analysis and tissue localization of members of the SLC6 family of transporters in adult *Drosophila melanogaster*

Matthew S. Thimgan<sup>1,\*</sup>, Jonathan S. Berg<sup>2</sup> and Ann E. Stuart<sup>1</sup>

<sup>1</sup>Department of Cell and Molecular Physiology, University of North Carolina School of Medicine, Chapel Hill, NC 27599, USA and <sup>2</sup>Department of Molecular and Human Genetics, Baylor College of Medicine, One Baylor Plaza Houston, TX 77030, USA

\*Author for correspondence at present address: Department of Anatomy and Neurobiology, Washington University in St Louis Medical School, Box 8108 660, S. Euclid Avenue, St Louis, MO 63110, USA (e-mail: thimgan@pcg.wustl.edu)

Accepted 15 May 2006

### Summary

The SLC6 family comprises proteins that move extracellular neurotransmitters, amino acids and osmolytes across the plasma membrane into the cytosol. In mammals, deletion of SLC6 family members has dramatic physiologic consequences, but in the model organism *Drosophila melanogaster*, little is known about this family of proteins. Therefore, in this study we carried out an initial analysis of 21 known or putative SLC6 family members from the *Drosophila* genome. Protein sequences from these genes segregated into either well-defined subfamilies, including the novel insect amino acid transporter subfamily, or into a group of weakly related sequences not affiliated with a recognized subfamily. Reverse transcription-polymerase chain reaction analysis and *in situ* hybridization showed that seven of these genes are expressed in the CNS. *In situ* hybridization revealed that two previously cloned SLC6 members, the serotonin and dopamine transporters, were localized to presumptive presynaptic neurons that previously immunolabelled for these transmitters. RNA for *CG1732* (the putative GABA transporter) and *CG15088* (a member of the novel insect amino acid transporter family) was localized in cells likely to be subtypes of glia, while RNA for *CG5226*, *CG10804* (both

members of the orphan neurotransmitter transporter subfamily) and *CG5549* (a putative glycine transporter) were expressed broadly throughout the cellular cortex of the CNS. Eight of the 21 sequences were localized outside the CNS in the alimentary canal, Malpighian tubules and reproductive organs. Localization for six sequences was not found or not attempted in the adult fly. We used the *Drosophila* ortholog of the mammalian vesicular monoamine transporter 2, *CG33528*, to independently identify monoaminergic neurons in the adult fly. RNA for *CG33528* was detected in a limited number of cells in the central brain and in a beaded stripe at the base of the photoreceptors in the position of glia, but not in the photoreceptors themselves. The SLC6 localization observations in conjunction with likely substrates based on phylogenetic inferences are a first step in defining the role of Na/Cl-dependent transporters in *Drosophila* physiology.

Supplementary material available online at <http://jeb.biologists.org/cgi/content/full/209/17/3383/DC1>

Key words: Na/Cl-dependent transporters, *in situ* hybridization.

### Introduction

The SLC6 family of transporters, also referred to as the neurotransmitter:sodium symporter family (NSS) or sodium neurotransmitter transporter family (SNF), is composed of integral membrane transporter proteins. These transporters are critical for maintaining physiological homeostasis in animals by transferring compounds such as neurotransmitters, amino acids or osmolytes across the plasma membrane. SLC6 members transport compounds against their concentration gradient by coupling substrate uptake to the energy built up in the transmembrane Na<sup>+</sup> gradient. Some of these transporters

also depend on the presence of extracellular Cl<sup>-</sup> (reviewed in Amara and Arriza, 1993; Nelson, 1998). Substrates for members of the SLC6 family include: neurotransmitters such as GABA, serotonin, norepinephrine and dopamine; the amino acids glycine and proline, which in mammals are also used as neurotransmitters; and the osmolytes taurine and betaine. Recently, the broad substrate profiles B<sup>0+</sup>, Bo and IMINO systems have been attributed to SLC6 transporters (Sloan and Mager, 1999; Broer et al., 2004; Takayama et al., 2005). The SLC6 family also includes a group of transporters for which no substrates have been identified, known as the 'orphan'

neurotransmitter transporters (Amara and Arriza, 1993; Nelson, 1998).

The SLC6 gene family is defined by conserved structural features, including a predicted twelve-transmembrane domain topology, highly conserved amino acid residues, particularly in transmembrane (TM) domains 1, 2 and 4–8, and a large extracellular loop between transmembrane domains 3 and 4 that is predicted to be glycosylated (Amara and Arriza, 1993; Nelson, 1998). Crystal structural data have shown that many of the conserved residues in TM1, TM6 and TM8 are involved in substrate and sodium binding in a bacterial leucine transporter (Yamashita et al., 2005). Mice null for individual SLC6 transporters show abnormal physiological phenotypes. For instance, disruption of the dopamine transporter (DAT) gene leads to persistence of dopamine in the extracellular space and a hyperlocomotive phenotype equivalent to the effects of cocaine and amphetamines, which are known to inhibit the DAT (Giros et al., 1996). When other SLC6 transporters lack function, it can result in death or disease (Gomez et al., 2003; Heller-Stilb et al., 2002; Quan et al., 2004; Tsai et al., 2004).

Despite their importance in mammalian physiology, there is little information about these proteins in *Drosophila*. Thus far, only four genes from the SLC6 family have been cloned from *Drosophila*: the genes responsible for the selective uptake of serotonin (*SerT*) (Corey et al., 1994; Demchyshyn et al., 1994) and dopamine (*DAT*) (Porzgen et al., 2001) and two orphan transporters *inebriated* (*ine*) (Burg et al., 1996; Soehnge et al., 1996) and *bloated tubules* (*blot*) (Johnson et al., 1999). The cellular pattern of expression of *SerT* RNA in the embryo is similar to that of serotonin immunolabelling (Demchyshyn et al., 1994); likewise *DAT* is expressed in a cellular pattern in the larva similar to that of dopamine immunolabelling (Porzgen et al., 2001). The other two sequences, *Ine* and *Blot*, do not yet have identified substrates but both are expressed in a variety of cells inside and outside the central nervous system (CNS) (Burg et al., 1996; Huang et al., 2002; Johnson et al., 1999; Soehnge et al., 1996).

To begin to address the role of SLC6 transporters in *Drosophila*, we used a bioinformatics approach to identify 21 *Drosophila* genes with similarity to known SLC6 transporters. Phylogenetic analysis revealed that these *Drosophila* SLC6 transporters segregated into four of the five previously recognized SLC6 subfamilies (Nelson, 1998) and helped to define a sixth subfamily, the insect amino acid transporters (IAAT) (Soragna et al., 2004; Boudko et al., 2005). Using *in situ* hybridization, we focused on localizing SLC6 transporters in the CNS of adult flies. Probes that did not label CNS cells were tested on a variety of other fly tissues as positive confirmation of the reagents. In the course of this work we also localized a vesicular monoamine transporter and, curiously, found that it labelled glial cells at the distal margin of the lamina cell body layer in addition to the expected localization in neurons in the brain in the monoaminergic pattern.

## Materials and methods

### Fly stocks

*Drosophila yw*<sup>67</sup> or *w*<sup>1118</sup> mutants, both of which lack eye pigment, were used in all *in situ* hybridizations.

### Bioinformatic analysis, multiple sequence alignment and phylogenetic analysis

To identify candidate *Drosophila* genes that could encode Na<sup>+</sup>/Cl<sup>-</sup>-dependent transporters, we initially performed BLASTP and TBLASTN (Altschul et al., 1997) searches of predicted proteins from the annotated *Drosophila* genome sequence (Adams et al., 2000). Well known Na<sup>+</sup>/Cl<sup>-</sup>-dependent monoamine transporters from several organisms, including *SerT*, *DAT* and the norepinephrine transporter, were used in sequence searches. A comprehensive group of known or predicted amino acid transporters from vertebrates, insects and worm were assembled, and the full peptide sequences were evaluated by multiple sequence alignment using CLUSTALX (Thompson et al., 1997). The alignment of 84 SLC6 family transporters was used to create neighbor-joining phylogenetic trees in order to depict relationships between various members of the SLC6 family across multiple species. One thousand bootstrap trials were performed to evaluate the significance of the branch node patterns. We used a bootstrap value of >75% to define subfamilies. A separate alignment of the 21 putative SLC6 from *Drosophila* and the leucine transporter from *Aquifex aeolicus* [for which the crystal structure was recently published (Yamashita et al., 2005)] was generated to highlight conserved regions in the transmembrane domains. The final alignments depicted herein were manually adjusted and shaded using GeneDoc software (Karl B. Nicholas and Hugh B. Nicholas, 1997).

### Riboprobe generation

Total RNA was isolated from *Drosophila* heads using the TRIzol reagent (Gibco, Carlsbad, CA, USA) and converted to cDNA using a poly-T primer and Superscript II reverse transcriptase (Gibco, Carlsbad, CA, USA). Portions of the cDNA for each candidate gene were amplified by polymerase chain reaction (PCR) and cloned into pGEM-T Easy (Promega, Madison, WI, USA). All clones were verified by sequencing (UNC sequencing facility). Digoxigenin (DIG)-labelled sense and anti-sense riboprobes were generated from linearized plasmids according to manufacturer's specifications using the DIG RNA labelling kit (Roche, Indianapolis, IN, USA). RNA probes averaging 1000 bp and ranging from 800–1200 bp were purified using three precipitations in 3× ethanol and 0.3 mol l<sup>-1</sup> LiCl incubated at -80 for 2 h between each precipitation and resuspended in DEPC-treated water or hybridization buffer.

### Tissue preparation

*Drosophila* were anesthetized with CO<sub>2</sub>. The back third of the abdomen was cut off and the proboscis was removed or, in the case of abdominal sections, the front half of the fly was cut off to allow complete penetration of fixative. The flies were submerged in ice-cold 4% paraformaldehyde (PFA) in

phosphate-buffered saline (PBS), transferred to 10%, then to 20% sucrose in PBS, and incubated at 4°C overnight in each solution. Flies were then suspended in Optimal cutting temperature (Tissue-Tek, Torrance, CA, USA) reagent, frozen in isopentane cooled by liquid nitrogen, and sectioned on a cryostat microtome at 10–15 µm. Sections were collected on room temperature Superfrost Plus slides and stored at 4°C until *in situ* hybridization or antibody labelling could be performed.

#### *In situ* hybridization

The DIG method (Roche, Indianapolis, IN, USA) of *in situ* hybridization was used following the procedure of Nowicki and Burke with minor modifications (Nowicki and Burke, 2000). Briefly, slides containing sections of *Drosophila* heads were washed and post-fixed in 4% PFA in PBS for 30 min then rinsed twice for 5 min in PBS. After two 2 min washes in 2× sodium chloride/sodium citrate pH 4.5 (SSC), sections were incubated for 30 min in Tris-glycine buffer. Riboprobes, suspended in hybridization buffer at 25–100 ng µl<sup>-1</sup>, were applied to the slides and incubated overnight at 65°C in a humidified chamber.

Sections were rinsed 3 times for 20 min each in 5× SSC at room temperature. Sections were transferred to a preheated solution of 20% formamide and 0.5× SSC (sol B) and incubated at 60°C for 40 min. Sol B was replaced with preheated sol B and the temperature of the solution was allowed to cool to 37°C. Sol B was again replaced with pre-heated sol B and the slides were incubated at 60°C for 30 min. Slides were transferred to 2× SSC at room temperature for 30 min. They were then incubated in 2% Boehringer Blocking Reagent in a maleic acid buffer (block) for a minimum of 10 min. Sections were then incubated overnight in a humidified chamber at 4°C in anti-DIG antibody diluted 1:5000 in block.

Antibody was rinsed away with four washes of 10 min and one wash of 20 min in Tris-buffered saline (TBS) at RT. Sections were then incubated for 10 min in 100 mmol<sup>-1</sup> Tris with 500 µg ml<sup>-1</sup> levamisole and 0.1% Triton X-100. Sections were placed in 0.131 mg ml<sup>-1</sup> 5-bromo-4-chloro-3'-indolylphosphate p-toluidine (BCIP) and 0.5 mg ml<sup>-1</sup> nitro-blue tetrazolium (NBT) in 10% polyvinyl alcohol for 1 h to 3 days. Sections were then rinsed in PBS and mounted with Glycergel (DAKO, Carpinteria, CA, USA). Sequential sections were probed by alternating between sense and anti-sense probes. Sense probes showed no specific label in the head but revealed non-specific labelling of the exterior margin of the eye.

#### Reverse transcription-polymerase chain reaction

Total RNA was isolated using the Qiagen RNeasy kit (Qiagen, Valencia, CA, USA) using the manufacturer's specifications. *w<sup>1118</sup>* flies were dissected into head, thorax and abdomen and immediately frozen on dry ice. Tissue was manually homogenized and further lysed using the QiaShredder spin column. Total RNA was treated with DNase using the DNA-free kit (Ambion, Austin, TX, USA). 2.5 µg of total RNA was used for reverse transcription reaction. PCRs were run for 30 cycles at optimal temperature for the respective

primer pairs. Exact primer sequences can be provided upon request.

#### Antibody staining

Flies were fixed as above and sectioned between 10–15 µm. Sections were permeabilized in PBS/0.5% Triton for 10 min and then washed in PBS. Endogenous fluorescence was quenched in 0.5% NaBH<sub>4</sub> in PBS for 10 min and the slides were washed in PBS. Sections were blocked in 5% normal goat serum (NGS) for 1 h, incubated overnight at 4°C in primary antibody in 5% NGS, rinsed in PBS, and incubated in an anti-goat secondary antibody Alexa 488 (Molecular probes, Carlsbad, CA, USA) at a dilution of 1:2000 in 5% NGS for 1 h. and rinsed again in PBS and mounted in Gel/Mount (Biomed, Forest City, CA, USA). Repo antibody (Developmental studies hybridoma bank, Ames, IA, USA) was used at 1:1 and Neurexin IV (NrxIV, generous gift of M. Bhat)

Fig. 1. Twenty-one sequences in the *Drosophila* genome are homologous to SLC6 transporters. (A) Schematic illustration of the structure SLC6 family transporters (based on the crystal structure of Yamashita et al., 2005). Transmembrane domains are represented by grey rectangles and are numbered according to Yamashita et al., while intracellular and extracellular loops are represented by a thick dark line. Many metazoan SLC6 transporters have large N-terminal and C-terminal extensions, represented by a broken thick line, and most metazoan SLC6 transporters have a 10–50 amino acid extracellular loop between TM3 and TM4, indicated by a thick broken line. Conserved intracellular domains are indicated by white rectangles, and conserved extracellular domains are indicated by black rectangles. The arrows indicate the position of the beta sheet conformation predicted by Yamashita et al. Members of the orphan neurotransmitter subfamily have divergent extracellular loops between TM7 and TM8, and between TM11 and TM12 (dotted lines). (B) Multiple sequence alignment generated in ClustalX using 21 putative *Drosophila* SLC6 transporters and the leucine transporter from *Aquifex aeolicus*, manually adjusted to maximize comparison to the *Aquifex aeolicus* transporter. Protein domains are annotated at the top of each sequence block, indicating transmembrane (TM) regions, intracellular linker regions (IL), extracellular linker regions (EL) alpha helical structure (α), and beta-sheets (arrows); designation of the protein domains is based on the published alignment of Yamashita et al., with minor changes. The names of each protein sequence are on the left (see Table S1 in supplementary material for accession numbers), asterisks are used to mark 10 amino acid intervals, and the residue numbers are indicated at far right. Amino acid residues are shaded according to the degree of conservation (black=100%, dark grey=80%, light grey=60%; amino acids with similar chemical properties are considered equivalent for the purposes of determining conserved residues). Gaps are represented by dashes and residues removed from the alignment for space reasons are indicated by numbers in parentheses. Selected residues that were considered important for transporter function by Yamashita et al. are indicated by symbols at the bottom of each sequence block (○, charged residues at extracellular and cytoplasmic entrances; †, ‡, residues important for coordinating sodium ions 1 and 2, respectively) and residues considered to be strictly conserved by Yamashita et al. are indicated as 'Invariant' on the bottom line of each sequence block to facilitate comparison between alignments.

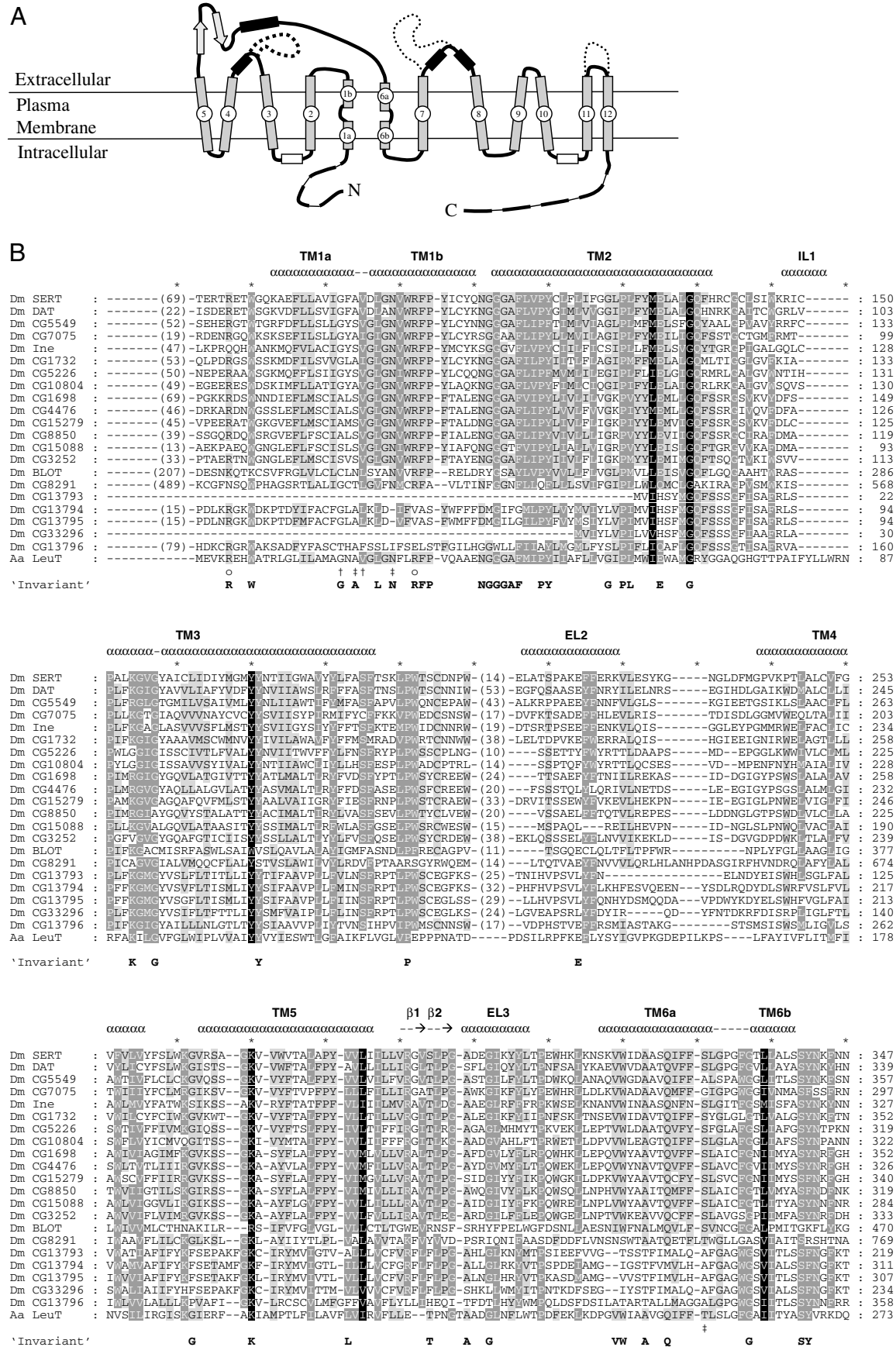


Fig. 1. For legend see previous page.

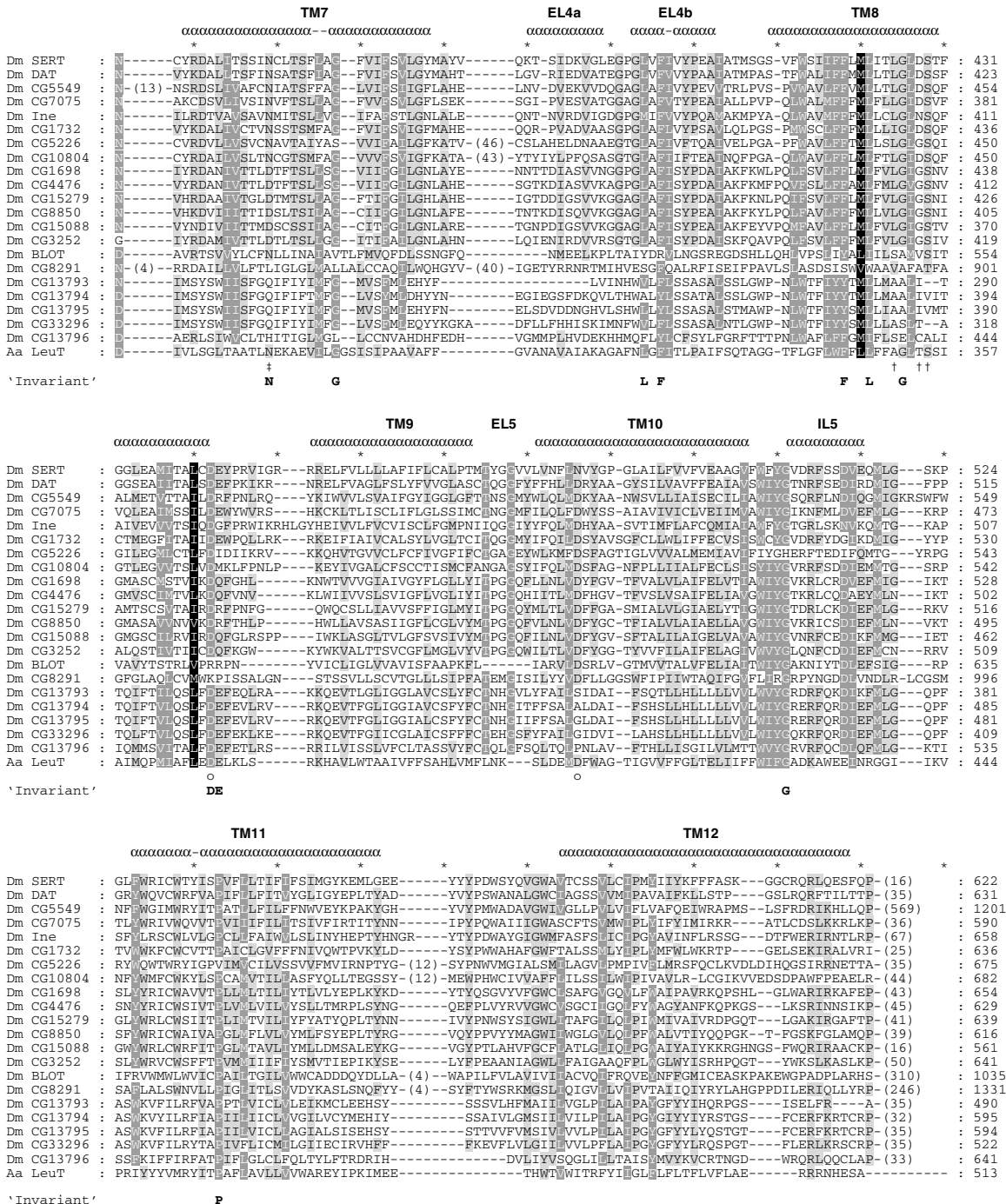


Fig. 1. Continued.

**Results**

*The Drosophila genome contains 21 genes with homology to SLC6 transporters*

We attempted to identify all SLC6 genes in the *Drosophila* genome using a bioinformatics approach with amino acid sequences for known SLC6 transporters as bait with which to search GenBank using the NCBI BLASTP and TBLASTN algorithms (Altschul et al., 1997). We identified a total of 21 genes that we consider to be putative members of the SLC6

was used at a dilution of 1:500. Sections labelled with fluorescent markers were visualized on a Nikon Eclipse E800 microscope with a 20× Plan Apo objective with a numerical aperture (NA) of 0.75 or a 40× Plan Apo objective with an NA of 0.95 and imaged with a Hammamatsu ORCA-ER camera (Bridgewater, NJ, USA) and visualized with the software package Metamorph (Universal Imaging, Downingtown, PA, USA). Images were optimized for visualization and publication using Adobe Photoshop (Adobe, San Jose, CA, USA).

family in *Drosophila* (listed in Fig. 1B and bolded and underlined in Fig. 2). All candidate sequences are predicted to have between ten and twelve TM domains (data not shown) and at least 20% identity to SerT or DAT. A multiple sequence alignment including each of the putative SLC6 family members in *Drosophila* with the amino acid sequence of a SNF6 family member from *Aquifex aeolicus*, LeuT<sub>AA</sub>, for which the crystal structure was recently published (Yamashita et al., 2005), reveals that each of the candidate sequences contains multiple residues that are absolutely or highly conserved among all of the *Drosophila* sequences (Fig. 1B). A complete alignment of all transporter sequences used in our

phylogenetic analysis is available (supplementary material, Fig. S1).

The highly conserved region in the vicinity of TM domains 1 and 2 (Lill and Nelson, 1998) was present in the majority of the candidate genes. For clarity in describing proteins occurring in multiple species, we will use a species prefix such as Dm for *Drosophila melanogaster*. Five of the candidate proteins (DmCG13793, DmCG13794, DmCG13795, DmCG13796 and DmCG33296) are more divergent at the N terminus, with DmCG13793 and DmCG33296 lacking TM1–TM2. The other three divergent candidate proteins have limited homology in the region of TM1 but have a significant degree of similarity

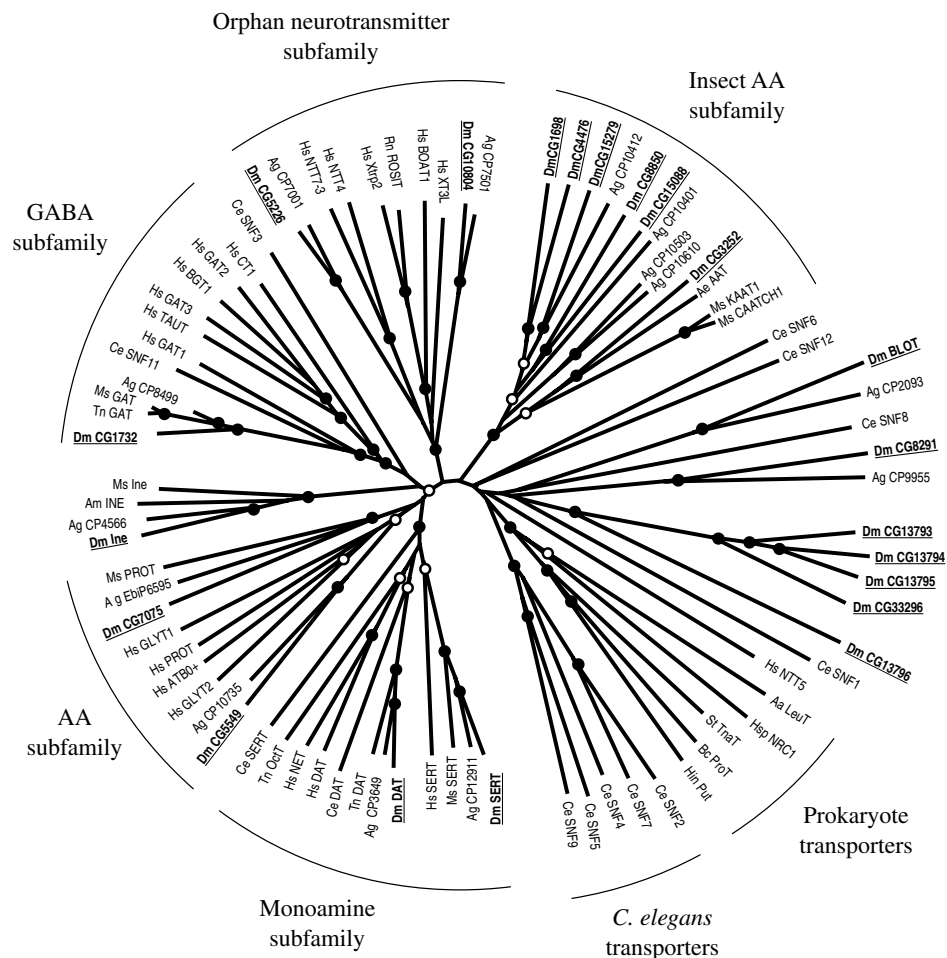


Fig. 2. Phylogenetic analysis assigns many *Drosophila* SLC6 proteins to previously identified subfamilies and a novel IAAT subfamily. An unrooted phylogenetic tree displaying *Drosophila* candidate amino acid (AA) sequences with cloned and predicted SLC6 transporter amino acid sequences from multiple organisms. Candidate *Drosophila* SLC6 transporters are labelled in bold and underlined. Prominently represented among the insect sequences are putative SLC6s from *Anopheles gambiae* (Ag), a mosquito known for carrying the malaria parasite and whose genome sequence was recently reported (Holt et al., 2002). We also have included known transporter sequences from insects such as the cabbage looper, *Trichoplusia ni* (Tn), the tobacco caterpillar *Manduca sexta* (Ms) and from mammals. See Table S1 in supplementary material for a list of the sequence names, abbreviations and accession numbers. Subfamilies (monoamine transporters, GABA transporters, orphan neurotransmitter transporters, insect amino acid transporters) are indicated. As expected, a number of sequences do not group with the previously identified subfamilies. Many of the *C. elegans* transporters are quite divergent and group with each other; likewise, the prokaryote transporters are most similar to each other but do not segregate into a recognized subfamily with eukaryote transporters. Several of the candidate *Drosophila* transporters form a small subgroup that does not convincingly group with any other transporters. Nodes that are identical in >90% of bootstrap trials are denoted by filled circles and nodes that are identical in 75–90% of bootstrap trials are denoted by empty circles. The complete multiple sequence alignment from which this phylogenetic tree was generated is available as Fig. S1 in supplementary material.

to SLC6 members throughout the remainder of their sequences (this is also the case for several of the more divergent *C. elegans* SNF genes). Based on the crystal structure of LeuT<sub>AA</sub>, the first transmembrane domain is likely to play a pivotal role in substrate binding (Yamashita et al., 2005). Therefore, these highly divergent sequences may lack functional transporter activity or have alternative roles. One possible explanation for the differences in the N-terminal region of the proteins is that most of the candidate genes are predicted *in silico* from genomic sequence and therefore are subject to the limitations of exon scanning algorithms. However, cDNA sequences for three of these candidate genes (DmCG13794, DmCG13795 and DmCG13796) have been deposited in the GenBank database and we have therefore included them in this analysis as tentative SLC6 family members.

Structural features identified in the candidate proteins may provide clues as to their function. For example, three candidates (Blot, Ine and DmCG8291) are predicted to have long intracellular N-terminal domains and three candidates (Blot, DmCG5549 and DmCG8291) are predicted to have large intracellular C-terminal domains. DmCG5549 has a short extension of the intracellular linker sequence between TM 6 and TM 7, while the orphan neurotransmitter subfamily members (including DmCG5226 and DmCG10804) are predicted to have extended extracellular linker 4a sequences between TM 7 and TM 8 as well as a short extension of the extracellular linker sequence between TM 11 and TM 12 (see Fig. 1A). Long intracellular terminal domains or loops may indicate intracellular regulation by protein-protein interactions or phosphorylation (Melikian, 2004) while extracellular loops are likely to be involved in substrate binding, the transport process and inhibitor binding (Zomot and Kanner, 2003).

*Drosophila* SLC6 transporters segregate into four previously recognized subfamilies and help define the novel insect amino acid transporter (IAAT) subfamily

In order to infer functional information about the putative *Drosophila* SLC6 family members, we compared them to a large number of known SLC6 sequences using phylogenetic analysis (Fig. 2). We aligned the 21 *Drosophila* sequences with an assembled collection of 63 putative neurotransmitter or amino acid transporters from other organisms: 25 sequences from other insects, 19 from vertebrates, 13 from *C. elegans* and 5 from prokaryotes. Fig. 2 displays a phylogenetic tree derived from the complete alignment of 84 SLC6 family members. Accession numbers for the sequences used are provided (supplementary material, Table S1) and the complete alignment is available (supplementary material, Fig. S1).

Some of the previously uncharacterized *Drosophila* sequences segregate into previously recognized subfamilies anchored by well-characterized transporters (Nelson, 1998). For example, DmCG1732 and its *Anopheles* homolog AgCP8499 are likely to function as GABA transporters as they are closely related to human GAT-1, *C. elegans* SNF11 [a recently identified GABA transporter (Mullen et al., 2006)],

and the GABA transporters from *Trichoplusia ni* (Gao et al., 1999) and *Manduca sexta* (Mbungu et al., 1995). DmCG5549 and its *Anopheles* homolog AgCP10735 are related to the amino acid transporter subfamily, which includes the amino acid transporters for glycine and proline and a transporter with a broader substrate capacity, B<sup>O+</sup> (Sloan and Mager, 1999). DmCG7075 and its *Anopheles* homolog, AgEbiG6595, are closely related to a proline transporter from *Manduca* (Sandhu et al., 2002), and these transporters segregate loosely with the glycine/proline subgroup and the monoamine transporters. Surprisingly, only the *Drosophila* DAT and SerT are present in the monoamine transporter subfamily. Since octopamine and histamine are also biogenic amines and the *Trichoplusia ni* octopamine transporter is present in the monoamine subfamily, we expected that *Drosophila* candidate genes with these transport activities would be found in this subfamily. However, it appears that there are no close monoamine homologs in this subfamily.

