

## SHORT COMMUNICATION

## Muscle membrane potential and insect chill coma

Jonas L. Andersen\*, Heath A. MacMillan and Johannes Overgaard

## ABSTRACT

Chill-susceptible insects enter a reversible paralytic state, termed chill coma, at mild low temperatures. Chill coma is caused by neuromuscular impairment, allegedly triggered by cold-induced depolarization of muscle resting membrane potential ( $V_m$ ). We used five *Drosophila* species that vary in cold tolerance (chill coma temperature spanning  $\sim 11^\circ\text{C}$ ) and repeatedly measured muscle  $V_m$  during a downward temperature ramp (20 to  $-3^\circ\text{C}$ ). Cold-tolerant species were able to defend their  $V_m$  down to lower temperatures, which is not explained by species-specific differences in initial  $V_m$  at  $20^\circ\text{C}$ , but by cold-tolerant drosophilids defending  $V_m$  across a broad range of temperatures. We found support for a previously suggested ‘critical threshold’ of  $V_m$ , related to chill coma, in three of the five species. Interestingly, the cold-tolerant *Drosophila* species may enter coma as a result of processes unrelated to muscle depolarization as their  $V_m$  was not significantly depolarized at their chill coma temperatures.

**KEY WORDS:** Chill tolerance, Critical thermal minimum, Cold exposure, Comparative, Inter-species

## INTRODUCTION

Many insects succumb to the effects of cold at temperatures considerably higher than those causing them to freeze (Hosler et al., 2000; Košťál et al., 2006; MacMillan and Sinclair, 2011a; Andersen et al., 2013, 2015b); however, some enter a reversible paralytic state at low temperature, termed chill coma, which is caused by neuromuscular impairment. Insects in chill coma may further develop chill injury and ultimately die if the cold exposure is severe (Košťál et al., 2004, 2006; MacMillan and Sinclair, 2011a). The onset temperature of chill coma (the critical thermal minimum,  $CT_{\min}$ ) and chill coma recovery time upon rewarming (CCRT) are both metrics that correlate closely to interspecific variance in insect distribution, and are widely used to assess insect cold tolerance (Hori and Kimura, 1998; MacMillan and Sinclair, 2011b; Andersen et al., 2015b).

The physiological mechanisms of chill coma remain largely unresolved. However, cold-induced neuromuscular impairment has been associated with chill coma and linked to a decrease of excitability due to a depolarization of either the central nervous system (CNS) and/or the muscle tissue (Wareham et al., 1974; Hosler et al., 2000; Rodgers et al., 2010; MacMillan et al., 2014). Low temperatures induce muscle depolarization in several chill-sensitive insects, including the hawk moth (Wareham et al., 1975), American cockroach (Wareham et al., 1974), vinegar fly, honey bee (Hosler et al., 2000) and migratory locust (MacMillan et al., 2014).

*In vitro*, such depolarization can directly decrease muscle force production, even at high temperatures (Findsen et al., 2014), possibly because of a decrease in voltage-sensitive  $\text{Ca}^{2+}$ -channel currents (Salkoff and Wyman, 1983). Accordingly, it has been suggested that cold-induced depolarization directly causes chill coma (Hosler et al., 2000; Findsen et al., 2014; MacMillan et al., 2014). Onset of chill coma has been repeatedly found to coincide with a ‘critical threshold’ muscle  $V_m$ , between  $-35$  and  $-45$  mV (Wareham et al., 1974; Hosler et al., 2000; MacMillan et al., 2014). Similarly, there seems to be a tight association between the recovery from chill coma and the recovery of membrane potential when the insect is returned to normal temperatures after cold exposure. Prolonged cold exposure causes increased  $[\text{K}^+]_{\text{ext}}$ , which depolarizes muscle cells, and insects that lose  $\text{K}^+$  balance during cold exposure only recover from chill coma when  $\text{K}^+$  balance is restored (MacMillan et al., 2012; Andersen et al., 2013; Findsen et al., 2013).

