

## RESEARCH ARTICLE

# The influence of PKA treatment on the Ca<sup>2+</sup> activation of force generation by trout cardiac muscle

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### SUMMARY

**β-Adrenergic stimulation of the mammalian heart increases heart rate, the strength of contraction as well as the kinetics of force generation and relaxation. These effects are due to the phosphorylation of select membrane and thin filament proteins by cAMP-activated protein kinase (PKA). At the level of the sarcomere, it is typically the phosphorylation of cardiac myosin binding protein C (cMyBP-C) and cardiac troponin I (cTnI) that is responsible for the change in the kinetics of contraction and relaxation. Trout cTnI (ScTnI) lacks two critical PKA targets within the N-terminus of the protein that, when phosphorylated in mammalian cTnI, cause a reduction in myofilament Ca<sup>2+</sup> affinity. To determine what role the contractile element plays in the response of the trout heart to β-adrenergic stimulation, we characterized the influence of PKA treatment on the Ca<sup>2+</sup> activation of skinned preparations dissected from ventricular trabeculae. In these experiments, isometric force generation and the rate of force development were measured over a range of Ca<sup>2+</sup> concentrations. The results demonstrate that PKA treatment does not influence the Ca<sup>2+</sup> sensitivity of force generation but it decreases maximum force generation by 25% and the rate of force re-development at maximal activation by 46%. Analysis of the trabeculae preparations for phosphoproteins revealed that PKA treatment phosphorylated myosin light chain 2 but not cTnI or cMyBP-C. These results indicate that the function of the trout cardiac contractile element is altered by PKA phosphorylation but in a manner different from that in mammalian heart.**

Supplementary material available online at <http://jeb.biologists.org/cgi/content/full/214/12/1989/DC1>

Key words: Ca<sup>2+</sup> activation, adrenergic stimulation, force generation, force redevelopment, phosphorylation, protein kinase A.

### INTRODUCTION

The response of the vertebrate heart to β-adrenergic stimulation is a critical component of the fight or flight response. In mammals, this reaction includes an increase in heart rate, an increase in the strength of contraction and increases in the rate of myocyte contraction and relaxation (Wehrens et al., 2006). The common modulator of these cellular functions is cAMP-activated protein kinase (PKA), which targets select proteins that are key regulators of each of these processes. For example, the phosphorylation of the ryanodine receptor and phospholamban increases the amount and rate of Ca<sup>2+</sup> cycling through the sarcoplasmic reticulum (Wehrens et al., 2006), while the increased kinetics of myocyte contraction and relaxation are due to the phosphorylation of cardiac myosin binding protein C (cMyBP-C) and cardiac troponin I (cTnI), respectively (Stelzer et al., 2007). Together, these changes in myocyte function, caused by the activity of PKA, result in an increase in cardiac output.

Using phylogenetic analysis we have recently shown that the number of PKA targets in cMyBP-C and cTnI increased in vertebrates after fish diverged from the vertebrate lineage (Shaffer and Gillis, 2010). For example, the two targets for PKA (Ser23 and Ser24) in human cTnI are within the N-terminal extension (Kobayashi and Solaro, 2005). This peptide, and associated PKA targets, is present in cTnI from all vertebrates, except fish (Shaffer and Gillis, 2010). It is also not found in any isoform of slow skeletal TnI (ssTnI) or fast skeletal TnI (fsTnI). This suggests that the phosphorylation of cTnI by PKA as a mechanism to regulate cardiac

function evolved in the lineage that led to amphibians, reptiles, birds and mammals after fish diverged (Shaffer and Gillis, 2010).

Previous work has demonstrated that the function of the teleost fish heart is sensitive to β-adrenergic stimulation (Farrell, 1985; Gesser et al., 1982; Graham and Farrell, 1989; Keen et al., 1993; Vornanen, 1998). The application of adrenaline to the trout heart increases heart rate and the strength of contraction (Aho and Vornanen, 2001), as well as the rate of contraction and relaxation (Shiels et al., 1998). These changes in cardiac function have been shown to be due, at least in part, to increased current (2.3-fold) through the L-type Ca<sup>2+</sup> channels (Vornanen, 1998). However, it is not known what role the regulatory proteins of the contractile element play in these effects of adrenergic stimulation on the trout heart.

The purpose of the current study was to characterize how PKA treatment influences the response of the trout contractile element to Ca<sup>2+</sup>. To do this, we developed a skinned cardiac preparation utilizing cardiac trabeculae dissected from the spongy ventricle wall of the trout heart. This preparation was then used to characterize the Ca<sup>2+</sup> sensitivity of force generation and the rate of force redevelopment. Ca<sup>2+</sup> activation was measured over a range of Ca<sup>2+</sup> concentrations, enabling the Ca<sup>2+</sup> sensitivity of force generation to be characterized. These measurements were made in control preparations as well as in preparations phosphorylated with PKA for 1 h. The sarcomere length of each preparation was set to 2.2 μm. This length was chosen to enable the results to be compared with previous studies of mammalian cardiac tissues that utilized similar methods.

## MATERIALS AND METHODS

### Cardiac trabecular mechanics

#### Trabecular dissection

Mechanical experiments were conducted on permeabilized trabeculae preparations dissected from the spongy ventricular wall of rainbow trout hearts. Adult rainbow trout, *Oncorhynchus mykiss* Walbaum 1792 (males and females, ~1 kg), were maintained in  $10 \pm 1^\circ\text{C}$  circulating ground water for the period of the study. Water was kept at normoxic levels with a 12h:12h photoperiod. All fish were fed *ad libitum* for the duration of the experiment and protocols were approved by the University of Guelph Animal Care Committee. Fish were removed from the tank by dip net and then rapidly killed by a blow to the head and severing of the cerebral spinal column. The time from netting of fish until removal of the heart was less than 1 min. Following removal, the heart was immediately placed in ice-cold physiological saline [in  $\text{mmol l}^{-1}$ : 94 NaCl, 24  $\text{NaCO}_3$ , 5 KCl, 1  $\text{MgSO}_4$ , 1  $\text{Na}_2\text{HPO}_4$ , 0.7  $\text{CaCl}_2$ , pH 7.6 at  $15^\circ\text{C}$ ]. All blood was washed from the heart and then the heart was placed onto a dissection plate on a chilled dissection stage containing ice-cold relaxing solution (in  $\text{mmol l}^{-1}$ : 100 KCl, 10 MOPS, 5  $\text{K}_2\text{EGTA}$ , 9  $\text{MgCl}_2$ , 4 NaATP, 20 2,3-butanedione monoxime (BDM), pH 7.0 at  $4^\circ\text{C}$ ) (Gillis et al., 2007). BDM inhibits cross-bridge formation and was used to reduce injury due to contracture. The protease inhibitors phenylmethanesulphonylfluoride (PMSF), leupetin and bezamidin were also present in all relaxing and skinning solutions. The ventricle wall was exposed and sheets of trabeculae ( $4.0 \times 2.0 \text{ mm}$ ) were dissected and then pinned out in a dissection dish. The trabeculae were skinned overnight, on ice, in relaxing solution containing glycerol (50% v:v) and Triton X-100 (1%) with one solution change, as previously described (Gillis et al., 2007). The following morning the skinning solution was replaced with the same solution minus the Triton X-100. The skinned trabeculae were then dissected on a chilled microscope stage to produce preparations that were  $\sim 1.7 \times 0.35 \text{ mm}$ . Preparations were only used if it was obvious that the muscle fibers ran in parallel down its length. These were visualized using polarized light. The ends of the preparations were wrapped in homemade aluminium foil 'T-clips'. These T-clips were 1.0 mm in length and 1 mm in width with a small triangular hole cut in one end to allow the preparation to be mounted to the experimental apparatus *via* steel hooks (Fig. 1A).