Two candidate genes, DmCG5226 and DmCG10804 (and their *Anopheles* homologs AgCP7001 and AgCP7501, respectively) group with the previously defined subfamily of orphan neurotransmitter transporters that includes NTT4 and NTT7-3 as well as two other newly predicted mammalian orphan transporters (Nash et al., 1998). Interestingly, this family also includes a neutral amino acid transporter (B<sup>OAT</sup>), mutations of which cause Hartnup disease (Broer et al., 2004; Kleta et al., 2004; Seow et al., 2004).

Our phylogenetic analysis identified a number of predicted genes that help define the novel insect amino acid transporter (IAAT) subfamily (Boudko et al., 2005), which to date is represented only in insects. Predicted sequences from *Drosophila melanogaster* (DmCG1698, DmCG3252, DmCG4476, DmCG8850, DmCG15088 and DmCG15279) and *Anopheles gambiae* (AgCP10401, AgCP10412, AgCP10503 and AgCP10610) are clearly related to the *Manduca sexta* transporters KAAT1 (Castagna et al., 1998) and CAATCH1 (Feldman et al., 2000), which are competent to transport substrate amino acids using either K<sup>+</sup> or Na<sup>+</sup> as the driving ion. Sequence similarities among KAAT1, CAATCH1 and the other members of the IAAT subfamily suggest that many of the members of this subfamily possess this unique functional property.

A large number of predicted *Drosophila* and *Anopheles* SLC6 transporters segregate into poorly defined subgroups. For example, DmIne and AgCP4566 do not segregate nicely into a recognized subfamily. DmIne mediates a response to hypertonic solutions (Chiu et al., 2000) and has been proposed to comprise its own subfamily (Boudko et al., 2005). Other sequences, including a number of orphan transporters from *C. elegans*, vertebrate NTT-5 and putative *Drosophila* transporters (DmBlot, DmCG8291, DmCG13793, DmCG13794, DmCG13795 and DmCG33296) make up a loose assemblage of outliers, which do not appear to be affiliated with defined subfamilies. We have identified *Anopheles* homologs for the *Drosophila* Ine and Blot orphan transporters (AgCP4566 and AgCP2093, respectively) as well

as for the other outlier sequences, suggesting a conserved, if still unknown, function for these transporters.

*Candidate gene expression in the head, thorax and abdomen of male and female flies*

We used RT-PCR to determine a relative segmental expression profile for the SLC6 homologs in adult flies. mRNA was isolated from the head, thorax and abdomen of either males or females and amplified by RT-PCR to determine the presence or absence of transcript in each segment (Fig. 3). As a control for consistent starting cDNA template across all segments we used primers for the gene *rp49*, and as a control for a gene with enriched expression in the head and thorax we used primers for the gene for the histamine synthesizing enzyme, *histidine decarboxylase (HDC)* (Burg et al., 1993; Pollack and Hofbauer, 1991).

Six candidates were not expressed evenly across all body segments (Fig. 3). Four candidates, *SerT*, *DAT*, *CG1732* and *CG10804*, showed enriched expression in the head and thorax of both males and females. The cell bodies of the CNS are found in the head and thorax, and cells that immunolabel for serotonin (Valles and White, 1988), dopamine (Budnik and White, 1988; Nassel and Elekes, 1992) and GABA (Buchner et al., 1988) reside within the CNS. Expression of the orphan transporter, *CG10804*, was only found in the head and thorax, suggesting that *CG10804* functions in the CNS. Two of the candidates, *CG7075* and *CG4476*, were expressed more abundantly in the abdomen of the male and female fly, respectively.

The remaining fifteen SLC6 transporters showed approximately equivalent expression across the three body segments in both genders. This result suggests that these transcripts are expressed either in organs in each of the segments or in an organ present in all three segments. Although twelve of these transcripts displayed robust expression, three transcripts (*CG8850*, *CG13793* and *CG33296*) revealed only weak expression.

*Genes involved in monoamine processing localize to a subset of cells in the adult CNS*

Previous immunocytochemistry has revealed the location of serotonergic and dopaminergic cells in the adult fly CNS; these

neurons are grouped in named clusters (Budnik and White, 1988; Valles and White, 1988). The riboprobes against *SerT* (Fig. 4A,B) and *DAT* (Fig. 4C,D) labelled CNS cells of roughly the same number and in the same general location as the immunolabelled cells. Due to limited resolution of the *in situ* hybridization technique and use of alternating sections between sense and anti-sense probes, we were only able to estimate the number of neurons that were labelled by each riboprobe. For the *SerT* probe, about 75 neurons labelled in cell clusters that approximated the position of the named clusters from previous serotonin immunolocalization studies in both the head and

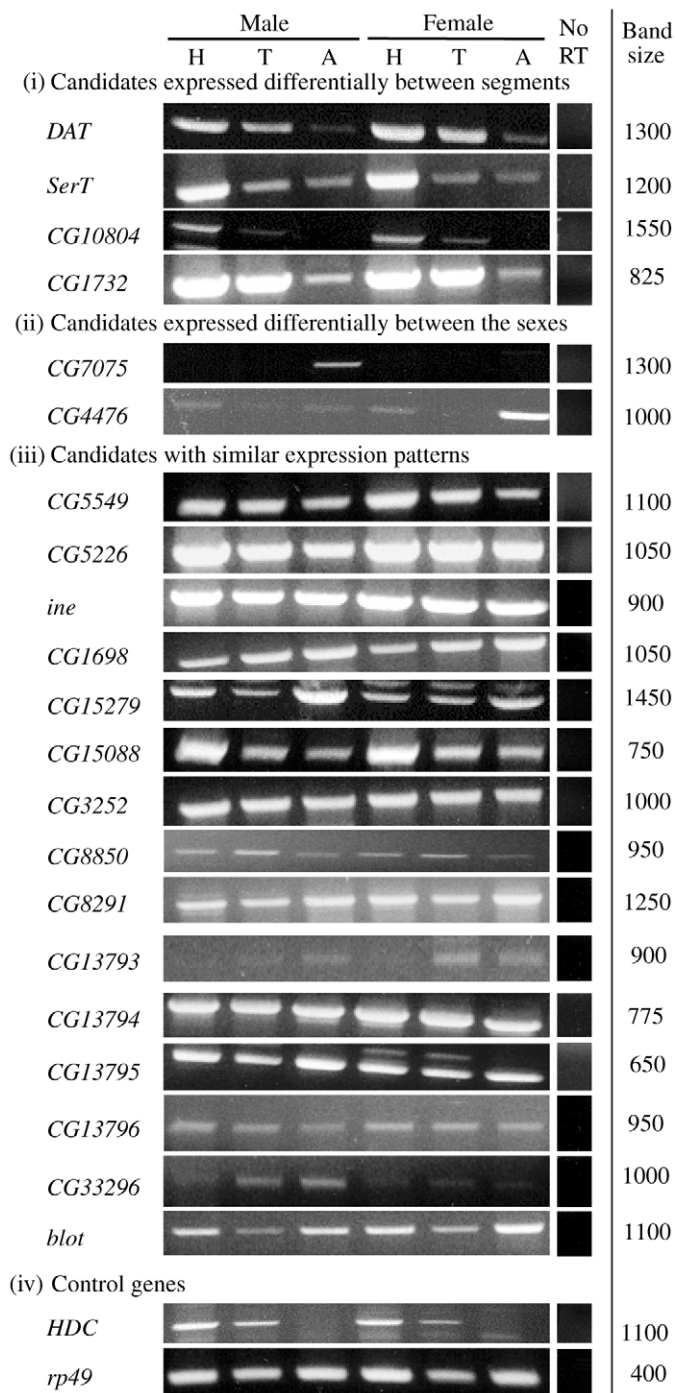


Fig. 3. The segmental mRNA expression profile of the SLC6 homologs in the head, thorax and abdomen of both male and female flies determined by RT-PCR. The head (H), thorax (T) and abdomen (A) were isolated from either male or female flies and RT-PCR was performed to determine presence or absence of transcript in a given body segment. Candidates are arranged from top to bottom as: (i) those expressed differentially in the three segments; (ii) those showing gender specificity; (iii) those expressed equally in the segments; (iv) control genes *HDC* and *rp49*. *HDC* is known to have enriched expression in the head and thorax and *rp49* controlled for equivalent starting template in the PCR reactions. The column labelled 'No RT' showed controls confirming that the amplicons were dependent on the presence of cDNA. Band size is measured in base pairs (bp).



thoracic ganglion. Labelled cells in the head (Fig. 4A) were found dorsally on either side of the midline in the position of the cluster called SP1 (arrows), ventrally in the position of the SE2 cluster (barbed arrow), and proximal to the optic lobe in the position of the LP2 cluster (notched arrow). The photoreceptor cells were never labelled. Cells in the thorax (Fig. 4B) are equivalent in position to Budnik and White's 'A1–7' neurons (arrows) (Budnik and White, 1988). Other sections showed neurons in positions of other named clusters and no cells were labelled in positions that substantially differed from the immunolabel. It is likely, therefore, that our *SerT* riboprobe is labelling serotonergic neurons.

The *DAT* riboprobe likewise labelled discrete cells in the pattern described for dopaminergic neurons using immunocytochemistry (Budnik and White, 1988). We identified up to 30 large cell bodies in a single preparation with alternating slides, approximately the number of large cell bodies labelled by the dopamine antibody. In the section shown in Fig. 3C, the riboprobe labelled cells in the positions of three named clusters: two dorsal clusters on either side of the midline corresponding to the DM cluster of neurons (arrows); neurons lateral to the dorsomedial (DM) cluster in the more dorsal DL1 cluster (white arrowheads); and cells more ventral in the DL2 cluster (carrot). We also detected *DAT* positive cells corresponding to the AbU (arrows) and ThL clusters (barbed arrow) in the thoracic ganglion (Fig. 4D). In other sections, we detected cells in other named clusters. Curiously, the riboprobe did not label a group of cells surrounding the medulla, the MC cell group that clearly labels for catecholamines [fig. 7A in Budnik and White (Budnik and White, 1988)], nor did it label a cluster of small cells along the midline referred to as the anteromedial (AM) neurons. Thus, as with the *SerT* riboprobe, the pattern of *in situ* label overlapped with that from dopamine immunohistochemistry, suggesting that, with the exception of the AM and MC cells, the *DAT* probe labels dopaminergic cells. It does not appear that the *DAT* probe labels octopaminergic in addition to dopaminergic cells, as we do not see *DAT* positive cells in the easily recognizable antennal lobe (AL) cluster of octopaminergic cells localized along the alimentary canal (Monastirioti et al., 1995). In addition, the *DAT* probe does not label octopaminergic cells in the larva and does not have a high affinity for octopamine in uptake experiments in *Xenopus* oocytes (Porzgen et al., 2001).

Although we did not find a *Drosophila* SLC6 homolog for either the plasmalemmal octopamine or histamine transporter, in the course of this study we localized an ortholog of a mammalian vesicular monoamine transporter 2 (VMAT2). In mammals, VMAT2 packages four biogenic amines (dopamine, serotonin, norepinephrine and histamine), and it is present in neurons using each of these four neurotransmitters (Peter et al., 1995). A *Drosophila* ortholog of the mammalian VMAT2 would be expected to label all monoaminergic cells, revealing cells that contain dopamine, serotonin, octopamine and histamine. The closest homologous sequence to VMAT2 in *Drosophila* is CG33528, which is 56% identical (Greer et al., 2005).

A riboprobe designed against CG33528 labelled on the order of 125 cells in the central brain (Fig. 4E,G) and thorax (Fig. 4F), which are in the anatomical position of named clusters of monoaminergic neurons. The riboprobes for *SerT* and *DAT* each labelled cells in the same location as the CG33528 label (compare Fig. 4A,C with 4E), but there were consistently more cells labelled by the CG33528 probe than for the *SerT* and *DAT* probes combined. This pattern may represent labelling of octopaminergic neurons since antibody labelling revealed that CG33528 was found in serotonergic, dopaminergic and octopaminergic (Greer et al., 2005) neurons, but was not found in the histaminergic photoreceptors (Chang et al., 2006). We were able to detect CG33528 label in both presumptive MC cells (open arrows in Fig. 4E), dopaminergic neurons present in the medullary cortex, and in the presumptive AL cells, octopaminergic cells that reside just lateral to the oesophagus (data not shown).

Curiously, we were unable to detect label in photoreceptors, which in the fly are histaminergic (Pollack and Hofbauer, 1991). To confirm that our *in situ* approach would work in the photoreceptors, we made a riboprobe to *HDC*, which is expressed in photoreceptors (Burg et al., 1993). The *HDC* riboprobe labelled the photoreceptor layer of adult head sections and in addition, about 20 discrete, bilateral cells in the central brain (Fig. 4J) as well as cells in the thoracic ganglion (data not shown) all in the anatomical positions consistent with immunolabelled histaminergic neurons (Pollack and Hofbauer, 1991).

Unexpectedly, the riboprobe for CG33528 also labelled cells in a punctuated arc at the fenestrated layer at the base of the photoreceptors (Fig. 4G–I), a distinct region where the photoreceptor cell bodies form the axons that ultimately synapse in the optic lobes. This band of beaded label is less than 5  $\mu\text{m}$  in width, the centers of the beads spaced 8–10  $\mu\text{m}$  apart, and the label is intermixed with the fenestrations at the base of the photoreceptors that are present at the distal-most edge of the lamina cortex. Fingers of label occasionally penetrate into the photoreceptor layer (block arrows in Fig. 4H).

The cell types of this layer have been described for *Drosophila melanogaster* (Eule et al., 1995) and ultrastructurally in *Musca domestica* (Saint Marie and Carlson, 1983). There are two glial subtypes that reside at this distal margin of the lamina cortex, the fenestrated and pseudocartridge glia. In both species, these cells have their somata spaced approximately 10  $\mu\text{m}$  apart; in *Musca*, both glial subtypes have been shown to wrap photoreceptor axons traversing this layer on their way to forming synapses in the optic lobes (Saint Marie and Carlson, 1983). The fenestrated glia, but not the pseudocartridge glia, have processes that invade the photoreceptor layers, similar to the CG33528 labelling. When alternating sections were labelled with CG33528 riboprobe (Fig. 4H) and an antibody to Neurexin IV (NrxIV; Fig. 4I), a component of septate junctions found on the plasma membrane of glia and not neurons (Baumgartner et al., 1996), the labelling pattern was similar. Like CG33528, the

NrxIV antibody labelled the base of the photoreceptors in a beaded fashion spaced approximately 10  $\mu\text{m}$  apart (Fig. 3I). This labelling pattern suggests that *CG33528* is present in the distal-most glia of the lamina cell body layer, in particular the spacing and the fingers penetrating into the retina suggest that that these are the fenestrated glia.

*Two SLC6 homologs label the CNS in a glial pattern*  
*CG1732* (the putative GABA transporter) and *CG15088* (a

member of the IAAT subfamily) are expressed in the CNS in a glial-like pattern (Fig. 5). Glia of the fly CNS form the barrier between hemolymph and the brain, wrap axons and isolate synaptic terminals in areas such as the lamina (Saint Marie and Carlson, 1983), among other roles. We have chosen to use the *Drosophila* glial terminology (Eule et al., 1995). In short, a layer of glia, called perineural glia (Fig. 5B, arrows), forms the outermost layer in the adult brain. Interior to the perineural glia, the somata of the subperineural glia (Fig. 4B, carrots) are

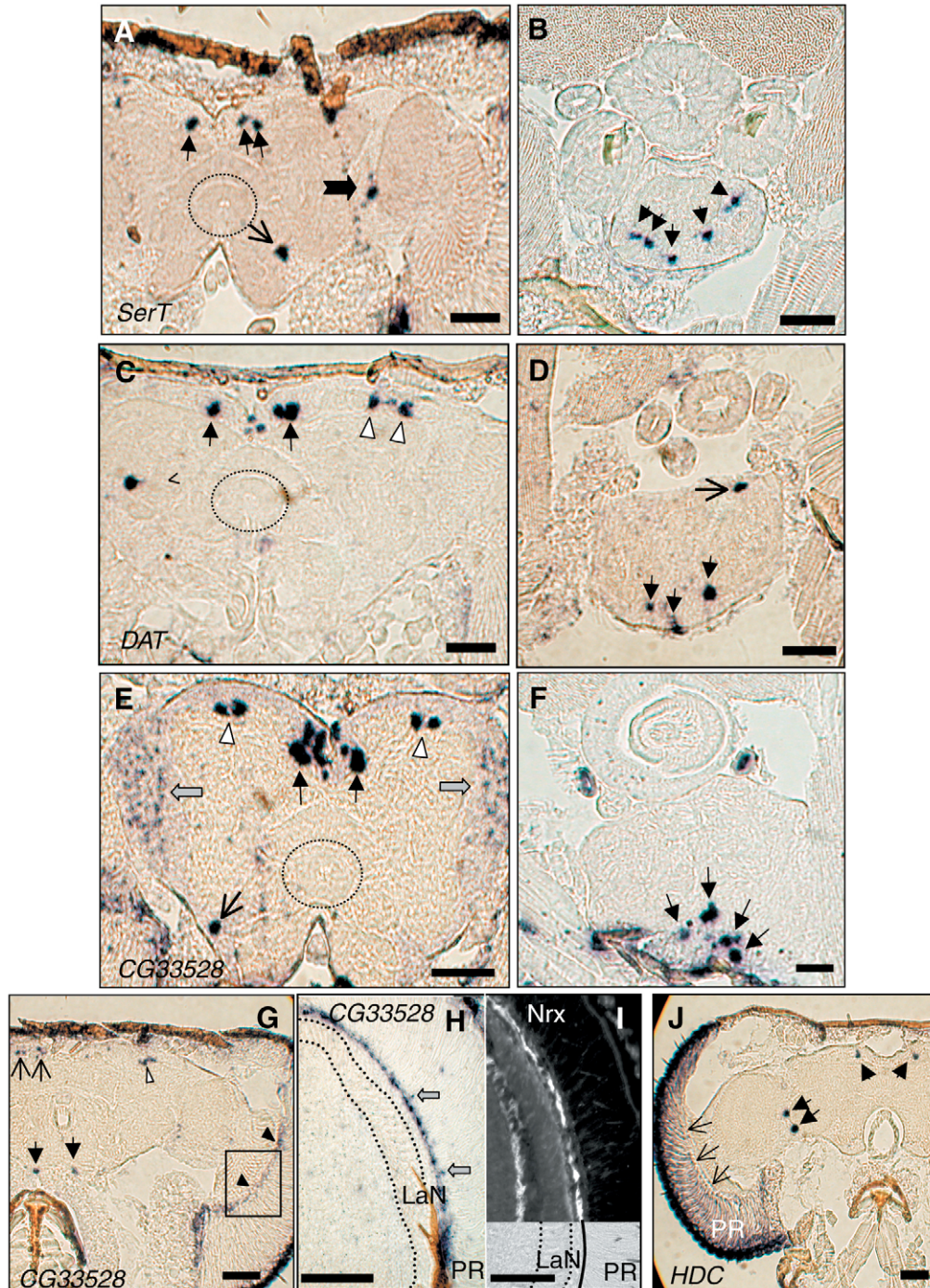


Fig. 4. For legend see next page.

interspersed with neuronal somata to form the cortex. Neuronal processes form the neuropil and neuropilar glia wrap the neuronal processes within the neuropil. As is true in general of invertebrate nervous systems, there are no neuronal somata within the neuropil.

The riboprobe *CG1732* labels a subset of regularly spaced cells that surround neuropil structures in the central brain (Fig. 5A,C) and ventral ganglion. Neuropil, including the subesophageal ganglion, the medulla and lobula (Fig. 5A), the antennal lobes and ventral nerve cord (data not shown), are surrounded by somata positive for *CG1732*. Between the medulla and lobula, *CG1732* labels a subset of giant glia at the inner chiasm (barbed arrow in Fig. 4A), a region that contains only glial and not neuronal cell bodies (Tix et al., 1997). This pattern is substantively different than that of GABA

immunolabelled cells, which have cell bodies throughout the cortex surrounding the medulla and in a large cluster next to the lobula (Buchner et al., 1988).

*CG1732* label (Fig. 5A,C) was compared in sequential sections to immunolabel for the glial nuclear marker, Repo (Fig. 5B,D) (Xiong et al., 1994). Repo labelled nuclei in positions of the perineural glia (arrows, Fig. 5B), the subperineural glia (carrots, Fig. 5B) and in regularly spaced intervals that form the outlines of several neuropilar structures. *CG1732* labelled only cells located between the cortex and neuropil (compare label surrounding the medulla and lobula in Fig. 5A with Fig. 5B). *CG1732* did not label perineural (arrows, Fig. 5A) or subperineural glia (carrots, Fig. 5A), which were labelled by Repo. Thus, the riboprobe for *CG1732* only labels the subset of glia that surround neuropil.

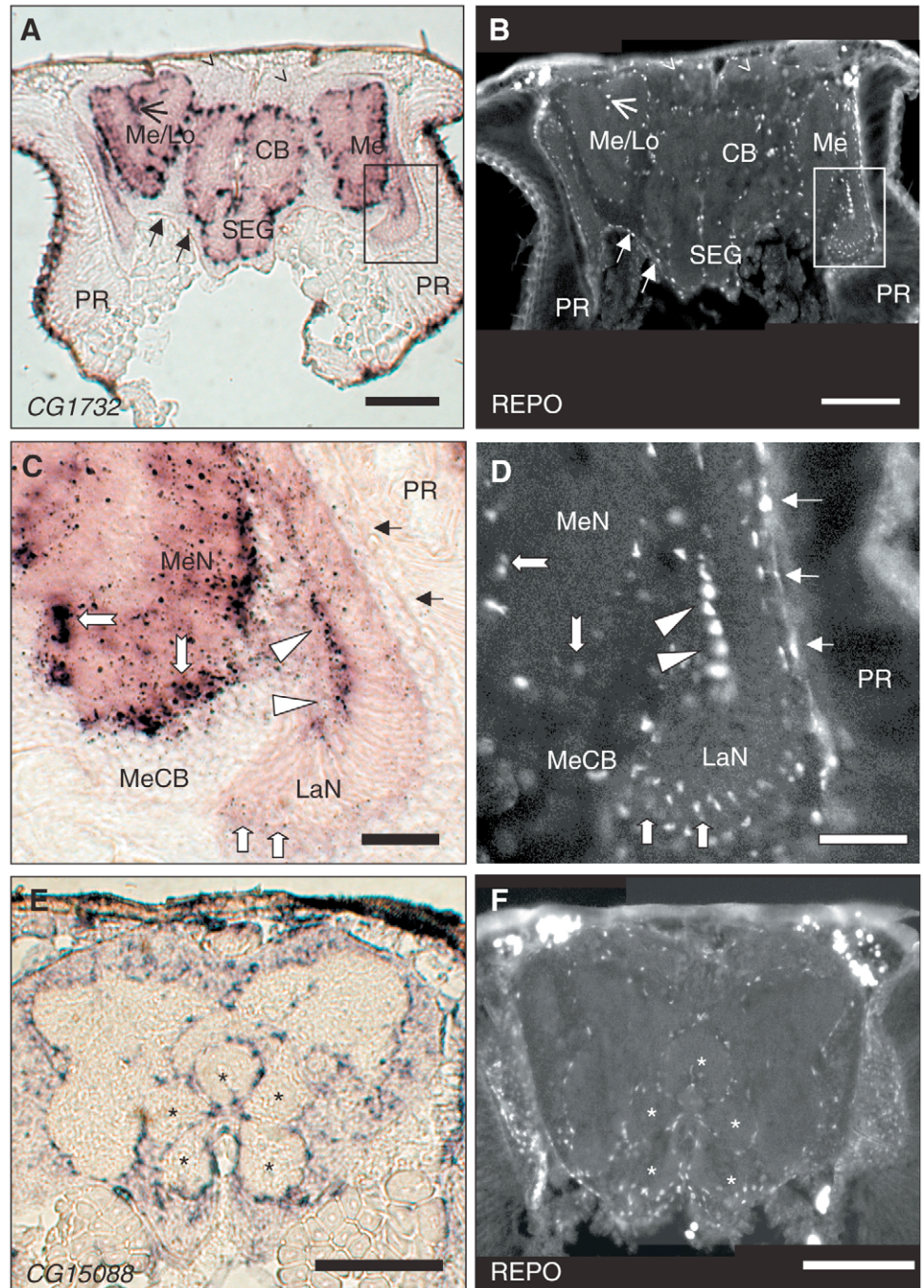
The label for *CG1732* had two additional, notable characteristics. First, there was a purple hue within the neuropil and not present throughout the cortex, which suggests that the mRNA for *CG1732* is possibly present within glial processes. Second, the purple hue is particularly dense in the distal one-third of the medulla (Fig. 5A,C), where the photoreceptors R7 and R8 make their first synapse. The dense label shows a banded pattern that runs parallel with the distal margin of the medulla and is visible in the left medulla (Fig. 5A). Repo-positive and *CG1732*-positive cells (notched open arrows in Fig. 5C,D) are located in similar positions and with similar spacing surrounding the neuropil. It appears that labelled processes emanate from regularly spaced cell bodies at the distal border of the medulla (Fig. 5C) and penetrate into the neuropil. The dense *CG1732*-labelling pattern ends abruptly at a consistent layer in the neuropil. Medulla neuropil glia are morphologically similar and localized similarly to these *CG1732* labelled cells [fig. 3A–C in Richardt et al. (Richardt et al., 2002)] and also send processes into the distal portion of the medulla, where the *CG1732* label is heaviest. *CG1732*-positive cells were not found in the cortex that surrounded the lamina or medulla.

Since *CG1732* is likely to be a GABA transporter and GABA uptake has been described in the lamina of the optic lobe (Campos-Ortega, 1974), we looked more closely at this region of the CNS. The optic lobes at higher magnification (boxed area of Fig. 5A,C) show that the lamina neuropil is lightly labelled with heavier label over a cluster of large cell bodies at the proximal border of the neuropil (arrowheads), where the outer chiasmatic and marginal glia both reside (Eule et al., 1995). The lamina neuropil has a weak purple tint that ends abruptly at the distal margin suggesting that cells that have processes that penetrate the lamina express this transporter. Epithelial glia would be the obvious candidate, as their processes surround the photoreceptor terminals in the lamina, and indeed in overdeveloped preparations, the cell bodies of the epithelial glia at the distal margin of the lamina appear to have faint label (data not shown). Nevertheless we were unable to definitively determine if the epithelial glia express *CG1732*.

The Repo antibody labels cells that both surround the neuropil and are intermixed with neuronal somata in the

Fig. 4. Localization of transcripts involved in monoaminergic neurotransmission using *in situ* hybridization. Representative frontal sections showing the labelling in the head (A,C,E,G–J) or the thorax (B,D,E). In (A,C,E) the ellipsoid body is circled with a dotted line to illustrate that sections are at approximately same depth in the head. (A,B) The *SerT* anti-sense riboprobe labels discrete cells in the head and in the thorax. Anatomical positions of the labelled cells in (A) are consistent with the SP1 (arrows), LP1 (notched arrow), and SE2 (barbed arrow) clusters of serotonergic neurons. Cells in (B) are present in the posterior portion of the ventral ganglion (arrows). (C,D) The *DAT* anti-sense riboprobe labels discrete cells in the head (C) and thorax (D). The anatomical position of the labelled cells suggests these cells are part of the DM (arrows), DL<sub>1</sub> (white arrowheads), and DL2 (carrot) clusters of neurons. The cells in (D) are present in the posterior portion of the ventral ganglion, and are likely to be part of the dorsal lateral cluster (barbed arrow) and the medial cluster (arrows). (E,F) The riboprobe for *CG33528* labels many cells (markers have remained consistent with A and C) throughout the head (E) and thorax (F) that are in similar anatomical locations as cells that label for the transporters, *SerT* and *DAT*. Gray open arrows denote labelling of the medullary layer (MC cells). (G) Wider view of *CG33528* labelling showing the cells of the brain that are labelled (arrows, barbed arrows and white arrowheads) and that a layer of cells at the base of the photoreceptors is also labelled (black arrowheads). Box denotes area of higher magnification in (H,I). (H,I) Serially sectioned preparation illustrating the region boxed in G but in a different preparation. (H) Section labelled with *CG33528* shows label right at the base of the photoreceptors (PR). Block arrows denote examples where the *CG33528* label has penetrated the photoreceptor layer. The dotted line represents the outline of the lamina neuropil (LaN). (I, top) Fluorescence image showing Nr<sub>x</sub>IV antibody labelling glial septate junctions at the base of the photoreceptors in the same anatomical position as the *CG33528* label in (H). (I, bottom) For clarity, the brightfield image of the fluorescence image has been presented so that the morphological features (i.e. the characteristic striations of both the photoreceptor layer and the lamina neuropil) that define the different layers in optic lobes can be distinguished. The broken line outlines the lamina neuropil and the solid line denotes the base of the photoreceptors. (J) The riboprobe for the histamine synthesizing enzyme, *histidine decarboxylase (HDC)*, labels cells in the central brain region (arrows) as well as the photoreceptor cell bodies (barbed arrows). In all panels dorsal is up. Scale bars, 50 μm.

Fig. 5. *CG1732* and *CG15088* are expressed in a glial-like pattern. The various glial subtypes are shown in (B) and the corresponding area in a sequential section is shown in (A). Examples of perineural (arrow) and subperineural (carrot) glia are marked [nomenclature according to Eule et al. (Eule et al., 1995)]. Inner chiasmatic glia are marked by the barbed arrow. Neuropil are labelled Medulla/Lobula (Me/Lo), suboesophageal ganglion (SEG), central brain (CB), and photoreceptors (PR). (A) Frontal section of the fly head showing the brain and optic lobes. Cells labelled with the anti-sense riboprobe for *CG1732* surround the neuropils of both structures. The label occurs in regularly spaced intervals; the neuropils have a purple hue suggesting that the mRNA may be present in penetrating processes. (B) Sequential section to A labelled with the glial marker, Repo. Cells that express *CG1732* are likely to be glia because they occupy the same anatomical position and approximate spacing as cells that label with the nuclear glia marker Repo. Boxed areas in A and B denote areas shown at higher magnification in C and D, respectively. (C) Higher magnification of the optic lobes in A. In both C and D, arrowheads indicate labelled cells in either outer chiasm giant glia or marginal glia at the proximal margin of the lamina neuropil. Medulla neuropil (MeN), Medulla cell bodies (MeCB), and lamina neuropil (LaN) are denoted. Arrows show the location of glia at the distal margin of the lamina; open arrows show position occupied by epithelial glia. Notched arrows mark positions of medulla neuropil glia. Glia distal to the lamina neuropil do not label with *CG1732* and epithelial glia at the distal margin may faintly label for *CG1732*. (D) Higher magnification of B showing Repo label in the large glial nuclei of the outer chiasm glia (arrowheads) at the proximal margin of the lamina. Arrows show glia at the distal border of the lamina and open arrows show the epithelial glia; both label for Repo. Notched arrows mark the nuclei of medulla neuropil glia. (E) Pattern of label of *CG15088* probe. (F) the Repo antibody labels nuclei in similar sections from different preparations. Asterisks denote comparable neuropils. Like Repo, *CG15088* appears to label glia throughout the cortex. All sections are frontal sections with dorsal up; scale bars, 200  $\mu\text{m}$  (A,B,E,F), 50  $\mu\text{m}$  (C,D).



cellular cortex of both the lamina and the medulla. Prominent among the Repo-labelled nuclei are cells at the proximal border of the lamina (arrowheads, Fig. 5D) that are in the same position as *CG1732*-positive cells (arrowheads, Fig. 5C). Outside the neuropil borders, there exist Repo-labelled glia at the base of the photoreceptors (Fig. 5D, arrows) that are not

labelled by *CG1732*, again indicating that that *CG1732* labels only a subset of glia.