Here, we test three hypotheses regarding chill coma onset and muscle membrane potential: (1) more cold-tolerant species defend  $V_m$  during cold exposure, either by having a more polarized baseline (initial)  $V_m$  or by an improved ability to maintain  $V_m$  when exposed to low temperatures. (2) Chill coma onset occurs at a critical depolarization of muscle  $V_m$  in insects. (3) Cold-sensitive species lose  $\text{K}^+$  balance during a cold exposure, which leaves the muscle membranes in a depolarized state after rewarming such that chill coma recovery is slowed. To test these hypotheses, we used five chill-sensitive *Drosophila* species with markedly different cold tolerance: *Drosophila birchii* (Dobzhansky and Mather 1961), *Drosophila equinoxialis* (Dobzhansky 1946), *Drosophila melanogaster* (Meigen 1830), *Drosophila persimilis* (Dobzhansky and Epling 1944) and *Drosophila montana* (Patterson and Wheeler 1942). For all species, we repeatedly measured muscle  $V_m$  while exposing flies to a temperature ramp from the rearing temperature ( $20^\circ\text{C}$ ) to  $-3^\circ\text{C}$  and following return to  $20^\circ\text{C}$ .

## RESULTS AND DISCUSSION

The five species of *Drosophila* examined varied widely in cold tolerance (Andersen et al., 2015b), and their chill coma onset temperatures ( $CT_{\min}$ ) were significantly different ( $H=93.38$ ;  $P<0.001$ , *post hoc* tests;  $P<0.001$ , in all cases) and ranged almost  $11^\circ\text{C}$  between the least and most cold tolerant (from  $8.8\pm 0.2$  to  $-2.0\pm 0.1^\circ\text{C}$ ; Fig. 1 and supplementary material Table S1). As hypothesized, we found that the more cold-tolerant species were better able to defend their  $V_m$  at lower temperatures. The rank order of significant depolarization was similar to the rank order of cold sensitivity (Fig. 1); a significant depolarization occurred at  $10^\circ\text{C}$  for *D. birchii*, followed by *D. equinoxialis* ( $5^\circ\text{C}$ ), *D. melanogaster* ( $0^\circ\text{C}$ ), *D. persimilis* ( $-3^\circ\text{C}$ ) and *D. montana* (which did not significantly depolarize, even at  $-3^\circ\text{C}$ ). The variation in  $CT_{\min}$  among species was not explained by a higher polarization of muscle  $V_m$  at  $20^\circ\text{C}$ . If chill coma occurs when the membrane potential passes a certain critical threshold, then possessing a more polarized ‘baseline’  $V_m$  would increase the change in  $V_m$  necessary to pass the

Department of Zoophysiology, Faculty of Science and Technology, Aarhus University, Aarhus DK-8000, Denmark.

\*Author for correspondence (jonas.andersen@bios.au.dk)

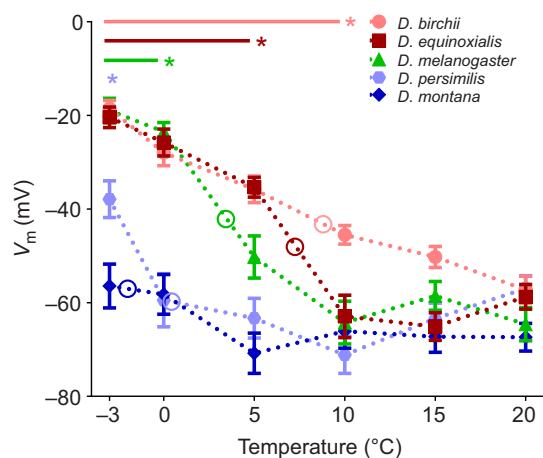
Received 15 April 2015; Accepted 11 June 2015

**List of symbols and abbreviations**

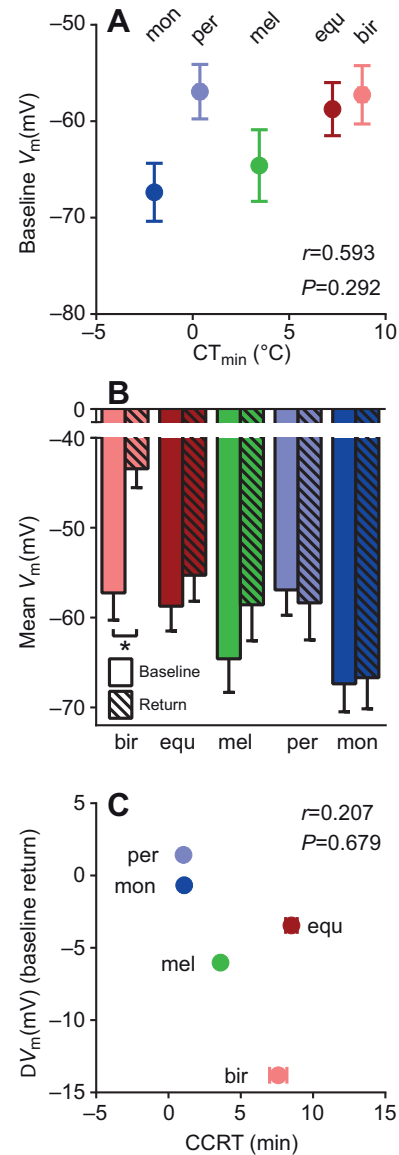
CCRT	chill coma recovery time
CNS	central nervous system
CT <sub>min</sub>	critical thermal minimum
V <sub>m</sub>	resting membrane potential
[K <sup>+</sup> ] <sub>ext</sub>	extracellular K <sup>+</sup> concentration