#### Treatment of trabecular preparations with PKA

Skinned and wrapped trabecular preparations were transferred into 2 ml of relaxing solution containing  $1 \text{ mmol l}^{-1}$  dithiothreitol (DTT) and  $100 \text{ U ml}^{-1}$  of the catalytic subunit of PKA from bovine heart (Sigma-Aldrich, St Louis, MO, USA). This concentration of PKA was selected to mimic conditions used by Patrick et al. (Patrick et al., 2010). The preparations were then incubated at  $15^\circ\text{C}$  for 1 h. Control preparations were incubated in the same solution without the PKA under the same conditions. Following incubation, the buffer

was replaced with 2 ml relaxing solution without PKA, then incubated at  $15^\circ\text{C}$  for 15 min with gentle agitation.

#### Muscle mechanics instrument

The mechanical measurements were made using a custom-designed instrument from Aurora Scientific (Aurora, ON, Canada) mounted on the stage of an inverted microscope (Model Eclipse TE 2000U, Nikon, Tokyo, Japan). The instrument is composed of a permeabilized fiber test system (Model 802B, Aurora Scientific), a force transducer (Model 400A, 2.0 kHz resonant frequency, Aurora Scientific) and a servo-motor (Model 308C, Aurora Scientific) tuned for a  $250 \mu\text{s}$  step response. The fiber test system consists of an 8-well stainless steel bath attached to two stepper motors. The stepper motors allow the preparation, mounted to the force transducer and servomotor, to be moved from bath to bath with an excursion time of 1.4 s. The temperature of the plate is regulated *via* three thermoelectric coolers and well temperature is measured *via* a thermocouple. Temperature is maintained within  $1^\circ\text{C}$  of the set temperature. Sarcomere length (SL) was measured by fast Fourier transform (FFT) analysis using the high-speed video sarcomere length system (Model 901A, Aurora Scientific) with a CCD camera (Model VGA-210-LMCN, Imperx, FL, USA) with  $640 \times 480$  resolution. The camera images the preparation through a  $40\times$  objective with a 3 mm working distance (Nikon).

#### Solutions for mechanical measurements

Solution composition was determined according to an iterative computer program that calculates the equilibrium concentration of ligands and ions based on published affinity constants. Activation solutions contained (in  $\text{mmol l}^{-1}$ ): 15 phosphocreatine, 15 EGTA, at least 40 MOPS, 1 free  $\text{Mg}^{2+}$ , 135  $\text{Na}^+\text{+K}^+$ , 1 DTT,  $250 \text{ U ml}^{-1}$  creatine phosphokinase (CPK; Sigma) and  $5 \text{ mmol l}^{-1}$  NaATP, pH 7.0 at  $15 \pm 1^\circ\text{C}$ . Ionic strength was  $0.17 \text{ mol l}^{-1}$ . The  $\text{Ca}^{2+}$  level (expressed as  $\text{pCa} = -\log[\text{Ca}^{2+}]$ ) was varied between pCa 9 and pCa 5.2 by adjusting  $\text{Ca}(\text{propionate})_2$  as previously described (Gillis et al., 2007).

#### Experimental protocol

The ability of the preparation to be activated by  $\text{Ca}^{2+}$  was measured by moving the preparation through the series of wells containing activation solutions with different pCa values. When the force trace for each pCa reached an asymptote, steady-state force and the rate of isometric tension redevelopment ( $k_{tr}$ ) were measured. The activation solutions had pCa values between 9 and 5.2. These pCa values were used because preliminary experiments demonstrated that the trabeculae were maximally activated by pCa 5.3 for both the control and PKA-phosphorylated preparations. In addition, the specific pCa values used were selected so that we could effectively describe the  $\text{Ca}^{2+}$  activation curve. Between each activation solution of different pCa, the preparation was exposed to the pre-activation solution (in

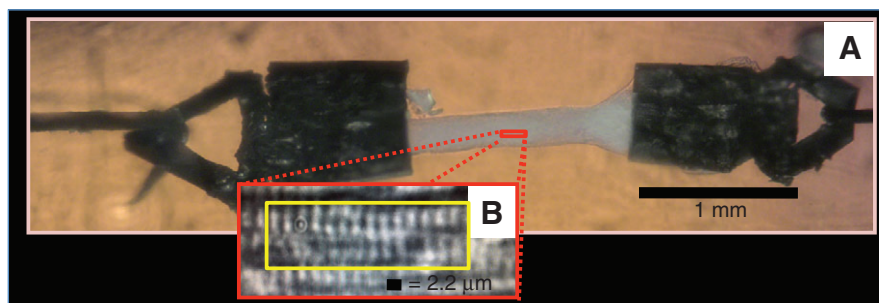


Fig. 1. Images of a skinned trout cardiac trabeculae preparation mounted to the muscle mechanics system. (A) Trabeculae preparation attached to force transducer and servomotor using aluminium foil clips. (B) Image of sarcomere pattern through  $40\times$  objective. The yellow box represents a typical sarcomere pattern that was analyzed using fast Fourier transformation to measure sarcomere length (SL).

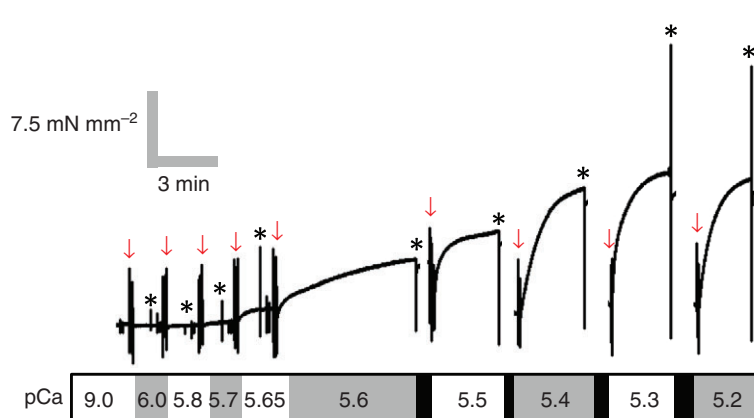


Fig. 2. Sample force trace resulting from the Ca<sup>2+</sup> activation of a trout cardiac trabeculae preparation at 15°C and SL 2.2 µm. The noise on the force trace is due to motor movements during rate of force redevelopment ( $k_{tr}$ ) measurements (\*) and movement of preparation between solution wells (↓). Force is displayed as mN mm<sup>-2</sup> and pCa values are indicated along the bottom of the trace.