*CG15088* labels a different set of glia from *CG1732* (Fig. 5E). This putative SLC6 transporter is found within the IAAT subfamily rather than in a neurotransmitter transporter subfamily. *CG15088* is present in both a punctate and diffuse

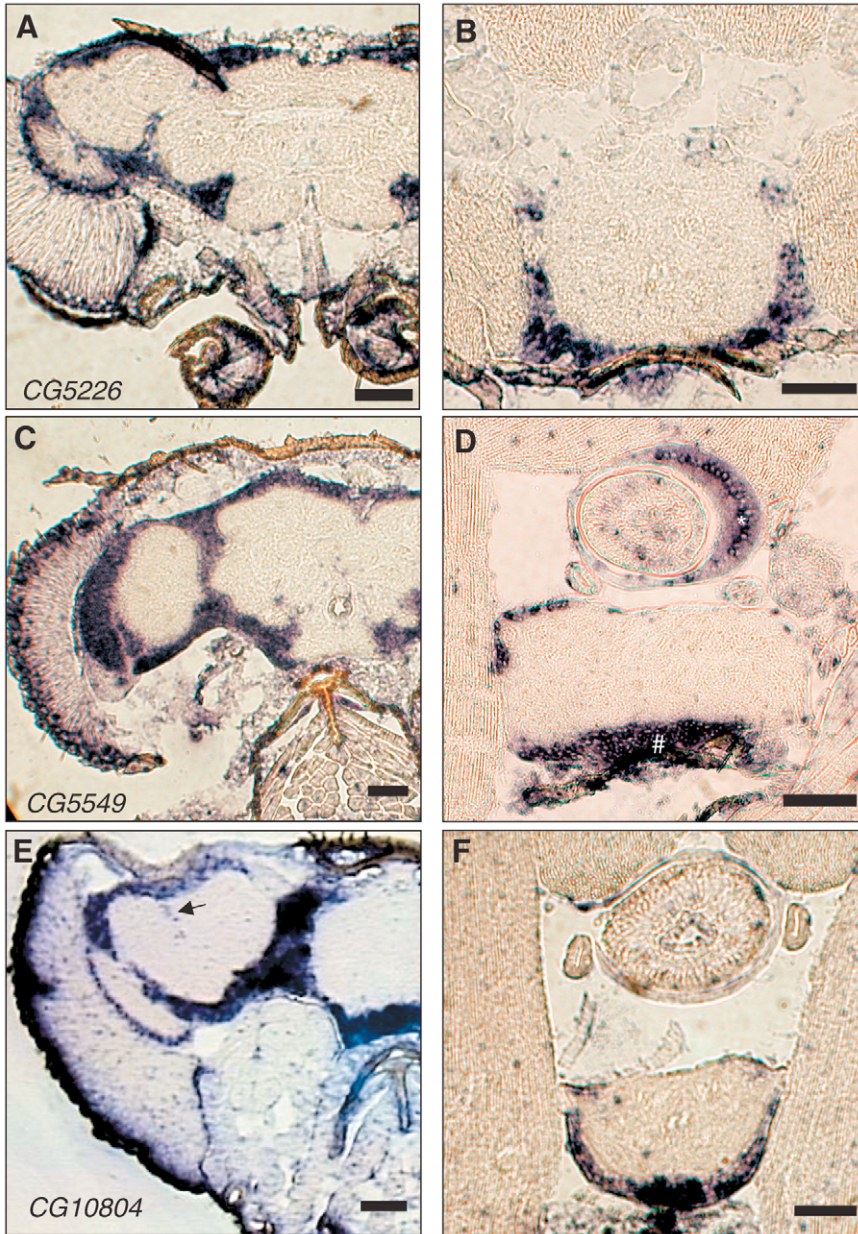


Fig. 6. *CG5226*, *CG5549*, and *CG10804* label cells broadly throughout the cell body layer in the adult head. (A) Representative horizontal section illustrating that the riboprobe for *CG5226* labels cells throughout the CNS cell body layer of the head (A) and representative frontal section showing labelling in the thorax (B). We did not detect glial labelling in the optic chiasm giant glia in addition to the cortical cell bodies. (C,D) Representative frontal sections showing the probe designed against *CG5549* labels the cellular cortex and the photoreceptors in the head (C) and CNS cells in the thoracic ganglion (#) and cardia (\*) in (D). (E,F) The riboprobe for *CG10804* labels cells throughout the adult *Drosophila* head, including inner chiasm giant glia (arrow) (E) and CNS cells in the thorax (F). All panels are frontal sections with dorsal up; scale bars, 50  $\mu\text{m}$ .

pattern throughout the cortex; the labelled cells line and define the neuropils at the inner border of the cortex and also are found throughout the cortex (Fig. 5E). This pattern is consistent with both that of subperineural and perineural glia. In contrast to *CG1732*, which labelled structures within the neuropil but not

the cortex, the riboprobe for *CG15088* labels the cortex and does not label neuropil. *CG15088* label has a similar pattern to that of Repo-positive cells (Fig. 5F). Label from both probes surround neuropil structures such as the antennal lobes, ventral bodies and the ellipsoid body shown in Fig. 5E,F (asterisks), and both reagents label cells throughout the cortex. Also, we have detected *CG15088* label in the inner chiasm giant glia where only glial cell bodies are found (data not shown). These results suggest that *CG15088* may be expressed in all glia of the *Drosophila* cortex where it may play a role in transporting nutrient amino acids within the CNS.

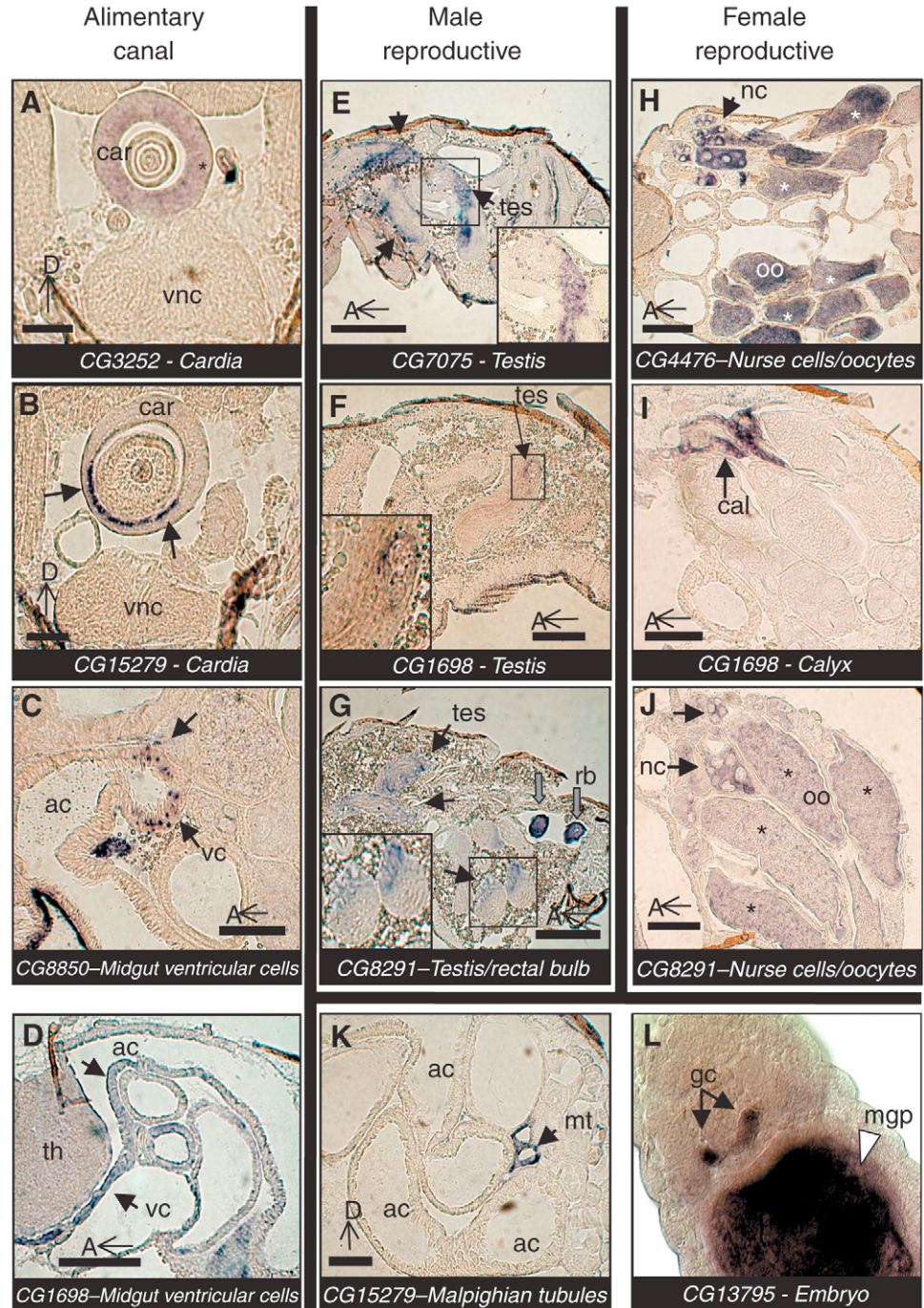
*Three Drosophila SLC6 homologs are broadly expressed in the CNS*

*CG5549*, a candidate in the amino acid subfamily, and *CG5226* and *CG10804*, both segregating with the orphan neurotransmitter transporter subfamily, are expressed broadly throughout the cellular cortex in the head (Fig. 6A,C,E) and the thorax (Fig. 6B,D,F). The cortex comprises primarily neurons with glial cell bodies intermixed, but dense label throughout the head prevented us from distinguishing whether both types of cells were labelled. In the region of the inner optic chiasm between the medulla and lobula neuropil, only glial cell bodies reside (Tix et al., 1997), and we used this region to determine if at least this subset of glia was labelled by the probes. The riboprobes for *CG5549* and *CG10804* (but not *CG5226*) labelled large cell bodies at the inner optic chiasm (Fig. 6E, filled arrow). Of these three candidates, only *CG5549* labelled the photoreceptors. *CG5549* also labelled a cell layer in the cardia (asterisk in Fig. 6D), an organ at the transition between the foregut and midgut. The cardia is responsible for the creation of the peritrophic membrane, a four-layered extracellular barrier between the contents of the gut and the gut epithelium (King, 1988).

*Eight SLC6 transporters were detected exclusively outside of the CNS*

SLC6 members have been shown to transport compounds other than neurotransmitters, including amino acids and osmolytes, such as betaine and taurine. For *Drosophila* SLC6 transporters that had no detectable CNS expression, we extended our analysis to verify that the riboprobe would bind to transcripts expressed in other organs.

Fig. 7. Eight candidate label cells exclusively outside of the CNS. Panels show representative sections of a variety of tissues labelled using *in situ* hybridization, and are organized vertically by type of tissue labelled: (A–D) alimentary canal, (E–G) male reproductive system, (H–J) female reproductive system, and (K,L) other tissues. Frontal sections through the gut demonstrate that the riboprobe for *CG3252* (A) and *CG15279* (B) both label cells in the cardia (car; \*), a structure at the transition between the foregut and the midgut. Ventral nerve cord is marked (vnc). (C,D) The anti-sense riboprobe for *CG8850* (C) and *CG1698* (D) both label ventricular cells (vc; filled arrows) that line the alimentary canal (ac) that runs from the thorax (th) into the abdomen. *CG8850* labels cells in a restricted domain of ventricular cells in the abdomen; neighbouring ventricular cells along the alimentary canal are not labelled. Sagittal sections reveal that *CG7075* (E), *CG1698* (F) and *CG8291* (G) label male reproductive tissue, likely the testis (tes) in the male abdomen (arrows). In addition, *CG8291* (G) labels the rectal bulb (rb; open arrows), a hindgut structure at the end of the alimentary canal. Insets show boxed areas at higher magnification. Horizontal (H) and sagittal (I,J) sections show that *CG4476* (H), *CG1698* (I), and *CG8291* (J) label structures in the female reproductive system. *CG4476* (H) and *CG8291* (I) label nurse cells (nc; arrows) and developing oocytes (oo; \*). *CG1698* (I) labels a structure called the calyx (cal; arrows). (K) *CG15279* (mt; arrows) also labels two closely apposed Malpighian tubules. (L) Midgut precursor cells (mgp; arrowhead) and the Garland cells (gc; arrows) were labelled by *CG13795* in a stage 14 embryo. At later stages, only the Garland cells are labelled. Anterior of the embryo is in the upper left corner and this is a dorsal view of the embryo. Scale bars, 50  $\mu\text{m}$  (A,B), 100  $\mu\text{m}$  (C,F,I–K) and 200  $\mu\text{m}$  (D,E,G,H). A, anterior; D, dorsal.



We determined if any of the candidate genes were expressed in other recognizable organs such as the alimentary canal, Malpighian tubules and/or reproductive organs.

Two candidates from the IAAT subfamily (*CG3252* and *CG15279*) were found in the cardia of the gut (Fig. 7A,B). The *Drosophila* cardia has been sub-divided longitudinally into 6 zones based on the morphological and intracellular characteristics of the constituent cells (King, 1988). *CG3252* is present in the long columnar cells that make up zone 5 of the

cardia (Fig. 7A) and the label persists through zone 6 and into the posterior ventricular cells of the midgut (data not shown) where nutrient uptake is thought to occur. *CG15279* (Fig. 7B) is expressed in cells near the junction of the cardia and the ventricular cells but these cells are much shorter than the zone 5 cells. These cells are likely to be the cells of zone 6. *CG15279* label does not persist beyond the transition of the cardia to the intestine.

Three other candidates are also detected in cells along the

alimentary canal. Two candidates from the IAAT subfamily, *CG8850* and *CG1698*, are present in the digestive portion of the midgut. *CG8850* (Fig. 7C) is found in cells that line the gut lumen in a restricted portion; adjacent cells do not express *CG8850*. Midgut epithelial cells are responsible for the uptake of nutrients from consumed food (Miller, 1950). The *CG1698* (Fig. 7D) riboprobe labels a broader domain of the midgut that stretches from the thoracic into abdominal segments. *CG8291*, a candidate that does not segregate into a distinct subfamily, is expressed in the rectal bulb of the alimentary canal (Fig. 7G, arrows).

Fig. 7E–J shows tissues labelled by four transcripts found in the reproductive organs. *CG7075* is found exclusively in the testis of the male fly (Fig. 7E). The testes are recognizable by the elongated, striated appearance of the developing sperm within the organ (Miller, 1950). The riboprobes for *CG1698* (Fig. 7F,I) and *CG8291* (Fig. 7G,J) both label the reproductive organs in both genders. *CG1698* labels the testis in the male (Fig. 7F) and a subset of cells in the female reproductive tract and potentially in the epidermis of the calyx, the region of the oviduct that meets the ovary (Fig. 7I). *CG8291* was detected in the testis (Fig. 7G, open arrows) and the nurse cells and oocyte in the female reproductive organ (Fig. 7J). Nurse cells furnish the developing oocyte with nutrients and maternally derived mRNAs (Mahajan-Miklos and Cooley, 1994). *CG4476* is expressed exclusively in the nurse cells and oocytes of the female reproductive system (Fig. 7H).

Only one candidate, *CG15279*, was found in the Malpighian tubules (Fig. 7K), the organ that carries out kidney-like function in the fly. The figure shows two closely apposed tubes

that fit the morphological description of the cells and anatomical description of the Malpighian tubules (Miller, 1950). This apposition, and the cell morphology of the labelled structures, suggests that these structures are the Malpighian tubules somewhere after the stalk splits from one to two tubes. We were able to detect positive cells with these same characteristics in multiple regions of the abdomen in frontal sections (data not shown). Since the Malpighian tubules comprise a total of four branches that wind through the abdomen, it seems likely that the tubular cross-sections we observe belong to this system.

We detected *CG13795*, an outlier candidate sequence, in the embryo but not in the adult (Fig. 7I). The probe for *CG13795* labels the midgut precursors and the Garland cells at stage 14 and 15; as the gut matures to stage 16 it loses the expression of *CG13795*, but labelling of the Garland cell cluster, with its characteristic U-shape (Miller, 1950), is retained. Garland cells encircle the proventriculus, are known for their high endocytosis activity, and are proposed to carry out a liver-like function in cleaning the hemolymph of the fly (Kosaka and Ikeda, 1983).

## Discussion

In this study, we set out to identify the full complement of *Drosophila* SLC6 transporters present in the *Drosophila* genome and determine their expression pattern in the adult CNS. Our search identified 21 sequences with sequence similarity to the SLC6 family, Table 1 lists these sequences and summarizes the overall findings. A majority of these sequences

Table 1. Summary of *Drosophila* SLC6 family transporters

Putative family	Transporter	Chromosome	Localization	RT-PCR expression
Serotonin	Dm SerT	60C8	CNS cells	Head/thorax
Dopamine	Dm DAT	53C7-C14	CNS cells	Head/thorax
Glycine	Dm CG5549	60A2-A3	Broad CNS	Equivalent
Proline	Dm CG7075	28C1	Male abdomen	Male abdomen
GABA	Dm CG1732	102D4	Glia	Head/thorax
'Orphan' NT transporters	Dm CG5226	55D4	Broad CNS	Equivalent
	Dm CG10804	3D4	Broad CNS	Head/thorax
	Dm Ine	24F4	Photoreceptors, CNS, digestive tract <sup>1</sup>	Equivalent
Insect AA transporters	Dm CG1698	46B3	Alimentary canal/reproductive cells	Equivalent
	Dm CG4476	67A2-A3	Female abdomen	Female abdomen
	Dm CG15279	35B6	Cardia/Malpighian tubules	Prominent in abdomen
	Dm CG8850	48F6	Ventricular cells	Equivalent
	Dm CG15088	55E11	Glia	Equivalent
Unclassified	Dm CG3252	4F9	Cardia	Equivalent
	Dm Blot	74B	Epithelium, CNS, Malpighian tubules <sup>2</sup>	Head/abdomen
	Dm CG8291	52D2	Reproductive cells	Equivalent
	Dm CG13793		Not done	Equivalent
	Dm CG13794		Not done	Equivalent
	Dm CG13795	28C2	Embryonic Garland cells/midgut	Equivalent
	Dm CG33296		Not done	Equivalent
Dm CG13796	28C2	No <i>in situ</i>	Equivalent	

<sup>1</sup>(Soehnge et al., 1996; Burg et al., 1996).

<sup>2</sup>(Johnson et al., 1999).

either segregated into four previously recognized subfamilies or helped to define a novel subfamily, the IAAT subfamily. Using *in situ* hybridization, we localized seven candidates to the CNS. Of these seven genes, two known genes (*SerT* and *DAT*) were expressed in a discrete pattern in the brain and thoracic ganglion predicted by known immunolabelling, two candidates (*CG1732* and *CG15088*) were expressed in a pattern consistent with glia, and three candidates (*CG5549*, *CG5226* and *CG10804*) were broadly expressed in the CNS. Neither phylogenetic analysis nor the *in situ* hybridization results suggested a candidate for an octopamine or histamine transporter despite the known presence of neurons containing these amines. Eight candidates labelled cells in the alimentary canal, reproductive organs and Malpighian tubules but were not detected in the CNS. We did not attempt or were unable to determine a pattern for the six remaining candidates for reasons discussed below.

*Drosophila SLC6 have similar structural features to the leucine transporter from Aquifex aeolicus*

The crystal structure of a bacterial Na<sup>+</sup>-dependent leucine transporter homologous with mammalian SLC6 was recently solved. Of particular interest, the structure reveals residues important for substrate binding, sodium binding and gating of the transporter. One major difference between our multiple sequence alignments and those of Yamashita et al. (Yamashita et al., 2005) is the inclusion of a larger number of sequences (see Fig. 1B and Fig. S1 in supplementary material). There are no absolutely invariant residues identified in our complete alignment of 84 putative transporters, and the only invariant residue in the *Drosophila* transporter alignment is a glycine residue at the end of TM2. We also find several residues that seem to be more conserved among metazoan transporters than would be suggested by the Yamashita alignment. The densest concentration of amino acid residues with direct contacts with the substrate and sodium were found in TM1, TM6 and TM8 with other critical residues scattered throughout the protein. Many of the known and candidate *Drosophila* SLC6 sequences contain these precise residues or have conservative amino acid substitutions, although there are clear differences among certain divergent transporter subgroups that may underlie differences in substrate specificity, ion binding and gating (see Fig. 1 and Fig. S1 in supplementary material).

Blot is perhaps one of the more divergent transporter sequences, yet it contains a conserved arginine responsible for gating and a conserved asparagine involved in sodium binding present in TM1, as well as many other conserved residues. In addition, Blot has been localized and the absence of Blot has a phenotype (Johnson et al., 1999), therefore Blot is a functional protein despite the disparity in sequence homology.

For CG33296 and CG13793–CG13796, the situation is not as clearcut, but our data suggests that they are expressed, as we have detected at least low expression by RT-PCR of each of these genes. Thus far, the predicted proteins for CG13793 and CG33296 are missing the critical TM1 and part of TM2. This could be a result of protein prediction errors or a common first

exon that is shared between these proteins (see below). Another possibility is that these proteins have adopted a slightly different method of functioning from the ancestral SLC6 transporter. CG13794, CG13795 and CG13796 contain divergent TM1 sequences (though they still retain several highly conserved residues) throughout this and other segments of the alignment. For CG13795, we have demonstrated discrete expression in the Garland cells in late stage embryos, suggesting a specific role in these cells.

*Sequences in the Drosophila genome help define a novel, large subfamily, IAAT*

Our results confirm and expand a sixth, novel subfamily of SLC6 transporters, referred to as IAAT (Boudko et al., 2005), which we show includes six putative transporters from *Drosophila* and seven putative transporters from *Anopheles*. With the exception of minor differences in branching geometry and branching order at nodes with low bootstrap values, the clusters of transporters in the two trees are largely the same (Boudko et al., 2005). The sequence differences between the IAAT and other SLC6 subfamilies may underlie physiological differences in the function of these transporters. There are three cloned members of this subfamily, the *Manduca sexta* proteins known as potassium-coupled amino acid transporter-1 (KAAT1) (Castagna et al., 1998), the cation-anion-activated amino acid transporter/channel-1 (CAATCH1) (Feldman et al., 2000) and the *Aedes aegypti* amino acid transporter (AeAAT1) (Boudko et al., 2005). All three are competent to transport amino acids using either K<sup>+</sup> or Na<sup>+</sup> as the driving ion, especially at highly negative membrane potentials. In contrast, other SLC6 transporters use only Na<sup>+</sup> as their driving ion. The molecular basis of this ion specificity may relate to residues in the transporter responsible for coordinating these ions. In this regard, it is intriguing that the IAAT transporters all contain an alanine or serine substitution at the site of the glycine residue in TM1a reported to be involved in coordinating sodium ion Na<sup>+</sup> (Yamashita et al., 2005). It should also be noted that other sodium coordinating residues or residues whose side chains reportedly interact with sodium ions in TM6a and TM8 are somewhat less conserved across the entire family of transporters.

The physiological difference in ion selectivity is thought to reflect the environment of the insect gut in which these transporters operate. In *Manduca*, goblet cells secrete high concentrations of potassium into the lumen of the gut; in addition, the columnar cells that take up the amino acids have an unusually high membrane potential across their luminal (apical) membrane (Harvey and Wieczorek, 1997). Nutrient uptake into the columnar cells is driven by both the high concentration of K<sup>+</sup> and the large negative membrane potential. All three of the cloned transporters are associated with the insect midgut cells: KAAT1 was localized to columnar cells, CAATCH1 was cloned from a library created from midgut epithelium, and AeAAT1 was cloned from a posterior midgut library and was localized to various structures in the gut, including the cardia, posterior midgut and Malpighian tubules (Boudko et al., 2005).



Our *in situ* data, however, demonstrate that *Drosophila* IAAT subfamily members are expressed in more varied tissues than the gut. Confirming this, RT-PCR data suggests that five out of six *Drosophila* members of the IAAT subfamily are expressed in all three segments of the fly, the exception being *CG4476*, which is only weakly expressed in tissues outside of the female abdomen. *In situ* hybridization revealed that IAAT members were expressed in such diverse tissues as CNS (*CG15088*), midgut (*CG1698* and *CG8850*), cardia (*CG3252* and *CG15279*) and the Malpighian tubules (*CG15279*). As the brain is not expected to have a high potassium environment or unusually high membrane potentials as in *Manduca* gut, it is possible that the *Drosophila* transporters from the IAAT subfamily could use either Na<sup>+</sup> or K<sup>+</sup> as their driving ion, depending on ion availability in a given environment.

We can compare our IAAT localization results with two independent sources of information: microarray expression studies being carried out on isolated tissues and the embryonic *in situ* hybridization project being carried out by the Berkeley *Drosophila* Genome Project (BDGP) (Tomancak et al., 2002). Microarray data from the adult Malpighian tubules demonstrated that only the SLC6 transcript, *CG15279*, was enriched (30-fold) compared against the rest of the fly (Wang et al., 2004). This agrees with our *in situ* hybridization results (Fig. 7K).

BDGP has partially completed a systematic determination of embryonic expression for each annotated *Drosophila* gene using *in situ* hybridization. Patterns for four of the IAAT subfamily members have been released. *CG3252* was found in the embryonic hindgut and the Malpighian tubules, whereas we detected it in the cardia of the adult alimentary canal. *CG4476* was detected in early embryos, probably due to maternal contribution, and in the embryonic stomatogastric nervous system, germ cells and endocrine system. In the adult, we detected *CG4476* in nurse cells and developing oocytes of the female reproductive system, which confirms that mRNA for *CG4476* is maternally contributed to the embryo. BDGP reported expression of *CG1698* in the embryonic proventriculus, frontal ganglion and stomatogastric nervous system, but we detected *CG1698* in the reproductive organs and midgut of the adult. *CG8850* was identified in the embryonic Malpighian tubules by BDGP, but we found it expressed in cells lining the adult ventriculus in the midgut. These expression differences between the adult and the embryo are likely due to the specific requirements of the cells at the adult and embryonic stage.

Discrepancies exist between our RT-PCR and *in situ* localization observations. Indeed such discrepancies might be expected. The areas of detection differed between *in situ* hybridization and RT-PCR. Our *in situ* hybridization focused on the CNS and on other tissues in which we could clearly distinguish a signal, while for RT-PCR an entire body segment was used to derive the cDNA. Therefore, the RT-PCR may have detected gene expression in a tissue that we did not examine using *in situ* hybridization. Also, RT-PCR is the more sensitive technique; a transcript expressed at low levels but

broadly throughout a tissue would be easier to pick up by RT-PCR.

#### *Monoaminergic neurotransmitter transporters, SerT and DAT are found in presynaptic neurons in the adult CNS*

We have localized *SerT* and *DAT* to neurons in the adult CNS that label in a pattern similar to the label for the respective neurotransmitter. Neurotransmitter transporters have been localized to presynaptic neurons, postsynaptic neurons and surrounding glia; *SerT* in the embryo (Demchyshyn et al., 1994) and *DAT* in the larva (Porzgen et al., 2001) have been localized to cells in the pattern of cells that immunolabelled for their respective neurotransmitter. Not every cell that labels for dopamine was positive for *DAT* labelling, indicating that all cells that release dopamine may not re-capture this compound. The localization of monoaminergic neurotransmitter transporters agrees with localization data from other species, which are found in the presynaptic neurons (Hoffman et al., 1998).

#### *Amino acid transporter subfamily*

There is only a single *Drosophila* SLC6 transporter (*CG5549*) that segregated into the amino acid subfamily. *CG5549* is expressed in each segment of the fly, as determined by RT-PCR, and is expressed broadly throughout the adult CNS, including the inner chiasm giant glia and the photoreceptors. Cloned members of this subfamily have substrates which include amino acids that double as neurotransmitters, such as proline or glycine (Malandro and Kilberg, 1996), or a broader substrate profile of cationic and neutral amino acids for the transporter B<sup>O+</sup> (Sloan and Mager, 1999). The broad expression of *CG5549* in both neurons and glia of the CNS suggests a basic role for this candidate, such as nutrient uptake.

#### *Orphan neurotransmitter transporter subfamily*

The two candidates that segregate with the orphan transporters, *CG5226* and *CG10804*, both displayed a broad localization pattern throughout the CNS. Within the CNS, both *CG5226* and *CG10804* were detected in neurons. *CG10804* expression was also detected in the inner chiasmatic glia but we were unable to detect *CG5226* in this subset of glia. RT-PCR revealed expression differences. *CG10804* was enriched in the head and thorax, segments containing cell bodies of the CNS, whereas *CG5226* was evenly expressed across all segments. These expression differences suggest that these two orphan transporters may be carrying out broad but different functions in the fly, such as importing a necessary compound for cellular function or clearing a compound with a wide extracellular distribution. The embryonic localization by BDGP of *CG5226* (*CG10804* has not been released) shows that *CG5226* is expressed in cells throughout the more mature CNS of embryonic stages 13–16, but not in earlier CNS stages when the CNS is still developing. The similar localization pattern for *CG5226* in the embryo and adult suggests a similar role in more mature CNS cells. It has been predicted that substrates for

members of the orphan neurotransmitter transporter family will include amino acids (Boudko et al., 2005), based on phylogenetic analysis, and our localization data is consistent with this prediction. Interestingly, these transporters possess highly divergent extracellular linker sequences 2, 4a and 6, consistent with a role for these sequences in substrate selectivity.

#### *GABA transporter subfamily*

In *Drosophila*, uptake of the neurotransmitter GABA has been demonstrated in the lamina (Campos-Ortega, 1974), but the gene responsible for this uptake has not been identified. There is compelling evidence that *CG1732* is a GABA transporter (GAT) in *Drosophila*. It is the only candidate that segregated into the GABA subfamily, and it has significant identity with other known GABA transporters: *Manduca sexta* GAT (80%), *Trichoplusia ni* GAT (81%), human GAT1 (59%) and human GAT3 (53%). By RT-PCR, *CG1732* is enriched in the head and thoracic segments but it is also present in the abdomen segment.

Evidence from other species suggests that GABA transporters can be present in both neurons and glia (Borden, 1996), but our data demonstrate that *CG1732* is expressed in a subset of glia. In *Drosophila*, the pattern of *CG1732* label only partially overlaps with the pattern of GABA immunolabelling in the optic lobes (Buchner et al., 1988). The GABA antibody labels approximately 1500 somata in the cortex of the medulla and, in the neuropil, there is faint label in the lamina, and denser label in the medulla. *CG1732* label is found in the neuropil but not in cell bodies of the cortex, suggesting that GABAergic neurons are not the primary source of *CG1732* expression in the medulla. Instead, the labelling pattern for *CG1732* is better represented by the immunolabelling pattern for the glial marker, Repo. In addition, glial expression of a *Drosophila* GABA transporter is supported by GABA uptake studies in the lamina of housefly and *Drosophila*, where <sup>3</sup>H-GABA was accumulated in the glia rather than neurons of the optic lobes (Campos-Ortega, 1974). These authors described uptake into glia of the distal border of the lamina (epithelial glia) and at the proximal border (marginal glia). We were not able to conclusively show expression of *CG1732* in these cell types, but the lamina does show weak labelling in the lamina neuropil. Finally, in the embryo BDGP localized *CG1732* expression to a subset of cells in the ventral nerve cord that are in a position consistent with the channel glia. Thus, our evidence, combined with published findings, indicates that expression of *CG1732* is in a subset of glia in *Drosophila*.

In another insect, *Manduca sexta* (Umesh and Gill, 2002), MasGAT immunoreactivity coincided with GABA immunoreactivity (Homberg et al., 1987) in the neuropil but did not appear to be in coincident locations in the cortex. In adult *Manduca*, somata immunoreactive for GABA appear throughout the cortex of the optic lobe (Homberg et al., 1987), as is the case for *Drosophila*. Somata in this same area of cortex are not labelled by the antibody against MasGAT (Umesh and Gill, 2002) but somata near the neuropil are clearly labelled and

often send a process into the nearest neuropil. In fact, some of these author's photographs reveal a band of dark staining surrounding the optic lobe neuropil, similar to *CG1732* labelling that we detect. These results suggest various possibilities. First, MasGAT is expressed in GABAergic neurons, but the final position of the MasGAT protein is at the terminals and not the cell bodies of GABAergic neurons. Therefore the antibody labelled only the terminals, leaving GABAergic cell bodies unlabelled. This explanation is possible though other somata were clearly labelled by the MasGAT antibody. It is also possible that only a small subset of neurons very near the neuropil express MasGAT. Another possibility is that MasGAT is expressed in glia similar to our conclusion in *Drosophila*.

#### *Candidates unaffiliated with a defined subfamily*

In our phylogenetic analysis, five candidates did not fall into any distinct subfamily, not even into the orphan subfamily. We have called these 'unclassified sequences' to distinguish these proteins from the previously defined orphan subfamily. We were able to localize these candidates outside the CNS.

Two unclassified candidates (*CG7075* and *CG8291*) are associated with the reproductive tissue. By *in situ* hybridization, *CG7075* is found in the testes of the male, and our RT-PCR results show that *CG7075* is expressed almost exclusively in the male abdomen, which supports our *in situ* localization results. In the adult, *CG7075* is likely to be a testes-specific gene because we did not detect it elsewhere in the fly. Also, *CG7075* was found as an EST (BF486171) from adult testes; testis expression was noted in a Flybase communication (Bazinet, 2000.7.10). *CG7075* was also found to be a testis-specific gene as determined by microarray analysis (Parisi et al., 2004).

*CG8291* is localized in the reproductive tissue in both genders, in nurse cells and oocytes in the female and in the testes in the male. In a microarray study, Parisi et al also found that *CG8291* had enriched expression in the testis (Parisi et al., 2004). The *CG8291* riboprobe also labelled a structure along the alimentary canal, the rectal bulb, of both males and females. The rectal bulb is thought to be responsible for the breakdown of the peritrophic membrane, which forms a barrier between the contents of the gut and the gut epithelium and is generated by the cardia (Miller, 1950). Unfortunately, there is no clear common feature among the tissues in which *CG8291* is expressed which could lead to a hypothesis regarding *CG8291* function.

Interestingly, several of our unclassified candidate genes are organized in a cluster at the genetic region of 28C. The open reading frames for *CG13793*–*CG13796*, *CG33296* and *CG7075* all reside in what appears to be a region of genetic duplications, with up to 98% identity in portions of the repeated regions. *CG13795* is 92% identical to *CG13793* and 80–89% identical to *CG33296*, so it was difficult to confidently distinguish one sequence from another in this area. The organization of this region may indicate a unique relationship between these genes. One possibility is that a common exon

could be spliced to any one of the genes to create differentially functioning proteins. Alternatively, the subtle sequence differences could be significant for the timing or localization of candidate expression. This genetic region could form some sort of locus that possibly functions similarly to the cholinergic locus in *Drosophila*, where two sequences with different functions share the first exons (Kitamoto et al., 1998). This region could also represent an area of gene duplication, in which transporters with different substrate profiles are generated as postulated by Boudko et al. (Boudko et al., 2005).