threshold. Although the baseline V<sub>m</sub> (at 20°C) was most polarized for the most cold-tolerant species (−67.4±2.9 mV, *D. montana*), the second most cold-tolerant species had the least-polarized V<sub>m</sub> of all the species at 20°C (−55.5±2.5 mV, *D. persimilis*). Accordingly, there was no significant tendency for the baseline V<sub>m</sub> to decrease with decreasing CT<sub>min</sub> among species ( $r=0.593$ ,  $P=0.292$ , Fig. 2A). Although there seems to be a tendency when ignoring *D. persimilis*, a more polarized baseline V<sub>m</sub> is clearly not a general strategy among all cold-tolerant drosophilids and an alternative strategy may therefore be to defend V<sub>m</sub> across a broad range of temperatures, as *D. persimilis* does. These two strategies are not mutually exclusive, and may contribute differently across the genus where cold tolerance has evolved several times (Kellermann et al., 2012; MacMillan et al., 2015).

Earlier observations have indicated a ‘critical threshold’ V<sub>m</sub> of −35 to −45 mV associated with onset of chill coma in insects (Wareham et al., 1974; Hosler et al., 2000; MacMillan et al., 2014). In the present study, we found support for such a critical threshold in three of the five species; *D. birchii*, *D. equinoxialis* and *D. melanogaster* did depolarize to around −40 to −50 mV at their CT<sub>min</sub>. These species may therefore experience a decrease or absence of action potentials as a result of the depolarization (Wareham et al., 1974; Salkoff and Wyman, 1983; Hosler et al., 2000; Finsen et al., 2014). By contrast, *D. persimilis* and *D. montana* suffered no significant depolarization at their CT<sub>min</sub>. Thus, cold-adapted *Drosophila* species may enter coma as a result of processes unrelated to depolarization of muscle V<sub>m</sub>. For these species, coma could result from a direct effect of temperature (not related to depolarization) on voltage-sensitive Ca<sup>2+</sup> channels,



**Fig. 1. The resting membrane potential of five species of *Drosophila* collected during cold exposure.** Flight muscle resting membrane potential collected for every 5°C from 20 to 0°C and at −3°C during a 0.2°C min<sup>−1</sup> downwards temperature ramp. Open circles indicate the species' mean critical thermal minimum (CT<sub>min</sub>) collected during a similar temperature ramp using different animals. The asterisk and solid lines aligned with the key cover the temperatures where the V<sub>m</sub> is statistically different ( $P<0.05$ ) from the baseline values (20°C) for each species. Data are means±s.e.m.



**Fig. 2. Initial muscle membrane potential at 20°C and after returning to 20°C following a cold exposure in five species of *Drosophila*.** (A) Mean baseline resting membrane potential (V<sub>m</sub>) plotted against CT<sub>min</sub> of the five species of *Drosophila*. (B) Baseline and return 20°C resting membrane potentials (V<sub>m</sub>); asterisk denotes statistical difference within species ( $P<0.05$ ). (C) The relationship between  $\Delta V_m$  and chill coma recovery time (CCRT).  $\Delta V_m$  is calculated as baseline mean V<sub>m</sub> minus return mean V<sub>m</sub>. The chill coma recovery time (CCRT) was collected after a 0.2°C min<sup>−1</sup> ramp from 20 to −3.5°C ( $N=10$  for all species). Data are means±s.e.m. (if possible). Species name abbreviations are shown in association with the data points and bars: *D. montana* (mon); *D. persimilis* (per); *D. melanogaster* (mel); *D. equinoxialis* (equ); *D. birchii* (bir).

hindering propagation of action potentials (Frolov and Singh, 2013) or to a failure of CNS conduction (Rodgers et al., 2010).