mmol<sup>-1</sup>: 100 KCl, 10 MOPS, 0.1 EGTA, 9.0 MgCl<sub>2</sub>, 4 NaATP) for ~5 s. The purpose of this Ca<sup>2+</sup>-free solution was to enable the rapid penetration of Ca<sup>2+</sup> into the muscle upon subsequent exposure and to prevent Ca<sup>2+</sup> contamination between wells. Prior to being mounted on the apparatus, each preparation was incubated on ice in 2 ml relaxation buffer without BDM for at least 15 min. The preparation was then mounted on the apparatus by attaching the T-clips on the muscle to steel hooks attached to the force transducer and servomotor (Fig. 1A). During this time the preparation was immersed in a well containing activation solution with a pCa of 9 and no BDM. The mean length (ML) and width of the preparation were then measured using an eyepiece reticle and the 4× objective. The mean length of the preparations used in this experiment was 1.7±0.1 mm and the mean width was 0.34±0.01 mm. To standardize force generation the cross-sectional area was calculated from the diameter assuming a circular geometry. To 'set' the clips on the hooks, the preparation was then activated using pCa 5.3 and allowed to generate 0.5 mN of force. The preparation was then moved back into pCa 9 to relax. This was done before the preparation was maximally activated. SL was then set to 2.2 µm by stretching the preparation accordingly using the XYZ micrometer translation stage, to which the servomotor was attached. Between each length adjustment, SL was measured by analyzing the sarcomere pattern at multiple locations (>5) throughout the preparation using FFT. An example of such a pattern is shown within the yellow box in Fig. 1B. The measurement generated, in µm, was a mean of the multiple sarcomeres (>10) in each analyzed pattern. If a consistent SL pattern could not be obtained throughout the preparation, the preparation was discarded. The passive force at pCa 9 was then measured as described previously (Gillis et al., 2007) using a release–restretch protocol. This value was subtracted from total force measured at higher Ca<sup>2+</sup> concentrations. The preparation was then maximally activated and force generation was measured. This value was used to characterize the effect of the subsequent activation protocols on muscle function. Steady-state isometric force and  $k_{tr}$  following rapid (<4 ML s<sup>-1</sup>) release–restretch (15% ML) was measured as described previously (Gillis et al., 2007). As the rate of force redevelopment is a function of cross-bridge formation, this measure is used as an estimate of the rate of cross-bridge cycling during Ca<sup>2+</sup> activation. The preparation was then returned to pCa 9 and allowed to relax. Once relaxed, the experiment began. Fig. 2 is a sample force trace of a preparation being activated over a range of pCa values.

#### Phosphorylation detection

Individual trabeculae were solubilized by sonicating them (3×10 s on ice) in 2× SDS loading buffer. Samples were then separated on

10% SDS-PAGE gels using the miniVE system (GE Healthcare, Piscataway, NJ, USA) and then stained with Pro-Q Diamond phosphoprotein gel stain following the manufacturer's instructions (Molecular Probes, Eugene, OR, USA). Briefly, gels were fixed overnight in 50% methanol, 10% acetic acid, and were stained for 75 min, in light-excluding containers. Gels were destained using 20% acetonitrile and 50 mmol<sup>-1</sup> sodium acetate (pH 4.0) and imaged using a Typhoon 9410 scanner (GE Healthcare) with an excitation wavelength of 532 nm and emission filter of 560 nm. Gels were then stained for total protein using SYPRO-Ruby protein stain (Bio-Rad, Hercules, CA, USA). To ensure that the Pro-Q stain was working and to compensate for non-specific binding, the 'peppermint stick' (Molecular Probes) protein ladder, which contains two phosphorylated proteins, was run on each gel according to the manufacturer's instructions. Once gels had been imaged for the Pro-Q signal, levels were reduced until only the two phosphorylated protein bands were visible in the ladder lane. Any bands that were still visible in the sample lanes were then characterized as phosphorylated. To determine whether PKA treatment altered the level of phosphorylation of individual proteins, the program ImageJ (National Institutes of Health, Bethesda, MD, USA) was used to analyze the density of the Pro-Q-stained protein bands. The Pro-Q signal was standardized to the SYPRO-Ruby signal for the same band to account for differences in protein loading between lanes. Individual myofilament proteins were identified on the SDS gel using mass spectroscopy and western blotting. Details of this analysis are available in the Appendix. Accompanying western blot images are available in the supplementary material Fig. S1.

#### Analysis

Each force–pCa curve was fitted with the Hill equation:

$$F = F_{\max} (1 + 10^{n_H(pCa - pCa_{50})})^{-1}, \quad (1)$$

where  $F_{\max}$  is the force at high [Ca<sup>2+</sup>] (low pCa), pCa<sub>50</sub> is the pCa needed to achieve 50% of  $F_{\max}$  (used here as a measure of the Ca<sup>2+</sup> sensitivity of force), and  $n_H$ , the Hill coefficient, reflects the steepness of the relationship. The reported values for each of these parameters represent the means of the values from the individual fits ±s.e. Student's *t*-tests were used for statistical comparisons of the different parameters between the two treatment groups.

The data produced by the  $k_{tr}$  protocol describing the increase in force following the rapid release–restretch was well fitted by a single exponential equation. The calculated error values, used as a goodness of fit indicator, were within an acceptable range. In addition, each data fit was examined by eye to confirm a good fit.

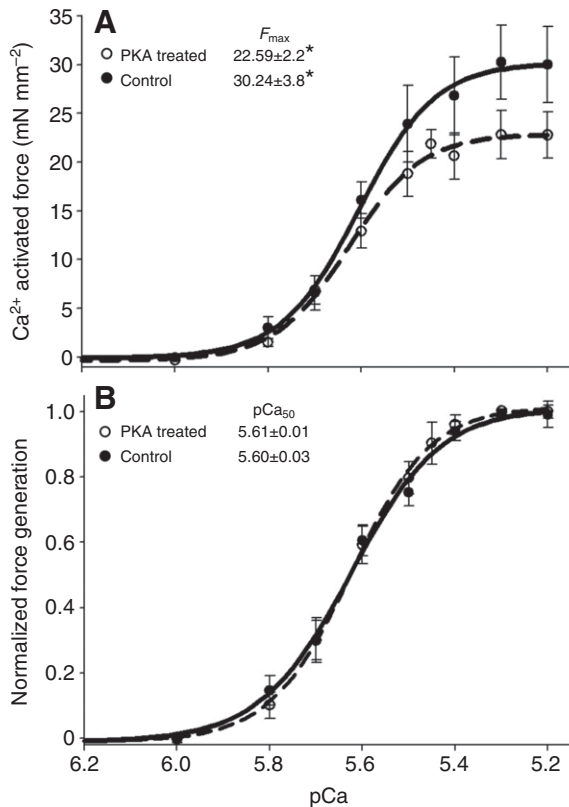


Fig. 3. PKA treatment reduces the force generated by skinned trout cardiac trabeculae during maximum activation but it does not change the  $\text{Ca}^{2+}$  sensitivity of force generation. (A) Actual force generated. (B) Relative force generated. Measurements made at  $15^\circ\text{C}$  and a SL of  $2.2\ \mu\text{m}$ . For control preparations,  $N=10$ ; for PKA-treated preparations,  $N=8$ . The difference in maximum force ( $F_{\text{max}}$ ) between the control and PKA-treated preparations is significant ( $P<0.05$ ). This is indicated by an asterisk. The difference between  $\text{pCa}_{50}$  values is not significant.