#### *Candidates not localized by in situ hybridization*

Five identified candidates (*ine*, *blot*, CG13793, CG13794 and CG33296) were eliminated from the *in situ* hybridization study. Two of the candidates, *blot* and *ine*, have already been cloned and localized (Burg et al., 1996; Huang et al., 2002; Johnson et al., 1999) and were not pursued further. Three candidates (CG13793, CG13794 and CG33296) were not pursued further because they did not have a predicted N terminus that agreed with other SLC6 homologs, and these candidates are located in a region likely with a genetic duplication. We approached CG13795 and CG13796 as representative sequences from this area. Neither CG13795 nor CG13796 were found in the adult, and therefore we did not pursue CG13793, CG13794 or CG33296 further. The fact that we were able to detect CG13795 in the embryo shows that the entire region cannot be attributed to pseudogenes and our RT-PCR results show that there is weak expression of each of these transcripts in the adult. Perhaps this genetic region represents an area where the genome has expanded and will generate an expansion of transporter population as described (Boudko et al., 2005).

Finally, one potential candidate with weak homology to SLC6 transporters (CG31904) was excluded from our study because homology extended only over a small fraction of the predicted protein. This small region is predicted to be fused with the adult cuticular protein 1 (which has no sequence similarity to SLC6 transporters), raising strong suspicions about an error in gene prediction.

#### *Monoamine neurotransmitter transporters not found in our analysis*

One surprising aspect of this study is that we did not find candidate sequences for the histamine and octopamine transporters in the monoamine subfamily. The absence of a *Drosophila* octopamine transporter (OAT) is perplexing since an OAT gene does exist in another insect, *Trichplusia ni* (Malutan et al., 2002), and, like *T. ni*, there are cells in *Drosophila* that immunolabel for octopamine (Monastirioti et al., 1995). Furthermore, octopamine influences behaviors such as grooming and locomotion (Yellman et al., 1997). The question remains whether *Drosophila* uses an octopamine transporter and if so how divergent the *Drosophila* sequence is from that of *T. ni*.

The absence of a histamine transporter sequence is similarly puzzling. Histamine is the neurotransmitter of *Drosophila*

photoreceptors, and genetic evidence indicates that a mechanism to accumulate histamine into photoreceptors exists in *Drosophila* (Melzig et al., 1998), as it does in other species (Battelle et al., 1999; Stuart et al., 2002; Stuart et al., 1996). Flies lacking the enzyme that synthesizes histamine, histidine decarboxylase (HDC), are blind and have lost histamine immunolabelling in the optic lobe. These flies can then be fed histamine, which restores vision and histamine immunolabelling, suggesting that in the fly there is a process by which histamine can be accumulated in the photoreceptors and restore function to those cells. Based on data from another species (Stuart et al., 2002; Stuart et al., 1996), one would predict that the histamine uptake is dependent on Na<sup>+</sup> and Cl<sup>-</sup>, as are the other SLC6 transporters.

Indeed, none of the SLC6 transporters were expressed in the pattern of histamine uptake, that is, in the photoreceptors and in cells located in the antennal lobe (Melzig et al., 1998), as would be expected in a direct uptake model. At the photoreceptor terminals, histamine may recycle in a more circuitous route through the surrounding glia (Borycz et al., 2002). Recent work on mutant flies has revealed that histamine may be conjugated to  $\beta$ -alanine into an intermediate compound, carcinine, by the enzyme Ebony. Ebony was localized by antibody to both epithelial and medulla neuropil glia that surround the photoreceptor terminals (Richardt et al., 2002). In this scheme, histamine released from the photoreceptors would be taken up into the glia and converted to carcinine. The carcinine then must be shipped to the photoreceptors where it would be catabolized into histamine and  $\beta$ -alanine by the enzyme tan. In this scheme the histamine transporter would reside in the surrounding glia and a separate transporter would exist in the photoreceptors to take up carcinine. However, we did not find an SLC6 homolog associated solely with either the photoreceptors or with their surrounding glia.

#### *VMAT/CG33528*

A sequence that is potentially involved with monoaminergic neurotransmission is the VMAT, CG33528 (Greer et al., 2005). It is 44% identical to the mammalian VMAT2, which packages histamine into vesicles (Erickson et al., 1995). Our anti-sense riboprobe decorated a small number of cells in the central brain that are likely to be monoaminergic neurons (Monastirioti, 1999) and, unexpectedly, glial cells in a thin layer at the base of the photoreceptors. But this riboprobe did not label the photoreceptors, which are histaminergic and would be expected to express such a transporter. Immunocytochemistry also fails to find CG33528 in the photoreceptors or other histaminergic neurons although it co-localized with dopamine, 5-HT and octopamine neurons as expected (Chang et al., 2006).

Curiously, the CG33528 label at the base of the photoreceptors is in glia, not neurons. Two types of glia (fenestrated and pseudocartridge) have been described in this region based on anatomy and enhancer traps (Eule et al., 1995; Saint Marie and Carlson, 1983). In the house fly, glia surround the photoreceptor axons as they traverse the lamina cortex.

Fenestrated glia contain large vesicles (~80–100 nm) and pseudocartridge glia contain even larger vacuoles (~0.2–2.0  $\mu\text{m}$ ); since these glia are thought to help form a type of blood–brain barrier (Auld et al., 1995), the vesicles and vacuoles may relate to this function. The monoamines histamine and serotonin are both present in the lamina, histamine in the photoreceptors and serotonin in nerve terminals that course amongst the monopolar cell bodies in the lamina cortex (Buchner et al., 1988; Pollack and Hofbauer, 1991). A vesicular transporter in glia is not without precedent. Mammalian astrocytes have been documented to express vesicular glutamate transporters and release glutamate from vesicles onto neurons in a regulated manner (Bezzi et al., 2004).

Is CG33528 involved in the recycling of histamine at the photoreceptor terminals? We think the anatomy argues against this possibility. First, neither the processes of the fenestrated nor pseudocartridge glia extend into the lamina neuropil where the photoreceptors synapse onto the monopolar cells (Saint Marie and Carlson, 1983). Epithelial glia reside in this region. Second, CG33528 has a much higher affinity for monoamines other than histamine (Greer et al., 2005). Given the juxtaposition of serotonin nerve endings with the glia expressing CG33528, perhaps this transporter is primarily concerned in some way with serotonin. Serotonergic terminals have been shown to penetrate into the lamina cortex (Buchner et al., 1988).

#### List of abbreviations

AA	amino acid
Ag	<i>Anopheles gambiae</i>
AL	antennal lobe
AM	midline cell
BDGP	Berkeley <i>Drosophila</i> genome project
blot	bloated tubules
CNS	central nervous system
DAT	dopamine transporter
Dm	<i>Drosophila melanogaster</i>
EST	expressed sequence tag
GABA	$\gamma$ -aminobutyric acid
GAT	GABA transporter
HDC	histidine decarboxylase
IAAT	insect amino acid transporter
ine	inebriated
LaN	lamina neuropil
MC	medulla cell
MeN	medulla neuropil
Ms	<i>Manduca sexta</i>
NA	numerical aperture
NGS	normal goat serum
NrxIV	Neurexin IV
NSS	neurotransmitter:sodium symporter family
OAT	octopamine transporter
PBS	phosphate buffered saline
PCR	polymerase chain reaction

PFA	paraformaldehyde
RT	room temperature
RT-PCR	reverse transcription-polymerase chain reaction
SerT	serotonin transporter
SNF	sodium neurotransmitter transporter family
TBS	Tris-buffered saline
TM	transmembrane
Tn	<i>Trichoplusia ni</i>
VMAT	vesicular monoamine transporter
VNC	ventral nerve cord

We would like to acknowledge the contributions of many people that made this work possible. Steve Crews provided support for this foray into fly biology. We thank David Krantz and Rapheal Romero for valuable discussions and sharing unpublished results. Antibodies were provided by Manzoor Bhat (Neurexin IV) and the Developmental Hybridoma Studies Bank (Repo; Iowa City, IA, USA). We appreciate the histology expertise of Kirk McNaughton and Barb Taylor. Alan Fanning and Jim Anderson allowed us use of their Nikon microscope and Husok Lee generously provided us with an HDC plasmid. This work was funded by the University Research Council and Medical Alumni Association of the University of North Carolina at Chapel Hill and NIH grant MH65501, both to A.E.S.

#### References

- Adams, M. D., Celniker, S. E., Holt, R. A., Evans, C. A., Gocayne, J. D., Amanatides, P. G., Scherer, S. E., Li, P. W., Hoskins, R. A., Galle, R. F. et al. (2000). The genome sequence of *Drosophila melanogaster*. *Science* **287**, 2185–2195.
- Altschul, S. F., Madden, T. L., Schaffer, A. A., Zhang, J., Zhang, Z., Miller, W. and Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.* **25**, 3389–3402.
- Amara, S. G. and Arriza, J. L. (1993). Neurotransmitter transporters: three distinct gene families. *Curr. Opin. Neurobiol.* **3**, 337–344.
- Auld, V. J., Fetter, R. D., Broadie, K. and Goodman, C. S. (1995). Gliotactin, a novel transmembrane protein on peripheral glia, is required to form the blood-nerve barrier in *Drosophila*. *Cell* **81**, 757–767.
- Battelle, B. A., Calman, B. G. and Hart, M. K. (1999). Cellular distributions and functions of histamine, octopamine, and serotonin in the peripheral visual system, brain, and circumesophageal ring of the horseshoe crab *Limulus polyphemus*. *Microsc. Res. Tech.* **44**, 70–80.
- Baumgartner, S., Littleton, J. T., Broadie, K., Bhat, M. A., Harbecke, R., Lengyel, J. A., Chiquet-Ehrismann, R., Prokop, A. and Bellen, H. J. (1996). A *Drosophila* neurexin is required for septate junction and blood-nerve barrier formation and function. *Cell* **87**, 1059–1068.
- Bazinet, C. (2000.7.10). FlyBase error report for CG7075 on Mon Jul 10 03:52:53 2000.
- Bezzi, P., Gundersen, V., Galbete, J. L., Seifert, G., Steinhauser, C., Pilati, E. and Volterra, A. (2004). Astrocytes contain a vesicular compartment that is competent for regulated exocytosis of glutamate. *Nat. Neurosci.* **7**, 613–620.
- Borden, L. A. (1996). GABA transporter heterogeneity: pharmacology and cellular localization. *Neurochem. Int.* **29**, 335–356.
- Borycz, J., Borycz, J. A., Loubani, M. and Meinertzhagen, I. A. (2002). *tan* and *ebony* genes regulate a novel pathway for transmitter metabolism at fly photoreceptor terminals. *J. Neurosci.* **22**, 10549–10557.
- Boudko, D. Y., Kohn, A. B., Meleshkevitch, E. A., Dasher, M. K., Seron, T. J., Stevens, B. R. and Harvey, W. R. (2005). Ancestry and progeny of nutrient amino acid transporters. *Proc. Natl. Acad. Sci. USA* **102**, 1360–1365.
- Broer, A., Klingel, K., Kowalczyk, S., Rasko, J. E., Cavanaugh, J. and Broer, S. (2004). Molecular cloning of mouse amino acid transport system

- B0, a neutral amino acid transporter related to Hartnup disorder. *J. Biol. Chem.* **279**, 24467-24476.
- Buchner, E., Bader, R., Buchner, S., Cox, J., Emson, P. C., Flory, E., Heizmann, C. W., Hemm, S., Hofbauer, A. and Oertel, W. H.** (1988). Cell-specific immuno-probes for the brain of normal and mutant *Drosophila melanogaster*. I. Wildtype visual system. *Cell Tissue Res.* **253**, 357-370.
- Budnik, V. and White, K.** (1988). Catecholamine-containing neurons in *Drosophila melanogaster*: distribution and development. *J. Comp. Neurol.* **268**, 400-413.
- Burg, M. G., Sarthy, P. V., Koliantz, G. and Pak, W. L.** (1993). Genetic and molecular identification of a *Drosophila* histidine decarboxylase gene required in photoreceptor transmitter synthesis. *EMBO J.* **12**, 911-919.
- Burg, M. G., Geng, C., Guan, Y., Koliantz, G. and Pak, W. L.** (1996). *Drosophila rosA* gene, which when mutant causes aberrant photoreceptor oscillation, encodes a novel neurotransmitter transporter homologue. *J. Neurogenet.* **11**, 59-79.
- Campos-Ortega, J. A.** (1974). Autoradiographic localization of <sup>3</sup>H-gamma-aminobutyric acid uptake in the lamina ganglionaris of *Musca* and *Drosophila*. *Z. Zellforsch. Mikrosk. Anat.* **147**, 415-431.
- Castagna, M., Shayakul, C., Trofii, D., Sacchi, V. F., Harvey, W. R. and Hediger, M. A.** (1998). Cloning and characterization of a potassium-coupled amino acid transporter. *Proc. Natl. Acad. Sci. USA* **95**, 5395-5400.
- Chang, H. Y., Grygoruk, A., Brooks, E. S., Ackerson, L. C., Maidment, N. T., Bainton, R. J. and Krantz, D. E.** (2006). Overexpression of the *Drosophila* vesicular monoamine transporter increases motor activity and courtship but decreases the behavioral response to cocaine. *Mol. Psychiatry* **11**, 99-113.
- Chiu, C., Ross, L. S., Cohen, B. N., Lester, H. A. and Gill, S. S.** (2000). The transporter-like protein inebriated mediates hyperosmotic stimuli through intracellular signaling. *J. Exp. Biol.* **203**, 3531-3546.
- Corey, J. L., Quick, M. W., Davidson, N., Lester, H. A. and Guastella, J.** (1994). A cocaine-sensitive *Drosophila* serotonin transporter: cloning, expression, and electrophysiological characterization. *Proc. Natl. Acad. Sci. USA* **91**, 1188-1192.
- Demchshyn, L. L., Pristupa, Z. B., Sugamori, K. S., Barker, E. L., Blakely, R. D., Wolfgang, W. J., Forte, M. A. and Niznik, H. B.** (1994). Cloning, expression, and localization of a chloride-facilitated, cocaine-sensitive serotonin transporter from *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. USA* **91**, 5158-5162.
- Erickson, J. D., Eiden, L. E., Schafer, M. K. and Weihe, E.** (1995). Reserpine- and tetrabenazine-sensitive transport of (3)H-histamine by the neuronal isoform of the vesicular monoamine transporter. *J. Mol. Neurosci.* **6**, 277-287.
- Eule, E., Tix, S. and Fischbach, K. F.** (1995). Atlas of glial cells in the adult optic ganglion.
- Feldman, D. H., Harvey, W. R. and Stevens, B. R.** (2000). A novel electrogenic amino acid transporter is activated by K<sup>+</sup> or Na<sup>+</sup>, is alkaline pH-dependent, and is Cl<sup>-</sup> independent. *J. Biol. Chem.* **275**, 24518-24526.
- Gao, X., McLean, H., Caveney, S. and Donly, C.** (1999). Molecular cloning and functional characterization of a GABA transporter from the CNS of the cabbage looper, *Trichoplusia ni*. *Insect Biochem. Mol. Biol.* **29**, 609-623.
- Giros, B., Jaber, M., Jones, S. R., Wightman, R. M. and Caron, M. G.** (1996). Hyperlocomotion and indifference to cocaine and amphetamine in mice lacking the dopamine transporter. *Nature* **379**, 606-612.
- Gomez, J., Hulsmann, S., Ohno, K., Eulenburg, V., Szoke, K., Richter, D. and Betz, H.** (2003). Inactivation of the glycine transporter 1 gene discloses vital role of glial glycine uptake in glycinergic inhibition. *Neuron* **40**, 785-796.
- Greer, C. L., Grygoruk, A., Patton, D. E., Ley, B., Romero-Calderon, R., Chang, H. Y., Houshyar, R., Bainton, R. J., Diantonio, A. and Krantz, D. E.** (2005). A splice variant of the *Drosophila* vesicular monoamine transporter contains a conserved trafficking domain and functions in the storage of dopamine, serotonin, and octopamine. *J. Neurobiol.* **64**, 239-258.
- Harvey, W. R. and Wiczorek, H.** (1997). Animal plasma membrane energization by chemiosmotic H<sup>+</sup> V-ATPases. *J. Exp. Biol.* **200**, 203-216.
- Heller-Stilb, B., van Roeyen, C., Rascher, K., Hartwig, H. G., Huth, A., Seeliger, M. W., Warskulat, U. and Haussinger, D.** (2002). Disruption of the taurine transporter gene (*taut*) leads to retinal degeneration in mice. *FASEB J.* **16**, 231-233.
- Hoffman, B. J., Hansson, S. R., Mezey, E. and Palkovits, M.** (1998). Localization and dynamic regulation of biogenic amine transporters in the mammalian central nervous system. *Front. Neuroendocrinol.* **19**, 187-231.
- Holt, R. A., Subramanian, G. M., Halpern, A., Sutton, G. G., Charlab, R., Nusskern, D. R., Wincker, P., Clark, A. G., Ribeiro, J. M., Wides, R. et al.** (2002). The genome sequence of the malaria mosquito *Anopheles gambiae*. *Science* **298**, 129-149.
- Homburg, U., Kingan, T. G. and Hildebrand, J. G.** (1987). Immunocytochemistry of GABA in the brain and suboesophageal ganglion of *Manduca sexta*. *Cell Tissue Res.* **248**, 1-24.
- Huang, X., Huang, Y., Chinnappan, R., Bocchini, C., Gustin, M. C. and Stern, M.** (2002). The *Drosophila* inebriated-encoded neurotransmitter/osmolyte transporter: dual roles in the control of neuronal excitability and the osmotic stress response. *Genetics* **160**, 561-569.
- Johnson, K., Knust, E. and Skaer, H.** (1999). bloated tubules (blot) encodes a *Drosophila* member of the neurotransmitter transporter family required for organisation of the apical cytocortex. *Dev. Biol.* **212**, 440-454.
- King, D. G.** (1988). Cellular organization and peritrophic membrane formation in the cardia (proventriculus) of *Drosophila melanogaster*. *J. Morphol.* **196**, 253-282.
- Kitamoto, T., Wang, W. and Salvaterra, P. M.** (1998). Structure and organization of the *Drosophila* cholinergic locus. *J. Biol. Chem.* **273**, 2706-2713.
- Kleta, R., Romeo, E., Ristic, Z., Ohura, T., Stuart, C., Arcos-Burgos, M., Dave, M. H., Wagner, C. A., Camargo, S. R., Inoue, S. et al.** (2004). Mutations in SLC6A19, encoding B0AT1, cause Hartnup disorder. *Nat. Genet.* **36**, 999-1002.
- Kosaka, T. and Ikeda, K.** (1983). Reversible blockage of membrane retrieval and endocytosis in the garland cell of the temperature-sensitive mutant of *Drosophila melanogaster*, shibirets1. *J. Cell Biol.* **97**, 499-507.
- Lill, H. and Nelson, N.** (1998). Homologies and family relationships among Na<sup>+</sup>/Cl<sup>-</sup> neurotransmitter transporters. *Meth. Enzymol.* **296**, 425-436.
- Mahajan-Miklos, S. and Cooley, L.** (1994). Intercellular cytoplasm transport during *Drosophila* oogenesis. *Dev. Biol.* **165**, 336-351.
- Malandro, M. S. and Kilberg, M. S.** (1996). Molecular biology of mammalian amino acid transporters. *Annu. Rev. Biochem.* **65**, 305-336.
- Malutan, T., McLean, H., Caveney, S. and Donly, C.** (2002). A high-affinity octopamine transporter cloned from the central nervous system of cabbage looper *Trichoplusia ni*. *Insect Biochem. Mol. Biol.* **32**, 343-357.
- Mbungu, D., Ross, L. S. and Gill, S. S.** (1995). Cloning, functional expression, and pharmacology of a GABA transporter from *Manduca sexta*. *Arch. Biochem. Biophys.* **318**, 489-497.
- Melikian, H. E.** (2004). Neurotransmitter transporter trafficking: endocytosis, recycling, and regulation. *Pharmacol. Ther.* **104**, 17-27.
- Melzig, J., Burg, M., Gruhn, M., Pak, W. L. and Buchner, E.** (1998). Selective histamine uptake rescues photo- and mechanoreceptor function of histidine decarboxylase-deficient *Drosophila* mutant. *J. Neurosci.* **18**, 7160-7166.
- Miller, A.** (1950). *Biology of Drosophila*. New York: John Wiley & Sons.
- Monastirioti, M.** (1999). Biogenic amine systems in the fruit fly *Drosophila melanogaster*. *Microsc. Res. Tech.* **45**, 106-121.
- Monastirioti, M., Gorczyca, M., Rapus, J., Eckert, M., White, K. and Budnik, V.** (1995). Octopamine immunoreactivity in the fruit fly *Drosophila melanogaster*. *J. Comp. Neurol.* **356**, 275-287.
- Mullen, G. P., Mathews, E. A., Saxena, P., Fields, S. D., McManus, J. R., Moulder, G., Barstead, R. J., Quick, M. W. and Rand, J. B.** (2006). The *Caenorhabditis elegans* snf-11 gene encodes a sodium-dependent GABA transporter required for clearance of synaptic GABA. *Mol. Biol. Cell* **17**, 3021-3030.
- Nash, S. R., Giros, B., Kingsmore, S. F., Kim, K. M., el-Mestikawy, S., Dong, Q., Fumagalli, F., Seldin, M. F. and Caron, M. G.** (1998). Cloning, gene structure and genomic localization of an orphan transporter from mouse kidney with six alternatively-spliced isoforms. *Recept. Channels* **6**, 113-128.
- Nassel, D. R. and Elekes, K.** (1992). Aminergic neurons in the brain of blowflies and *Drosophila*: dopamine- and tyrosine hydroxylase-immunoreactive neurons and their relationship with putative histaminergic neurons. *Cell Tissue Res.* **267**, 147-167.
- Nelson, N.** (1998). The family of Na<sup>+</sup>/Cl<sup>-</sup> neurotransmitter transporters. *J. Neurochem.* **71**, 1785-1803.
- Nowicki, J. L. and Burke, A. C.** (2000). *Hox* genes and morphological identity: axial versus lateral patterning in the vertebrate mesoderm. *Development* **127**, 4265-4275.
- Parisi, M., Nuttall, R., Edwards, P., Minor, J., Naiman, D., Lu, J., Doctolero, M., Vainer, M., Chan, C., Malley, J. et al.** (2004). A survey of ovary-, testis-, and soma-biased gene expression in *Drosophila melanogaster* adults. *Genome Biol.* **5**, R40.
- Peter, D., Liu, Y., Sternini, C., de Giorgio, R., Brecha, N. and Edwards, R. H.** (1995). Differential expression of two vesicular monoamine transporters. *J. Neurosci.* **15**, 6179-6188.

- Pollack, I. and Hofbauer, A.** (1991). Histamine-like immunoreactivity in the visual system and brain of *Drosophila melanogaster*. *Cell Tissue Res.* **266**, 391-398.
- Porzgen, P., Park, S. K., Hirsh, J., Sonders, M. S. and Amara, S. G.** (2001). The antidepressant-sensitive dopamine transporter in *Drosophila melanogaster*: a primordial carrier for catecholamines. *Mol. Pharmacol.* **59**, 83-95.
- Quan, H., Athirakul, K., Wetsel, W. C., Torres, G. E., Stevens, R., Chen, Y. T., Coffman, T. M. and Caron, M. G.** (2004). Hypertension and impaired glycine handling in mice lacking the orphan transporter XT2. *Mol. Cell Biol.* **24**, 4166-4173.
- Richardt, A., Rybak, J., Stortkuhl, K. F., Meinertzhagen, I. A. and Hovemann, B. T.** (2002). Ebony protein in the *Drosophila* nervous system: optic neuropile expression in glial cells. *J. Comp. Neurol.* **452**, 93-102.
- Saint Marie, R. L. and Carlson, S. D.** (1983). The fine structure of neuroglia in the lamina ganglionaris of the housefly, *Musca domestica* L. *J. Neurocytol.* **12**, 213-241.
- Sandhu, S. K., Ross, L. S. and Gill, S. S.** (2002). Molecular cloning and functional expression of a proline transporter from *Manduca sexta*. *Insect Biochem. Mol. Biol.* **32**, 1391-1400.
- Seow, H. F., Broer, S., Broer, A., Bailey, C. G., Potter, S. J., Cavanaugh, J. A. and Rasko, J. E.** (2004). Hartnup disorder is caused by mutations in the gene encoding the neutral amino acid transporter SLC6A19. *Nat. Genet.* **36**, 1003-1007.
- Sloan, J. L. and Mager, S.** (1999). Cloning and functional expression of a human Na<sup>+</sup> and Cl<sup>-</sup>-dependent neutral and cationic amino acid transporter B(0+). *J. Biol. Chem.* **274**, 23740-23745.
- Soehnge, H., Huang, X., Becker, M., Whitley, P., Conover, D. and Stern, M.** (1996). A neurotransmitter transporter encoded by the *Drosophila inebriated* gene. *Proc. Natl. Acad. Sci. USA* **93**, 13262-13267.
- Soragna, A., Mari, S. A., Pisani, R., Peres, A., Castagna, M., Sacchi, V. F. and Bossi, E.** (2004). Structural domains involved in substrate selectivity in two neutral amino acid transporters. *Am. J. Physiol.* **287**, C754-C761.
- Stuart, A. E., Morgan, J. R., Mekeel, H. E., Kempter, E. and Callaway, J. C.** (1996). Selective, activity-dependent uptake of histamine into an arthropod photoreceptor. *J. Neurosci.* **16**, 3178-3188.
- Stuart, A. E., Gebhardt, K. A., Vogel, S. N. and Rodriguez, O.** (2002). Does the neurotransmitter transporter underlie adaptation at a histaminergic photoreceptor synapse? *Vis. Neurosci.* **19**, 307-319.
- Takanaga, H., Mackenzie, B., Suzuki, Y. and Hediger, M. A.** (2005). Identification of mammalian proline transporter SIT1 (SLC6A20) with characteristics of classical system imino. *J. Biol. Chem.* **280**, 8974-8984.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. and Higgins, D. G.** (1997). The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.* **25**, 4876-4882.
- Tix, S., Eule, E., Fischbach, K. F. and Benzer, S.** (1997). Glia in the chiasm and medulla of the *Drosophila melanogaster* optic lobes. *Cell Tissue Res.* **289**, 397-409.
- Tomancak, P., Beaton, A., Weiszmam, R., Kwan, E., Shu, S., Lewis, S. E., Richards, S., Ashburner, M., Hartenstein, V., Celniker, S. E. et al.** (2002). Systematic determination of patterns of gene expression during *Drosophila* embryogenesis. *Genome Biol.* **3**, RESEARCH0088.
- Tsai, G., Ralph-Williams, R. J., Martina, M., Bergeron, R., Berger-Sweeney, J., Dunham, K. S., Jiang, Z., Caine, S. B. and Coyle, J. T.** (2004). Gene knockout of glycine transporter 1, characterization of the behavioral phenotype. *Proc. Natl. Acad. Sci. USA* **101**, 8485-8490.
- Umesh, A. and Gill, S. S.** (2002). Immunocytochemical localization of a *Manduca sexta* gamma-aminobutyric acid transporter. *J. Comp. Neurol.* **448**, 388-398.
- Valles, A. M. and White, K.** (1988). Serotonin-containing neurons in *Drosophila melanogaster*: development and distribution. *J. Comp. Neurol.* **268**, 414-428.
- Wang, J., Kean, L., Yang, J., Allan, A. K., Davies, S. A., Herzyk, P. and Dow, J. A.** (2004). Function-informed transcriptome analysis of *Drosophila* renal tubule. *Genome Biol.* **5**, R69.
- Xiong, W. C., Okano, H., Patel, N. H., Blendy, J. A. and Montell, C.** (1994). repo encodes a glial-specific homeo domain protein required in the *Drosophila* nervous system. *Genes Dev.* **8**, 981-994.
- Yamashita, A., Singh, S. K., Kawate, T., Jin, Y. and Gouaux, E.** (2005). Crystal structure of a bacterial homologue of Na<sup>+</sup>/Cl<sup>-</sup> dependent neurotransmitter transporters. *Nature* **437**, 215-223.
- Yellman, C., Tao, H., He, B. and Hirsh, J.** (1997). Conserved and sexually dimorphic behavioral responses to biogenic amines in decapitated *Drosophila*. *Proc. Natl. Acad. Sci. USA* **94**, 4131-4136.
- Zomot, E. and Kanner, B. I.** (2003). The interaction of the gamma-aminobutyric acid transporter GAT-1 with the neurotransmitter is selectively impaired by sulfhydryl modification of a conformationally sensitive cysteine residue engineered into extracellular loop IV. *J. Biol. Chem.* **278**, 42950-42958.

```

DmCG8291 : MSSKEQQAAGRDFKRNSNAYS SL PPTGTGAGCSGAALGSGTGTGRKRSKQHLEHQFGLSNLSSESIRQACAYYDDE : 80
          *          20          *          40          *          60          *          80
AmINE : -----MNATDSNSGVNNGDSNNTIAAKTNAHGNSVRLQSVIGKQSSC : 42
DmCG8291 : LSDEDLIEIQTPQAFHQTKQLQLQPI SFRLGDELGERSAFCSGQEMQVPLSTPIKQAAETDEALCVLSELDAILD : 160
          *          100          *          120          *          140          *          160
AmINE : VRKMSTMP EEEDASDDMDPDVIRIGNQHYYQHHDADSDSPVEERGRLLSITDSTITAHHRKIAITPSSSSNLHHCNGLK : 122
MsINE : -----MNKVE : 5
DmCG8291 : VHDVSQLNGTCCSSGVSNGSDDDKVEDYLDMLDNLNLEEMDNALNREDSLI IIDGHTSLKREPRTRTLP LSRKKKSSKKKS : 240
          *          180          *          200          *          220          *          240
          *          260          *          280          *          300          *          320
CeSERT : -----MLRWH : 5
TnOctT : -----MATLSGTG : 8
DmCG5549 : -----MGQSSKSGSAGVG : 13
HsGLYT2 : -----MDCSAPKEMNKL PANSPEAAA : 21
HsATB0+ : -----MDKCLKPS : 8
HsGLYT1 : -----MSGGDTRAAIARPRMAAAHGVPV : 22
AgCP4566 : -----Q : 1
AmINE : KAI PRNDSGNSIHERAYSRLQQRSGDQVQPEARLLNRDNPQGI LRSSARGIDSHSGGRYRCSSVRSVKSNTMDEHHH : 202
MsINE : SSTEAAAAPSVAIHVEQHD-----DEQDKENSKLLSAHSPAPSITP-----SGQMRKVKFSFSDTHKIRDVTTASG : 69
DmCG8291 : EDQEAAQGFQREHQRLKRTFSCSLRPTSQIASSSGSLETSTPEVMDVWRRQSMRRALEQEDTKATVAMEREEDDIP TLLV : 320
DmBLOT : -----MSSSYKPRSDEALAEQFSLARAGSGTKPPATPTYRQI : 38
CeSNF12 : -----MNGEWKSALRLQIEALAKRNLHRKSSVDEIKKRT : 35
HsNTT5 : -----MKTEAQ : 6
          *          340          *          360          *          380          *          400
DmSERT : -----MDRSGSSDFAGAAATGRSNPAWSDDKESPNNEDDSNEDDGHHTPAKVTDPLAPK--LANNE : 62
AgCP12911 : -----E : 1
MsSERT : -----MPPSDAPPAPTAPPDLPAT--TAQKS : 25
HsSERT : -----METTPLNSQKLSACEDGEDCQENGLVQKVVPTPGDKVESGQISNGYSAVPSGAGDD--TRHSI : 63
CeSERT : SVRRKQHQQQLAELSSGAASMLSAPESSRRVSRMSVKAPTASEYMP LSVADKPLTLTVSTSHSIDNEPIAALGLLPTK : 85
DmDAT : -----MSPTGHISKSK : 11
AgCP3649 : ----- : -
TnDAT : -----MALK : 4
CeDAT1 : -----MQLVPTDDPDEKIGRTSNGMQNAT : 24
HsNET : -----MLLAR-----MNPQVQPENNGADTGPQPLRARKTAE LLVKERNGVQC-- : 44
HsDAT : -----MSK-----SKCSVGLMSVVPAKEPNVAVGPKVEVELLLVKEQNGVQLTS : 44
TnOctT : VVHTHTAPSYEEQRANPALLSRGASGTPGGRSVRDDGYCSASSTPRAF DNKSTKGSVVTLSICYKPKIQIEECPYSEPN : 88
DmCG5549 : G-----VGGGGGVVCPSTAIS : 29
AgCP10735 : ----- : -
HsGLYT2 : AQGHDPGCPARTSPEQELPAAAAAPPPRVRPASTGAQTFQSADARACEAERPVG SCKLSSPRAQAASAALRDLREAQ : 101
HsATB0+ : -----FPKCREKE : 16
HsPROT : -----MKKLQGAH---LRKPVTPD : 16
HsGLYT1 : APSSPEQVTL LFPVQRFFLPFSGATPSTSLAES-----VLKVVHGAY---NSGLLPQ : 72
DmCG7075 : -----MANNQPPT : 8
AgEbiP6595 : ----- : -
MsPROT : ----- : -
DmINE : -----MPNRQDYDAQSSK-----HSEQSFFRFNR-VAKVPLGVDDDDSY : 38
AgCP4566 : QPQTTIARSGSYIFSDVG--GTPRLFSDATSIRSLASIGMGSTDGRRMVIRRV PNSPNELLTMIN-PPTPDETYENESY : 78
AmINE : QYHPPAASWASIVFSDNGQTHVRTL PDSVSRSLASIGMGSSNGRKLTI RRVLTSPSELLNMVHPPPSFEDDL SVCTSF : 282
MsINE : AASARSLRPIEIVNTYPEGSESGTNNYGAPSVRSLASIGMGCTDGRKMVIRRV PTPTELFHLVR-PPTPPDEDSASHES : 148
TnGAT : -----MDTKNDG-----RSDDIELS-----AQGTSNK : 22
MsGAT : -----METKNDS-----RSDDIELS-----AQGSGNK : 22
AgCP8499 : ----- : -
DmCG1732 : -----MYTNSASDGGGDVCKHESIEMSKELGHT--TSLQSSSPT : 39
CeSNF11 : -----MCHYNLIHPAH-----WFFFLQFFR : 20
HsGAT1 : -----MATNGSKVADG---QISTEVSEAP-----VANDKPK : 28
HsGAT3 : -----MTAEKALPLGNGKAAEEARESEAPGGGCSGGAAPA-----RHPR : 40
HsGAT2 : -----MDSRVS---GTTSNGETKPV-----YPV : 20
HsBGT1 : -----MDGKVAVQERGP PPAVSWVPPE-----GK : 24
HsTAUT : -----MATKEKLQCLKDFHKDLLKPS-----GKSPG : 27
HsCT1 : -----MAKSAENGIYSVSGDEKKGPLIAPGPDGAPAK-----GDGPV : 38
CeSNF3 : -----MGTSEHVPLPT-----DEAKA : 17
DmCG5226 : -----MAATKI IDAP---RNGHEMAPLNTRARGDGTGVTIVLTAP : 38
AgCP7001 : -----MTTKVDGVGPRNNGHEMAPLNTRARGDGTGVTI MLSP : 40
HsNTT7-3 : -----MPKNSKVVKRELD--DDVTE SVKDLLSNEDAADDAFKTSELIVDQGE : 45
HsNTT4 : -----MPKNSKVTQREHSSEHVTE SVADLLALEEPVD--YKQSVLNVAGEA : 44
HsXtrp2 : -----MAHAPEPDPAAAC----- : 12
RnROSIT : ----- : -
HsBOAT1 : -----MVRLVLPNPGLDARIPSLAELETIEQ----- : 26
HsXT3L : ----- : -
DmCG10804 : -----MSDAFDESSELEQDPPFPFGSEASTSKAKAELVAIATA : 37
AgCP7501 : ----- : -
DmCG1698 : -----METTINNNDNGYMNQAVVGLNASTISIPG-----AYKPNESNGKKT LSESGSAPP PGHGD : 60
DmCG4476 : -----MKDLKT-----EILVSG-----GLPKLETS---RCAGSGEALLKPSLLPM : 37
DmCG15279 : -----MDT--TQRN---HQNLAFVGDGGRSTAST-----VEISTN-----SPALRN--SD : 39
AgCP10412 : -----MEG--RDNNGFIGDNSPSIAGYRWTTPAAPNGVHVTHGGLALTDVELAVGKTVPRPTPATGED--AG : 65
DmCG8850 : -----MQQAQGSREHGP GSNDDG-----ISTVIYSAEGEEL : 30
DmCG15088 : -----MTQLEISKK----- : 9
AgCP10401 : -----MATSNPAFENDEPVLTSDRTSHTATKPS-----LAAVTAKSSAKVNQFQQPPEQTGGNT : 55
AgCP10503 : ----- : -
AgCP10610 : ----- : -
DmCG3252 : -----MELKGVQPS-----NGSSNGSGNGATNAA : 24
AeAAT : -----MPEIATISYPE-----SKKNDEANS SHGNGNGVQLN : 32
MsKAAT1 : -----MNDGQVNGGFESSEPKMEP-----KRSSQISLPPANLKA TMD : 37
MsCAATCH1 : -----MNDGQVNGGFESSEPKMEP-----KRSSQISLPPANKAALD : 37
BcProT : ----- : -