The cold-induced depolarization of muscle tissue in insects can be caused by a number of factors, including reduced activity of electrogenic pumps, direct temperature effects on membrane permeability and conductance as well as increases in [K<sup>+</sup>]<sub>ext</sub> following cold exposure (Košťál et al., 2004; MacMillan and Sinclair, 2011a; Andersen et al., 2013; Finsen et al., 2013; MacMillan et al., 2014). The prevailing data suggest that this occurs during a two-step process where a direct temperature-related

depolarization is followed by a further gradual depolarization caused by perturbation of ion and water balance (MacMillan et al., 2014). In the present study, we examined the contribution of ionic perturbation to  $V_m$  by measuring membrane potential after the temperature ramp, when the flies were returned to 20°C. Reverting the temperature-dependent depolarization meant that any remaining difference ( $\Delta V_m$ ) between the baseline  $V_m$  at 20°C and the measurement after cold exposure (also 20°C) is likely to be caused by an increased  $[K^+]_{ext}$ . We related  $\Delta V_m$  to the CCRT, since this is considered to be a good measure of an animal's ability to recover from increased  $[K^+]_{ext}$  caused by cold exposure (MacMillan et al., 2012; Andersen et al., 2015b). Although not significant, we noted that *D. persimilis* and *D. montana* were very similar or slightly hyperpolarized when returned to 20°C while the remaining (less cold tolerant) species tended to be slightly depolarized (Fig. 2B). This depolarization was only significant in *D. birchii* ( $t=-3.74$ , d.f.<sub>19,52</sub>,  $P=0.001$ ), and there was no overall relationship between CCRT and  $\Delta V_m$  among species ( $r=0.207$ ,  $P=0.679$ , Fig. 2C). It is possible that the association between  $\Delta V_m$  and CCRT is obscured by the experimental design, where  $V_m$  during recovery was measured 13 min after the return to 20°C. All species had recovered from chill coma at this time (Fig. 2C), and any  $K^+$ -dependent depolarization (which we infer from  $\Delta V_m$ ) may have been partially recovered.

In conclusion, more cold-tolerant *Drosophila* species are able to maintain muscle  $V_m$  at lower temperatures than their cold-sensitive congeners. Also, cold-tolerant species may have evolved the ability to circumvent the 'critical threshold' of muscle depolarization related to chill coma in the chill-sensitive *Drosophila* (and other insect species). Some chill-sensitive species experience an increased  $[K^+]_{ext}$  during cold exposure that augments the cold-induced depolarization, whereas the cold-tolerant flies avoid this.

## MATERIALS AND METHODS

### Experimental animals

Five *Drosophila* species were provided from laboratory cultures: *D. montana* (Anneli Hoikkala, University of Jyväskylä, Finland); *D. melanogaster* (Volker Loeschcke, Aarhus University, Denmark); *D. equinoxialis* and *D. persimilis* (the *Drosophila* Species Stock Center, San Diego, USA) and *D. birchii* (Ary Hoffmann, University of Melbourne, Australia) (Table 1). Experimental flies were raised as described in Andersen et al. (2015a). All experimental flies were 6- to 9-day-old non-virgin female flies, raised under low-density conditions at 20±1°C.

### Measurements of chill coma onset and recovery

To associate the species' physiological differences to their cold tolerance phenotype we used two measures associated with chill coma, because both CCRT and  $CT_{min}$  strongly correlate with lethal temperature in these species (Andersen et al., 2015b). In a parallel study, conducted on the same populations, we measured the critical thermal minimum ( $CT_{min}$ ) (Andersen et al., 2015a). Briefly,  $CT_{min}$  was scored by submerging individual flies in 5 ml vials ( $N=20$  per species) in an ethylene glycol and water solution (1:2) and progressively cooling at a rate of 0.2°C min<sup>-1</sup> from 20°C. Once

spontaneous movement ceased, flies were motivated to move by tapping the vial, and the  $CT_{min}$  was recorded when all capacity for movement stopped. Another measure of cold tolerance, chill coma recovery time (CCRT), was assessed from the time it took the flies to recover from chill coma and regain standing position following a similar temperature ramp. Flies ( $N=10$  per species) were placed individually in 5 ml sealed containers and submerged in a cooling bath for the duration of the temperature ramp (20 to -3.5°C at 0.2°C min<sup>-1</sup>), after which they were quickly returned to room temperature to allow for visual recording of CCRT, while tapping the vials every 30 s to motivate standing as fast as physiologically possible.