## RESULTS

### $\text{Ca}^{2+}$ activation of trabeculae

The exposure of the trout cardiac trabeculae preparations to increasing levels of  $\text{Ca}^{2+}$  resulted in increased levels of force generation (Fig. 2). When the force produced was plotted against pCa, the resulting curve was sigmoidal in shape and reached an asymptote by pCa 5.3 (Fig. 3A). The  $\text{pCa}_{50}$  calculated for control preparations was  $5.60\pm 0.03$ . This  $\text{pCa}_{50}$  value is lower (greater  $\text{Ca}^{2+}$  concentration) than that previously reported ( $6.28\pm 0.06$ ) for the  $\text{Ca}^{2+}$  activation of force generation by skinned single trout cardiac myocytes at a SL of  $2.3\ \mu\text{m}$  (Patrick et al., 2010). It is also higher than the amplitude of intracellular  $\text{Ca}^{2+}$  concentrations measured during myocyte activation using Fura-2 at  $14^\circ\text{C}$  (pCa 6.9). However, it is very difficult to compare measurements made between labs because of differences in experimental conditions and buffer

design/composition. For example, the study by Patrick and colleagues (Patrick et al., 2010) was completed at  $22^\circ\text{C}$ , where the sensitivity of the preparation to  $\text{Ca}^{2+}$  would be greater due to the influence of temperature on the  $\text{Ca}^{2+}$  affinity of cTnC (Gillis et al., 2000). In addition, the pCa of a solution for force activation or the calibration of a  $\text{Ca}^{2+}$  binding dye is calculated using an iterative process that takes into account ionic strength, total  $\text{Ca}^{2+}$  concentration, solution pH, experimental temperature, and the concentration of  $\text{Ca}^{2+}$  chelators. If any of these vary within the solution from what was used to calculate pCa, due to differences in moisture content or purity of the chemical components, the actual pCa will be significantly different. This in turn would translate into a difference in the  $\text{pCa}_{50}$  determined when the solutions are used in an experiment. For example, the  $\text{pCa}_{50}$  of rat trabeculae measured at  $15^\circ\text{C}$  with a SL of  $2.2\ \mu\text{m}$  using the same recipes and techniques as in the current study was  $5.41\pm 0.01$  (Regnier et al., 2004), while that of single rat myocytes at  $22^\circ\text{C}$  and a SL of  $2.3\ \mu\text{m}$  measured by Patrick and colleagues (Patrick et al., 2010) was  $6.05\pm 0.04$ . Both of these  $\text{pCa}_{50}$  values are for cardiac tissue from the same species but measured at different temperatures with different solutions. Therefore, while the  $\text{pCa}_{50}$  reported in the current study is different from what might be considered physiological, this difference is likely due to variations in experimental conditions and buffer composition between studies.

As mentioned above, each preparation was maximally activated prior to the protocol used to generate the force–pCa curve to allow for any ‘run-down’ to be characterized. During this initial activation, the maximum force generated was measured, as was the  $k_{\text{tr}}$ . These initial values were then compared with those collected during the maximal activation at the end of the  $\text{Ca}^{2+}$  activation protocol. For the control preparations, the rundown was equal to 9% of the mean initial value, while for the phosphorylated preparations the rundown was equal to 7% the mean initial value. There were also relatively minor, non-significant, differences in  $k_{\text{tr}}$  for both the treatment and control between the first maximal activation and the second. For the control preparations, the initial  $k_{\text{tr}}$  at maximum activation was  $2.79\pm 0.7\ \text{s}^{-1}$  and that during the final maximal activation was  $3.14\pm 0.5\ \text{s}^{-1}$  (Table 1). For the phosphorylated preparation, the initial  $k_{\text{tr}}$  at maximum activation was  $2.37\pm 0.4\ \text{s}^{-1}$  and that during the final maximal activation was  $1.70\pm 0.3\ \text{s}^{-1}$  (Table 1). Together, these limited changes in preparation function indicate that there was minimal ‘run down’ during the duration of the experiment.

$k_{\text{tr}}$  was measured at each  $\text{Ca}^{2+}$  activation and these values were plotted against pCa (Fig. 4). Interestingly, the highest  $k_{\text{tr}}$  values were measured at a relatively low  $\text{Ca}^{2+}$  concentration when the amount of force being generated was less than half maximal (Fig. 4). For the control preparation this value was  $5.25\pm 0.9\ \text{s}^{-1}$  at pCa 5.6 while that for the phosphorylated trabeculae was  $6.5\pm 0.9\ \text{s}^{-1}$  at pCa 5.7. These values were greater than those obtained during the initial maximum activation at pCa 5.3. When the preparations were subsequently exposed to higher  $\text{Ca}^{2+}$  concentrations, causing force generation to increase towards a maximum,  $k_{\text{tr}}$  decreased (Fig. 4). In the control tissue the  $k_{\text{tr}}$  at maximal activation was  $3.14\pm 0.5\ \text{s}^{-1}$

Table 1. The influence of PKA treatment on the  $\text{Ca}^{2+}$  activation of trout cardiac trabeculae

	$F_{\text{max}}$ ( $\text{mN mm}^{-2}$ )	$\text{pCa}_{50}$	$n_{\text{H}}$	$k_{\text{tr}}$ at $F_{\text{max}}$ ( $\text{s}^{-1}$ )	Passive force ( $\text{mN mm}^{-2}$ )
Control ( $N=10$ )	$30.24\pm 3.8^{\text{a}}$	$5.60\pm 0.03^{\text{a}}$	$6.22\pm 0.90^{\text{a}}$	$3.14\pm 0.51^{\text{a}}$	$1.10\pm 0.18^{\text{a}}$
PKA treated ( $N=8$ )	$22.59\pm 2.2^{\text{b}}$	$5.61\pm 0.01^{\text{a}}$	$6.72\pm 0.70^{\text{a}}$	$1.70\pm 0.25^{\text{b}}$	$1.11\pm 0.15^{\text{a}}$

$F_{\text{max}}$ , maximum force generated;  $\text{pCa}_{50}$ ,  $\text{Ca}^{2+}$  concentration measured as pCa at half-maximum force generation;  $n_{\text{H}}$ , Hill coefficient;  $k_{\text{tr}}$  at  $F_{\text{max}}$ , rate of force generation at maximum activation. Significant differences between values in the same column are indicated by different superscripts ( $P<0.05$ ).



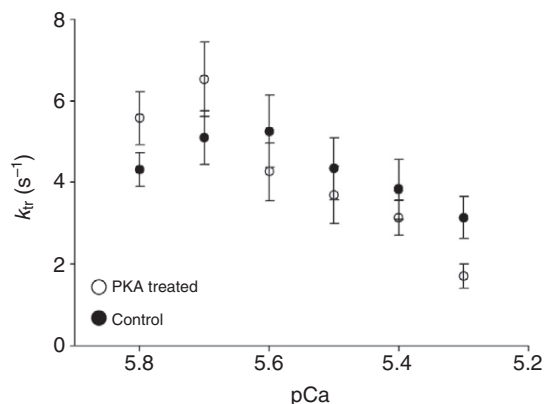


Fig. 4. PKA treatment reduces the  $k_{tr}$  in skinned trout cardiac trabeculae preparations at maximum activation. For control preparations,  $N=10$ ; for PKA-treated preparations,  $N=8$ . The difference in  $k_{tr}$  between the control and PKA-treated preparations at maximum activation (pCa 5.3) is significant ( $P<0.05$ ) at pCa 5.3.

(Table 1). This is significantly less ( $P<0.05$ ) than that at pCa 5.6. The same result was seen in the phosphorylated preparations, with the  $k_{tr}$  at maximal activation ( $1.7\pm 0.9 s^{-1}$ ) being significantly less than that at pCa 5.7 ( $P<0.05$ ). It is unlikely that the decrease in  $k_{tr}$  at higher rates of force generation is due to run down as the  $k_{tr}$  values obtained during the initial maximum activation were not statistically different from those obtained during the second maximum activation. It is also unlikely that the decrease in  $k_{tr}$  was due to limiting concentrations of ATP in the solutions. All solutions contained  $5 mmol l^{-1}$  ATP as well as  $15 mmol l^{-1}$  phosphocreatine and  $250 U ml^{-1}$  CPK. This ensured that any ADP produced was recycled back to ATP. It is likely, therefore, that this decrease in the rate of cross-bridge cycling is a physiologically relevant phenomenon.