```

```

HinSnfT : ----- : -
StTnaT : ----- : -
CeSNF2 : -----MPTPVSTKKTIA PSTMKTIVAPPTMRTTMAPTMTKTIVAPPTMKTTRSPSTMKT : 55
CeSNF7 : -----MAQSNALTTTTGSSAPSVIPPTAPTASVTKKAP----TKGP--VQPQ : 42
CeSNF4 : -----MSTSNPDNSTDSPDVSKTGDYKSGTQTNTDLSLETKLSDDGPKKTVDLSLIPT : 55
CeSNF5 : -----MADSGSNEEAMKRQAPSVKFDSPKPDQVVSQQSNQLSS----QKSIQQSTQSK : 51
CeSNF9 : -----MTSP-----TQRRSPS---DSSAAEDQRIATTIVRDSGS---SKEIEE---SQ : 39
DmCG13794 : -----MIYETS : 6
DmCG13795 : -----MVYETS : 6
DmCG13793 : ----- : -
DmCG33296 : ----- : -
DmCG8291 : ELP PRRDAEMRRCF SQGDCQASVAPT VGSQMLTEAHIFDNL LQT NARASSEEP RPRQYGRRL EGPPTVVRPL LLAQSRPQ : 400
AgCP9955 : ----- : -
DmBLOT : SGAMATLPRHDNHNHNDNSSRIRSDSTATSRVCYIGGAPTGGGNGNEQISVTLDRRQPQRMSWLNMRRAKANETDLPT : 118
AgCP2093 : ----- : -
CeSNF6 : -----MSVSNNDPEQRNGRGMASGNNVMSLYPPFIKQDAKLPDYTREGD : 45
CeSNF12 : VDMDKRIVKLRELVGSSANDAALFYLECVCHADETERLLNRGSSVGEKKKWKVKRKTSSSVAPPLSRTISSLPVSSPD : 115
AaLeuT : ----- : -
HsNFT5 : PSTSLLANTSWTGTVISDSVPGSQTWEDKGSLTRASATSWTSEAQVSAARVAEAQARTSQPKQISVLEALTASALNQKPTH : 86
DmCG13796 : -----MKGICGELQNLNMLLGDCEHSEHGTLPDGFIGEPVETESGGGMGSLALARRIRSRQVHSMAVRTGS : 69
HspSnfT : ----- : -
CeSNF1 : -----MWFEESSWSN : 10
CeSNF8 : -----MKPK : 4

          *          420          *          440          *          460          *          480
DmSERT : RILVSV----- : 69
AgCP12911 : RSLVVAL----- : 8
MsSERT : RSVVSL----- : 32
HsSERT : PATTTTLVAE----- : 73
CeSERT : EGRVAALRR----- : 94
DmDAT : TPTPHDNDNN----- : 21
AgCP3649 : ----- : -
TnDAT : TPTPG----- : 9
CeDAT1 : LPIDGPVNTE----- : 34
HsNET : --LLAPRDG----- : 51
HsDAT : STLTNPRQSP----- : 54
TnOctT : KRLRTNSIKT----- : 98
DmCG5549 : GCASGT-----ALIQLDATGPGAGG----- : 49
AgCP10735 : ----- : -
HsGLYT2 : GAQASPPPGSSGPNALHCKI PSLRGPEGD---ANVSVGKGT LERNNTPVVGVWNMSQSTVVLGTDGITSVLPGSVATV : 177
HsATB0+ : KVSASS-----ENFHVGE----- : 29
HsPROT : LLMTPS-----DQGDVLLDV----- : 31
HsGLYT1 : LMAQHSLAMA-----QNGAVPSEAT----- : 92
DmCG7075 : TNATRDKKRI----- : 18
AgEbiP6595 : ----- : -
MsPROT : ----- : -
DmIne : LDMSDET----- : 45
AgCP4566 : GGMSDDSD----- : 86
AmINE : DDAEDDAQD----- : 291
MsIne : DCEEEED----- : 156
TnGAT : TSEVATKS----- : 30
MsGAT : PSDVAVKS----- : 30
AgCP8499 : ----- : -
DmCG1732 : TTDTSNKQQL----- : 49
CeSNF11 : VIMVEATS----- : 28
HsGAT1 : TLVVVKVQK----- : 36
HsGAT3 : VKRDK----- : 45
HsGAT2 : MEKKEED----- : 27
HsBGT1 : LDQEDED----- : 31
HsTAUT : TRPEDEAEG----- : 36
HsCT1 : GLGTPGGRL----- : 47
CeSNF3 : ELEQSQHS----- : 25
DmCG5226 : QRNSVQSV----- : 47
AgCP7001 : DRSSIGSREGA----- : 51
HsNFT7-3 : E-KDTPDV----- : 51
HsNFT4 : GGKQKAV----- : 51
HsXtrp2 : ----- : -
RnROSIT : ----- : -
HsBOAT1 : ----- : -
HsXT3L : ----- : -
DmCG10804 : SADQQDI----- : 44
AgCP7501 : ----- : -
DmCG1698 : SPSGV----- : 65
DmCG4476 : ALESS----- : 42
DmCG15279 : DQEAA----- : 44
AgCP10412 : RVPTT----- : 70
DmCG8850 : TINCE----- : 35
DmCG15088 : ----- : -
AgCP10401 : SRTSALPEMNL----- : 66
AgCP10503 : ----- : -
AgCP10610 : ----- : -
DmCG3252 : STEKT----- : 29
AeAAT : AS----- : 34
MsKAAT1 : NIDDM----- : 42
MsCAATCH1 : NIDDT----- : 42
BcProT : ----- : -
HinSnfT : ----- : -
StTnaT : ----- : -
CeSNF2 : KEFSMTQTTKTVQSTITNPVSFLDS-----EVTAKYVVKDGGGAAR : 95

```



```

CeSNF7 : KKP-----TTVDN-----GR : 52
CeSNF4 : VSQNMKDKD--NTTLD-----DRKFDGTGVGIA : 82
CeSNF5 : IDPSRIDTKN--TIVTT-----TR : 68
CeSNF9 : ADPC : 43
DmCG13794 : YDSGHKPF : 15
DmCG13795 : YDTGHKPF : 15
DmCG13793 : : -
DmCG33296 : : -
DmCG8291 : SAPTRVQMREPQLQDTPHTPIMSTCSELSSARSSRMPSPVSLPSDSSSSGSSSAEHDQEPDPVQTTTMCASSTTTPLEPL : 480
AgCP9955 : : -
DmBLOT : VTPSPQTTTRSCISTVSSVVDSGGGRTTATSGRISSSGIVTSLGSSNNTLSEIQGGYHDMDDAVSKNFISASAHNITTAS : 198
AgCP2093 : : -
CeSNF6 : IEYPFEITG : 55
CeSNF12 : VIQTTIDVSALEDQTP-----QRPHWWDQFQLYR : 143
AaLeuT : : -
HsNNT5 : EKVMTEKKES : 97
DmCG13796 : AYDTLERP : 77
HspSnfT : : -
CeSNF1 : LRLTD : 15
CeSNF8 : KEVR : 8

```

```

* 500 * 520 * 540 * 560
DmSERT : -----T-ERT ET GQ-KAE AV GFA D G ICYQN GA C F IFG FY LA FHR : 139
AgCP12911 : -----TGERR ET SQ-KAE AV GFA D G ICYQN GA C M LFG FY LA FHR : 79
MsSERT : -----TPARQ ET AK-KAE AV GFA D G ICYQN GA C M LFG FF LA YHR : 103
HsSERT : -----LHQGE ET GK-KVD SV GYA D G ICYQN GA T MAIFG FY LA YHR : 144
CeSERT : -----RSSMV DK AT-KME AV GYA D G SVCYKH GA F M MIG FY LV FHR : 165
DmDAT : -----SISDE ET SG-KVD SV GFA D A LCYKN GA G M VVG FY LA HNR : 92
AgCP3649 : -----EE ET SG-KVD SV GFA D A LCYKN GA G M VVG FY LA FNR : 68
TnDAT : -----VGE ET GK-KVD SV GFA D A LCYKN GA C M VVG FY LA FHR : 78
CeDAT1 : -----PKDPA EQ SG-KLD SV GFA D G LCFKN GV S M LLT FY LC YHR : 105
HsNET : -----DAQD ET GK-KID SV GFA D A LCYKN GA T F IIA FY LA YNR : 121
HsDAT : -----VEAQD ET GK-KID SV GFA D A LCYKN GA L F VIA FY LA FNR : 125
TnOctT : -----EADDG ET GT-GAD SI GFA D A LCYRN GA T M VFPA FY LI YNR : 169
DmCG5549 : -----AGESEHE GT TG-RFD SL GYS G G LCYNN GA T M VIA MF LSF YAA : 122
AgCP10735 : -----K GS TG-RYD SL GYS G G LCYSN GA T M VIA MF LS A VVG : 67
HsGLYT2 : ATQEDEQGDENKA GN SS-KLD SM GYA G G LAFQN GA L M ALA FF VS FAS : 256
HsATB0+ : -----NDENQD GN SK-KSD SM GYA G G LTYSN GA A M ALA FF CS FAS : 101
HsPROT : -----DFAAH GN TG-KLD SC GYC G G RAYTN GA F M AIC FF LS FSS : 102
HsGLYT1 : -----KRDQNLK GN GN-QIE TS GYA G G LCYRN GA F F M IPC FF LSF FAS : 165
DmCG7075 : -----ERDEN GQ KS-KSE SL GYA G G LCYRS AA L M IIA FY IL FSS : 89
AgEbiP6595 : -----A L M VLC FF TC FSG : 29
MsPROT : -----MSGATQDR GS-QLE SC GYA G G LCYRN GA F T IIC VY TT FAS : 71
DmIne : -----AQLKPRQOH AN-KMQ AC GYS G G MCYKS GV C I FICS LF LS YTG : 117
AgCP4566 : -----LDNLKPR QH AN-KMQ AC GYS G G LCYKS GV F I LIC LF LA YTG : 159
AmINE : -----PGDYRQSR PH AN-KMQ AC GYS G G LCYKS GV F I IVC LY LS PTR : 365
MsIne : -----AAVHLKPR PF AN-KIQ AC GYS G G LCYKS GA F I LIC LF LA YTA : 230
TnGAT : -----DLPE GS GS-KLD SV GLA G G LCYKN GA F T FLA FF LA MLT : 100
MsGAT : -----NLPE GS AS-KLD SV GLA G G LCYKN GA F T FLA FF LA MLT : 100
AgCP8499 : -----LD SV GLA G G LCYKN GA F T FLA FF LA MLT : 59
DmCG1732 : -----VKIEQLPD GS SS-KMD SV GLA G G LCYKN GA I T FLA FF LA MLT : 123
CeSNF11 : -----VAE EQ SS-WAD SC GYA G G LCYQN GA C S VFC A FI TSW LMS : 97
HsGAT1 : -----KAADLPD DT KG-RFD SC GYA G G LCGKN GA F T IFA FL CS YTS : 109
HsGAT3 : -----AVHE GH NN-KVE SVAGEI G G LCYKN GA V FFICC FF TA FTS : 115
HsGAT2 : -----GTLE GH NN-KME SVAGEI G G LCYKN GA F L F FTC FL TA YTS : 97
HsBGT1 : -----QVKD GQ TN-KME SVAGEI G G LCYKN GA F F FFFVC FF VA YTS : 101
HsTAUT : -----KPPQ EK SS-KID SVAGGF G G LCYKN GA F F FGS FF II YTS : 106
HsCT1 : -----AVPP ET TR-QMD SC GFA G G LCYKN GV V IALVG FF IS FMK : 117
CeSNF3 : -----EPPD GQ TG-KFD SM AYA G G LCYKN GS V M FFCLAA FL VT YLQ : 95
DmCG5226 : -----IPGNEPE AA SG-KMQ F SI GYS G G LCQQN GA M M ILE FL LG RMR : 120
AgCP7001 : ---GALPDGDPD AA SG-KMQ F SI GYS G G LCQQN GA M M ILE FL LG RMR : 126
HsNNT7-3 : -----EEGSEVEDE PA NS-KLQ AQ GFS G G LCQKN GA L L MVI FF LS RIR : 126
HsNNT4 : ---EEELDAED- PA NS-KLQ AQ GFS G G LCQKN GA L L III FF LA RIR : 125
HsXtrp2 : -----DLGDE PK DN-KAQ SC GFA G G LCQTY GA V A VPE FH LA RLR : 83
RnROSIT : -----IEDE PK DN-KLQ SC GFA G G LCHTH GA F A VFE FY LA RLR : 70
HsBOAT1 : -----EEASS PK DN-KAQ TC GFC G G LCQSH GA L L VLE LY FA RLR : 97
HsXT3L : -----MEKA PL AN-SLQ FAC SYA G G LCQMY GS I M IVE LY LA RMR : 70
DmCG10804 : ---VEGEKEGEE ES DS-KIM AT GYA G G LAQKN GA F M CIQ FY LA RLR : 119
AgCP7501 : -----DE ES DS-KWT AT GYA G G LAQKN GA F M LLQ FY LA RLR : 68
DmCG1698 : -----EAGQPGKK DS NN-DIE SC ALS G G TALEN GA L V ILV K YY ML FSS : 139
DmCG4476 : -----ELSPDRKA DN GS-SLE SC ALS G G TALEN GA L V FVV K YY ML FSS : 116
DmCG15279 : -----KVPE---E AT GK-GVE SC AMS G G TALDN GA L V FLI K YY MV FSS : 115
AgCP10412 : -----IPEPNAD DQ GK-GVE SC AMS G G VALEN GA I V LLV K YY MI FSS : 144
DmCG8850 : -----AESSESSGQ DQ SR-GVE FSC ALS G G IALEN GA V V LLI R YY VI FSS : 109
DmCG15088 : -----SEKPAEKPAEQ GN-GLE FSC SLS G G IAFQN GT L A LVI R YY IS FTG : 83
AgCP10401 : -----MKNIHKVQ DK GK-DIE SC ALS G G TALEN GA L V LLV R YY ML S FSS : 140
AgCP10503 : -----EK TS-NIE T SC AYS GFG TALDN GA L V FLI R YY MA FCS : 66
AgCP10610 : -----TAFKN GA L V FII R YY MV FSN : 44
DmCG3252 : -----DAEKPTAE TN GN-GLE SC SVS G G TAYEN GA I V FLI K YY MI FTS : 103
AeAAT : -----QPENAAQN PE SN-KLE SC SMS G G TAYEN GA L V FVI R YY MA FTS : 108
MsKAAT1 : -----DLEAEPPE MV SN-NIE SC ATS G G IAYQN GA I V LLV K YY CV FSS : 116
MsCAATCH1 : -----DLEAEPPE MV SN-NIE SC ATS G G IAYQN GA V V LLV K YY CV FSS : 116
BcProT : -----MET QQ GT-RAG FAA GSA G G TAYEN GA F LFA LTT S LAF FA HRHR : 69
HinSnfT : ---MAHSAPKAQK ET SG-RRR AA GSA G G TTYEN GA L A LTA LF DYA HRHR : 76
StTnaT : -----MEAQ DQ KS-RSG FAT GAA G G F MAYQN GA F A LTA MI FGF HKMR : 70
CeSNF2 : KAYRDAIQVENTD QA RG--IES SS GQA G G TVAYQN TS V I CAFVFA AIH FA YAA : 173
CeSNF7 : RAYRNQVVENVND QA RG--IESF SS GQA G G TTAYKN LS A V CGILFA AIH FA YAA : 130
CeSNF4 : DILRDQAKRAAMQ GSCETPVTNSANLC GMS G G SRAYEN SV ITCALLF GYV FA YQG : 162
CeSNF5 : PTLDTPEEEEEK DG GN-SFE TS GLA G G TRAYNN SA LTCALFL AVYF FLT YQG : 147

```

```

CeSNF9 : ----- GA GN-QIE ST GMA G G TRAYNN SA VACALLF AVY FL YHR : 109
DmCG13794 : -----PDLK GK DK-PTD FACFGLA K D- VAS WFFPDM IFG L Y VIYL MV HSF FSS : 84
DmCG13795 : -----PDLN GK DK-PTD FACFGLA K D- VAS WMFFDM ILG F Y SIYL MV HSF FSS : 84
DmCG13793 : -----MV HSY FSS : 12
DmCG33296 : -----VIYL LV HSF FSS : 20
DmCG8291 : HQQLLREKCGFNSQ PH-AGSRT AL GCT G F C AVLTINF N- Q L LSVIF LW MC AKIR : 588
AgCP9955 : -----AR PH-AISRS ATTFCT G F S AVFSVHF AN- Q L FSLLF LW MV ARIR : 64
DmBLot : NAKLLPAVEDESN QTKCS-VFRG LC CLN SYA V -RELDY SA V L FLV VL IS FLG : 276
AgCP2093 : -----RSLPQSRSS-IFRG LC CLN TFA V -RELQLH SA V L LLV VL IS FLG : 68
CeSNF6 : -----VGDENRI GN SN-KSD AV GFTAG GSF LVFQH AA LCM CLAS FF MV FSS : 128
CeSNF12 : RTDLLRFKQNDER EL RT-QKD F SC GFM G GHTM AKVYQH GV F LFS IFF VF HLS YTG : 222
AaLeuT : -----MEVK EH AT-RLGL AMAGNA G G FL VQAAEN GA I AFLLV MW WA YGG : 70
HsNtT5 : -----EVLLA PF SS-KTE AQ GFS KPSC A LWLNS CS AAI IFM FLV LF MAA SMR : 168
DmCG13796 : -----YRHDKC GR AKSADF FASCTHAFSS IFSELST GILHG WLL A L G LFYS FL AF FSS : 150
HspSnfT : -----MA DV HS-RLG AA GSA G G MTSN AA V LGV LCV GLLA FV RRSR : 68
CeSNF1 : -----LVEEW DL PS-KIE SA AYLFA T FLNL KLILEN VA AAA A V GVLC TMV MS VTG : 86
CeSNF8 : -----KDALP PE KS-WYDL FSV NLC G S FLI LAKVHEYR GA A G I IML Y LY LI FHR : 79

```

```

* 580 * 600 * 620 * 640
DmSERT : C C SI KR C ALK ----- YAICLIDYI GM NT GWA Y LFA FT-----SK TS DNPWNTENCMQ- : 206
AgCP12911 : C C SI KR C ALK ----- YAICLIDYI GM NT GWA Y LFA FS-----SE TK GNPWNTENCSP- : 146
MsSERT : C C TL KR C ALK ----- YAICMIDIY GM NT GWA Y LIA LASIN---SV TS DNEWNTPLCTP- : 173
HsSERT : N C SI KR C IFK ----- YAICIIAFY AS NT AWA Y LIS FT-----DQ TS KNSWNTGNCTN- : 211
CeSERT : S C SI KR C LFR ----- YGICCCICTF AI NA AQA Y AIV LSKIW--DSE AS GNPWNTPRCSD- : 236
DmDAT : K A TC GR V LFK ----- YAVVLIAPY DF NV AWS R FFA FT-----NS TS NNIWNTPNCRPF : 160
AgCP3649 : K A TC GR V LFK ----- YAVVLIAPY DF NV AWS R FFA FT-----DS TH SNAWNTVECKPF : 136
TnDAT : K A TC GR V LFK ----- YAVVLIAPY DF NV AWA R FFA FT-----TM TN DNEWNTPACKPF : 146
CeDAT1 : K A TT GR C LFK ----- YCVILTAPY DF NV AWG H LYT FS-----FN AS NNSYNPAC--Y : 171
HsNET : E AATV K- C FFK ----- YAVILIALY GF NV AWS Y LFS FT-----LN TD GHTWNSPNCTDP : 188
HsDAT : E AAGV K- C ILK ----- FTVILISLY GF NV AWA H LFS FT-----TE IH NNSWNSPNCSDA : 192
TnOctT : Q P TL K- C LFK ----- FCAVMVAFY SF NV GWAFY LVS AR-----SE VH DNSWNTDQCWDS : 236
DmCG5549 : L P AV RRFC LFR ----- TGMILVSAI ML NL AWT F MFA FA-----PV QN EPAWSTKYCFYSY : 190
AgCP10735 : L P ALL KR A ISE ----- YGMILVSGM ML NV AWT F MVV FE-----DP RG QHEWNTDPCFYSY : 135
HsGLYT2 : Q P SV K-AI ALQ -----C IAMLIISVL AI NV CYT F LFA FV-----SV GS NNPWNTPECKDK : 323
HsATB0+ : L P SV R- L LFK ----- ITMVLISIF TI NV AYS Y MFA FQ-----SE KN S-SWSDKNCRSR : 167
HsPROT : L P AV K- S LFK ----- AAMLLVLGL AI NM AVV F LFA LT-----SD EH GNMWNTTELC-- : 166
HsGLYT1 : Q C GV R- S MFK ----- YGMVVVSTY GI NV CIAFY FFS MT-----HV AY NNPWNTHDCAG- : 231
DmCG7075 : T CTGM R- T LLK -----T IAQVVVNAVCV SV SYP RMIFYCFF-----KK ED SNSWNTDCCVTA : 156
AgEbiP6595 : T C TV K- V LLK -----A IAIVLNLICNA IA SYP L LWR LQ-----AQ ES GNAWNTPRCJLE : 96
MsPROT : A C SV N- N LFK -----A YAVIVLNVIASI SA SYP L IYH MS-----SP QS GNSWNTVNCTEI : 138
DmIne : R P GALGQ C LFK -----A LASVVVSFL ST SV GYS Y FFT FK-----TE ID NNRWNTPDCWVP : 185
AgCP4566 : R P GALGQ C LFK -----T LASVVVSFL ST SV AYA Y FFT FR-----PE TD SHRWNTPDCWIP : 227
AmINE : R P GALGQ C LFK -----A LSSVVVSFL ST HNV AYA Y FFTAFR-----AKQ SD ENSWNTLACWLP : 433
MsIne : H P GALGQ C LFK -----A LASVVVSFL ST AV AWA Y FFT FK-----TE AS SNRWNTDQCWVP : 298
TnGAT : I G GV K- A IFK ----- YAAAVMSCW NV IV AWA F FFM MR-----SD RN DNYWNTATCVNP : 167
MsGAT : I G GV K- A IFK ----- YAAAVMSCW NV IV AWA F FFM MR-----SD RN DNYWNTATCVNP : 167
AgCP8499 : I G GV K- A IFK ----- YAAAVMSCW NV IV AWA F FFM LR-----AD RT DNANWSINCVP : 126
DmCG1732 : I G GV K- A IFK ----- YAAAVMSCW NV IV AWA F FFM MR-----AD RT NNWNTVNCVSP : 190
CeSNF11 : V G GM K- C IFK ----- IAAAVMAFW NI IV SWAAT LYN FTM-----SD KN DHAWNTPNCRSE : 165
HsGAT1 : I G GV K- A MFK ----- LAAAVLSFW NI IV SWA Y LYN FT-----TT KQ DNPWNTDRCFSN : 176
HsGAT3 : E G TC RK C LFE ----- YATQVIEAH NV II AWA F LSNCF-----TE AT GHEWNTENCVE- : 182
HsGAT2 : Q G TA RK C IFE ----- YASQMVIL NV II AWA F LFS FT-----ID GG YHEWNTHECME- : 164
HsBGT1 : Q S TA RK C LFO ----- LASVVIESY NV II AWA F LFS FT-----SE TT NNFWNTHECTD- : 168
HsTAUT : E G TC EK C LFS ----- YASVVIVSL NV IV AWATY LFO FQ-----KE AH NHSWNTPHCMED : 174
HsCT1 : A S NV N- C LFK ----- YASMVIVFCYNT IM AWGFY LVK FT-----TT AT GHTWNTPDCVEI : 184
CeSNF3 : K A EM L- C LFR ----- IGVVIAFMCIA CV AWA F MIS IA-----WVF ET NNYWNTATCVTG : 162
DmCG5226 : L A GV NT H WLK ----- ISSCIVTLF AL NV TWVFF LFN FR-----YP SS P-LNGTG---FE : 184
AgCP7001 : L A GV NT H WLK ----- ISSCIVTLF AI NV TWCFY FFN FR-----YP AV PQLNGT-----D : 189
HsNtT7-3 : R S GV NY S KLG ----- FASCVVCFY AL NV GWS F FSQ FQ-----QP DQ P-LVKNAHSTFV : 193
HsNtT4 : R S GV HY C RLG ----- FSSCIVLFL GL NV GWS F FFK FQ-----YP SE P-VVRNGSVAVV : 192
HsXtrp2 : K S GV TA S YLS ----- LGCVTLSFL SL NT AWW W LLN FQ-----HP SS P-PDLN---RTGF : 148
RnROSIT : R S GV KT S YLG ----- LGCFVSFSL SL NT LWV W FLN FQ-----HP ST P-LDLN---RTGF : 135
HsBOAT1 : R S GV SS H ALK ----- LASMLTFSM GL NT SWI W LFN FQ-----EP SD P-LNEN---QTGY : 162
HsXT3L : Q S GA RT S YLS ----- VASVVVSFF SM NV NAWAFW LPH FQ-----DP SV P-LNGN---HTGY : 135
DmCG10804 : K A GV SQ S YLG ----- ISSAVVSYI AL NT AWC I LLH FE-----SP AD PTRYLKNFTYDH : 187
AgCP7501 : K A GV HE SAYLG ----- ISSAFVSYI AL NT AWC I LLH FE-----TP AE PKRLFKNFTYDI : 136
DmCG1698 : R S KV D-FS IMR ----- YQVVLATGI TT AT ALT R FVD FY-----PT SY REEWGTECLDSG : 206
DmCG4476 : R I QV D-FA LMR ----- YAQLLALGV AT AS ALT R FPD FA-----SE SF REEWGDCGCVS-- : 181
DmCG15279 : R S KV D- C AMK ----- AQQAFQVFM ST AA AIIGR FIE FR-----NP ST RAEWGIHCINSA : 182
AgCP10412 : R S KV D- A IMR ----- YQLFSVTA IT SS ALIAR MID FM-----NP AH RSEWQPCNIDSV : 211
DmCG8850 : R C RA D- A IMR ----- AYQVYSTALATT AC ALT R LVA FS-----EV TY LVEWGKSCVATG : 176
DmCG15088 : R V KA D- A LLK ----- ALGQVLATAASIT SS ALT R WLA FG-----SE SR WESWGTDCHDGN : 150
AgCP10401 : R C NV D-AS AMR ----- VGQTYSTFI MT AS AVT R LIA FG-----DP SE NEAWNATCIDSR : 207
AgCP10503 : R C KI D- A AMR ----- VQGSVAMVVAMS TP AIT R LLL FS-----SE SK DHSWSRCSIRTF : 133
AgCP10610 : R C KI D- A AMR ----- VGQTVAIFT IT AS AVT R LVA FN-----PE AK DPTWPDVCBSSR : 111
DmCG3252 : Q T KI S- V GFV ----- YQAFGTIC IS SS ALT Y LFW FQ-----SE SY RDEWNTN-CVNSR : 169
AeAAT : RSS KI E- S LFK ----- IQGLVGTTS VS AV ALT H IFA FA-----SE AT KDNWADNCVDS- : 174
MsKAAT1 : RNS KI S- S AMK -----T YAQAVGCGY LS VV CGLC F LAM FQ-----AT AI QPEWEN-CVPSD : 182
MsCAATCH1 : RNS KV S- S AMK -----T YAQAAGCGY LS VV CGLC Y LAM FQ-----AT AI QPEWEN-CVPSD : 182
BcProT : GSAPLT FR S RAEF----- WQQMVCVTFI ST AV AWS S TYFAITG-----A GK----- : 122
HinSnfT : G APLS RRFSS HFEV-----F WQQMVNVI GL AV GWAAS TYF FTG-----A GD----- : 129
StTnaT : TAT TA KK NRRFEW----- WQQITVPVV VT SV SWS R LIF FTQ-----A GD----- : 123
CeSNF2 : KSPPAV RR M ALE ----- WMTCTIVGAV GV MI SWI L IINIFYV-----NMMGR DNPWQNNCVDG : 240
CeSNF7 : KSPPAA RR M ILE ----- WMTCLVGA I GV MI SWI L LFNIIYA-----TKIS----- : 183
CeSNF4 : RSPPFV RR M ALE -----F WVA----- : 183
CeSNF5 : KSPPVI RR R ILE ----- WMGVFAVLA AI IV SWISI MINICRG-----HFAL SH NNDWNNGTSCIT : 216
CeSNF9 : TTAPII RRAFA ILQ ----- WMAVAVSSL AI II GWSTV MASIVMG-----HTSK NR GNSWNNAESCVT : 178
DmCG13794 : S F SA R- S FFK ----- YVSVFLTIS LI SIFAAPV L MIN FR-----PT SCEGFKSWYNESDYG : 151
DmCG13795 : S F SA R- S FFK ----- YVSGFLTIS LI SIFAAPV L IIN FR-----PT SCEGLSSWHNGSATL : 151