### Resting membrane potential

Single flies were placed at 0°C for 25 s, to induce a brief paralysis allowing us to place the flies directly on a custom-built glass plate connected to a programmable cooling bath preset at 20°C. For each experimental round, 16 flies were mounted and immobilized on the glass plate using a thin layer of sports resin. To obtain muscle resting membrane potential ( $V_m$ ) a reference electrode (diameter, 0.05 mm; 99.99% hard platinum, Pg Metal Shop, London, UK) was placed in the hemolymph by puncturing the carapace directly. Immediately before measuring, a small window was cut in the dorsolateral anterior half of the thorax with the tip of a 0.45×12 mm syringe needle, and a glass micro electrode (Clark borosilicate glass microelectrodes, GC100TF; Warner Instruments, Hamden, CT, USA) was then inserted into the flight muscle (supplementary material Fig. S1), both electrodes were mounted on micro manipulators (World Precision Instruments Inc., Berlin, Germany). The glass electrode was pulled to a tip resistance of 5–10 MΩ using a Flaming-Brown P-97 electrode puller (Sutter Instruments Co., Novato, CA, USA) and both the reference and glass electrode were connected to an Electro 705 differential electrometer (World Precision Instruments Inc., Sarasota, FL, USA) such that data could be obtained through a 1401 Micro3 data acquisition system connected to a PC running Spike2 (v8, Cambridge Electronic Design, Cambridge, UK). The glass microelectrode was gently moved inward at a 25–40 deg angle aiming at the dorsolateral flight muscles. A flight muscle fiber was assumed to be penetrated once an instant drop in the measured potential was registered on the screen. Each fly was used to assess  $V_m$  at one temperature and for each fly a minimum of three 'repeatable' (all within a 10 mV span) membrane potentials were collected in succession (assumed to be from the same muscle fiber). After a minimum of three repeatable measurements, the electrode was moved deeper into the flight muscle and the practice was repeated on a different muscle fiber if possible (i.e. we obtained  $V_m$  from one or two muscle fibers per animal). In cases where we succeeded in measuring two fibers we used the most negative  $V_m$  from the animal. In cases where we were unable to obtain repeatable measurements, the fly was rejected from the dataset. After measuring  $V_m$  at 20°C (baseline) the temperature ramp was initiated (-0.2°C min<sup>-1</sup>), and  $V_m$  was measured in two new flies for every 5±0.5°C down to 0°C, and at -3±0.5°C. After the measurement at -3°C the temperature bath was quickly reset to 20°C and 15 min later (after 13 min at 20°C) two more flies were measured (20°C return). We were unable to decrease temperatures further to -3.5°C, because this exposed the experimental setup to the risk of ice formation. The  $\Delta V_m$  was calculated for each species as the difference between the mean of the baseline  $V_m$  (at 20°C) minus the mean of the return  $V_m$  (also at 20°C). We collected  $V_m$  from between 8 and 14 animals per temperature per species (5 species×7 temperatures, resulting in 35 species-temperature combinations).

**Table 1. Details of the five *Drosophila* species used for experiments**

Species (abbreviation)	Origin	Collection year	Distribution profile	$CT_{min}$ (°C)	CCRT (min)	Source laboratory
<i>D. birchii</i> (bir)	Australia	2008	Tropical	8.79±0.19	7.6±0.6	Hoffmann, Australia
<i>D. equinoxialis</i> (equ)	Honduras	<1984	Tropical	7.25±0.13	8.5±0.4	DSSC, USA
<i>D. melanogaster</i> (mel)	Denmark	2011	Widespread	3.46±0.07	3.6±0.1	Loeschcke, Denmark
<i>D. persimilis</i> (per)	Canada	Unknown	Temperate	0.38±0.14	1.0±0.1	DSSC, USA
<i>D. montana</i> (mon)	Finland	2008	Temperate	-1.99±0.05	1.1±0.1	Hoikkala, Finland

All species were kept in a 20±1°C room for at least a year before experiments. Table shows species name (abbreviation), country of origin, year of collection, species distribution profile, critical thermal minimum ( $CT_{min}$ , means±s.e.m.), the chill coma recovery time (CCRT, means±s.e.m.) along with the source where the flies have been kept since collection, before being brought to our lab (see Materials and methods).