#### Influence of PKA on trabeculae preparation function

The treatment of the trabeculae preparations with PKA caused the maximum force generated to decrease from  $30.3\pm 3.8$  to  $22.59\pm 2.2 mN mm^{-2}$  (Table 1, Fig. 3A). This represents a 25% decrease in maximum force generation. There was, however, no difference in the amount of force generated by the control and treatment preparations at their pCa<sub>50</sub> values of  $5.60\pm 0.013$  and  $5.61\pm 0.01$ , respectively. In addition, there was no effect of PKA treatment on the Ca<sup>2+</sup> sensitivity of the preparation. This is evident when the force–pCa curves for the control and treatment groups are standardized and then superimposed (Fig. 3B). In addition, the pCa<sub>50</sub> values for the PKA-treated and control preparations were virtually identical (Table 1). There was also no influence of PKA treatment on the Hill equation generated from the force–pCa curves, indicating that the co-operativity of the contractile element was not altered by the treatment (Table 1). PKA treatment was found to decrease the  $k_{tr}$  at maximal activation (Fig. 4). In the phosphorylated preparations  $k_{tr}$  was 46% less than that of the control fibers (Table 1). Finally, there was no effect of PKA treatment on the passive force of the preparations. These values were  $1.11\pm 0.15$  and  $1.10\pm 0.18 mN mm^{-2}$  for the control and PKA-treated preparations, respectively.

#### Influence of PKA treatment on phosphorylation state of contractile proteins

The treatment of the trabeculae with PKA increased the phosphorylation level of myosin light chain 2 (MLC-2) (Fig. 5).

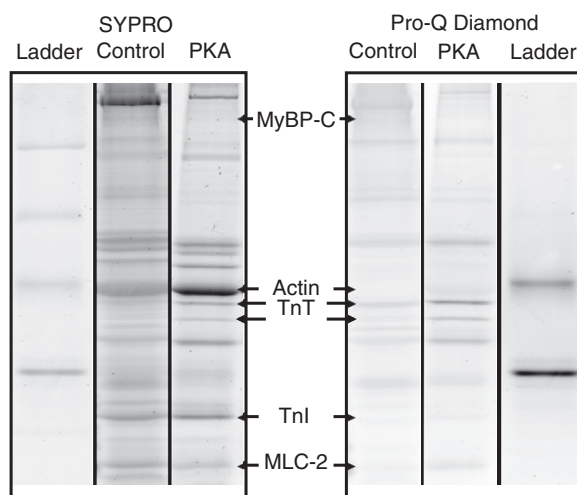


Fig. 5. PKA treatment of skinned cardiac trabeculae increases the phosphorylation state of myosin light chain 2 (MLC-2). (A) SYPRO-Ruby-stained gel showing total protein content of control and PKA-treated preparations. (B) Pro-Q Diamond-stained gel indicating presence of phosphorylated proteins in control and PKA-treated preparations. The positions of myosin binding protein C (MyBP-C), actin, troponin I (TnI), troponin T (TnT) and MLC-2 are indicated on the gel. The bands representing MyBP-C, TnI, TnT and MLC-2 were identified using western blotting or mass spectrometry. The position of actin is inferred from its highly conserved mass and the fact that it is the most prevalent protein in muscle. The peppermint stick ladder was used, which contains two phosphorylated proteins at 23.6 and 45 kDa. The level of phosphorylation of each protein was compared between treatment and control samples via the ratio of the density of the Pro-Q/SYPRO-stained bands using ImageJ.

The standardized density of the MLC-2 band stained with Pro-Q Diamond increased from  $0.78\pm 0.09$  to  $1.00\pm 0.07$ ; this is equal to a ~28% increase in the level of phosphorylation. A Pro-Q band was detected for cTnI and cMyBP-C but these did not increase with PKA treatment, indicating that there was no change in the level of phosphorylation (Fig. 5).

#### DISCUSSION

The limited effect of PKA phosphorylation on the contractility of trout cardiac tissue is in significant contrast to what occurs when mammalian cardiac tissue is similarly treated (Fentzke et al., 1999; Holroyde et al., 1979; Matsuba et al., 2009; Mope et al., 1980; Solaro et al., 2008; Stelzer et al., 2007). For example, the exposure of porcine ventricle fiber bundles to PKA shifted the pCa<sub>50</sub> by 0.1 pCa units (Matsuba et al., 2009) while the exposure of mouse cardiac myocytes to PKA shifted the pCa<sub>50</sub> of force generation by 0.2 pCa units (Fentzke et al., 1999). This change in the Ca<sup>2+</sup> sensitivity of force generation in skinned mammalian cardiac preparations is due to the phosphorylation of the N-terminal extension of cTnI (Matsuba et al., 2009; Yasuda et al., 2007). As trout cTnI lacks these phosphorylation sites it is not surprising that treatment of the trout trabeculae with PKA had no impact on the Ca<sup>2+</sup> sensitivity of force generation. These results indicate that the Ca<sup>2+</sup> sensitivity of the trout heart cannot be regulated to the same degree as the mammalian heart.

The reduction in force generation and cross-bridge cycling rates at maximal activation that occurred in the trout preparations following PKA treatment is not predicted by previous studies with skinned mammalian tissue. In mammalian tissue following PKA

treatment there is no change in force generation (Zhang et al., 1995) while the rate of force development increases (Stelzer et al., 2007). The physiological significance of this finding is limited as it is unlikely that the trout heart would ever be maximally activated. There was also no influence of PKA treatment on force generation at sub-maximal levels.

In mammalian hearts the accelerated rate of force development with PKA treatment is caused by the phosphorylation of cMyBP-C (Stelzer et al., 2007). This is thought to be in response to changes in the interaction between this protein and the myosin head (Stelzer et al., 2007). In the current study we did not detect a change in the phosphorylation state of cMyBP-C but we did for MLC-2. Previous work has also reported that MLC-2 is phosphorylated in trout cardiac tissue (Patrick et al., 2010). MLC-2 is located at the neck region of the myosin molecule and, together with the essential light chain, stabilizes the  $\alpha$ -helical neck of the myosin head (Szczena-Cordary et al., 2004). The phosphorylation of this regulatory protein influences the flexibility of the myosin head and therefore alters its ability to bind to actin, thereby affecting the kinetics of cross-bridge formation (Buck et al., 1999; Olsson et al., 2004). MLC-2 is thought to be primarily targeted by myosin light chain kinase; previous work has, however, demonstrated that rat MLC-2 is phosphorylatable by protein kinase C (PKC) (Noland and Kuo, 1993; Venema and Kuo, 1993; Venema et al., 1993) and PKA (Venema and Kuo, 1993). In mammalian tissue the phosphorylation of MLC-2 increases the activity of actin–myosin ATPase (AM ATPase) (Scruggs et al., 2009) and the  $\text{Ca}^{2+}$  sensitivity of force generation (Morano et al., 1988; Morano and Ruegg, 1986; Olsson et al., 2004; Sweeney and Stull, 1986). Comparison of the amino acid sequence of rat cardiac MLC-2 with that from a salmonid heart reveals that there are phosphorylatable residues in the fish protein at the same positions as known phosphorylation targets in rat MLC-2. However, the amino acid sequences of the two proteins are only 69% identical. It is not too surprising, therefore, that the phosphorylation of trout MLC-2 has a different effect on contractile function from phosphorylation of MLC-2 in the mammalian heart. The changes in contractile function caused by the phosphorylation of MLC-2 in the trout trabeculae are likely caused by changes in the functional characteristics of the myosin heads, influencing the formation of force-generating cross-bridges.