```

DmCG13793 : S F SA R- S LFK ----- YVSLPLTIT LI TIFAAMP L VLN FR-----PT SCEGFKSQWQNDTIK : 79  
DmCG33296 : S F AA R- A LFK ----- YVSIFLTFPTLI SMF AIP L LIN FR-----PT SCEGLKSWHNRDTDEY : 87  
DmCG8291 : A P SM K- S ICA ----- IALVMQQCF AL STVS AWI V LRDVFPFTA--ARSGYR QEMAFPPRYDASNAT : 629  
AgCP9955 : G P TM R- S ICK ----- IALLLAQTL AL SAIS AWV V FRDAFIMR--NEK-YR QEPFEAYHSGRDNQS : 134  
DmBLOT : Q AAHT R-AS IFK -----ACMISRFASWLSAI VSLQAVLA A IGMFAS-----ND RE AGPVKLR----L : 339  
AgCP2093 : Q SAHM R-AA FLK -----ASLVGRIASWLAAI TSMQSVIA L IGLMFLAF-----KS LRE SKSVTINPQAIY : 135  
CeSNF6 : SAA SV K- V LFK ----- FAQVTISGFFAV NI SAWT F LIN FS-----FS SN ANSWSGENCFTLG : 195  
CeSNF12 : QANFTA QR M IGS ----- WALVVIAP AV NI AWA H FPQ AKGLLLG-DE ET RDEWQLDNRCN : 295  
AaLeuT : AQGHGTTPA FYLLWRNRFAKI VFGLWIPLV AI VY ESWT G AIKFLVG-----L EPPNATDPDSILR-- : 142  
HsNTT5 : Q G GV KI A WIG ----- YSSFMVCFI GL NV NSWI F MSQ FQ-----FP EK PLTMNSSGFDE : 236  
DmCG13796 : S T SA R- A IFK ----- YAIIILLNLGLTLT SIAAVP I TVN IH-----PV MS NNSWNTQECSLH : 217  
HspSnfT : RSP GA SS SDSTLWR----VF HLSVVIADV LS SV GGWI R FGA LTG----- : 119  
CeSNF1 : RAP LA YH F VFR ----- VSQILFTMV LACMTKF STLCL LYYYFWTFHAGRSG LN KAFPEFVSQPCR : 160  
CeSNF8 : CSPWFIFIRRA ILQ -----F FMALVSAVT LYP QYS ARAFK LLSLARYRS---QD ST GNWNTTETGHIV : 150

\*            660            \*            680            \*            700            \*            720

DmSERT : -----VTS--ENFTEL-A-----TSPAKE : 222  
AgCP12911 : -----VTGRVNETVTS--T-----VATATT : 164  
MsSERT : -----VTS--PQTNPV--S-----STPAKE : 189  
HsSERT : -----YFSEDNITWTLHS-----TSPAEE : 230  
CeSERT : -----DLNVTISRNGTPLT-----TPSEE : 255  
DmDAT : ESQNASRVFVIG--NYSDLYAMGNQ-----SLLYNETYMNGSSLDTSVAVGHVE-----GFQSAASE : 214  
AgCP3649 : GYINSTATVTVN-----RTG-----VLAANVTSTVSTINDT-----KFASAASE : 175  
TnDAT : EAIWESSANKSR-----VRQSSSSTLGSPPPT-----PFTSAASE : 182  
CeDAT1 : EPHWSEDTAMC-----RSANQSVSAEK-----ISAEE : 200  
HsNET : KLLNG-----SVLGNHTKYSKYK-----FTPAEE : 212  
HsDAT : HPGD-----SSGDSGLNDTFG-----TTPAAE : 215  
TnOctT : GRDNA-----TNRTDVRVYQGPLSH-----FTPASE : 261  
DmCG5549 : A--QADQCEAT-----NGTYLRTCHNATSAEENITLALGALKR----PPAEE : 234  
AgCP10735 : E--EEDNCLAT-----NRSYMANCMSRE--QYRLNLVTLRTPRK---PPAEE : 176  
HsGLYT2 : TKLLLDSCVIS-----DHPKIQIKNSTFCMTAYPNVTMNVNFTSQANKTFVSGSEE : 373  
HsATB0+ : P--IVTHCNVSTV-----NKGIQEIIQMNKSWVDINNFTCINGSEIYQPGQLP--SEQ : 216  
HsPROT : ---LEHRVSK-----DGN--GALPLNLCTCTV-----SPSEE : 192  
HsGLYT1 : ---VLDASNLT-----NGSRPAALPSNLSHLLNHSLQRT-----SPSEE : 267  
DmCG7075 : S-----DMGKQ-----NSS--DVFKTS-----DE : 174  
AgEbiP6595 : RG----DQHVLEFLN-----NSAMAI GDRYRTPA-----DE : 123  
MsPROT : TGNS--SFFTS-----NGS-----ITTPE-----DE : 157  
DmIne : QRKGIN-----ASAPDTS-----RTPSEE : 204  
AgCP4566 : ERLKHN-----LTRPDMS-----RTPTEE : 246  
AmINE : TYSGIDN-----RTRPNAS-----RTPAEE : 453  
MsIne : N--HN-----HTKPNGS-----QTPTEQ : 314  
TnGAT : YDRKNLTCWSTM---DMTTFCT-----LNGKNLSKAVLS-----DPVKE : 203  
MsGAT : YDRKNLTCWSSLG---DMSTFCT-----LNGRNVSKAVLS-----DPVKE : 204  
AgCP8499 : YDRKDLLCWETIGVNGTLTKICS-----LNSNMVMSMDLA-----DPVKE : 166  
DmCG1732 : YERKNLHCWDKLIIN-GTFQKVC-----VSALNITSLELT-----DPVKE : 229  
CeSNF11 : YVKIPCDNRTIAEFFNVKVLTHDHIHEYKQKQFVGEK---MNWTVCSAADLSV-----VSPVKE : 222  
HsGAT1 : YS-----MVNTNMT-----SAVVE : 191  
HsGAT3 : FQKLNVSNY-----SHVSLQAT-----SPVME : 205  
HsGAT2 : FQKTNGS-----LNGTSENAT-----SPVIE : 185  
HsBGT1 : FLNHSAG-----TVTPFENFT-----SPVME : 190  
HsTAUT : TMRKNKSVW-----ITLSSINFT-----SPVIE : 197  
HsCT1 : FRHEDCANASLANL-----TCDQLADRR-----SPVIE : 212  
CeSNF3 : KENFTELAR-----IKALVASAGGHT-----QTSVEQ : 189  
DmCG5226 : LEECAKS-----SETTY : 196  
AgCP7001 : VEECERS-----SETAY : 201  
HsNTT7-3 : EPECEQS-----SATTY : 205  
HsNTT4 : EAEECKS-----SATTY : 204  
HsXtrp2 : VEECQGS-----SAVSY : 160  
RnROSIT : VQECQSS-----GTVSY : 147  
HsBOAT1 : VDECARS-----SPVDY : 174  
HsXT3L : DEEKEKA-----SSTQY : 147  
DmCG10804 : EPECVAS-----SPTQF : 199  
AgCP7501 : EPECVVS-----SPTKY : 148  
DmCG1698 : P-----QEASRAT--SLAG-----SGVR--TTSAEF : 228  
DmCG4476 : -----ASGGQ--PLQG-----QLSRNFSSSTQL : 202  
DmCG15279 : P-----DASNWSQLESNDQRPQNYTMK-----SQNDRVITSSSEW : 216  
AgCP10412 : A-----GSA-----SPNDS-----SSNRTLTSSEL : 232  
DmCG8850 : A-----TAANDSS-----IVQGVSSAEL : 194  
DmCG15088 : A-----QNSSGK-----MSPAQL : 163  
AgCP10401 : L-----ITNMAEN-----STATAVSSAEL : 226  
AgCP10503 : L-----DMRTLRL-----MDWPRIAGTF : 150  
AgCP10610 : L-----GSIALGP-----NVTQPKTSADL : 130  
DmCG3252 : PQEYVDNLLTGV-----SLANESARNLSGI-----VANDETEKLQSSSEL : 209  
AeAAT : -----SLVNVEMRDS-----NSTSGQKVSSSQI : 197  
MsKAAT1 : P-----TLAAG-----VGNITNGTSSAEL : 201  
MsCAATCH1 : P-----TLAAS-----VNNITNGTSSAQL : 201  
BcProT : -----DTQS : 126  
HinSnfT : -----KPID : 133  
StTnaT : -----DPGT : 127  
CeSNF2 : SEQSRCKNGLYSNMATANASSFLNNT-----MAPLMSGSKRIL---YINGKQDATNMSVEVGTQ : 298  
CeSNF7 : ----- : -  
CeSNF4 : ----- : -  
CeSNF5 : MADQYLCKNHTKVMAN--STLWN-----SSLPIPKMVFYFNGACQD-----ANGTDVSTATEQ : 267  
CeSNF9 : SGLQPLCASFNATGNETSPPVWAN-----LSSEGRGKFYIYLNTSCYDKVDLEFNKTKMIAATEQ : 237  
DmCG13794 : MTTCNMTIS-----EDLTDNSDNNTDFHR-----PHFVPSVL : 184  
DmCG13795 : SSSCKNTL-----LLEDFSEYNRSHQ-----LLHVPSVL : 180  
DmCG13793 : WT-----EEVAKETENDTYFYI-----TNIHVPSVL : 105  
DmCG33296 : PTLCHP-----TIYKQHEN---YT-----LGVEAPSRL : 112  
DmCG8291 : GN-----LTQTVAE : 638

AgCP9955 : YR-----LPDVTAD : 143  
 DmBLOT : SGYLLTG-----TSGQE : 351  
 AgCP2093 : NGYDVQQ-----SSGQE : 147  
 CeSNF6 : TRIQCKEMN-----GTLVNGSCIVEHASSNETTVIPLHDLGSIPLSK : 238  
 CeSNF12 : LHLNHCFCFNSTN-----SITAPEA : 314  
 AaLeuT : -----PFKE : 146  
 HsNNT5 : CER-----TTPSIY : 245  
 DmCG13796 : ENYDVDDYRVDP-----HSTVE : 234  
 HspSnfT : -----A : 120  
 CeSNF1 : EAGSITN-----ITQINNRLN-----TIQAESSMM : 185  
 CeSNF8 : FQNKCVN-----ETKIK-----MEAVESAEIQ : 172

	*	740	*	760	*	780	*	800											
DmSERT	:	ERKV	ESYKG-----	NGLDFMGPVKPT	ALC	FGV	V	VYFS	W	RS--A	VV	VTALA	V	I	LLV	G	:	292	
AgCP12911	:	TAMRQV	EQYKS-----	NGLDFMGPVKPS	ALC	FGV	V	VYFS	W	RS--A	VV	VTALA	V	L	LLA	G	:	234	
MsSERT	:	ERNV	EQHKS-----	NGLDDMGPIKPS	ALC	FGV	V	VYFS	W	RS--A	VV	VTALA	V	L	LLA	G	:	259	
HsSERT	:	TRHV	QIHR-----	KGLQDLGGISWQ	ALC	MLI	T	YFS	W	KT--S	VV	VTATF	I	S	LLI	GA	:	300	
CeSERT	:	LYKV	EVQKS-----	TGFDDLGGVKT	AVC	LAV	I	VYFA	W	PQS--S	IV	VTATA	I	S	LLI	G	:	325	
DmDAT	:	NRVI	ELNRS-----	EGIHDLGAIKWD	ALC	LIV	L	CYFS	W	ST--S	VV	FTALF	A	L	LLI	G	:	284	
AgCP3649	:	NRVI	ELDKS-----	EGIHDLGAIKWD	ALC	LAV	L	CYFS	W	ST--S	VV	FTALF	A	L	LLI	G	:	245	
TnDAT	:	NRAI	ELQGS-----	EGHDLGPIKWD	ALC	FAV	I	CYFS	W	ST--S	VV	FTALF	A	L	LLV	G	:	252	
CeDAT1	:	YKGF	GLHEANAPNSHVIRSVTDLGNVRWD	ALS	FVV	L	CYFS	W	HT--S	VV	FTALF	V	G	LFI	G	:	278		
HsNET	:	ERGV	HLHES-----	SGIHDLGLPQWQ	LAC	MVVVI	LYFS	W	KT--S	VV	ITATL	F	F	LLVHG	:	282			
HsDAT	:	ERGV	HLHQS-----	HGIDDLGPPRWQ	LTC	LVIV	LYFS	W	KT--S	VV	ITATM	V	TALLL	G	:	285			
TnOctT	:	HRAV	EMQNS-----	EGLNDLGFPPKWQ	AIC	GLV	VTLYLS	F	KS--S	VV	MTATM	V	S	LLA	G	:	331		
DmCG5549	:	NNFV	GL--S-----	KGIEETGSIKLS	AAC	FLA	T	VFLC	S	QS--S	VV	FTALF	V	V	LFV	G	:	302	
AgCP10735	:	KNYF	QI--S-----	AGIEETGMINQR	ALC	LVA	A	VFLC	S	KS--S	VV	FTALF	V	AALFI	G	:	244		
HsGLYT2	:	KYFV	KI--S-----	AGIEYPGEIRWP	ALC	FLA	V	VYAS	A	KT--S	VV	FTATF	V	V	LLI	G	:	441	
HsATB0+	:	NKVA	QR--S-----	SGMNETGVIVVY	ALC	LLA	L	VGAA	F	KS--S	VV	FTALF	V	L	LLV	GA	:	284	
HsPROT	:	SRVY	HIQGS-----	QRIGVSGEIRWN	CLC	LVA	V	VFLC	L	KS--S	VV	FTATF	L	L	LLV	G	:	262	
HsGLYT1	:	RLVY	KL--S-----	DDIGNFGEVRLP	LGC	GVS	L	VFLC	I	KS--S	VV	FTATF	V	T	LFV	G	:	335	
DmCG7075	:	HLEV	RI--S-----	TDISDLGGMVWEQLTA	IIT	I	IYFC	A	KS--V	VV	FTVPF	L	F	LLI	GA	:	242		
AgEbiP6595	:	HNEI	HI--S-----	EGIEQMGIVVW	LLCDVIA	I	IFGC	A	QS--V	VV	CTATF	A	A	LLV	G	:	191		
MsPROT	:	HRHL	QV--S-----	PNINHIGSIVAP	FWCNLIC	I	VYLC	CN	KS--V	IV	FTVLF	V	S	LFV	G	:	225		
DmIne	:	ENKV	QISGG-----	LEYPGMMRWE	FAC	ICA	L	VYFATW	S	KS--SA	VR	FTATF	V	I	LMV	A	:	272	
AgCP4566	:	ENKV	QISHG-----	IEYPGAMRWE	VAC	VCA	I	VYFA	W	S	KS--SA	VR	LTATL	V	V	FLG	S	:	314
AmINE	:	DNKV	QISNG-----	IESGPLRWE	IAC	ITA	I	VYFS	W	S	KS--SAQVR	LTATL	L	V	FLM	S	:	521	
MsIne	:	ERKV	NMSAG-----	IEYPGMMRWE	AAC	VCA	V	VYFA	W	S	KS--SA	VR	ITTTL	L	I	FLG	S	:	382
TnGAT	:	ERRA	QISSG-----	IEHIGNIRWE	AGT	LLV	V	CYFC	W	RW--T	VV	FTALF	F	T	LLI	G	:	271	
MsGAT	:	ERRA	QISSG-----	IEHIGNIRWE	AGT	LLV	V	CYFC	W	RW--T	VV	FTALF	F	T	LLI	G	:	272	
AgCP8499	:	ERRT	MISSG-----	IDQVGSIRWE	AGT	LLV	I	CYFC	W	KW--T	VV	FTALF	F	T	LLI	G	:	234	
DmCG1732	:	ERRA	QISHG-----	IEEIGNIRWE	AGT	LLV	I	CYFC	W	KW--T	VV	FTALF	V	T	LLV	G	:	297	
CeSNF11	:	NHRV	GISSG-----	LENPGGIRWD	ALF	LLV	I	CYLC	F	KW--T	VV	ITASF	M	FCLLI	G	:	290		
HsGAT1	:	ERNMHQMTDG	-----	LDKPGQIRWP	AIT	AIA	I	VYFC	W	GW--T	VV	FSATY	I	I	LFV	G	:	259	
HsGAT3	:	EHRV	AISDG-----	IEHIGNLRWE	ALC	LAA	T	CYFC	W	TKS--T	VV	VTATF	I	L	LLI	G	:	273	
HsGAT2	:	ERRV	KISDG-----	IQHLGALRWE	ALC	LLA	V	CYFC	W	KS--T	VV	FTATF	L	V	LLI	G	:	253	
HsBGT1	:	ERRV	GITSG-----	IHDGLSLRWE	ALC	LLA	V	CYFC	W	KS--T	VV	FTATF	L	V	LLI	G	:	258	
HsTAUT	:	ERNV	LSSPG-----	IDHPGSLKWD	ALC	LLV	L	CFFC	W	RS--T	VV	FTATF	A	L	LLV	G	:	265	
HsCT1	:	ENKV	RLSGG-----	LEVPALNWE	TLC	LAC	V	VYFC	W	KS--T	IV	FTATF	V	V	LLV	G	:	280	
CeSNF3	:	EKRK	HDTGD-----	ISEPGGIQWE	FFI	AAA	L	VYFA	W	TQ--AR	FV	FCALF	V	F	LLI	G	:	257	
DmCG5226	:	YRFT	DAAPS-----	MDEPGGLKWW	VLC	MLS	T	VFFI	M	QS--S	VV	FTSLF	I	T	FFI	G	:	264	
AgCP7001	:	YRFT	DAAPS-----	IDETGGFKWW	VLC	MLS	T	VFFI	M	QS--S	VV	FTSLF	I	T	FFI	G	:	269	
HsNNT7-3	:	YREA	NISSS-----	ISESGGLNWK	TIC	LAA	V	VCLA	I	QS--S	II	FSSLF	V	ICFLI	AF	:	273		
HsNNT4	:	YREA	LISDS-----	ISESGGLNWK	TLC	LVA	S	VGMA	V	QS--S	VM	FSSLF	V	ACFLV	G	:	272		
HsXtrp2	:	YRQT	NITAD-----	INDSGSIQW	LIC	AAS	A	VYMC	I	ET--T	VI	FTALF	L	T	FLI	G	:	228	
RnROSIT	:	YRQT	NITSD-----	ISNTGTIQW	FLC	VAC	TTVYLC	I	ES--T	VI	FTALF	L	T	FLI	G	:	215		
HsBOAT1	:	YRET	NISTS-----	ISDSGSIQW	LIC	ACA	S	LYMCTI	ET--T	AV	ITSTL	V	T	FLI	G	:	242		
HsXT3L	:	YRKT	NISPS-----	LQENGQVWEPALC	LLA	L	VYLC	L	TES--T	VV	FTASL	C	I	YLI	G	:	215		
DmCG10804	:	YRFT	QCSSES-----	VDMPENFNH	AIA	IVS	F	VYIC	VQ	TS--S	IV	MTAIF	V	I	FFF	G	:	267	
AgCP7501	:	YRET	QVSPS-----	VNEPEQIINYT	ALA	ITA	S	VYLC	VQ	TE--SS	IV	ITAIF	V	I	FFF	G	:	216	
DmCG1698	:	TNII	REKASID-----	DGIGYPSWS	ALA	AVA	I	IAGI	F	KS--S	AS	FLALF	V	L	LLV	A	:	297	
DmCG4476	:	LQRIV	NETDSLE-----	EGIGYPSGS	ALM	GIS	LTVTLI	I	KS--S	AA	VLALF	V	F	LLA	A	:	271		
DmCG15279	:	VKEV	HEKPNIE-----	EGIGLPNWE	VIG	FIA	SCVFFI	R	KS--S	AS	FLALF	V	G	LLV	A	:	285		
AgCP10412	:	TKVV	KELDGID-----	DGIGLPLDR	TLF	VLS	S	VFLT	I	KS--S	AS	FLALF	V	T	LLV	AC	:	301	
DmCG8850	:	TQTV	REPESLDD-----	NGLGTPSWD	VLC	LAT	V	IGTI	S	RS--S	AS	FLALF	V	I	LLA	A	:	264	
DmCG15088	:	---REI	HEVPNID-----	NGLSLPNWQ	VAC	AIA	L	IGGV	I	KS--S	AA	FLGVF	V	L	LLA	A	:	229	
AgCP10401	:	VKDV	KEASTIH-----	DGIGTPDWR	VLC	LVP	TCICLT	I	KS--S	VA	FLAIF	V	L	LLI	AC	:	295		
AgCP10503	:	PLSCTLQ-----	GLGWPDGK	VLC	LVS	S	LVVI	I	RS--T	AA	FLAIF	V	F	LLAHS	:	211			
AgCP10610	:	RNTI	HRAPLAE-----	GLGWPDGK	VLC	LVS	S	LVVI	I	RS--T	AA	FLAIF	V	F	LLAHS	:	198		
DmCG3252	:	LNVV	KEKLDIS-----	DGVGDPPDK	TLA	FVA	V	IFLV	M	KS--S	AA	FLALF	V	F	LLI	A	:	278	
AeAAT	:	LDIV	KEKDSID-----	DGIGAPDVK	TLW	SLA	V	IFLV	V	KS--S	VA	FLAIF	V	I	ILV	A	:	266	
MsKAAT1	:	LRTV	QSDGIE-----	GGLGAPIWY	VLC	FIA	L	VFGV	A	KS--S	AA	FLALF	V	ITLFIT	:	270			
MsCAATCH1	:	LRTV	QSDGIE-----	GGLGAPIWY	VLC	FIA	L	VFGV	A	KS--S	AA	FLALF	V	ITLFIT	:	270			
BcProT	:	LFKEY	HVAD-----	KPGQFGLVPE	LIP	ALV	I	VLGVAF	KKG	IEVUNRIFIPLLVV	FL	IVV	A	:	195				
HinSnfT	:	IGEF	KMADI-----	KNGISFEFVGM	TAP	IAM	I	ALGV	SM	QKG	IA	VSSVLMPLVV	FM	LVIYS	:	203			
StTnaT	:	SSDF	GITS-----	GPLELGLRWG	FAA	AVV	FANYISAO	SGG	IE	ACKIMTPLLVA	L	FVI	G	:	196				
CeSNF2	:	SKSV	RPSAG-----	FTDFNSINLP	LGA	ALC	V	AADV	I	KA--I	LS	VTVIL	F	V	LLICG	:	366		
CeSNF7	:	--KNFV	KPSTG-----	FTDFNSINVP	LAA	AVC	V	AIIS	I	KA--I	LS	YTVLL	A	I	ALMI	G	:	249	
CeSNF4	:	--ANYI	KPSKG-----	LLDINSINWP	FIA	S	-----	FI	-----	IS	VTVIL	I	I	LLI	G	:	234		
CeSNF5	:	MTYI	QPSG-----	MLDFGGFNWP	FAG	SVC	L	TGLG	L	AKI--M	IS	VSVLV	V	V	LFING	:	335		
CeSNF9	:	FDVY	KPTDG-----	FLEFSSINFKSFFG	NAC	L	VVLI	W	GY--M	AS	VIVTL	F	I	FLG	:	305			
DmCG13794	:	LKHVESVQEENY	-----	SDLRQDYDLRWFVSLFVLV	A	VAFIFY	FSETAMF	FIRYVMVIGTLI	L	CFG	F	:	258						
DmCG13795	:	QNHYSMQQDAV	-----	PDWYKYDELSWHFVGLFALL	V	IAFIFY	FSETAKF	LIRYVMVITLV	L	CFV	F	:	254						
DmCG13793	:	N-----	ELNDYEISWH	SGLFALV	ATIAFIFY	FSEPAKF	CIRYVMVIGTVA	L	CFV	F	:	166							
DmCG33296	:	DYIR	-----	QDY-----	FNTDKRFDISR	IGLFTLS	A	IATIFYHFSEPAKF	CIRYVMVITTMV	V	CFV	F	:	181					
DmCG8291	:	NVVV	QRHLANHPDAS	--GIRPHVNRD-Q	AFY	ALI	AAVFLI	C	KS--L	LA	IYTLV	LVA	A	VTA	F	:	713		
AgCP9955	:	NGVV	QRYLHAHQAVSS	--SLPTGYAIRFO	AFN	AII	TFVFA	C	RS--L	ITIGVFA	AA	TAVCA	F	:	219				
DmBLOT	:	CLQLTF	TPFWR-----	NPLYFGL	AAG	IGL	I	VMLCTHNAKIL	---R	SI	VFLVGLV	CTLTGWE	:	415					
AgCP2093	:	CLKLTF	RPVWR-----	SSLYFGL	ALS	ILL	V	SMVCTHS	KIN---R	SI	LFGVALV	IFETGWE	:	211					

CeSNF6 : QNDV MLSKG-----VDDFGTLNMY GLC LAC IAVFLC FQ KS--S VV VAVIV I T LLL L : 306  
 CeSNF12 : HSEV SLSTFG-----DFALGPLQSH VLS AAA L VFFG F GS--IAQTMNVATV L S LLL G : 383  
 AaLeuT : LYSYIGVPGD-----EPILKPSLFAYIVFLITMF NVSI I SKG-IE FAKIAMPTL I AVFLVI VF : 214  
 HsNNT5 : YQQA KASDR-----IEDGGSPVYS VLPFFLC C VGAF IN KS--T VI VLVLV CF VGFFI T : 313  
 DmCG13796 : RSMIASTAKG-----STSMSISWS LIG LSI L VLAL L P AF--I VLRCSVLM GFFVAVFLYL : 301  
 HspSnfT : SNPGQYFATVS-----YGLDAVGFHLLFLA TGLI V KRG-IENATKLMMPAVVV I LAAWAS : 184  
 CeSNF1 : Q LTLERLPSSE-----IADPKDFQYP LIAQGAI IAIFVA CF RF--V TVAFVLM LA SS L FFL SA : 253  
 CeSNF8 : LRVNHRIGVS-----HMSFTNFNYG AYS FII LTISFVQN VNTR--TTPLI IPTVFA L CHPLLV S : 240

		*	820	*	840	*	860	*	880		
DmSERT	: S P	----	DE KY YLT E	-----	WHK KNSK ID AS	----	LGP GF T ALS	NKFNN CY	-----	: 350	
AgCP12911	: T P	----	AE RY YLT Q	-----	WDK NNSR ID AS	----	LGP GF T ALS	NKFNN CY	-----	: 292	
MsSERT	: T P	----	TE RY YLT E	-----	WHK QNSK ID AS	----	LGP GF T ALS	NKFNN CY	-----	: 317	
HsSERT	: T P	----	WR LF YLK N	-----	WQK LETG ID AA	----	LGP GF V AFA	NKFNN CY	-----	: 358	
CeSERT	: L P	----	KN YYYV D	-----	FEK KDPA SA AT	----	LGP GF V ALS	NDFNN CY	-----	: 383	
DmDAT	: T P	----	SFL QY YLT N	-----	FSA YKAE VD AT	----	LGP GF V AYA	NKYHN VY	-----	: 342	
AgCP3649	: T P	----	SAE LY YLR N	-----	FDV YNAE VD AT	----	LGP GF V AYA	NKYHN VY	-----	: 303	
TnDAT	: T P	----	SAT QY YLS N	-----	FEA TQPQ VD AT	----	LGP GF V AYA	NKYHN VY	-----	: 310	
CeDAT1	: T P	----	WQN EY YLR N	-----	FEM KRPS QD AT	----	LGP GF V AYS	NDFHN VY	-----	: 336	
HsNET	: T P	----	SN NAYLHID	-----	FYR KEAT ID AT	----	LGAGF V AFA	NKFDN CY	-----	: 340	
HsDAT	: T P	----	ID RAYLSVD	-----	FYR CEAS ID AT	C	----	LG VGF V AFS	NKFTN CY	-----	: 343
TnOctT	: L P	----	TR AYYLQ E	-----	LTR KDTQ VD AV	----	VGAGF VH SYA	NTFHN CY	-----	: 389	
DmCG5549	: T P	----	ST LF YLT D	-----	WKQ ANAQ GD AV	A	----	LSPAW G TLS	NKFSN CYNCGTFE	: 366	
AgCP10735	: T P	----	YD LY YVR D	-----	WDK RTAQ GD AV	A	----	LSPAW G TLA	NRFDN CY	-----	: 302
HsGLYT2	: T P	----	GA WYFIT K	-----	WEK TDAT KD AT	----	LSAAW G TLS	NKFHN CY	-----	: 499	
HsATB0+	: T E	----	SK SY YIGAQS	-----	NPTK KEAK KD AT	----	LSVAW G ALS	NKFKN CF	-----	: 344	
HsPROT	: T P	----	WK QFYLT Q	-----	FHH LSSK IE AL	----	LG VGF G TFA	NTFHQ IY	-----	: 320	
HsGLYT1	: T E	----	FD MYYLT Q	-----	WDK LEAK GD AS	----	LGC AW G TMA	NKFHN CY	-----	: 393	
DmCG7075	: T P	----	WK KFYLY E	-----	WHR LDLK AD AV	G	----	IGPGW G NMA	SSFRN AK	-----	: 300
AgEbiP6595	: T P	----	LE KFYIM Q	-----	WSQ LNLR AD AI	----	LGPGW G NMA	NQFKN TK	-----	: 249	
MsPROT	: T P	----	WK MFYIL D	-----	WAQ AKPK AD AT	----	LGPGW G SMS	NKFHY NL	-----	: 283	
DmIne	: T D	----	AE RFFFR K	-----	WSE KNAN IN AS	N N	----	LGITF S SFA	NKYNN IL	-----	: 330
AgCP4566	: T E	----	DK HYFFR N	-----	WEE GRAN IN AA	N N	----	IGIAF S SFA	NKYNN IL	-----	: 372
AmIne	: T E	----	DK QFFPH R	-----	WEL ADAK IY TA	N	----	VGIAF S CFA	NKFHNTIL	-----	: 579
MsIne	: T D	----	DG RFFFK D	-----	WEL KQSRP VN AS	N	----	IGIAF S MFA	NRFDN FL	-----	: 440
TnGAT	: T P	----	MQ KFYVM N	-----	MSK MESE ID VT	----	YGLGL T ALG	NKFTN VY	-----	: 329	
MsGAT	: T P	----	ME KFYVM N	-----	MSK LESE ID VT	----	YGLGL T ALG	NKFTN VY	-----	: 330	
AgCP8499	: T P	----	FE KFYVS N	-----	LSK AESE ID VT	----	YGLGL T ALG	NKFNN VY	-----	: 292	
DmCG1732	: T P	----	LE KFYII N	-----	FSG TNSE ID VT	----	YGLGL T ALG	NKFTN VY	-----	: 355	
CeSNF11	: T E	----	GV EFYLK D	-----	FSK LESK VD VT	----	YGLGL A ALG	NKFNN VY	-----	: 348	
HsGAT1	: T P	----	KE LFYIT N	-----	FRK SDSE LD AT	----	YGLGL S ALG	NSFHN VY	-----	: 317	
HsGAT3	: T P	----	SE KFYLY D	-----	LSR SDPQ VD GT	----	YAICL C TALG	NNYNN CY	-----	: 331	
HsGAT2	: T P	----	AQ QFYLY N	-----	LTR WDPQ MD GT	----	FAICL C TALG	NKYHN CY	-----	: 311	
HsBGT1	: T P	----	YQ IYYLK D	-----	LFR KDPQ MD GT	----	FAICQ C TALG	NKYHN CY	-----	: 316	
HsTAUT	: T P	----	GA KFYLY D	-----	TR EDPQ ID GT	----	YAICL A TSLG	NKYKY SY	-----	: 323	
HsCT1	: L P	----	LD IYYLK D	-----	WSK GSPQ ID GT	----	YAIGL A TALG	NRFNN CY	-----	: 338	
CeSNF3	: T E	----	GT YFYLK N	-----	ATR LDPA KD GT	----	YGVGF A ALG	HNKFNH CF	-----	: 315	
DmCG5226	: T R	----	GA MHMYT K	-----	VEK LEPT LD AT	----	FGLAF S AFG	NTPKN CV	-----	: 322	
AgCP7001	: T K	----	GA MHMYT K	-----	VEK LEPT LD AT	----	FGLAF S AFG	NTPKN CV	-----	: 327	
HsNNT7-3	: L N	----	SID RHMPF K	-----	LEI LEPK RE AT	A	----	LGLGF G AFS	NKRDN CH	-----	: 331
HsNNT4	: L R	----	VD LHMPF K	-----	LDK LDQP RE AT	A	----	LGLGF G AFS	NKQDN CH	-----	: 330
HsXtrp2	: T P	----	TK IYLF T N	-----	MHI QNPR LD AT	----	LSLAF GH AFA	NSPRN CQ	-----	: 286	
RnRNSIT	: T P	----	TE TYLFT N	-----	MKI QNSR LD AT	----	LSLAF GH AFA	NQPRN CE	-----	: 273	
HsBOAT1	: T K	----	TN VF LFT N	-----	VTE AQDPT LD GA	----	FSLAF G SFS	NSVHN CE	-----	: 300	
HsXT3L	: T H	----	TN MYMFT K	-----	IEQ ANPKA IN AT	----	LGLGF S AFA	NEPSN CQ	-----	: 273	
DmCG10804	: T K	----	AD AHLFT R	-----	WET LDPV LE GT	----	LGLAF G AFS	NPANN CY	-----	: 325	
AgCP7501	: T K	----	SD AHLFT R	-----	WES LEPV LE GT	----	LGLAF G AFS	NPANN CY	-----	: 274	
DmCG1698	: T P	----	FD LYFLR Q	-----	WHK LEPQ YA VT	----	LAICF N MYA	NRFHG IYR	-----	: 356	
DmCG4476	: T P	----	YD MYFLT Q	-----	WEK LEPQ YN VT	----	LAVCF V MYA	NRFHG VYR	-----	: 330	
DmCG15279	: T P	----	SID YFYFK Q	-----	WKG LDPK YA VT	C	----	LSVCF N MYA	NKFGH VHR	-----	: 344
AgCP10412	: T P	----	VD VYFLK Q	-----	WDK YDPK YA VT	C	----	LSICF N MYA	NKFRH VYR	-----	: 360
DmCG8850	: T P	----	WQ VYFLK Q	-----	WSQ LNPH YA IT	----	LAICF T MYA	NDFNK VHK	-----	: 323	
DmCG15088	: T P	----	ID YFYFK Q	-----	WRE LNPL YA VT	----	LAICF T TYA	NNFNR VYN	-----	: 288	
AgCP10401	: T E	----	AD LYFIK Q	-----	WDR LEAK YA VT	----	LTICF N MYA	NRFHN VYRFVNAR	-----	: 359	
AgCP10503	: S E	----	SLE KFFLT K	-----	WES FSAK ME VT	C	----	LSICF G AYS	NNFSN VYRDAMII	: 275	
AgCP10610	: S E	----	SLE KFFLT K	-----	WES FSAK ME VT	C	----	-----	-----	: 233	
DmCG3252	: T E	----	RD LFFLE Q	-----	WGE LNPT KE VV	C	----	LAVGS P MFA	NRFDHGIYR	-----	: 337
AeAAT	: T E	----	VD IFFIK Q	-----	WGE LNPX YS TT	----	LSVGM S MFS	NNFHH IYR	-----	: 325	
MsKAAT1	: L P	----	TD LFFVT Q	-----	WNK LELG YA IT	----	LSVCT A MFS	NGFKQ VYR	-----	: 329	
MsCAATCH1	: I P	----	TD LFFVT Q	-----	WAK LELG YS VT	----	LTVCT P MFS	NGFRH IYR	-----	: 329	
BcProT	: T E	----	MQ DAFPK D	-----	WSR FDKG IA YG	----	LSLAF I TYA	LPKNS TTN	-----	: 254	
HinSnfT	: F P	----	AQ DALFT D	-----	WTK SNPS LA YG	----	LSIGF I TYA	LKES LGT	-----	: 262	
StTnaT	: T P	----	TY NYFLN D	-----	FSK MDPG VA YS	----	TLAV V AYA	VPEDS LAN	-----	: 255	
CeSNF2	: S P	----	SD YHLFG	-----	QTFDNY YKRET TA LQ	C	----	LSIGQ G NIA	NKKTY WYR	-----	: 426
CeSNF7	: T E	----	KT HFLFWGKPNEYGHYDFAG	YDPT	IA LM C	----	LSIGQ G NIG	NKKTY WYR	-----	: 316	
CeSNF4	: T E	----	GD KYFLA	-----	RTDWVA YNFKT SA LT	C	----	LSIGF V NTA	NKQNHRCFR	-----	: 294
CeSNF5	: FQD	----	SGV EMYFG	-----	TPNYTK YEQDT TE LK	C	----	LSVGH G SLS	SPKRN IFR	-----	: 395
CeSNF9	: T D	----	ED YYYLG	-----	NPDWSK FMPAT GE LK	C	----	LGIGY G GMA	SKPNN CFR	-----	: 365
DmCG13794	: F P	----	LL RKYVT S	-----	PDEIA GIGSTFV VLHA	----	FGAG S ALS	NGFKT IMS	-----	: 315	
DmCG13795	: F P	----	LN HRYVT K	-----	ASDMA GVVSTFI VLHA	----	FGAG S TLS	NGFKT IMS	-----	: 311	
DmCG13793	: F P	----	HL KNYMT S	-----	EEFV GTSSFI ALQA	----	FGAG S TLS	NGFKT IMS	-----	: 223	
DmCG33296	: F P	----	SHKL WMYIT N	-----	TKDFSEGIYSTFI ALQA	----	FGAG S ALS	NGFKT IMS	-----	: 238	
DmCG8291	: Y VD	----	PSR QNIPAAASD	-----	FFDF VNSNS TA TQ T L	----	WGLLGAS AIT	RSHTN	-----	: 768	
AgCP9955	: T FID	----	YDS QNIPFATD	-----	WQDFLNSRS TS AQ T L	----	WGLLGVS YALNCRSNRKGSCNV	-----	: 279		
DmBLOT	: RNSF	----	SRHYFPELWGFDS	-----	NL AESN FN LM L	----	VNCGF A PMITGKFLYKG	-----	: 470		
AgCP2093	: T KSI	----	NEQYFPDLWPFHP	-----	ESPADST FN LV I	----	LNIGF A PVLTKGFLYKG	-----	: 266		
CeSNF6	: T D	----	SLAA FYFLT K	-----	WEI MDLH GE AV	A	----	VSCCS G FTIA	SRFHN IYK	-----	: 365
CeSNF12	: S P	----	NK TFLFTVDS	-----	TK WKWQ KS AE	E	----	LGIDA P SMAA	SRYRN IYR	-----	: 442
AaLeuT	: L ETPNGT	AD	NFLWT D	-----	FEK KDPG IA VG	----	LSLGF A TYA	VRKDQ	-----	: 273	