### Statistics and analyses

Data were deemed normally distributed by investigating the boxplots with superimposed data points for the different experimental groups. This was generally confirmed as Shapiro–Wilk tests only rejected normality in 4 of the 35 groups. Differences in  $V_m$  were investigated using a generalized linear model, examining the effect of temperature on  $V_m$  for each species individually. To test for differences in critical thermal minimum ( $CT_{min}$ ) among species [data previously published in Andersen et al. (2015a)], we used a non-parametric Kruskal–Wallis test (H test) and a Dunn's multiple *post hoc* comparison test. Initial (baseline) and return to 20°C  $V_m$  values were analyzed by Welch two-sample unpaired *t*-tests. The regression between the baseline  $V_m$  and  $CT_{min}$  as well as the relationship between CCRT and  $\Delta V_m$  were conducted using a linear regression. All statistics were done in R 3.1.2 (R Core Team, 2014) and values reported as means $\pm$ s.e.m. unless otherwise stated.

### Acknowledgements

We would like to thank Kirsten Kromann for her help with rearing flies and help during the experiments, Jens Christian Kondrup and John Svane Jensen for technical assistance, and the Loeschcke laboratory for supplying fly medium.

### Competing interests

The authors declare no competing or financial interests.

### Author contributions

All authors designed the experiments, J.L.A. performed the experiments and analyzed the data, and all authors wrote the manuscript.

### Funding

The research was funded by a Carlsberg foundation equipment grant [to H.A.M.], a Sapere Aude Starting Grant, Danish Council for Independent Research Natural Sciences [to J.O.] and a grant from the Faculty of Science and Technology of Aarhus University [to J.L.A.].

### Supplementary material

Supplementary material available online at <http://jeb.biologists.org/lookup/suppl/doi:10.1242/jeb.123760/-DC1>