In a related study, we have demonstrated that the phosphorylation of a cardiac troponin (cTn) complex containing trout TnI (ScTnI, where S refers to the fact that trout are salmonids), rat cTnT and human cTnI caused a reduction in the  $\text{Ca}^{2+}$  affinity of the complex (Kirkpatrick et al., 2011). The only protein that was phosphorylated by this treatment was ScTnI. In the current study, PKA treatment did not alter the phosphorylation state of ScTnI. This suggests that when the proteins are functioning within an intact contractile element the phosphorylation sites are not accessible to the kinase because of the protein conformation.

The treatment of the trabeculae preparations with PKA did not alter the passive force measured at pCa9. Passive force was measured by a release–restretch protocol that measures the force difference before and after all the cross-bridges are knocked off under relaxing conditions. Previous work by Patrick and colleagues (Patrick et al., 2010) has shown that the passive tension of single trout cardiac myocytes is determined by the content of titin, a phosphorylatable protein. As a multi-cellular preparation was used in the present study, if a decrease in tension was caused by the phosphorylation of titin it was likely dampened by the connections between myocytes.

Experiments with rat cardiac trabeculae completed at the same temperature and at a SL of 2.25  $\mu\text{m}$  generated a maximum force of  $20.2 \pm 4.0 \text{ mN mm}^{-2}$  (Regnier et al., 2004). In the current study, this value for the control preparation was equal to  $30.24 \pm 3.8 \text{ mN mm}^{-2}$ . This suggests that the force being generated by these preparations is at least equal to that generated by mammalian tissue under the same conditions. The  $k_{\text{tr}}$  values generated during maximal activation of the control trout preparations are less than those obtained during the maximal activation of rat cardiac trabeculae. In the current study, the fastest  $k_{\text{tr}}$  value ( $5.25 \pm 0.4 \text{ s}^{-1}$ , Fig. 4) corresponded to sub-maximal activation of force at pCa5.6. Using similar methods, Regnier and colleagues (Regnier et al., 2004) have reported  $k_{\text{tr}}$  values in rat trabeculae as high as  $10.6 \pm 4.0 \text{ s}^{-1}$  during maximal activation. This indicates faster cross-bridge cycling rates for the rat heart than for the trout heart (Brenner, 1988). The higher level of steady-state force production in the trout than in the rat corresponds to a greater cross-bridge binding and force production in the trout. This increased force response would also correlate with a slower myosin detachment rate in the trout, if the attachment rate was similar for the two preparations (Hancock et al., 1997). In contrast with observations of mammalian  $k_{\text{tr}}$ , which increases with  $[\text{Ca}^{2+}]$ , the trout  $k_{\text{tr}}$ –pCa relationship decreased with increased  $\text{Ca}^{2+}$  activation and force generation. This implies that, in the trout preparations, an increase in  $\text{Ca}^{2+}$  caused a decrease in the rate of cross-bridge detachment.

One factor to consider is that the higher rates of cross-bridge cycling in the trout preparations were obtained at sub-maximal levels of activation. As it is not expected that an intact preparation would ever be maximally activated, it is likely that the differences between the preparations at these levels of activation are more meaningful. The higher  $k_{\text{tr}}$  values obtained at the lower  $\text{Ca}^{2+}$  concentrations likely reflect the fact that the preparation functions optimally at these concentrations. The rate of cross-bridge cycling also appeared to be higher at these sub-maximal levels of activation in the PKA-treated preparations but the difference was not statistically significant. This does, however, suggest that PKA has a different impact on cross-bridge cycling rate at sub-maximal levels of activation than at maximal levels of activation.

#### Physiological significance

The results of the current study suggest that the contractile element plays a comparatively smaller role in the functional changes caused by  $\beta$ -adrenergic stimulation in the trout heart. These functional changes are an increase in heart rate, an increase in the strength of contraction (Aho and Vornanen, 2001), as well as increases in the rate of contraction and relaxation (Shiels et al., 1998). The increase in heart rate is caused by increased chronotropy regulated by the sinoatrial node, while the increased force generation is due to increased current through the L-type  $\text{Ca}^{2+}$  channels (Aho and Vornanen, 2001; Vornanen, 1998). In the trout heart the L-type  $\text{Ca}^{2+}$  channels play a comparatively significant role, delivering activator  $\text{Ca}^{2+}$  for myocyte contraction (Vornanen, 1998). Any increase in the current through this channel, therefore, has significant potential to increase contractility. In the current study we demonstrated that there was a decrease in force generation by the skinned preparation at maximum activation in response to PKA treatment. In the intact myocyte, the parallel increase in activator  $\text{Ca}^{2+}$  entering through the L-type channel would likely compensate for this loss of myofilament function that occurs with PKA phosphorylation.

In the mammalian heart the decrease in  $\text{Ca}^{2+}$  sensitivity of the contractile element caused by the phosphorylation of cTnI by PKA translates into faster rates of muscle relaxation following contraction

and an increase in the kinetics of cross-bridge cycling (Kentish et al., 2001; Li et al., 2004; Rarick et al., 1999; Solaro and Rarick, 1998; Yasuda et al., 2007). The result of the current study demonstrating that PKA treatment does not affect the Ca<sup>2+</sup> sensitivity of trout cardiac tissue illustrates the fact that the contractile element does not play a role in this effect of PKA phosphorylation on contractility of the trout heart. This limited response of the trout contractile element to PKA treatment correlates with the results of a recent phylogenetic study where we demonstrated that the number of PKA targets in cTnI and cMyBP-C increased during vertebrate evolution (Shaffer and Gillis, 2010). For example, the N-terminal peptide, containing two PKA sites, did not appear in vertebrate cTnI until after fish diverged from the vertebrate lineage. Its absence in fish cTnI is therefore a consequence of the evolutionary history of the cardiac proteins in this vertebrate group. Accordingly, the isoforms of cTnI and cMyBP-C found in the trout heart can be viewed as ancestral or primitive forms of the proteins and their limited role in the regulation of the trout cardiac contractile element is a consequence of this.

## APPENDIX

### Identification of contractile proteins on protein gels

#### Methods

##### Western blotting

Western blotting was used to identify cTnI, cTnT and cMyBP-C. For western blot analysis, native trout cardiac muscle proteins were separated on an SDS-PAGE gel. The SDS-PAGE gel was then transferred onto Hybond-P PVDF membranes using an ECL Semi-Dry Transfer Unit (GE Healthcare). The primary antibodies used were sc-8118 for cTnI and sc-19 for cTnT (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA). These are polyclonal goat anti-human antibodies synthesized using the C-terminus of each protein as the epitope. The primary antibody used for cMyBP-C was a customized polyclonal rabbit antibody for rat cMyBP-C, kindly donated by Dr Samantha Harris (University of California Davis). For the detection of cTnI and cTnT, the secondary antibody was a rabbit anti-goat IgG conjugated to horseradish peroxidase (AP106P; Millipore, Etobicoke, ON, Canada). For cMyBP-C, the secondary antibody was goat anti-rabbit IgG conjugated to horseradish peroxidase (G21234; Invitrogen, Eugene, OR, USA).