CeSNF1	:	-----SIFLSVNHLLISFVQTMSSIICI	YLSMK-----	: 343
CeSNF8	:	-----VATLAVIL TGLSIIISATV SI	YTRFILG-----	: 331
		* 980 * 1000 * 1020 * 1040		
DmSERT	:	-----QKTSIDK GLE-----	-GPG V	: 399
AgCP12911	:	-----QNKSIIE GLE-----	-GPG V	: 341
MsSERT	:	-----QNKSIIE GLE-----	-GPG V	: 366
HsSERT	:	-----RNEDVSE AKDA-----	-GPS L	: 408
CeSERT	:	-----TNKPINE VGEH-----	-DAS I	: 433
DmDAT	:	-----LGVRIED ATE-----	-GPG V	: 391
AgCP3649	:	-----SQSQIQD ATE-----	-GPG V	: 352
TnDAT	:	-----SGRDIQD ATE-----	-GPG V	: 359
CeDAT1	:	-----SGKPIEA AQE-----	-GPG V	: 385
HsNET	:	-----HKVNIED ATE-----	-GAG V	: 389
HsDAT	:	-----HSVPIGD AKD-----	-GPG I	: 392
TnOctT	:	-----QGVPISS ATE-----	-GPG V	: 438
DmCG5549	:	-----LNVVVEK VDQ-----	-GAG A	: 422
AgCP10735	:	-----LDTSVDS IDQ-----	-GAG A	: 351
HsGLYT2	:	-----RKVNIEN ADQ-----	-GPG A	: 548
HsATB0+	:	-----SGKEVSK VKS-----	-GPD A	: 393
HsPROT	:	-----LGVVVDQ AKA-----	-GPG A	: 369
HsGLYT1	:	-----LGVVDSR ADH-----	-GPG A	: 442
DmCG7075	:	-----SGIPVES ATG-----	-GAG A	: 349
AgEbiP6595	:	-----TGLPVASAAATG-----	-GPG A	: 298
MsPROT	:	-----TNVPSVQ ATA-----	-GPG A	: 332
DmIne	:	-----QNTNVRD IGD-----	-GPG I	: 379
AgCP4566	:	-----QNTTVEN ISG-----	-GPG I	: 421
AmINE	:	-----QNMSVED LTD-----	-GPG V	: 628
MsIne	:	-----QNTPVKD IAD-----	-SPG L	: 489
TnGAT	:	-----QORPVAE AAS-----	-GPG A	: 378
MsGAT	:	-----QORPVAE AAS-----	-GPG A	: 379
AgCP8499	:	-----QORPVAE AAS-----	-GPG A	: 341
DmCG1732	:	-----QORPVAD AAS-----	-GPG A	: 404
CeSNF11	:	-----QEKVSAE AQA-----	-GPG L	: 397
HsGAT1	:	-----TKRSIAD AAS-----	-GPG A	: 366
HsGAT3	:	-----QGVPIAE AES-----	-GPG A	: 380
HsGAT2	:	-----QGVPISE AES-----	-GPG A	: 360
HsBGT1	:	-----QGVPISE AES-----	-GPG A	: 365
HsTAUT	:	-----QGVDIAD AES-----	-GPG A	: 372
HsCT1	:	-----QGVHISK AES-----	-GPG A	: 387
CeSNF3	:	-----AQKDISE VKP-----	-GVG A	: 364
DmCG5226	:	SNTEILVKNKLLGLND-----TQG--YEQAMSLNLTGTELLR	QLSE----CSLAHELDNAEAGTG A	: 418
AgCP7001	:	VNKHILVKHNLPSMD-----VDQSE-YEHAMAGLNETMTKDWFEK----	CSLEDQLSSAAEAGTG A	: 425
HsNFT7-3	:	QNSSETIMKFLKMGNISQDIIPHHI--NLSTVTAED-YHLVYDIIQKVKEEFPALH--LNSCKIEEELNKAVQGTG A	: 444	
HsNFT4	:	ENAEKILGYLNTNVLRSRDLIPPHV--NFSLHTFKD-YMEMYVIMTKVEDQFSALG--LDPCLLEDELKDSVQGTG A	: 443	
HsXtrp2	:	RNLSLINDFDPEQS-----ISRDD-YPAVLMHLNATWPKR	AQLP--LKACLEDLFDKDSASGPG A	: 387
RnROSIT	:	RNLSLINDFDPELS-----ISRDE-YPSVLMYLNATQPER	ARLP--LKTCHLEDLFDKDSASGPG A	: 374
HsBOAT1	:	TNILLINGFDLPEGN-----VTQEN-FVDMQQRCSNADPAAYALV--FQTCDINAFLEAVEGTA	: 401	
HsXT3L	:	KVSLLLTNTFDLEDGF-----LTASN-LEQVQGYLASAYPSKYSEMFPQIKNCSLESELDTAVQGTG A	: 376	
DmCG10804	:	ERNGLVAQNKTHN-LP-----VCDLQTELANVSSLA	CLHG----IYTYIYLPFQSASGTA	: 418
AgCP7501	:	ERSEMIRLNKSHDLLP-----VCDLQKELENVTPRR	HRSSSFTIISVFPHLLQASGTA	: 372
DmCG1698	:	-----NNVTTDIAS VNG-----	-GPG A	: 405
DmCG4476	:	-----SGTKDIAS VKA-----	-GPG A	: 379
DmCG15279	:	-----IGTDDIGS VKG-----	-GAG A	: 393
AgCP10412	:	-----TGKTDVGN VKS-----	-DAG A	: 409
DmCG8850	:	-----TNTKDISQ VKG-----	-GAG A	: 372
DmCG15088	:	-----TGNPDIGS VKG-----	-GAG A	: 337
AgCP10401	:	-----LEAPDIKH VRG-----	-GAG A	: 411
AgCP10503	:	-----TGNR-VED QLQ-----	-GPQ T	: 366
AgCP10610	:	-----	-	: -
DmCG3252	:	-----LQIENIRD VRS-----	-GTG A	: 386
AeAAT	:	-----LGIEDISK VKS-----	-GTG A	: 374
MsKAAT1	:	-----LNSVDGD IGSG-----	-GTS A	: 378
MsCAATCH1	:	-----LNSEVGD VGAG-----	-GTS A	: 378
BcProT	:	-----MGVPVDK ASAG-----	-VG A	: 302
HinSnfT	:	-----QGQEVSE AKGG-----	-IG A F	: 310
StTnaT	:	-----AGVPFEE AVAG-----	-PG A	: 303
CeSNF2	:	-----RGINSTD DAFN-----	-HIVKEGHA A	: 480
CeSNF7	:	-----RNISISNPKGFN-----	-EVVQEGHA A	: 370
CeSNF4	:	-----RGVTVD-----	-KVVTDGTT A	: 342
CeSNF5	:	-----TGQDVK-----	-DVVKSGLS A	: 443
CeSNF9	:	-----KGCSSVA-----	-EVVNDGFS A	: 413
DmCG13794	:	-----EGIEGSPDKQ LTH-----	-WAL	: 362
DmCG13795	:	-----ELSDVDDNGH LSH-----	-W L	: 358
DmCG13793	:	-----L INH-----	-W L	: 260
DmCG33296	:	-----ADFLFLHHISK MNF-----	-W L	: 288
DmCG8291	:	SPENPDCYTSIYSLQSNTPYLLS--YPRSLIPHYSSFIGETYRRNRMTIH	ESG-----FQALRFISE	: 875
AgCP9955	:	SYENLGTNVFLRPNQPLPPQHAT--MPNRWLLRYSTVVGESFKRSYADPSRESG-----	YQALR VTE	: 386
DmBLOT	:	-----SNGFQNMEEK-----	-PLTA	: 516
AgCP2093	:	-----TEAHFPYPELS-----	-ALT	: 313
CeSNF6	:	-----FAISLDFHIRD-----	-GFH V	: 414
CeSNF12	:	-----SNSNVND LKHDP-----	-LY S T	: 491
AaLeuT	:	-----VANAVAIKAG-----	-AFN G	: 325
HsNFT5	:	RNAEILLKILNGLKLPDPAKPPVNLNPTSIYNAWLSGLPQHIKSMVLRE	TECN-----IETQFLKASEGPKFA	: 482
DmCG13796	:	-----HVGMMPLH DEKH-----	-HMQFL	: 411
HspSnfT	:	-----ACTG-----	-GPGAL	: 286
CeSNF1	:	-----LGIHPTE LETG-----	-EDQ	: 359
CeSNF8	:	-----RKSVDYLT LPN-----	-GLL THS	: 350

DmSERT : V PE ATM-----SGSVF SI L IT D TFGGLEAMI A C EYP--RVIGR---RRELFVLLLLAFIFLCA : 468
AgCP12911 : V PE ANM-----KGSVF SI L IT D TFGGLEAMI A C EYP--RTIGR---RRELFVLLLLGLIYICA : 410
MsSERT : V PE ATM-----TGSVF AI L IT D TFGGLEAVT A C EYP--RVLGR---HREVFVAVLLLFYIYICA : 435
HsSERT : T AE ANM-----PASTF AI L IT D TFGGLEAVT A L EFP--HVWAK---RRERFVAVVITCFPFIS : 477
CeSERT : V PQ ATM-----DYSCEF SF V IT D TFGGLEAVT A S ESR---FLSKN---RKWFVVLVICIYYFIS : 501
DmDAT : V PA ATM-----PASTF AL M LT D SFGGSEAI A S EFP---KIKRN---RELFVAVGFLSIFYFVG : 459
AgCP3649 : V PA ATM-----PGSIF AL M LT D SFGGSEAI A S EFP---KIGRN---REIFVAVFLSIFYFVG : 420
TnDAT : V PA ATM-----PGSTF AL M LT D SFGGSEAI A S EFP---PIGRH---REIFVAVFLSIFYFVG : 427
CeDAT1 : V PE STM-----PYAPF SV L MT D SFGGSEAI G S EFP---ILKKN---REVVFVAVGFLSIFYFVG : 453
HsNET : L PE STL-----SGSTF AV V LA D SMGGMEAVI G A DFQ---VLLKR---HRKLFVAVGFLSIFYFVG : 457
HsDAT : I PE ATL-----PLSSA AV I LT D SMGGMESVI G I EFG---LLHR---HRELFTLVAVGFLSIFYFVG : 460
TnOctT : V PE ATL-----PGASL AM F IM D GMGGLECVI G L QAR---ACGATWLRREHFTLVAVGFLSIFYFVG : 509
DmCG5549 : V PEV TRL-----PVSPL AV V LT D QFALMETVT A L RFP---NLRQY---KIWVVLVAVGFLSIFYFVG : 490
AgCP10735 : V PEL AKL-----QMPRL SV F LT D QFALMETVT A L TFP---ALRRR---KIAVAVGFLSIFYFVG : 419
HsGLYT2 : V PE TRL-----PLSPF AI L LT D MFATIETIV S S EFP--KYLRT---KPVFTLGGCICFFIMG : 617
HsATB0+ : A PE AQL-----PGSIF AI F LT D QFASLETIT T Q LFP---KVMKM---RVPTLGGCICFFIMG : 462
HsPROT : V PQ TML-----PLSPF SF F LT D QFAPLETIV A T EFP---YLRPK---KAVFSGLICVAVYLMG : 438
HsGLYT1 : A PE TLL-----PISPL SL F IL G QFCLLETIV A V EVGN-EWILQ---KTYVTLGAVAGFLG : 512
DmCG7075 : T PE ALL-----PVPQL AC F FL D VFVQLEAVI S L EWY---WVRSH---KCKLTLISCLIFLGLS : 417
AgEbiP6595 : T PE SML-----PLSPL AL S FF D VFVQLEAVI S M VLP---RLRPM---KLVQVATGCTFLFLLS : 366
MsPROT : T PAT TMM-----PAPNL AIT V FF D MFVTIEIAG L EFP---RFKSR---KRIIAFITCVVLSFIS : 400
DmIne : V PQ AKM-----PYAQL AV F LC N QFAIVEVVV S Q GFP---RWIKRHLVYHEIVLVAVGFLSIFYFVG : 451
AgCP4566 : V PQ AKM-----PAAQL AV F LC N QFAIVEVVV S Q GFP---RWIKRHLVYHEIVLVAVGFLSIFYFVG : 493
AmINE : L PQ AKM-----PASQV AV F VC S N QFAIVEVVV S Q GFP---KWVRRHLTCHTMLVLLICVISFLG : 700
MsIne : V PQ AKM-----PASQL AV F FLC N QFAIVEVVV S Q GFP---DMIRKRLVYHEIVLVAVGFLSIFYFVG : 561
TnGAT : A PS LQL-----PGAPL SC F LL D QFCTMEGFI A I EWP---KLLRR---KEIFIAITCVLSYLVG : 447
MsGAT : A PS LQL-----PGAPL SC F LL D QFCTMEGFI A I EWP---KLLRR---KEIFIAITCVLSYLVG : 448
AgCP8499 : A PS LQL-----PGAPL SC F LL D QFCTMEGFI A I EWP---HLLRR---KEIFIAITCVLSYLVG : 410
DmCG1732 : V PS LQL-----PGSPM SC F LL D QFCTMEGFI A I EWP---QLRRR---KEIFIAITCVLSYLVG : 473
CeSNF11 : A PSG LQL-----PYTQF SC L LF D QFCTMEGFI A I EFP---QIRKKY---GREIFVGVICVISYLVG : 468
HsGAT1 : A PE TQL-----PISPL AI S LM D QFCTMEGFI A V EYP---RLLRN---RELFIAAVCVLSYLVG : 435
HsGAT3 : A PK TMM-----PLSPL AT M IF D QFVVEVSLV A V MYP---KVFRRGY---RRELLLALSIVSYLVG : 451
HsGAT2 : A PR VML-----PFSPL ACC F VL D QFVVEVSLV A V MYP---HVFRKN---RREVLILGVSIVSYLVG : 431
HsBGT1 : A PK TMM-----PLSQL SC I IF D QFVVEVSLV ASI MFP---RQLRKS---RRELLLTIAVMVCYLVG : 436
HsTAUT : A PK TMM-----PLPTF SI I LL D QFVVEVSLV S V LYP---SFLRKY---RREIFIAVAVCVLSYLVG : 443
HsCT1 : A PR TLM-----PVAPL AA F LL D QFVVEVSLV G L LLP---ASYFRF---QREISVALCCALCFVID : 458
CeSNF3 : A PEVANS---PMKQV AV L TI D QVICMEGLF A E AFD---ILRKY---KQSLGIFCLFFFCIG : 432
DmCG5226 : V TQ VEL-----PGAPF AV T LS G QVIGLEMLCT F ID-----IPKIRKPYVTVGVVAVGFLSIFYFVG : 486
AgCP7001 : V TE VEL-----PGSFP AV T LS G QVIGLEMLCT F ID-----IPKIRKPYVTVGVVAVGFLSIFYFVG : 493
HsNFT7-3 : A TE THF-----PASPF SV L VN G MFGTIEGIV P V T-----FKVREKILTVICCLLAFICG : 509
HsNFT4 : A TE THF-----PASPF SV L IN G MFGTIEGIV P I T-----FKVREKILTVICCLLAFICG : 508
HsXtrp2 : V TETDLHM---PGAPV AM G FT S MFGTIEGIV P L VGV-----LPRWPKEALTVLAVGFLSIFYFVG : 456
RnROSIT : V TE LHM-----PGASV SV G FT S MFGTIEGIV P F MGI-----LPKGVPEKMTVGVVAVGFLSIFYFVG : 443
HsBOAT1 : V TE TKM-----PLSPL SV I FC S MFGTIEGIV P LRV-----IPKPKVKEVTLGLICLGTFLIG : 470
HsXT3L : V TE KNM-----EVSQV SV F LM G MLGNTAAIL P T SKI-----ISSHLPEKISGLVAVGFLSIFYFVG : 445
DmCG10804 : I TE NQF-----PGAQL AV L FT D QFQTLGEGV S V MK-----LFPNLPKIEYVAVGFLSIFYFVG : 486
AgCP7501 : I TE NQF-----PAAQL AV L FT D QFQTLGEGV S M MK-----LFPNVPKEMITGALCMSCCVLS : 440
DmCG1698 : S PD AKFK-----WLPQL SV L FV G NVGMASCMV K QFG---HLKNWT---VVVAVGFLSIFYFVG : 472
DmCG4476 : S PD AKFK-----MFPQV SL A FM G NVGMASCMV K QFV---NVKLWI---IVVAVGFLSIFYFVG : 446
DmCG15279 : S PD AKFK-----NLPQI SV L FV G NIAMTSCSV A R RFP---NFGQWQ---CSLLIAVAVGFLSIFYFVG : 460
AgCP10412 : S PE AKFE-----VLPQA SV L FV G NVAMTSCVM V K QFP---RVRNMQ---AATIIAICVGLL : 476
DmCG8850 : S PE AKFK-----YLPQL AV F LV G NIGMASAVVNV K RF---THLPHWL---LAVSASIGFLSIFYFVG : 439
DmCG15088 : S PE AKFE-----YVPQM AV L FV G TVGMGSCILLR V QFGL-RSPPIWK---LASGLTVLGFVSG : 406
AgCP10401 : T PD AKFT-----FWPQF AIA L FV G NVGMATTIM V R RFP---QLQPAL---VAVGIAVGYGIG : 478
AgCP10503 : T PD AKFD-----AAQNV AV L PFL G NTGIVTTIV A R RFP---ELPNWK---VVISIAVGYGCC : 433
AgCP10610 : ----- : -
DmCG3252 : S PD SKFQ-----AVPQL SV F FV G IVALQSTIV I C QFK---GWKYW---VALTTSVCGFLMG : 453
AeAAT : S PD AKFD-----IVPQL SV F FV G AVALLGAAI AFW AFP---KRKYWH---LALLSTIGFCTG : 441
MsKAAT1 : S PD AKT-----FQVQL SV L SV G SVALLSTLN V M AFP---RVPTVY---MSALSTIGFCTG : 444
MsCAATCH1 : S PD AKT-----FQVQL SV L SV G SVALLSTFN LAM AFP---RVPTVY---MSAMTSCGFLG : 444
BcProT : V PQI NELP-----MSPV LG LS TGA T LISLAEVCFAA SEKFS-----LSREKTIIGMGLLVLVLS : 367
HinSnfT : A PTI NKAP-----PGEVLGM GA TFAA T FISVIEVII A Q KIR-----ISRKGVTIVGVPMMLVS : 375
StTnaT : A PK SMLPGPT---WLSQL GI SA LLA S SISQMESFA A I RFG-----VDRKLLGWVPSLIGFAFS : 372
CeSNF2 : V TE AEMP-----VPYL YA I FL S EVVIVEIVC A RFR-----YLRERRWLVLSVAVVFLG : 548
CeSNF7 : V TE AQMP-----YFPL YA V FL S EVVIEIVC C A RFH-----YLRQHRKMTVTVVSTTFLLG : 438
CeSNF4 : A PD NQMP-----FPGL SF F FL S LLVIVEVMC C KFP-----RLREKRVVAVGFLSIFYFVG : 410
CeSNF5 : V PE TRMP-----VPWL CF L FL AS EIALVDVFC C Y QYP-----RFRNRKVIIVAVGFLSIFYFVG : 511
CeSNF9 : A PE GRMP-----LPEL SF L FF S EVAYVNVFC S C QFV-----NLRKRKLVAVAVGFLSIFYFVG : 481
DmCG13794 : SSAT SSLG-----WPNL TF T LMAA IVITTIQIFTVLQS F EFEVLRVRKQE-----VTFGLIGGIAVCS : 430
DmCG13795 : SSAS STMA-----WPNL TF S LIAA IVMTTIQIFTVLQS F EFEVLRVRKQE-----VTFGLIGGIALCS : 426
DmCG13793 : SSAS SSLG-----WPNL TF T LMAA I---TQIFTVLQS F EFEGLRAKQE-----VTFGLIGGIAVCS : 326
DmCG33296 : SSAS NTLG-----WPNL TF S LLAS T---ATQIFTVLQS F EFEKLERKQE-----VTFGLIGGIALCS : 354
DmCG8291 : I PAV SLAS---D---SISW GA A ATPAF GQLCVMMKPIA G S TGS-----SVLLSCVTLGLLS : 937
AgCP9955 : LLA A AATP-E-HIAP AM G LA VLF GQLCVMMKPIA G A PS-----SILLSCVTLGLLS : 449
DmBLOT : I DRV NGSREGDSHLLQHLVPS I A ILSA V ITVAVYTSTR LPRPN-----YVICLIGLVVA : 580
AgCP2093 : I DR S-VQESD-PTLQRLIPA S T FISS V IYIYITSTR I RHPN-----YTCLAGLVVA : 375
CeSNF6 : FLAE AGVS-----VAPL AG I LL VHA QMVFVETIV S C EYP-----ERLRRNRHVLTVVAVGFLSIFYFVG : 483
CeSNF12 : V PVGTSFMY-----WGGL AT G VLAA DAEFAWLEMIA AFM HFS-----MKNKAVENRLLAFVAVGFLSIFYFVG : 560
AeAeuT : TLPAIFSQTAG-----GTFLG F FFA T SAIMQPMIAF E ELK-----LSRKHAVLWTAIVVFAFS : 390
HsNFT5 : S VE SFLP-----PSVF SF L LA S AIGIMQII P Q TFS-----FFRKHKLLIVGFLSIFYFVG : 550
DmCG13796 : C SYLFGFRFT---TTPNL AF G FLSE CALIIOQMSVI A F EFE-----TLRSRRLVAVGFLSIFYFVG : 479
HspSnfT : GLAN FVAVP-----YGRAVGA A SLAA S SISMLELVP F V EYG-----VSRRRATLAVGFLSIFYFVG : 345
CeSNF1 : W ML VVSY-----LPKL SG LLT CICTMFSVITLALSVL TFE AFG---ENWSKCCRRFLLAVGFLSIFYFVG : 429
CeSNF8 : T PVL RQVA-----ANSWL VF ST VV WK CALIMKTITVTA IFN---IKMKTYPFDCLCIFVSMVIATSG : 421
DmSERT : - PTMTYG VVLVNFV L GPG-LAILFVV--FV AAG F F VDRFSS EQ L SKPGLF----- : 527
AgCP12911 : - PTMTYG VVLVNFV L GPG-LAILFVV--FV AAG F F VENFSA EQ L KPSLFF----- : 469

DmSERT : - PTMTYG VVLVNFV L GPG-LAILFVV--FV AAG F F VDRFSS EQ L SKPGLF----- : 527
AgCP12911 : - PTMTYG VVLVNFV L GPG-LAILFVV--FV AAG F F VENFSA EQ L KPSLFF----- : 469