### References

- Andersen, J. L., Findsen, A. and Overgaard, J. (2013). Feeding impairs chill coma recovery in the migratory locust (*Locusta migratoria*). *J. Insect Physiol.* **59**, 1041–1048.
- Andersen, J. L., MacMillan, H. A. and Overgaard, J. (2015a). Temperate *Drosophila* preserve cardiac function at low temperature. *J. Insect Physiol.* **77**, 26–32.
- Andersen, J. L., Manenti, T., Sørensen, J. G., MacMillan, H. A., Loeschcke, V. and Overgaard, J. (2015b). How to assess *Drosophila* cold tolerance: chill coma temperature and lower lethal temperature are the best predictors of cold distribution limits. *Funct. Ecol.* **29**, 55–65.
- Findsen, A., Andersen, J. L., Calderon, S. and Overgaard, J. (2013). Rapid cold hardening improves recovery of ion homeostasis and chill coma recovery time in the migratory locust, *Locusta migratoria*. *J. Exp. Biol.* **216**, 1630–1637.
- Findsen, A., Pedersen, T. H., Petersen, A. G., Nielsen, O. B. and Overgaard, J. (2014). Why do insects enter and recover from chill coma? Low temperature and high extracellular potassium compromise muscle function in *Locusta migratoria*. *J. Exp. Biol.* **217**, 1297–1306.
- Frolov, R. V. and Singh, S. (2013). Temperature and functional plasticity of L-type Ca<sup>2+</sup> channels in *Drosophila*. *Cell Calcium* **54**, 287–294.
- Hori, Y. and Kimura, M. T. (1998). Relationship between cold stupor and cold tolerance in *Drosophila* (Diptera: Drosophilidae). *Environ. Entomol.* **27**, 1297–1302.
- Hosler, J. S., Burns, J. E. and Esch, H. E. (2000). Flight muscle resting potential and species-specific differences in chill-coma. *J. Insect Physiol.* **46**, 621–627.
- Kellermann, V., Loeschcke, V., Hoffmann, A. A., Kristensen, T. N., Fløjgaard, C., David, J. R., Svenning, J.-C. and Overgaard, J. (2012). Phylogenetic constraints in key functional traits behind species' climate niches: patterns of desiccation and cold resistance across 95 *Drosophila* species. *Evolution* **66**, 3377–3389.
- Košťál, V., Vambera, J. and Bastl, J. (2004). On the nature of pre-freeze mortality in insects: water balance, ion homeostasis and energy charge in the adults of *Pyrhocoris apterus*. *J. Exp. Biol.* **207**, 1509–1521.
- Košťál, V., Yanagimoto, M. and Bastl, J. (2006). Chilling-injury and disturbance of ion homeostasis in the coxal muscle of the tropical cockroach (*Nauphoeta cinerea*). *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* **143**, 171–179.
- MacMillan, H. A. and Sinclair, B. J. (2011a). The role of the gut in insect chilling injury: cold-induced disruption of osmoregulation in the fall field cricket, *Gryllus pennsylvanicus*. *J. Exp. Biol.* **214**, 726–734.
- MacMillan, H. A. and Sinclair, B. J. (2011b). Mechanisms underlying insect chill-coma. *J. Insect Physiol.* **57**, 12–20.
- MacMillan, H. A., Williams, C. M., Staples, J. F. and Sinclair, B. J. (2012). Reestablishment of ion homeostasis during chill-coma recovery in the cricket *Gryllus pennsylvanicus*. *Proc. Natl. Acad. Sci. USA* **109**, 20750–20755.
- MacMillan, H. A., Findsen, A., Pedersen, T. H. and Overgaard, J. (2014). Cold-induced depolarization of insect muscle: differing roles of extracellular K<sup>+</sup> during acute and chronic chilling. *J. Exp. Biol.* **217**, 2930–2938.
- MacMillan, H. A., Ferguson, L. V., Nicolai, A., Donini, A., Staples, J. F. and Sinclair, B. J. (2015). Parallel ionoregulatory adjustments underlie phenotypic plasticity and evolution of *Drosophila* cold tolerance. *J. Exp. Biol.* **218**, 423–432.
- R Core Team (2014). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. R-project.org.
- Rodgers, C. I., Armstrong, G. A. B. and Robertson, R. M. (2010). Coma in response to environmental stress in the locust: A model for cortical spreading depression. *J. Insect Physiol.* **56**, 980–990.
- Salkoff, L. B. and Wyman, R. J. (1983). Ion currents in *Drosophila* flight muscles. *J. Physiol.* **337**, 687–709.
- Wareham, A. C., Duncan, C. J. and Bowler, K. (1974). The resting potential of cockroach muscle membrane. *Comp. Biochem. Physiol. A Physiol.* **48**, 765–797.
- Wareham, A. C., Duncan, C. J. and Bowler, K. (1975). The resting potential of the muscle membrane of moths. *Comp. Biochem. Physiol. A Physiol.* **52**, 295–298.

**Table S1. Details of the five *Drosophila* species used for experiments.** All species were kept in a  $20\pm 1^\circ\text{C}$  room for at least a year before experiments. Table contains; Species name (abbreviation), country of origin, year of collection, species distribution profile, critical thermal minimum ( $\text{CT}_{\text{min}}$ , means  $\pm$  s.e.m.), the chill coma recovery time (CCRT, means  $\pm$  s.e.m.) along with the source where the flies have been kept since collection, before being brought to our lab (see methods).

Species (abbreviation)	Origin	Collection year	Distribution profile	$\text{CT}_{\text{min}}$ ( $^\circ\text{C}$ )	CRRT (min)	Source laboratory
<i>D. birchii</i> (bir)	Australia	2008	Tropical	$8.79 \pm 0.19$	$62.5 \pm 4.6$	Hoffmann, AU
<i>D. equinoxialis</i> (equ)	Honduras	<1984	Tropical	$7.25 \pm 0.13$	$42.3 \pm 2.0$	DSSC, US
<i>D. melanogaster</i> (mel)	Denmark	2011	Widespread	$3.46 \pm 0.07$	$15.7 \pm 0.9$	Loeschcke, DK
<i>D. persimilis</i> (per)	Canada	Unknown	Temperate	$0.38 \pm 0.14$	$1.1 \pm 0.2$	DSSC, US
<i>D. montana</i> (mon)	Finland	2008	Temperate	$-1.99 \pm 0.05$	$1.8 \pm 0.4$	Hoikkala, FI



**Fig. S1. Photo showing the placement of the electrodes.** The reference electrode were inserted directly through the carapace (left) and glass electrode inserted into a flight muscle through a small window in the carapace cut with the tip of a syringe-needle (circled).