##### Mass spectroscopy

To identify MLC-2, the band corresponding to the correct mass was cut from the gel then exposed to trypsin in-gel digestion. The resulting peptides were analyzed using nano-LC quadrupole time-of-flight (QTOF) mass spectrometry. This identification was completed at the Mass Spectrometry Facility at the University of Guelph.

#### Results

##### Western blotting

This analysis successfully identified cTnI, cTnT and cMyBP-C. Supplementary material Fig. S1 is a composite image created from the results of the western blots for each muscle protein. This identified single bands for cTnI and cMyBP-C. The cTnT antibody identified two different protein bands indicating that there are two isoforms of TnT in the trout heart.

##### Mass spectroscopy

Comparison between the peptide sequences obtained from the trypsin digestion of native myosin regulatory light chain 2 and the amino acid sequence of Atlantic salmon myosin regulatory light

chain 2 resulted in a display score of 79% and 48% coverage, indicating a positive match (supplementary material Fig. S2).

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## REFERENCES

- Aho, E. and Vornanen, M. (2001). Cold acclimation increases basal heart rate but decreases its thermal tolerance in rainbow trout (*Oncorhynchus mykiss*). *J. Comp. Physiol. B* **171**, 173-179.
- Brenner, B. (1988). Effect of Ca<sup>2+</sup> on cross-bridge turnover kinetics in skinned single rabbit psoas fibers: implications for regulation of muscle contraction. *Proc. Natl. Acad. Sci. USA* **85**, 3265-3269.
- Buck, S. H., Konyn, P. J., Palermo, J., Robbins, J. and Moss, R. L. (1999). Altered kinetics of contraction of mouse atrial myocytes expressing ventricular myosin regulatory light chain. *Am. J. Physiol.* **276**, H1167-H1171.
- Farrell, A. P. (1985). A protective effect of adrenaline on the acidotic teleost heart. *J. Exp. Biol.* **116**, 503-508.
- Fentzke, R. C., Buck, S. H., Patel, J. R., Lin, H., Wolska, B. M., Stojanovic, M. O., Martin, A. F., Solaro, R. J., Moss, R. L. and Leiden, J. M. (1999). Impaired cardiomyocyte relaxation and diastolic function in transgenic mice expressing slow skeletal troponin I in the heart. *J. Physiol.* **517**, 143-157.
- Gesser, H., Andresen, P., Brams, P. and Sund Laursen, J. (1982). Inotropic effects of adrenaline on the anoxic or hypercapnic myocardium of rainbow-trout and eel. *J. Comp. Physiol.* **147**, 123-128.
- Gillis, T. E., Marshall, C. R., Xue, X. H., Borgford, T. J. and Tibbits, G. F. (2000). Ca<sup>2+</sup> binding to cardiac troponin C: effects of temperature and pH on mammalian and salmonid isoforms. *Am. J. Physiol.* **279**, R1707-R1715.
- Gillis, T. E., Martyn, D. A., Rivera, A. J. and Regnier, M. (2007). Investigation of thin filament near-neighbour regulatory unit interactions during force development in skinned cardiac and skeletal muscle. *J. Physiol.* **580**, 561-576.
- Graham, M. S. and Farrell, A. P. (1989). The effect of temperature acclimation and adrenaline on the performance of a perfused trout heart. *Physiol. Zool.* **62**, 38-61.
- Hancock, W. O., Huntsman, L. L. and Gordon, A. M. (1997). Models of Ca<sup>2+</sup> activation account for differences between skeletal and cardiac force redevelopment kinetics. *J. Muscle Res. Cell Motil.* **18**, 671-681.
- Holroyde, M. J., Potter, J. D. and Solaro, R. J. (1979). The calcium binding properties of phosphorylated and unphosphorylated cardiac and skeletal myosins. *J. Biol. Chem.* **254**, 6478-6482.
- Keen, J. E., Vianzon, D.-E., Farrell, A. P. and Tibbits, G. F. (1993). Thermal acclimation alters both adrenergic sensitivity and adrenoceptor density in cardiac tissue of rainbow trout. *J. Exp. Biol.* **181**, 27-47.
- Kentish, J. C., McCloskey, D. T., Layland, J., Palmer, S., Leiden, J. M., Martin, A. F. and Solaro, R. J. (2001). Phosphorylation of troponin I by protein kinase A accelerates relaxation and crossbridge cycle kinetics in mouse ventricular muscle. *Circ. Res.* **88**, 1059-1065.
- Kirkpatrick, K. P., Robertson, A. S., Klaiman, J. M. and Gillis, T. E. (2011). The influence of trout cardiac troponin I and PKA phosphorylation on the Ca<sup>2+</sup> affinity of the cardiac troponin complex. *J. Exp. Biol.* **214**, 1981-1988.
- Kobayashi, T. and Solaro, R. J. (2005). Calcium, thin filaments, and the integrative biology of cardiac contractility. *Annu. Rev. Physiol.* **67**, 39-67.
- Li, M. X., Wang, X. and Sykes, B. D. (2004). Structural based insights into the role of troponin in cardiac muscle pathophysiology. *J. Muscle Res. Cell Motil.* **25**, 559-579.
- Matsuba, D., Terui, T., O-Uchi, J., Tanaka, H., Ojima, T., Ohtsuki, I., Ishiwata, S., Kurihara, S. and Fukuda, N. (2009). Protein kinase A-dependent modulation of Ca<sup>2+</sup> sensitivity in cardiac and fast skeletal muscles after reconstitution with cardiac troponin. *J. Gen. Physiol.* **133**, 571-581.
- Mope, L., McClellan, G. B. and Winegrad, S. (1980). Calcium sensitivity of the contractile system and phosphorylation of troponin in hyperpermeable cardiac cells. *J. Gen. Physiol.* **75**, 271-282.
- Morano, I. and Ruegg, J. C. (1986). Calcium sensitivity of myofilaments in cardiac muscle-effect of myosin phosphorylation. *Basic Res. Cardiol.* **81** Suppl. **1**, 17-23.
- Morano, I., Bachle-Stolz, C., Katus, A. and Ruegg, J. C. (1988). Increased calcium sensitivity of chemically skinned human atria by myosin light chain kinase. *Basic Res. Cardiol.* **83**, 350-359.
- Noland, T. A., Jr and Kuo, J. F. (1993). Phosphorylation of cardiac myosin light chain 2 by protein kinase C and myosin light chain kinase increases Ca<sup>2+</sup>-stimulated actomyosin MgATPase activity. *Biochem. Biophys. Res. Commun.* **193**, 254-260.
- Olsson, M. C., Patel, J. R., Fitzsimons, D. P., Walker, J. W. and Moss, R. L. (2004). Basal myosin light chain phosphorylation is a determinant of Ca<sup>2+</sup> sensitivity of force and activation dependence of the kinetics of myocardial force development. *Am. J. Physiol.* **287**, H2712-H2718.
- Patrick, S. M., Hoskins, A. C., Kentish, J. C., White, E., Shiels, H. A. and Cazorla, O. (2010). Enhanced length-dependent Ca<sup>2+</sup> activation in fish cardiomyocytes permits a large operating range of sarcomere lengths. *J. Mol. Cell. Cardiol.* **48**, 917-924.
- Rarick, H. M., Tang, H. P., Guo, X. D., Martin, A. F. and Solaro, R. J. (1999). Interactions at the NH<sub>2</sub>-terminal interface of cardiac troponin I modulate myofilament activation. *J. Mol. Cell. Cardiol.* **31**, 363-375.
- Regnier, M., Martin, H., Barsotti, R. J., Rivera, A. J., Martyn, D. A. and Clemmens, E. (2004). Cross-bridge versus thin filament contributions to the level and rate of force development in cardiac muscle. *Biophys. J.* **87**, 1815-1824.