MsSERT : - PTTYG VYLV L V GPG-LAILFVV--FA AAG C V VDRFSE RT L HTPGWF----- : 494  
HsSERT : - VTLTFG AYVVK LEE ATG-PAVLTV--LI A A S F ITQFCR KE L FSPGWF----- : 536  
CeSERT : - FPAISYG QVFIPL E GVS-LSVLFI--TC M A C F VDQFSK RA L FYPGIY----- : 560  
DmDAT : - ASCTQG FYFFH L R AAG-YSILVAV--FF A A S I TNRFS E RD I FPPGRY----- : 518  
AgCP3649 : - ASCTQG FYFFQ L R AAG-YSILIAV--FF A A S I TERFCN KD I FAPGIY----- : 479  
TnDAT : - TSCTKG FYFFQ L R AAG-YSILVAV--FF A A S I TERFCE RD I FRPGLY----- : 486  
CeDAT1 : - AMCTEG IIMEWLII GTT-WGLLIAV--FC A V A I LRQFVH KE M FRPGNY----- : 512  
HsNET : - FCITKG IYVLT L T AAG-TSILFAV--LM A G S F VDRFSN QQ M FRPGLY----- : 516  
HsDAT : - FCVTNG IYVFT L H AAG-TSILFGV--LI A G A F VGQFSD QQ T QRPSLY----- : 519  
TnOctT : - CINVTGP IYMFH L T AAG-ISLLCSA--LF A A S F LKRFS E EE L FRPGLY----- : 568  
DmCG5549 : - GFTTNS MYWLQ M K AAN-WSVLLIA--IS C L A I SQRFLN QG I KRSPFWN----- : 550  
AgCP10735 : - IFVTNS MYWFO F K AAN-WSVLLIA--IA C L G V CERFIS EE L KRSPFWN----- : 479  
HsGLYT2 : - FPMITQG IYMFQ V T AAS-YALVIA--IF L G S V LQRFCE EM I FQPNIF----- : 676  
HsATB0+ : - VCVTQA IYVWH I H CAG-WGLIAA--IL L G I I GNRFIE TEM I AKRWIFW----- : 522  
HsPROT : - ILTTDG MYWLV L D SAS-FGLMVVV--ITTC A TRV IQRFCR HM L FKPGLY----- : 497  
HsGLYT1 : - PLTSQA IYWLL M N AAS-FSLVVIS--CIMC A M I HRNYFQ QM L FPPPLF----- : 571  
DmCG7075 : - SIMCTNG MYLQ F W SSA-IAVIVIC--LV I M A I IKNFML EF L KRPTLY----- : 476  
AgEhiP6595 : - ACVTQG MYLVQ L S WGS-ISVILVC--IV V T G I CEQFVR EF IDRKVERF----- : 425  
MsPROT : - ICNTEG LHVIG L SHVAI-LCVPLVC--AL I AAV T --ENFSF LF T RPLRRI----- : 457  
DmIne : - PNIIQG IYFQ M H AAS-VTIFLFA--FC M A A F TGRLSK KQ T KAPSFY----- : 510  
AgCP4566 : - PNLIQG IYFQ F H AAS-VSIMFLA--FF T A A F INRLSK KQ T YRPSFY----- : 552  
AmINE : - PNISQG IYFQ I H AAS-ISIMFLA--FF V A S F VRRLCN KE T RVPSSY----- : 759  
MsIne : - PHIIHS IYVFO M Y AAS-LSITYLA--FF V A A F VGRLSR KQ T RQPSLY----- : 620  
TnGAT : - SCISEG MYVFO L S AVSGFCLLFLI--FF C S S A VNRFFYD KE I YYPTIW----- : 507  
MsGAT : - SCISEG MYVFO L S AVSGFCLLFLI--FF C S S A VNRFFYD KE I YYPTIW----- : 508  
AgCP8499 : - TCITEG MYVFO L S AVSGFCLLFLM--FF C S S A VDRFYD KD I YPLSW----- : 470  
DmCG1732 : - FCITQG MYIFQ L S AVSGFCLLWLI--FF C S C C VDRFYD KD I YYPTVW----- : 533  
CeSNF11 : - TTVTEG FYVFO F F AASGWALLWLL--FF C A S SL IDRWEH KS I YPSAW----- : 528  
HsGAT1 : - SNIITQG IYVFK F Y SASGMSLLFLV--FF C S S F VNRFYD QE V SRPCIW----- : 495  
HsGAT3 : - VMLTEG MYIFQ F S AASGMCLLFVA--IF C C G V SNRFYD ED I YRPSLY----- : 511  
HsGAT2 : - IMLTEG MYVFO F Y AASGMCLLFVA--IF S C A V AKRFYD ED I YRPWPL----- : 491  
HsBGT1 : - FLVTEG MYIFQ F Y ASSGICLLFLS--LF V C S V ADRFYD ED I YRPWPL----- : 496  
HsTAUT : - TMTVEG MYVFO F Y AASGVCLLWVA--FF CFV A I GDNLYD ED I YRPGW----- : 503  
HsCT1 : - SMVTDG MYVFO F Y SASGTTLLWQA--FW C V A V ADRFMD AC I YRCPW----- : 518  
CeSNF3 : - PMVTHS SHWLT F A GASGYALLFVV--FF V G A G AHNIRKA HE I VTLPKG----- : 492  
DmCG5226 : - FIFCTGA EYWLK F S AGT-IGLVVVA--LM M A I I HERFTE FQ T YRPGRY----- : 545  
AgCP7001 : - IFTTGA EYWLK F S AGT-IGLVVVA--LM M A I I HEKFTE YQ T IRPGLY----- : 552  
HsNTT7-3 : - IFVQRS NYFVT F D SAT-LPLLVV--IL N A C V IDKFME KD L FAPSRY----- : 568  
HsNTT4 : - LRVQRS NYFVT F D SAT-LPLLVV--IL N A A I TKKFMQE TE L FRPYRF----- : 567  
HsXtrp2 : - TCFTLQS NYWLE F N AAS-LNLLMLA--FL V G V V MKRFCD AW T RRPSPY----- : 515  
RnROSIT : - CFTLQS NYWLE F S AAS-LNLLMLA--FM V G I HV IKRFCD EW T RRPSPY----- : 502  
HsBOAT1 : - FIFTLNS QYWLS L S AGS-IPLLIA--FC MFS V V VDRFNE EF I HKPNIF----- : 529  
HsXT3L : - VFTMEA NYWFD F D AAT-LSLLLV--LV T A C V LRRFES KA T RAVSWY----- : 504  
DmCG10804 : - CFANGA SYIFQ M S AGN-FPLLLIA--LF C S S I VRRFSD EM T SRPNFY----- : 545  
AgCP7501 : - CFANGA SYIFQ M S AGS-YTLLIA--FF C G S I LKRFD EL T SRPSLY----- : 499  
DmCG1698 : - LYITPG QVLLN V Y GVT-FVALVLA--IF L T A I VKRLCR EF I IKTSLY----- : 531  
DmCG4476 : - IYITPG QHII T M FHGVT-FVSLVSA--IF L A G I TKRLCQ AEY LNIKTSNY----- : 505  
DmCG15279 : - MYITPG QVMLT V F GAS-MIALVLG--IA LYT G I TDRLCK EF L RKVGLY----- : 519  
AgCP10412 : - SIYVTPG QYVVK V Y GAS-SIALVLA--IA L A G V VDRLCK TEF L HRPNLY----- : 535  
DmCG8850 : - VYMTPG QVPLN V F GCT-FIALVLA--IA L A G I VKRIS EF LNVKTSFY----- : 498  
DmCG15088 : - VYMTPG QVPLN V F GVS-FTALILA--IG L A A I VNRFCE KF M IETGWY----- : 465  
AgCP10401 : - IYVTPG QVLDL F GAS-FVALVLA--VF MFTFA I VSRICR EF L IKTGly----- : 537  
AgCP10503 : - VYITP----- : 439  
AgCP10610 : ----- : -  
DmCG3252 : - VYVTPG QWILT V F GGT-YVVFILA--IF LAG V V LQNFCD EF CNRRVSLY----- : 512  
AeAAT : - VYITPG QWILD V H GGT-FLIYVLA--II M A F I LDNWCN EF VQRRVGLY----- : 500  
MsKAAT1 : - VYCTPG QFILE V H GGT-FLVLFCA--IS LAG F I LENLCL EF M KKTGFY----- : 503  
MsCAATCH1 : - VYCTPG QYILE V H GGT-FLVLFCA--IS LAG F I LENLCL EF L KKTGAY----- : 503  
BcProT : - VFATRG LMFLY G Y ANN-FGLITIA--LA V T GLILRRLPVYQ HANFVSDIKLRT----- : 426  
HinSnfT : V LFGTPT LPMLD F K VNY-FGIVAVA--FASL A -VANEKLGLLG HLNETSDFKVG----- : 434  
StTnaT : - ALFATGA VHILD V H VGS-YAIAIILG--LV A V G IM TARIR-EHVN TSDIRVGM----- : 430  
CeSNF2 : - VMTTDA FYWFD F E STG-ISATFGT--AIMC T F C T NHFRA AV W EPESWLATCL-----GPA : 614  
CeSNF7 : - VMTSDA FYWFD Y E SAG-VSATFGT--AIMC T C C LDNFRA RE WEEEPESRLDKYL-----GPG : 504  
CeSNF4 : - ILTTDA IYWFA F E GSG-FGALISA--TSMC I S I MRNFRA AS L RG-TRISNIF-----GHN : 475  
CeSNF5 : - VFSTRA YWFE F E AAG-FSSVCTV--VC L V M I FRNVRD TE V HARNKFTGAI-----GAH : 577  
CeSNF9 : - VMVTDG FYWFI F E AAG-VSSCCAV--TA V I A V RRNQLA KE F ESKNKCTSWI-----GQH : 547  
DmCG13794 : - FYFCTNH ITFFSALALDAIF--SHSLH--LLLL V L I RERFQR EF L QPFASWK----- : 489  
DmCG13795 : - FYFCTNH IIFFSALGLDAIF--SHSLH--LLLL V L I RERFQR EF L QPFASWK----- : 485  
DmCG13793 : - YFCTNH VLYFA LSIDAIF--SQPLLH--LLLL V L V RDRFQK KF L QPFASWK----- : 385  
DmCG33296 : - FFFCTEH SFYFA LGIDVIL--AHSLH--LLLL V L I QKRFQR EF L QPFASWK----- : 413  
DmCG8291 : - PFATEM ISILYV FLLGGSWFPII--TA IFG FLIR RPYNG VNDLRLCGSMS----- : 997  
AgCP9955 : - PLATEG IHIVHYL TILGAAWLLLLLW--IGHI A FLVR RPFSTDI VNDLQMLQSF----- : 509  
DmBLOT : - ISFAAPKFLIAR L SRLVG-TMVVTAL--VF L A T I AKNIYT EFSI RPIFRV----- : 639  
AgCP2093 : - AGLLCPNYIFPR L TRIVG-ALAVCAM--IF I L I V SKNLYT EFSL RPVLKA----- : 434  
CeSNF6 : - FBCLSS LFWME LTQ VLT-WPLVVA--FL C A N V VDNMLD AKW V YWPCY----- : 542  
CeSNF12 : - PLCAQG IFVPHAIENLNAN-WNSFSLA--LLSVAI C V IDNYLT SA LRVPRIQISKATRLKEKLIYFFGPG : 636  
AaLeuT : H VMPLNKS---LDEM F AGT-IGVVFFG--LT L IF F I ADKAWEE NRGGIKVPRI----- : 447  
HsNTT5 : - FTRTPS SYFIR LSD WIVFPIIVVVV--F T A S A ARFLA TI L HPISPI----- : 609  
DmCG13796 : S YFCTQL FSQTLQPLNLA--THLLISG---ILV MTT V RVRFOC QF L KTISSPK----- : 539  
HspSnfT : VPTVTSATLSPG F FVAGTLDVLLMTAGLIGL FTG VV QNAVTEYELGAGAVASALSTP----- : 410  
CeSNF1 : - YFATNA RHAYE ATRSIKYCTIYFILT--A LFATA F CAHKLKG HN MRSRCCSCFG----- : 490  
CeSNF8 : - FLCFES PLYLT IQDIQTVATGIFITA---LFI LEL LDITYG HE M QPSNDQWSR-----AYT : 485

\*                    1220                    \*                    1240                    \*                    1260                    \*                    1280

DmSERT : ---WRIC TYIS VFLLT FIFSIMG-----YK-----EMLGEEY DWSYQVGWA TCSSV C MYI : 584  
AgCP12911 : ---WRIC KYIS TFLFC LIFSLG-----YE-----EMLGEEY EWSVAAGWA TLSSV C TYM : 526  
MsSERT : ---WRTC SYIS VELLV FVFSVLA-----HE-----EMLGGEYT SWSITVGMV TGTVSC LYI : 551  
HsSERT : ---WRIC VAIS LFLLF ICSFLMS-----PP-----QLRFLQYN YWSIILGYC GTSSF C TYI : 593  
CeSERT : ---WRVC T-CS VFISV FIMTVYNS---FK-----PIQMASYT WWSVILGWLF RLSSV A VFA : 618



CeDAT1 : IYKFNARG-----N--TISEKWQVMTMPYKRPN----- : 597  
HsNET : IYKFLSTQ-----G--SLWERLAYGITPENEHLL----- : 600  
HsDAT : AYKFCSLP-----G--SPREKLAYAIAPKEDREL----- : 603  
TnOctT : IYKLIISTP-----G--TFREVRACCISPESEHEA----- : 652  
DmCG5549 : FQEIWRAPMS-----L--SFRDRIKHLLQPTAEWGPA-----G : 640  
AgCP10735 : VKNM----- : 542  
HsGLYT2 : VIKMHLAPG-----RFIERLKLVCSPQPDWGP----- : 760  
HsATB0+ : IIKIQAKG-----NIFQRLISCCRPASNWGP----- : 608  
HsPROT : LVAVLREEG-----SLWERLQQASRPAMDWGP----- : 581  
HsGLYT1 : MFRLCRTGD-----TLLQRLKNAATKPSRDWGP----- : 656  
DmCG7075 : FYIMIRKRA-----TLCDSLKKRLKP-LDWTP----- : 559  
AgEbiP6595 : SYKLLHTKG-----SL----- : 493  
MsPROT : AKVLLRAEG-----SLLERIRASCRPSNDWGP----- : 528  
DmIne : VINFLRSSGD-----TFWERIRNTLRPN-IYECKI----- : 598  
AgCP4566 : VYQIVRAEGN-----TFGEKLLNLTLPN-IYECKI----- : 640  
AmINE : IYEFIRANGN-----TCAEKLRSNIKPH-FEACKI----- : 847  
MsIne : VIVIRAPGD-----SLREKLRYSIQPTSIKCEGV----- : 707  
TnGAT : VYLWRVTPG-----TWQEKFNKIVRIP-EDVPS----- : 591  
MsGAT : IYLWRVTPG-----TWQEKFKIVRIP-EDVPS----- : 592  
AgCP8499 : ----- : -  
DmCG1732 : FWLWKRTPG-----ELSEKIRALVRID-EDVTR----- : 617  
CeSNF11 : IWVWFKTPG-----TVQEKIKLLCRPD-IEIKG----- : 614  
HsGAT1 : AYMFLLTKG-----SLKQRIQVMVQPS-EDIVR----- : 579  
HsGAT3 : CITVWKTEG-----TLEKQLKLTTPS-TDLKM-----RGKLGVSPRM : 606  
HsGAT2 : LYRLGTLKG-----PFRERIRQLMCPA-EDLPQ-----RNPAGPS-- : 583  
HsBGT1 : VTLLKTRG-----PFRKRLRHVITPD-SSLPQ-----PKQHPCLDGSAGRN-- : 595  
HsTAUT : VIRLQCTEG-----PFLVRVKYLLTTPR-EPNRW-----AVEREGAT-- : 596  
HsCT1 : LGCLLRAG-----TMAERWQHLLTQPI-WGLHH-----LEYRAQDADVRG-- : 615  
CeSNF3 : IYVLFNTKH-----L--TLKERVRKGLNLD-GSFES----- : 579  
DmCG5226 : LMRSPQCLK-----VDLDIHQGSIRRNETTASTKE----- : 644  
AgCP7001 : LLRRFQILK-----VDLDIHQGSIRRNETTASTKE----- : 651  
HsNFT7-3 : IVRRFNLDLDD-----SSGNLASVTYKRGVLEKPEVNL-ED- : 672  
HsNFT4 : VLRHFPLLSD-----GS-NTLSSVSYKGRMKDINSLEND----- : 671  
HsXtrp2 : LA-----QLTRRRRTWRDRDARPDTD----- : 608  
RnROSIT : LA-----QLLFQYRQWKN----- : 588  
HsBOAT1 : IY-----KLRNHCQKPGDHQ----- : 618  
HsXT3L : LG-----TFVQRLLK----- : 584  
DmCG10804 : VLRLCGIKVV-----ED--SDPAWFPPEALRELVHIVPHE----- : 647  
AgCP7501 : ILRLCGINVI-----ED--SEPAWFPPEALRELVHIVPHE----- : 601  
DmCG1698 : IPAVRKQPSH-----L--GLWARIRKAFPLPNWGP----- : 617  
DmCG4476 : GYANFKQPKG-----S--LKSRIINNSIKPHSDWGP----- : 590  
DmCG15279 : IVAIVRDPGQ-----T--LGAKIRGAFTPKKNWGP----- : 604  
AgCP10412 : AYAVYKQSGK-----S--LNEKIKNAFKPTAAWGP----- : 620  
DmCG8850 : LVTIYQPGK-----T--FGSKFGLAMQPTANWGP----- : 583  
DmCG15088 : IYAIYKRGH-----NGS--FWQIRAAKCPSDTWGP----- : 552  
AgCP10401 : IYTFLLKRKEP-----D--WRDRLLHCFKPTHWGP----- : 622  
AgCP10503 : ----- : -  
AgCP10610 : ----- : -  
DmCG3252 : LWYISRHP-Q-----GT--YWKSLKASLKPSDRWGP----- : 597  
AeAAT : VWTVTGHSTK-----ES--VWKITKAAKPSQWGP----- : 586  
MsKAAT1 : VVTLKYR-T-----GN--FRETVKKAFHSPKSWGP----- : 589  
MsCAATCH1 : GFSLYKYR-T-----GT--FSETIKKAFHSPKSWGP----- : 589  
BcProT : GLKKWDAQI-----HSDSAKLEKEWDRGVS----- : 508  
HinSnfT : SRLKWKSET-----KLTIDETKGE----- : 508  
StTnaT : ---QWRSQ-----LDVGTGVEG----- : 501  
CeSNF2 : IAAVINTRRM-----NIPIRGAFMLQKQHPHYDRISEQWPEWK----- : 713  
CeSNF7 : GFAIKRFKDL-----NIPIRGVFMVQKQHPAYDRISEKWPEWK----- : 603  
CeSNF4 : LCTYMDFKKR-----GPIRGMFLQKSHPSYPRISRGWSEEK----- : 573  
CeSNF5 : LLNFIRCRNR-----GHSYRTLFLMLQKQHASYRRIAANYSKDQ----- : 675  
CeSNF9 : FCAFVEFKRK-----GHDLRALFMKQRLKSFGRITYQTMASKD----- : 648  
DmCG13794 : IYIYRSTG-----SFCERFKRTRCPDWDYPVEM----- : 571  
DmCG13795 : FYYLYQSTG-----TFCERFKRTRCPDWDYPVEM----- : 567  
DmCG13793 : FYYIHQRPG-----SISELFR--ATDWPVEM----- : 463  
DmCG33296 : FYYLRQSPG-----TFLERLKRSCRPTDWDYPVEM----- : 495  
DmCG8291 : IIQIYRYLAH-----GPPDILLERIQLLYRPPPEEGEPRRPSARQTASQSRNALGQTTEGGQ : 1117  
AgCP9955 : VIQIYRYLCR-----GPPDILLTRRAAVSSPLQRTNRNGGAAGGRDSSEP--TITISMDSRS : 623  
DmBLOT : QVEYNFFGMI-----CEASKPAKEWGPADPLARHSWKQWRSVQDTRGRDFTLR----- : 744  
AgCP2093 : QVDYNIFSMI-----HGATKPSKDWGPADPLVRHAWKQWKSVCEDTGERDFTLR----- : 539  
CeSNF6 : IWQFCIAKG-----TITQKWVRYLYPDDAWGPAMAHR----- : 634  
CeSNF12 : FMQIRQIRNEGKLLKSLFDTSSEWESQDSDLEPKDLYMRQSGKFESPPNRRRTPTIFTHRENTYMYIDSRGPTVRSRVFP : 776  
AaLeuT : FLTFLVFLAE-----RRRNHESA----- : 513  
HsNFT5 : VYCRHRIFF-----RPSKSGDPMTASTSLPLSHQLTP----- : 711  
DmCG13796 : VYKVCRTNG-----DWRQRLQQLAPHDWHPVDAD----- : 617  
HspSnfT : VIAVVSFLG-----LRRPASVA----- : 477  
CeSNF1 : FMTILCACCK-----GKDHAKTCKRIRTVFAARLQNHHR----- : 585  
CeSNF8 : YYSYLKMMWHFKEFEKEKQFNSSILLWHRQLHLRSWNRIEKEMSEEQRNAFHQHLPEK----- : 601

\* 1380 \* 1400 \* 1420 \* 1440  
DmSERT : ----VVPQQ--GTSV----- : 622  
AgCP12911 : ----AIPGQLYGTAV----- : 565  
MsSERT : ----IPPADSTLCNL----- : 587  
HsSERT : ----EIPCGDIRLNAV----- : 630  
CeSERT : ATSLAADPTQIIDSSLLD--PIHTLTPV----- : 671  
DmDAT : MVLNG-VTTEVTVRLT--DTETAKEPVDV----- : 631  
AgCP3649 : MAVNG-VSIEPAQVRLT--HSDEGEE----- : 585  
TnDAT : TVHNGMVVSEGGVRLASTVQTPTSQPPPTPLASAPALV----- : 612  
CeDAT1 : ----QTEYIPIPTT----QPHSDIML----- : 615  
HsNET : ----VAQRD-IRQFQ--LQHWLAI----- : 617  
HsDAT : ----VDRGE-VRQFT--LRHWLKV----- : 620

TnOctT : -----IRGGAPVSRFS----WRHWLYV----- : 670  
DmCG5549 : RPCVNLHAERYQSQNYDGVNTNKILIEEESPKQAANKKDRQLTLDVDVDECLPLATVQKRPAGPKSMGKQSSGNLQT : 720  
AgCP10735 : ----- : -  
HsGLYT2 : -----FLAQHRGERYKMMIDPLGT-----SSLGKLPVKDLELGTQC----- : 797  
HsATB0+ : -----YLEQHRGERYKDMVDPK-----KEADHEIPTVSGSRKPE----- : 642  
HsPROT : -----SLEENRTGMYVATLAGSQSPKPLMVHMRKYGGITSFENTAIEVDREIAEEEEESMM----- : 636  
HsGLYT1 : -----ALLEHRTGRYAPTIIAPSPEDGFVQPLHPDK-----AQIPIVGSNGSSRLQDSRI----- : 706  
DmCG7075 : -----ADPQDRAEYEAFFRRQR-----RMPGFMSDMDMESSK----- : 590  
AgEbiP6595 : ----- : -  
MsPROT : -----SEPEKRREWELLLKQK-----ADIFPLNDLDKY----- : 556  
DmIne : -----CGEHHCEHDYPEQEQFMAQEMATVYKPTNPHLLNLGQCKGYNAMQASPSHAEAGGPCGQ----- : 658  
AgCP4566 : -----CGEHHCDHDFDDE----- : 654  
AmINE : -----CGQEYCNESMHNFEEDMIKEQQD---PNSPIQLSISPLSKSQT----- : 888  
MsIne : -----NGCDICCESDPPDKTVIN----- : 727  
TnGAT : -----LRTKMQAEEQAKHGCKA----- : 608  
MsGAT : -----LRTKM----- : 597  
AgCP8499 : ----- : -  
DmCG1732 : -----LREKMLREAYAKEIEFNLSL----- : 636  
CeSNF11 : -----AMENAENLELVEDFQNAI----- : 632  
HsGAT1 : -----PENGEQPPQAGSSTSKEAYI----- : 599  
HsGAT3 : VTVNDCDAKLKSDGTIAAITTEKETHF----- : 632  
HsGAT2 : -----APATPRTSLRLTELESHC----- : 602  
HsBGT1 : -----FGPSPTREGLTAGEKETHL----- : 614  
HsTAUT : -----PNSRTVMNGALVKPTHIIVETMM----- : 620  
HsCT1 : -----LTTLPVSESSKVVVSVSM----- : 635  
CeSNF3 : -----PAKKNLVNNAEELFKIESSSQ----- : 600  
DmCG5226 : ---MIDNDDNMSDMPDPPQDSLAAATRFITIGDFEN----- : 675  
AgCP7001 : ---MID-EDDVVSPDLPPQSLAATRFITIGDFDV----- : 681  
HsNTT7-3 : ---DTSLIHGKIPSEMPSPNFKNIYRKQSGSPTLDTA--PNRGYIGYLMADIMPDPESDL----- : 730  
HsNTT4 : ---ETRFLLSKVPEEAPSPMPTHRSYLGPGSTSPLETSGNPNRGYSGYLLAS---TPESEL----- : 727  
HsXtrp2 : ---MRPDTDRPDTDMRPDTDMR----- : 628  
RnROSIT : -----THLESALKPQESRGC----- : 603  
HsBOAT1 : -----LVSTLSTASMGDLKY----- : 634  
HsXT3L : -----RGDADPVA----- : 592  
DmCG10804 : ---PTELEERSIFCFNMDGTGEMCCPKYGLPEKSLEEEE----- : 682  
AgCP7501 : ---PTDIEISLFCIRADGSEGLCCPIYGPREQPLDEEE----- : 636  
DmCG1698 : -----SDPQTLKRYQLFVQ---EGNANALFRSSIWHKIYDNIIFG----- : 654  
DmCG4476 : -----SDPKKLMYQMFRLRNKDEENSKNVNRR--CVCYTAGDRIFG----- : 629  
DmCG15279 : -----SDPLLREQYHKEIE---NELTPKRGQGIWAAIKQNIIFG----- : 639  
AgCP10412 : -----IDPATHYEYKFFID---ED----- : 636  
DmCG8850 : -----LQTKQFEAYIIHRK-----READFKSPRGYGLFDNIIFG----- : 616  
DmCG15088 : -----ANVQ-LDATI----- : 561  
AgCP10401 : -----EDPELNAKYHESVY---KHEQSLPRDRSLVRMFDNVFS----- : 658  
AgCP10503 : ----- : -  
AgCP10610 : ----- : -  
DmCG3252 : -----ANPEIRREWVIFKN--QKAAQRATQKDTSKLGFWRKVNFCGSNK----- : 641  
AeAAT : -----CTEHTRKAWLKYKE--EAKSRRDDIIYAKNHSSIVRKLCLVFGMYDDFIQLDSTRL----- : 640  
MsKAAT1 : -----RSPRLREWMQFRI--EAKALR--QKMNTSRVKHLWYSITGAYRRNINK----- : 634  
MsCAATCH1 : -----RSPRERREWQFKA--EAKALR--QKMNTSRVKHLWYSITGAYRRNIN----- : 633  
BcProT : ----- : -  
HinSnfT : ----- : -  
StTnaT : ----- : -  
CeSNF2 : -----KAI GDKLPEQEPGDDDLNMDIGLETKETWSFLYKPLSIRVFNKAKI VIETGKYLK----- : 768  
CeSNF7 : -----K I GDKLPELEPEGEDDL--EDGLEGSDD-DLFYEY----- : 635  
CeSNF4 : -----RSVGAQLPSFEPGEEEFKSQEPGYGPV----- : 600  
CeSNF5 : -----LALQDQLPDKEPWEDE---NVDLTDSESESRNAASGDVPIDDVATIDTSSTYHQVY----- : 728  
CeSNF9 : -----RAKQKILPDREPWDDAPTGHQFVASPDFQSSSQSTRS-ERPERSNYLNSAASVHSKTQ----- : 706  
DmCG13794 : -----EHRQQYEEAVSNTDMTQQLTAGPN----- : 595  
DmCG13795 : -----EHRQQYEEAVDSTEMTHQLFEVTEEVN----- : 594  
DmCG13793 : -----EYRQQYGELVQKSEMTHQLFDRTEEVN----- : 490  
DmCG33296 : -----KHRQYHGDVGNIEMTDHLFNGIEEVN----- : 522  
DmCG8291 : HDAQNADAPPKYTPPPSYTTATGARLAKLLRQSI RRSVRRVLGDSSRTRPVLSLDAESASPAAPPDYLTILNTPAGSSFNA : 1197  
AgCP9955 : VRDADDAPPKYTPPPSYTTATGARLAKLLRNSIRRSVRRVLGGETSGSAP--SVRQRTALPQTAPDSL----- : 688  
DmBLOT : RRGTRDYTHSIKKGQYSSATKYGVQNGGQTQTPMHQHQHWSSTPGNSSPNYSGSMFGDSAI EEDISVDFPFGITQQQYVP : 824  
AgCP2093 : RRGTKDYTNSIKKGQYTHSHKYGASS-----NRNLSTGTSGNSPNYSGSVFGDSAI EEDMSVEKYP----- : 599  
CeSNF6 : AEKFLQIPEARRLLPPEVEIASRGLVQEEMPMYSYDNTSSAADVRSNRSTGHGATDVRVAATNNTIPKFERETAI- : 713  
CeSNF12 : LGASLDPYGWKAGRLDRDQQQI EETASNYSEEDSATNNSFMATVKHNDMELTLFGSPPAI LGDDEKIMTTRFSESMPV : 856  
AaLeuT : ----- : -  
HsNTT5 : -----SKEVQKEEILQVDETKYPSTCNVTS----- : 736  
DmCG13796 : -----NRRFYEEIMGISEMLVIDANANTT----- : 641  
HspSnfT : ----- : -  
CeSNF1 : -----DEKAQPIPRYTSNAPGYLLLPQAPLAPEPEIYA----- : 617  
CeSNF8 : ---ESEDLEEFWKLVRPFPEFTHSETVMMPIIPKSKEIVKPRQASKRKRKSKKDEFLTFETYKKEGK----- : 669

\* 1460 \* 1480 \* 1500 \* 1520  
DmCG5549 : NAKNGTHKSPTEISPSKQPLIPSEKDNKSANI KVLAAGTQSSGRECTTMTSTFAGGGEKSKTKVNLATGATSAAPLASS : 800  
DmCG8291 : DPSPASSSSPESIEIDQRPVGYQSRQSLGRKLRHRSAGSTLERRPYTAEDVVTILRSSVRRHQSQGGNLTASTLPRPP : 1277  
DmBLOT : FQASDTKQSRYSQRIRQTAQPQTQTMRRPRESEVKEHREVVYIRRLSDGGTGTGHATRIEISPSNESITYGNGKSNMNS : 904  
CeSNF12 : NYKCRNVEVPRI PNKLQNMERMARKTRKRSSPASDPPVPTSLPPPQLQHCSEPPMNSKESHSP EITPGDDSP : 936

\* 1540 \* 1560 \* 1580 \* 1600  
DmCG5549 : NIATAATAVAATKSPPAAGVAPTTLVGIATGTTVAAAANGKATLKTITISSGAVATAAGGNKAASVAAATVAAGKTP : 880  
DmCG8291 : TAAMSTHLEDASFRSIEENLVNAEPPDRTPGVLELELEAGQAEECARNNTSVI----- : 1331  
DmBLOT : SAMSRLPGRSTGCLAAPASRRRACMSSGRPARWSMLTHTSCHLQRRPVERRITFAGGSSTSIKRSIPRSC EIRGGPGV : 984  
CeSNF12 : SISNSSDDSSDDCFRRATVIRKRTSDDDAFTHFSTATAESISITPLDFPQRQSLSSVAIYDQEQNGRSKVLSQLKRPK : 1016

\* 1620 \* 1640 \* 1660 \* 1680  
DmCG5549 : AAAMALNVAASVKVSTDKTAPLKTITISASVATAPGGTKASTAPSATAAVVSVSSGATVPAASKAAAPL KATAPPPSL : 960  
DmBLOT : GNEAFTFKTKCICQNKRICICNPYCIYGTPNYKLRITNHVHTESVSSSCEL----- : 1035

CeSNF12 : IDMPPK----- : 1022

                  \*      1700          \*      1720          \*      1740          \*      1760

DmCG5549 : AAVNATQAAKSTPVAAASALVASAPPAKSTSASVAKTNTNAGSKPTATLSTCKSDAPAAAASVGVINGKVTPSGGATSTA : 1040

                  \*      1780          \*      1800          \*      1820          \*      1840

DmCG5549 : QPAVNQKATAAAAAQKSQGATATATPTKKTALNQKTIVPSAGSGKGDVPSSRSTDTPPPAKGAASSNISAKSTKAGKTQAN : 1120

                  \*      1860          \*      1880          \*      1900          \*      1920

DmCG5549 : ETSPTKSAVKPIKAGSTSPTKAVVSKPGASAAKTAASAATATATAAGSSSNKKAATSTAATLGSQAKTKGNASGAGTK : 1200

DmCG5549 : K : 1201

Table S1. Sequence names, abbreviations, and accession numbers used for phylogenetic analysis

Species	Sequence name	Abbreviation	GenBank accession no.	
<b>INSECTS</b>				
<i>Drosophila melanogaster</i>	Serotonin transporter	Dm SERT	NP_523846	
	Dopamine transporter	Dm DAT	NP_523763	
	Inebriated	Dm Ine	NP_523763	
	Bloated tubules	Dm Blot	CAB53640	
	CG1698	Dm CG1698	NP_610517	
	CG1732	Dm CG1732	NP_651930	
	CG3252	Dm CG3252	NP_572219	
	CG4476	Dm CG4476	NP_648293	
	CG5226	Dm CG5226	NP_611340	
	CG5549	Dm CG5549	NP_611836	
	CG7075	Dm CG7075	NP_609135	
	CG8291	Dm CG8291	NP_611064	
	CG8850	Dm CG8850	NP_610755	
	CG10804	Dm CG10804	NP_726868	
	CG13793	Dm CG13793	NP_609136	
	CG13794 (orf)	Dm CG13794	AAAY55020	
	CG13795 (orf)	Dm CG13795	AAQ22494	
	CG13796 (orf)	Dm CG13796	AAAN71401	
	CG33296	Dm CG33296	NP_995644	
	CG15088	Dm CG15088	NP_611364	
	CG15279	Dm CG15279	NP_609726	
	<i>Anopheles gambiae</i>	EbiP6595	Ag Ebi6595	XP_316980.2
		CP2093	Ag CP2093	XP_315020.2
		CP3649	Ag CP3649	XP_308462.2
		CP4566	Ag CP4566	XP_319694.2
		CP7001	Ag CP7001	XP_309367.2
		CP7501	Ag CP7501	EAA06912.3
		CP8499	Ag CP8499	XP_318854.2
		CP9955	Ag CP9955	XP_320247.2
		CP10401	Ag CP10401	XP_309846.2
		CP10412	Ag CP10412	XP_309840.2
		CP10503	Ag CP10503	XP_309844.2
CP10735		Ag CP10735	XP_311463.2	
CP10610		Ag CP10610	XM_309845.2	
CP12911		Ag CP12911	XP_310113.2	
<i>Manduca sexta</i>		Potassium AA transporter 1	Ms KAAT1	AAC24190.1
		CAATCH1	Ms CAATCH1	AAF18560.1
		GABA transporter	Ms GAT	AAA92342.1
	Proline transporter	Ms PROT	AAP94997.1	
	Serotonin transporter	Ms SERT	AAAN59781.1	
	Inebriated homolog	Ms Ine	AAF21642.1	
<i>Trichoplusia ni</i>	GABA transporter	Tn GAT	AAF70819.1	
	Octopamine transporter	Tn OctT	AAL09578.1	
	Dopamine transporter	Tn DAT	AAAN52844.1	
<i>Apis mellifera</i>	Inebriated homolog	Am INE	XP_395197.2	
<i>Aedes aegypti</i>	Amino acid transporter	Ae AAT	AF543193.1	
<b>MAMMALS</b>				
<i>Homo sapiens</i>	GABA transporter 1 (SLC6A1)	Hs GAT1	NP_003033.2	
	Norepi transporter (SLC6A2)	Hs NET	NP_001034.1	
	Dopamine transporter (SLC6A3)	Hs DAT	NP_001035.1	
	Serotonin transporter (SLC6A4)	Hs SERT	NP_001036.1	
	Glycine transporter 2 (SLC6A5)	Hs GLYT2	NP_004202.1	
	Taurine transporter (SLC6A6)	Hs TAUT	NP_003034.2	
	Proline transporter (SLC6A7)	Hs PROT	NP_055043.1	
	Creatine transporter (SLC6A8)	Hs CT1	NP_005620.1	
	Glycine transporter 1 (SLC6A9)	Hs GLYT1	NP_964012.2	
	GABA transporter 3 (SLC6A11)	Hs GAT3	NP_055044.1	
	Betaine transporter (SLC6A12)	Hs BGT1	NP_003035.2	
	GABA transporter 2 (SLC6A13)	Hs GAT2	NP_057699.2	
	AA transporter B0+ (SLC6A14)	Hs ATB0+	NP_009162.1	
	NT transporter 7-3 (SLC6A15)	Hs NTT7-3	NP_877499.1	
	NT transporter 5 (SLC6A16)	Hs NTT5	NP_054756.2	
	Transporter Xtrp2 (SLC6A17)	Hs Xtrp2	NP_872438.1	
	B0 AA transporter 1 (SLC6A19)	Hs BOAT1	NP_001003841.1	
	Transporter XT3L (SLC6A20)	Hs XT3L	NP_064593.1	
	<i>Rattus norvegicus</i>	ROSIT	Rn ROSIT	U12973
	<b>WORM</b>			
<i>Caenorhabditis elegans</i>	Dopamine transporter	Ce DAT1	NP_499043	
	Serotonin transporter	Ce SERT	AAK84832	
	SNF family transporter 1	Ce SNF1	NP_491491	
	SNF family transporter 2	Ce SNF2	NP_492396	
	SNF family transporter 3	Ce SNF3	NP_493910	
	SNF family transporter 4	Ce SNF4	NP_496730	
	SNF family transporter 5	Ce SNF5	NP_496735	
	SNF family transporter 6	Ce SNF6	NP_497416	
	SNF family transporter 7	Ce SNF7	NP_499702	
	SNF family transporter 8	Ce SNF8	NP_502271	
	SNF family transporter 9	Ce SNF9	NP_503077	
	SNF family transporter 11	Ce SNF11	NP_505873	
	SNF family transporter 12	Ce SNF12	NP_509527	
<b>PROKARYOTES</b>				
<i>Aquifex aeolicus</i>	Leucine transporter	Aa LeuT	NP_214423.1	
<i>S. thermophilum</i>	NaCl-dep. transporter	St TnaT	YP_074271.1	
<i>Bacillus cereus</i>	Na/Proline symporter	Bc ProT	NP_833789.1	
<i>H. influenzae</i>	SNF family transporter	Hin SnfT	ZP_00154505.1	
<i>Halobacterium sp.</i>	SNF family transporter	Hsp SnfT	AAG19816.1	