- Scruggs, S. B., Hinken, A. C., Thawornkaiwong, A., Robbins, J., Walker, L. A., de Tombe, P. P., Geenen, D. L., Buttrick, P. M. and Solaro, R. J.** (2009). Ablation of ventricular myosin regulatory light chain phosphorylation in mice causes cardiac dysfunction in situ and affects neighboring myofilament protein phosphorylation. *J. Biol. Chem.* **284**, 5097-5106.
- Shaffer, J. F. and Gillis, T. E.** (2010). Evolution of the regulatory control of vertebrate striated muscle: the roles of troponin I and myosin binding protein-C. *Physiol. Genomics* **42**, 406-419.
- Shiels, H. A., Stevens, E. D. and Farrell, A. P.** (1998). Effects of temperature, adrenaline and ryanodine on power production in rainbow trout *Oncorhynchus mykiss* ventricular trabeculae. *J. Exp. Biol.* **201**, 2701-2710.
- Solaro, R. J. and Rarick, H. M.** (1998). Troponin and tropomyosin: proteins that switch on and tune in the activity of cardiac myofilaments. *Circ. Res.* **83**, 471-480.
- Solaro, R. J., Rosevear, P. and Kobayashi, T.** (2008). The unique functions of cardiac troponin I in the control of cardiac muscle contraction and relaxation. *Biochem. Biophys. Res. Commun.* **369**, 82-87.
- Stelzer, J. E., Patel, J. R., Walker, J. W. and Moss, R. L.** (2007). Differential roles of cardiac myosin-binding protein C and cardiac troponin I in the myofibrillar force responses to protein kinase A phosphorylation. *Circ. Res.* **101**, 503-511.
- Sweeney, H. L. and Stull, J. T.** (1986). Phosphorylation of myosin in permeabilized mammalian cardiac and skeletal muscle cells. *Am. J. Physiol.* **250**, C657-C660.
- Szczesna-Cordary, D., Guzman, G., Ng, S. S. and Zhao, J.** (2004). Familial hypertrophic cardiomyopathy-linked alterations in Ca<sup>2+</sup> binding of human cardiac myosin regulatory light chain affect cardiac muscle contraction. *J. Biol. Chem.* **279**, 3535-3542.
- Venema, R. C. and Kuo, J. F.** (1993). Protein kinase C-mediated phosphorylation of troponin I and C-protein in isolated myocardial cells is associated with inhibition of myofibrillar actomyosin MgATPase. *J. Biol. Chem.* **268**, 2705-2711.
- Venema, R. C., Raynor, R. L., Noland, T. A., Jr and Kuo, J. F.** (1993). Role of protein kinase C in the phosphorylation of cardiac myosin light chain 2. *Biochem. J.* **294**, 401-406.
- Vornanen, M.** (1998). L-type Ca<sup>2+</sup> current in fish cardiac myocytes: effects of thermal acclimation and beta-adrenergic stimulation. *J. Exp. Biol.* **201**, 533-547.
- Wehrens, X. H., Lehnart, S. E., Reiken, S., Vest, J. A., Wronska, A. and Marks, A. R.** (2006). Ryanodine receptor/calcium release channel PKA phosphorylation: a critical mediator of heart failure progression. *Proc. Natl. Acad. Sci. USA* **103**, 511-518.
- Yasuda, S., Coutu, P., Sadayappan, S., Robbins, J. and Metzger, J. M.** (2007). Cardiac transgenic and gene transfer strategies converge to support an important role for troponin I in regulating relaxation in cardiac myocytes. *Circ. Res.* **101**, 377-386.
- Zhang, R., Zhao, J. and Potter, J. D.** (1995). Phosphorylation of both serine residues in cardiac troponin I is required to decrease the Ca<sup>2+</sup> affinity of cardiac troponin C. *J. Biol. Chem.* **270**, 30773-30780.



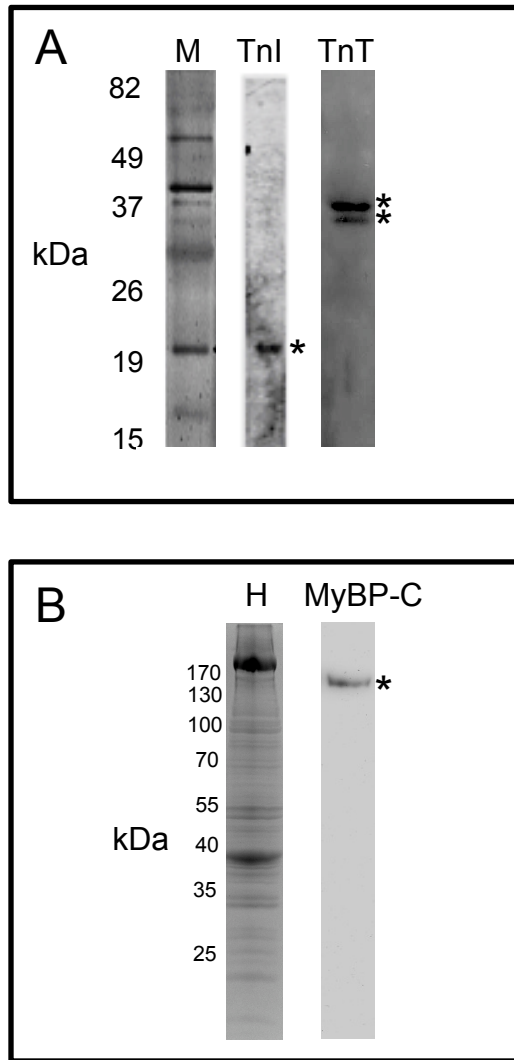


Figure 1. **Composite image showing results of western blotting to identify cTnI, cTnT and MyBP-C.** Panel A: lane 1 is trout cardiac myofilament proteins (M) stained with coomassie blue for total protein, lane 2 is western blot of trout myofilament proteins probed with antibody for mammalian cardiac troponin I, lane 3 is western blot of trout myofilament proteins probed with antibody for mammalian cardiac troponin T. Panel B, lane 1 is whole trout heart homogenate (H) stained with coomassie blue, lane 2 is a western blot of whole trout heart homogenate with antibody for rat cardiac myosin binding protein C. \* is used to identify protein band

MAPKKAKR **RG AAAEGGSSNV FSMFEQSQIQ EYKEAFTIID QNRDGII**SKD 51  
DLRDVLASMG QLNVK **NEELE AMVKEASGPI NFTVFLTM**FG EKLKGADPED 101  
VIVSAFKVLD PEATGFIKKE FLQELLTTQC DRFSAEEMK **N LWAAFPPDVA** 151  
**GNVDYKQICYV**THGEEKEE

**Figure 2. The results of Nano-LC-QTOF mass spectroscopy sequence analysis of in gel trypsin digested native trout cardiac myosin regulatory light chain 2 mapped onto the amino acid sequence of Atlantic salmon cardiac myosin regulatory light chain 2.** The boxes indicate peptides that matched between the sample and the sequence. A display score of 79% with 48% coverage was obtained indicating a positive match.